Acellular Dermal Matrix: General Principles for the Plastic Surgeon

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Abstract

Acellular dermal matrix (ADM) is a recently-developed, biologically-derived product with many useful applications in plastic surgery, in both cosmetic and reconstructive procedures. While the use of ADM initially outpaced quality literature, within the past 10 years the literature on ADM has rapidly expanded. Some of these data show promising results in treating historically-challenging problems within our field; thus, an effort to clarify and summarize existing work with ADM is indicated. While subsequent articles in this supplement focus on specific applications, this article provides a general overview of the biology of, applications for, and existing literature on ADM.

Keywords

attenuated tissue, breast reconstruction, complications, flaps, radiation, tissue expanders

Derived from the Greek word plastikos, meaning "to mold or give form," plastic surgery has a long-standing history of creativity and innovation for the purpose of reshaping traumatized, congenitally-malformed, or aestheticallydispleasing abnormalities. The methods and techniques employed for this purpose have varied over the years, including autologous and alloplastic options. As our understanding of wound healing, tissue biology, and manufacturing/materials science has progressed in recent decades, the substrates available to the surgeon for application in a multitude of procedures have expanded rapidly. While synthetic materials are helpful in a variety of surgical settings, permanent prostheses suffer by nature of their inability to become completely incorporated into the human form—a permanent synthetic implant is always at risk for infection, extrusion, and invasion into surrounding tissue. Biological implants, however, carry the unique ability to become integrated into the native tissue, which aids in wound strength and offers a more biocompatible solution. Specifically, acellular dermal matrix (ADM) has revolutionized our approach to difficult clinical scenarios, including head and neck, breast, abdominal wall, and extremity surgery. In this introduction to the supplement "Acellular Dermal Matrix: Fundamentals and Expanding Applications in Plastic Surgery," we review the basic science behind, general principles about, and applications of ADM in plastic surgery. Subsequent articles detail indications and describe techniques to illustrate the breadth of ADM application in plastic surgery.

WOUND HEALING

To fully understand the structure and function of ADM, one must first become familiar with the wound healing process. Whether the injury is a result of surgery or trauma, before the healing process may begin, the body's initial response is one of hemostasis. This is achieved via platelet activation in response to endothelial injury, exposed collagen, and thrombin; it is aided by initiation of the coagulation cascade. In addition, local blood flow is slowed via prostaglandin- and catecholamine-induced vasoconstriction.

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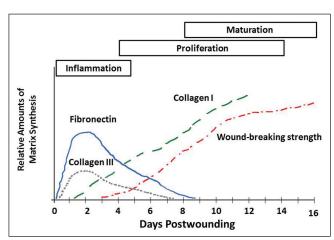


Figure 1. Deposition of wound matrix components over time. Although fibronectin and collagen Type III constitute the early matrix, collagen Type I accumulates later, corresponding to the increase in wound-breaking strength. Reprinted from Witte MB, Barbul A. General principles of wound healing. *Surg Clin North Am* 1997;77(3), with permission from Elsevier, Inc.

Platelet aggregation in turn causes release of a whole host of cytokines (including adenosine diphosphate, serotonin, and thromboxane A2), which ultimately increases hemostasis and vasoconstriction. While vasoconstriction initially controls blood loss, inflammatory mediators such as kinins, histamines, prostaglandins, and leukotrienes soon cause local vasodilation, allowing for the influx of inflammatory cells necessary for the healing process to begin. Within eight hours of injury, polymorphonuclear cells begin cleaning the wound and preventing infection. This process continues for the first three days following injury and is augmented by the infiltration of macrophages, which help to remove debris. This orchestration is directed by cytokines, including TGF, PDGF, FGF, IL-1, and TNF.^{2,3}

Following completion of the inflammatory stage of wound healing, the next phase begins, characterized by proliferation. By Days 5 through 7, fibroblasts become the primary cell of interest, with an overall goal of collagen synthesis. Type III collagen predominates in early production but is replaced in the long term by Type I collagen for maturation of the scar (Figure 1). This collagen framework is embedded in a milieu of fibronectin and glycosaminoglycans, including heparan sulfate, hyaluronic acid, chondroitin sulfate, and keratan sulfate, which all aid in the creation of a matrix for wound structure.3 Overall, this matrix is ultimately responsible for wound structure and the reestablishment of wound continuity—characteristics that make it ideal for a biological implant. Fibroblasts also secrete matrix metalloproteinases, which help shape the surrounding matrix to allow for fibroblast migration.³ The resultant matrix structure also provides a framework for angiogenesis by providing a support network for budding vessels, stimulated by HIF-1, FGF, PDGF, TGF-β, TNF-α, and VEGF.⁴ Superficially, the wound is simultaneously being covered by epithelial cells where appropriate. This process occurs via rapid mitosis of epithelial cells from the periphery of the wound and adnexal structures.

Once all key components of the wound are in place, the final phase of wound maturation and remodeling begins. Collagen is degraded and reformed to render a more uniform and balanced structural array, which maximizes wound strength. The overall quantity of collagen does not change; however, Type III collagen is replaced by Type I collagen, which is stronger and more durable. Overall, wound strength continues to improve until approximately 12 weeks, at which point the wound has reached its final strength. Overall wound strength reaches approximately 80% of its initial strength but never regains its original function.³ Elastin, another structural protein, provides for elasticity and stretching within native tissue and healing wounds. Myofibroblasts aid in wound contraction, especially in open wounds healing by secondary intention.³

THE CLINICAL DILEMMA

With any surgical wound, the goal is rapid healing with a minimal scar that will have a long-term natural contour and provide lasting tensile strength. Wound healing is ideally begun with primary wound closure. The body provides the natural substrate for healing and offers the best chance for positive results. However, tension is often too great for primary closure, and additional tissue is needed to complete the repair. Local flaps—pedicle or free flaps can be utilized in these cases to provide additional soft tissue coverage and the necessary ingredients for wound healing. Unfortunately, autologous tissue is neither always available nor free of donor morbidity. While synthetic materials such as Marlex mesh, polytetrafluoroethylene, titanium implants, or methylmethacrylate have been useful for implantation and structural support, they all suffer from several limitations. They do not bring any of the basic wound healing units (eg, glycosaminoglycans, fibronectin) into the field to amplify the wound healing process. In addition, they become only minimally integrated in the final wound. Although scar tissue may adhere to the implant, it is never truly an integrated implant. Finally, because they are permanent foreign bodies, they are all subject to infection and extrusion. Therefore, an ideal prosthesis is one that augments the body's natural efforts, provides structural support, allows for ingrowth, and is eventually replaced or fully integrated. Many of these characteristics are found in ADM.

ACELLULAR DERMAL MATRICES

The dermis is a layer of the body rich with wound matrix, composed of collagen, elastin, fibrillin, and glycosaminoglycans. It provides a semirigid yet elastic support system to ward off injury from trauma as well as provide structural and nutritional support to the epidermis above. While the dermis is composed primarily of a matrix for structural

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Table 1. ADM Products

Proprietary Name	Manufacturer	Origin	Unique Attributes
AlloDerm, Repliform	Lifecell Corp.	Human dermis	
Cymetra	Lifecell Corp.	Human dermis	Micronized for injection
NeoForm	Mentor	Human dermis	
Flex HD	Musculoskeletal Transplant Foundation and Ethicon	Human dermis	Prehydrated and ready for use
DermaMatrix	Musculoskeletal Transplant Foundation and Synthes	Human dermis	
SureDerm	Hans Biomed Corp.	Human dermis	
Glyaderm	Euroskinbank	Human dermis	Relatively inexpensive
Collamend	Bard	Porcine dermis	
Permacol, Enduragen, Pelvicol, Zimmer Collagen	Tissue Science Laboratories	Porcine dermis	Available in large sheets for abdominal wall use (Permacol); crosslinked
Strattice	Lifecell Corp.	Porcine dermis	Premade geometric shapes
Surgisis (SIS)	Cook Biomedical	Porcine intestinal submucosa ^a	Variety of products for specific indications
Surgimend	TEI Biosciences	Bovine dermis	
Veritas, Peri-Guard	Synovis	Bovine pericardium ^a	Noncrosslinked
Integra	Integra LifeSciences Corp.	Bovine Achilles tendon and shark cartilage ^a	Semipermeable silicone layer over matrix
Matriderm	Skin and Healthcare	Bovine dermis	Designed for skin defects
Pelnac	Gunze Corp.	Bovine dermis	Designed for skin defects
Renoskin	Groupe Perouse Plastic	Bovine dermis	Silicone outer layer; designed for skin defects

^aNot a true ADM.

support of the skin, it serves a major role in thermoregulation through vasoconstriction, dilation, and evaporation via sweat glands.

Also within the dermis lie multiple nerve endings designed for special functions, such as temperature sensation and proprioception. Thus, although the dermis is full of the necessary elements for wound healing, the cells within this layer (eg, sweat glands, nerve endings) are rich in antigenic material, making routine allotransplantation or xenotransplantation impossible without rapid rejection. Multiple proprietary methods have been devised to strip the dermis of its cellular components after harvest from human cadaver, pig, or cow dermis, as well as intestinal submucosa. These methods help the ADM materials to retain many of the structural elements (matrices) necessary for wound healing without the concern of rejection (since they are acellular). This has allowed for the routine placement of ADM for many cases that require structural support or a scaffold for improved wound healing. There are many names and types of ADM products—all purporting to be the best formula but all based upon the same principles (Table 1).

ADM is made by taking a full-thickness section of skin from a donor source—which in most cases is human cadaver, porcine, or bovine in origin. In the case of human donors, the tissue is screened for infectious diseases such as HIV and hepatitis. The tissue is run through a series of steps, with each company having its own proprietary process. AlloDerm (LifeCell Corp., Branchburg, New Jersey), one of the most ubiquitous products on the market, has a manufacturing process that begins by immersing the tissue in a buffered salt solution to separate and eliminate the dermis. Next, a series of mild detergents are applied to eliminate all cellular elements from the tissue. The tissue is extensively tested for any contamination to ensure a sterile product, then freeze-dried to prevent crystallization and allow for stable packaging and storage.

Of note, several materials in this category are not entirely biological. In particular, Integra (Integra Life Sciences Corp., Plainsboro, New Jersey), which is designed for superficial wounds, has an adherent layer of silicone sheeting on its outer surface that serves as an additional temporary barrier to bacterial infiltration and helps avoid desiccation by preventing evaporation from the wound. In

essence, Integra mimics skin in that the silicone sheet acts as the epidermis while the matrix component serves as replacement dermis.

An attribute of certain ADM products involves crosslinking, whereby adjacent collagen is linked chemically. While crosslinking occurs within normal human tissue, the degree to which it occurs during preparation of ADM can be controlled. Theory and research suggest that noncrosslinked substrates suffer from faster degradation due to collagenases within the body,⁵ whereas too much crosslinking may lead to limitations in native tissue ingrowth with subsequent poor tissue integration.⁶⁻⁸ Agents such as glutaraldehyde or hexamethylene diisocyanate can be utilized to crosslink the collagen within the matrix. Due to the poor integration of noncrosslinked ADM, these implants may show promise in applications where adjacent adhesion formation is undesirable, such as with intra-abdominal placement adjacent to the bowel and when a temporary barrier is indicated.⁹ Additionally, in aesthetic surgery where softer contours and subtle results are needed, noncrosslinked agents are likely more appropriate due to increased pliability. In contrast, properly crosslinked matrices likely offer more lasting strength in applications such as abdominal wall reconstruction and hernia repair.¹⁰

GENERAL APPLICATIONS

Due to the unique characteristics of biological implants and the variety of available products, ADM can be tremendously helpful in a range of applications within the field of plastic surgery. Although there are, to date, little if any prospective randomized data comparing ADM to the current standard of care in each field, there are several areas of plastic surgery that have yielded a significant amount of outcomes data, the results of which tend to be excellent. Despite numerous reports of novel ways of placing ADM in surgery, the following areas are the best studied.

Burns and Wounds

While grafting has long been a staple for the replacement of lost skin, ADM has become increasingly popular in these cases over the past 20 years for coverage of open soft tissue defects. While skin grafting is significantly less expensive, ADM placement can help prevent a painful and displeasing donor site. AlloDerm has been reported in the literature as a dermal substitute in full-thickness burn wounds. 11,12 In addition, ADM has been used in conjunction with split-thickness grafts for treatment of full-thickness wounds. 13-15 For significant full-thickness burns crossing the joint, ADM products are effective in wound coverage and allow for sufficient elasticity to prevent contracture over the joint.¹⁶ Aside from burn wounds, ADM has been shown to be effective in the treatment of diabetic foot ulcers, with one prospective randomized study demonstrating improved healing efficacy following coverage with dermal matrices. 17,18 While the cost of ADM may limit widespread use, early results seem promising for many types of wound management.

Abdominal Wall Procedures

As many surgeons have found themselves operating in the abdomen with poor fascia for closure, combined with contamination from enteric sources, there has always been interest in an implant material that would add strength to the abdominal closure while resisting infection. Permanent mesh is unusable in these situations, and temporary synthetic meshes often do not provide adequate or lasting tensile strength to prevent hernia formation. ADM has been shown to be effective in decreasing hernia recurrence rates in infected fields when compared to primary closure without mesh.19,20 While AlloDerm (human ADM) was initially used for this application, many find its overall strength lacking; furthermore, its elasticity makes it prone to bulging over the long term. 21-23 Therefore, its placement in the abdominal wall should be abandoned. More recently, porcine ADM products have been placed with success, preventing recurrence and providing for closure that is resistant to chronic infection.^{9,24} For best results, these complex abdominal wall closures are coupled with component separation, which provides additional native coverage of the implant where there has otherwise been loss of sufficient fascia or even abdominal domain (Figure 2). Studies have shown an exceedingly high recurrence rate when the mesh serves as a bridge rather than a bolster.^{25,26} Overall, this highlights an important principle: ADM should be placed in a position that maximizes surface area contact with vascularized native tissue to facilitate wound healing and decrease hernia recurrence.

Similarly, in the setting of permanent mesh infection, removal of the infected implant often leaves a large defect in a contaminated operative field. Biological mesh provides an opportunity to bolster the closure without the risk of permanent infection requiring reoperation. Not all studies have had overwhelmingly positive results with ADM for abdominal wall reconstruction, due to high recurrence rates, extrusion, and infection.²⁷⁻²⁹ It is important to recognize that this material is in the early stages of use and that further prospective randomized studies are necessary to clearly delineate outcomes.

Reconstructive Breast Procedures

Reconstruction with tissue expanders and implants is a multistage process that can be exhausting to patients already coping with a difficult diagnosis. The limitation of immediate one-stage reconstruction has always been the inability to completely cover an implant with sufficient soft tissue to ensure a tension-free closure and avoid implant extrusion. Typically, with placement of a subpectoral expander, the taut muscle is the limiting factor for implant size and thus requires expansion. ADM has been increasingly used as an internal support and cover for single-stage operations. In this approach, the pectoralis major is allowed to "window-shade" upward,

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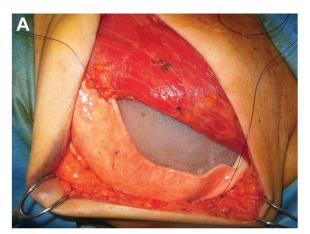
Figure 2. (A, B) This 45-year-old woman presented with loss of abdominal domain, in need of concomitant bowel surgery. (C) Component separation ventral hernia repair with the placement of ADM bolster (Permacol) in a retrorectus position. Note a small midline area that could not be covered entirely. (D) This 58-year-old man also presented with similar loss of domain and is shown undergoing a colostomy reversal. (E, F) Even with component separation, the ADM could not be covered in the patient featured in part D. (G, H) One year postoperatively, the patient in parts D, E, and F, with no evidence of hernia recurrence of bulge.

and an AlloDerm sling is fixed from the inferior aspect of the pectoralis down to the inferior mammary fold (Figure 3). Thus, the implant is completely covered and supported by the AlloDerm. In many cases, this allows for single-stage reconstruction with placement of the final implant at the time of initial reconstruction. While few data exist directly comparing different products, at least one retrospective study did show similar side effect profiles between AlloDerm and DermaMatrix (Synthes, Inc., West Chester, Pennsylvania). However, there are some data suggesting that implant infection and extrusion rates in alloplastic reconstruction are higher with

AlloDerm.³⁵⁻³⁷ Other ADM products have not been studied sufficiently to draw any similar conclusions. Overall, human-derived products with a high elastin content (such as AlloDerm) are desirable in this application, due to their ability to stretch.

Cosmetic Breast Procedures

Although ADM products have been utilized extensively in breast reconstruction, similar techniques have been applied in cosmetic breast surgery. ADM placement can yield a natural





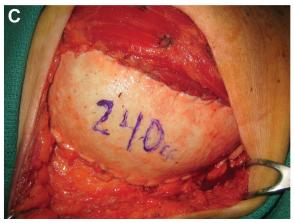


Figure 3. (A) In this two-stage breast reconstruction, an AlloDerm sling has been placed for additional coverage of the implant. Note that the pectoralis major has been released inferiorly. (B) Upon expander exchange, the ADM is visible but has been incorporated into a healthy capsule for adequate implant coverage. (C) In sum, 240 cc was placed into the expander at the time of initial placement.

Photos courtesy of Joseph J. Disa, MD—Memorial Sloan-Kettering Cancer Center.

contour for significant irregularities. In addition, significant implant malposition can be corrected with the support of a sling to lift and hold an implant in proper position.^{38,39} Similar in concept, an internal support can be placed to help prevent bottoming out with inferior pedicle breast reduction.⁴⁰ One significant deterrent from widespread application in cosmetic procedures is the need for patients to absorb the cost of the ADM.⁴¹

Head and Neck Procedures

In eyelid and periorbital surgery, the surgeon is often in need of a biological construct that will provide enough structure for support while simultaneously offering an aesthetically natural, soft contour. ADM has been used successfully in these cases to bolster lid structure and fill volume deficits. 42,43 Similarly, ADM can act as an off-the-shelf material for reconstructing the tarsal plate. 44,45

Secondary rhinoplasty is often plagued by insufficient structural support to establish the desired optimal cosmetic outcome. In these patients, many surgeons have found ADM useful for filling space while maintaining a soft contour in either saddle nose deformities or irregularities of the alar rim. 46-48 In addition, nasal septal perforations have been successfully treated with ADM. 49

In children with cleft palate, there is often insufficient soft tissue to provide stable, tension-free closure. ADM has been useful for adding bulk to this closure, whether primarily, in revisional surgery, or for treatment of a fistula. 50-53 In addition, ADM has been useful in the healing process of nasoalveolar bone grafting by providing additional nasal-oral mucosal lining where it is otherwise deficient. 54 Similarly, ADM has been successfully placed to line the oral cavity following intraoral cancer extirpation with similar quality of life to split-thickness skin grafts, at a lower cost, and without donor defects. 55

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CONCLUSIONS

The production of biological mesh for various surgical applications is an ever-burgeoning market with potential for great improvement in patient care. The ideal implant for surgical placement would be a biological replacement of lost native structure that, over time, is either replaced by normal human tissue or at least completely integrated. While there is no perfect ADM for all applications, there are many available products that work well with proper technique and patient selection. While there have been numerous publications on ADM, it does seem that the application of these materials has far outpaced the evidence in support of their use. That is not to say that no evidence exists; rather, we must recognize that these materials are very expensive and that just because they can be useful in a given situation does not mean that they always offer improved clinical results.

Disclosures

Dr. Serletti is a stockholder with Johnson and Johnson, Inc. None of the other authors have anything to disclose.

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REFERENCES

- 1. Furie B, Furie BC. Mechanisms of thrombus formation. *N Engl J Med* 2008;359:938-949.
- 2. Velnar T, Bailey T, Smrkolj V. The wound healing process: an overview of the cellular and molecular mechanisms. *J Int Med Res* 2009;37:1528-1542.
- 3. Teller P, White TK. The physiology of wound healing: injury through maturation. *Surg Clin North Am* 2009;89:599-610.
- 4. Przybylski M. A review of the current research on the role of bFGF and VEGF in angiogenesis. *J Wound Care* 2009;18:516-519.
- de Castro Bras LE, Proffitt JL, Bloor S, Sibbons PD. Effect of crosslinking on the performance of a collagenderived biomaterial as an implant for soft tissue repair: a rodent model. *J Biomed Mater Res B Appl Biomater* 2010; 95:239-249.
- 6. Kang NH YY, Woo JS, Ahn JH, Kim JY. Effects of crosslinking agents on the stability of human acellular dermal matrix. *J Korean Soc Plast Reconstr Surg* 2008;35:248-254.
- Kelley P, Gordley K, Higuera S, Hicks J, Hollier LH. Assessing the long-term retention and permanency of acellular cross-linked porcine dermal collagen as a softtissue substitute. *Plast Reconstr Surg* 2005;116:1780-1784.
- 8. Liang HC, Chang Y, Hsu CK, Lee MH, Sung HW. Effects of crosslinking degree of an acellular biological tissue on its tissue regeneration pattern. *Biomaterials* 2004;25:3541-3552.
- 9. Burns NK, Jaffari MV, Rios CN, Mathur AB, Butler CE. Non-

- cross-linked porcine acellular dermal matrices for abdominal wall reconstruction. *Plast Reconstr Surg* 2010;125:167-176.
- 10. Gaertner WB, Bonsack ME, Delaney JP. Experimental evaluation of four biologic prostheses for ventral hernia repair. *J Gastrointest Surg* 2007;11:1275-1285.
- 11. Wainwright DJ. Use of an acellular allograft dermal matrix (AlloDerm) in the management of full-thickness burns. *Burns* 1995;21:243-248.
- 12. Wainwright D, Madden M, Luterman A, et al. Clinical evaluation of an acellular allograft dermal matrix in full-thickness burns. *J Burn Care Rehabil* 1996;17:124-136.
- 13. Livesey SA, Herndon DN, Hollyoak MA, Atkinson YH, Nag A. Transplanted acellular allograft dermal matrix: potential as a template for the reconstruction of viable dermis. *Transplantation* 1995;60:1-9.
- 14. Han LH, Lu RR, Huo R, Yuan DL, Guo X. [Histologic analysis and long-term effect of acellular dermal matrix combined with autologous thin split-thickness skin graft]. *Zhonghua Zheng Xing Wai Ke Za Zhi* 2010;26:272-274.
- 15. Jiong C, Jiake C, Chunmao H, et al. Clinical application and long-term follow-up study of porcine acellular dermal matrix combined with autoskin grafting. *J Burn Care Res* 2010;31:280-285.
- 16. Yim H, Cho YS, Seo CH, et al. The use of AlloDerm on major burn patients: AlloDerm prevents post-burn joint contracture. *Burns* 2010;36:322-328.
- 17. Reyzelman A, Crews RT, Moore JC, et al. Clinical effectiveness of an acellular dermal regenerative tissue matrix compared to standard wound management in healing diabetic foot ulcers: a prospective, randomised, multicentre study. *Int Wound J* 2009;6:196-208.
- 18. Winters CL, Brigido SA, Liden BA, Simmons M, Hartman JF, Wright ML. A multicenter study involving the use of a human acellular dermal regenerative tissue matrix for the treatment of diabetic lower extremity wounds. *Adv Skin Wound Care* 2008;21:375-381.
- 19. Diaz JJ Jr, Conquest AM, Ferzoco SJ, et al. Multi-institutional experience using human acellular dermal matrix for ventral hernia repair in a compromised surgical field. *Arch Surg* 2009;144:209-215.
- 20. Candage R, Jones K, Luchette FA, Sinacore JM, Vandevender D, Reed RL 2nd. Use of human acellular dermal matrix for hernia repair: friend or foe? *Surgery* 2008;144:703-709.
- 21. de Moya MA, Dunham M, Inaba K, et al. Long-term outcome of acellular dermal matrix when used for large traumatic open abdomen. *J Trauma* 2008;65:349-353.
- 22. Bluebond-Langner R, Keifa ES, Mithani S, Bochicchio GV, Scalea T, Rodriguez ED. Recurrent abdominal laxity following interpositional human acellular dermal matrix. *Ann Plast Surg* 2008;60:76-80.
- 23. Gupta A, Zahriya K, Mullens PL, Salmassi S, Keshishian A. Ventral herniorrhaphy: experience with two different biosynthetic mesh materials, Surgisis and AlloDerm. *Hernia* 2006;10:419-425.
- 24. Hsu PW, Salgado CJ, Kent K, et al. Evaluation of porcine dermal collagen (Permacol) used in abdominal wall reconstruction. *J Plast Reconstr Aesthet Surg* 2009;62:1484-1489.

- 25. Blatnik J, Jin J, Rosen M. Abdominal hernia repair with bridging acellular dermal matrix: an expensive hernia sac. *Am J Surg* 2008;196:47-50.
- 26. Jin J, Rosen MJ, Blatnik J, et al. Use of acellular dermal matrix for complicated ventral hernia repair: does technique affect outcomes? *J Am Coll Surg* 2007;205: 654-660.
- 27. Maurice SM, Skeete DA. Use of human acellular dermal matrix for abdominal wall reconstructions. *Am J Surg* 2009:197:35-42.
- 28. Schuster R, Singh J, Safadi BY, Wren SM. The use of acellular dermal matrix for contaminated abdominal wall defects: wound status predicts success. *Am J Surg* 2006;192:594-597.
- 29. Lee EI, Chike-Obi CJ, Gonzalez P, et al. Abdominal wall repair using human acellular dermal matrix: a follow-up study. *Am J Surg* 2009;198:650-657.
- 30. Namnoum JD. Expander/implant reconstruction with AlloDerm: recent experience. *Plast Reconstr Surg* 2009; 124:387-394.
- 31. Topol BM, Dalton EF, Ponn T, Campbell CJ. Immediate single-stage breast reconstruction using implants and human acellular dermal tissue matrix with adjustment of the lower pole of the breast to reduce unwanted lift. *Ann Plast Surg* 2008;61:494-499.
- 32. Gamboa-Bobadilla GM. Implant breast reconstruction using acellular dermal matrix. *Ann Plast Surg* 2006;56:22-25.
- 33. Breuing KH, Warren SM. Immediate bilateral breast reconstruction with implants and inferolateral AlloDerm slings. *Ann Plast Surg* 2005;55:232-239.
- 34. Becker S, Saint-Cyr M, Wong C, et al. AlloDerm versus DermaMatrix in immediate expander-based breast reconstruction: a preliminary comparison of complication profiles and material compliance. *Plast Reconstr Surg* 2009; 123:1-6.
- 35. Nguyen MD, Chen C, Colakoglu S, Morris DJ, Tobias AM, Lee BT. Infectious complications leading to explantation in implant-based breast reconstruction with AlloDerm. *Eplasty* 2010;10:e48.
- 36. Lanier ST, Wang ED, Chen JJ, et al. The effect of acellular dermal matrix use on complication rates in tissue expander/implant breast reconstruction. *Ann Plast Surg* 2010;64:674-678.
- 37. Chun YS, Verma K, Rosen H, et al. Implant-based breast reconstruction using acellular dermal matrix and the risk of postoperative complications. *Plast Reconstr Surg* 2010;125:429-436.
- 38. Spear SL, Seruya M, Clemens MW, Teitelbaum S, Nahabedian MY. Acellular dermal matrix for the treatment and prevention of implant-associated breast deformities. *Plast Reconstr Surg* 2011;127:1047-1058.
- 39. Maxwell GP, Gabriel A. Use of the acellular dermal matrix in revisionary aesthetic breast surgery. *Aesthet Surg J* 2009;29:485-493.

- 40. Brown RH, Izaddoost S, Bullocks JM. Preventing the "bottoming out" and "star-gazing" phenomena in inferior pedicle breast reduction with an acellular dermal matrix internal brassiere. *Aesthetic Plast Surg* 2010;34:760-767.
- 41. Hartzell TL, Taghinia AH, Chang J, Lin SJ, Slavin SA. The use of human acellular dermal matrix for the correction of secondary deformities after breast augmentation: results and costs. *Plast Reconstr Surg* 2010;126:1711-1720.
- 42. Rubin PA, Fay AM, Remulla HD, Maus M. Ophthalmic plastic applications of acellular dermal allografts. *Ophthalmology* 1999;106:2091-2097.
- 43. McCord C, Nahai FR, Codner MA, Nahai F, Hester TR. Use of porcine acellular dermal matrix (Enduragen) grafts in eyelids: a review of 69 patients and 129 eyelids. *Plast Reconstr Surg* 2008;122:1206-1213.
- 44. Lee EW, Berbos Z, Zaldivar RA, Lee MS, Harrison AR. Use of DermaMatrix graft in oculoplastic surgery. *Ophthal Plast Reconstr Surg* 2010;26:153-154.
- 45. Hayek B, Hatef E, Nguyen M, Ho V, Hsu A, Esmaeli B. Acellular dermal graft (AlloDerm) for upper eyelid reconstruction after cancer removal. *Ophthal Plast Reconstr Surg* 2009;25:426-429.
- 46. Gryskiewicz JM. Waste not, want not: the use of Allo-Derm in secondary rhinoplasty. *Plast Reconstr Surg* 2005; 116:1999-2004.
- 47. Romo T 3rd, Sclafani AP, Sabini P. Reconstruction of the major saddle nose deformity using composite alloimplants. *Facial Plast Surg* 1998;14:151-157.
- 48. Meyers S, Rohrer T, Grande D. Use of dermal grafts in reconstructing deep nasal defects and shaping the ala nasi. *Dermatol Surg* 2001;27:300-305.
- 49. Lee D, Joseph EM, Pontell J, Turk JB. Long-term results of dermal grafting for the repair of nasal septal perforations. *Otolaryngol Head Neck Surg* 1999;120:483-486.
- Kirschner RE, Cabiling DS, Slemp AE, Siddiqi F, LaRossa DD, Losee JE. Repair of oronasal fistulae with acellular dermal matrices. *Plast Reconstr Surg* 2006;118:1431-440.
- 51. Cole P, Horn TW, Thaller S. The use of decellularized dermal grafting (AlloDerm) in persistent oro-nasal fistulas after tertiary cleft palate repair. *J Craniofac Surg* 2006;17:636-641.
- 52. Helling ER, Dev VR, Garza J, Barone C, Nelluri P, Wang PT. Low fistula rate in palatal clefts closed with the Furlow technique using decellularized dermis. *Plast Reconstr Surg* 2006;117:2361-2365.
- 53. Clark JM, Saffold SH, Israel JM. Decellularized dermal grafting in cleft palate repair. *Arch Facial Plast Surg* 2003;5: 40-44.
- 54. Clavijo-Alvarez JA, Vecchione L, DeCesare G, et al. Autologous bone grafting with adjunctive use of acellular dermal matrix for alveolar cleft defects: early outcomes. *Cleft Palate Craniofac J* 2010;47:116-121.
- 55. Girod DA, Sykes K, Jorgensen J, Tawfik O, Tsue T. Acellular dermis compared to skin grafts in oral cavity reconstruction. *Laryngoscope* 2009;119:2141-2149.