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Systematic significance of fruit morphology and anatomy in tribes Persicarieae and Polygoneae (Polygonaceae)

LOUIS P. RONSE DECRAENE FLS^{1*}, SUK-PYO HONG FLS² AND ERIK SMETS FLS1

¹Laboratory of Plant Systematics, Institute for Botany and Microbiology, Katholieke Universiteit Leuven, B-3001 Leuven, Belgium

²Laboratory of Plant Systematics, Department of Biology, Kyung Hee University, Seoul 130-701, Korea

Fruits of Polygonaccae have a basically similar construction of indehiscent nuts or achenes. Sections of fruits, coupled with surface patterns were studied with SEM and LM in all genera of the tribes Persicaricae and Polygoneae (Polygonoideae-Polygonaceae). The outer layer of the pericarp is usually thickened and its anatomy can be used consistently to delimit genera more than any other character of the fruit. Cells are most often puzzle piece-shaped in surface view, but the shape of the cells may become polygonal with straight anticlinal walls towards the endocarp. The primary sculpture of the cells is highly variable and has value at the specific level, rarely at the generic level. No strict correlation exists between the external surface patterns and the anatomy. Two main cell types can be recognized in cross- and longitudinal section, correlated with the straight or undulating outline of the anticlinal walls. No distinction can be made between sections Persicaria, Tovara, Echinocaulon, and Cephalophilon of the genus Persicaria; all share narrow rectangular cells with undulating anticlinal walls. Aconogonon and Bistorta can be delimited by the square to rectangular cells with a narrow dichotomously branching lumen and straight anticlinal walls; both genera are best grouped as a single genus with two sections. A similar arrangement is found occasionally in species of Polygonum s.s., Polygonella, Atraphaxis, Fallopia and Calligonum. Fruit anatomy of Pteropyrum is distinctive. The genus Polygonum s.s. shows a wide range of integrating patterns, ranging from straight to undulating anticlinal walls and cannot be separated from Polygonella. Fagopyrum is aberrant in having a parenchymatic exocarp and a thickened mesocarp; other evidence supports its isolated position. Different fruit anatomical patterns have arisen several times in evolution and have a limited value at tribal level but are useful at generic level. It is suggested that an arrangement with straight anticlinal walls and a broad lumen, eventually with dendritic branching towards the periphery, is ancestral.

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ADDITIONAL KEYWORDS:- achene anticlinal walls exocarp - generic delimitation pericarp - surface patterns - tribal classification.

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^{*} Author for correspondence: Louis.RonseDeCraene@bio.kuleuven.ac.be

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INTRODUCTION

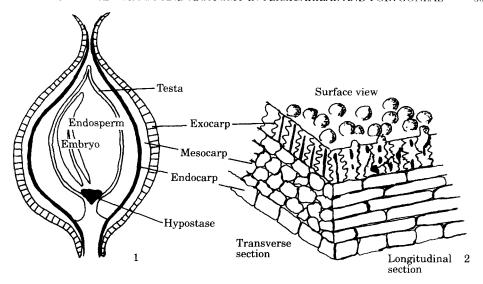
Subdivision of the large, heterogeneous genus *Polygonum* into a number of smaller, more natural units is a current necessity with a number of recent proposals (cf. Haraldson, 1978; Ronse Decraene & Akeroyd, 1988; Hong, 1992). This paper is the second in a series aiming to critically analyse and improve the delimitation and infrageneric relationships of tribes Persicarieae and Polygoneae presented by Ronse Decraene & Akeroyd (1988). In a previous investigation, Hong, Ronse Decraene & Smets (1998) studied the adaxial tepal epidermis of a number of species belonging to both tribes. Two main types could be recognized, corresponding to the tribes Persicarieae and Polygoneae, plus a third type of papillose cells, shared by some species of *Fagopyrum* and *Oxygonum*.

Classification of *Persicaria* at the sectional level is still not fully clarified, and recognition of genera such as *Aconogonon*, as accepted by Hong (1992), and *Bistorta*, separated from *Persicaria*, needs further investigation. The affinities of *Polygonella* to *Polygonum s.s.* have been recently investigated by Ronse Decraene, Hong & Smets (in press); palynology, fruit anatomy, and the general flower structure are overwhelming evidence for treating *Polygonella* as a section of *Polygonum*.

The taxonomic value of fruit morphology has been recognized early (see e.g. Marek, 1954, 1958; Kanai, Takeno & Taniguchi, 1983; Wolf & McNeill, 1986; Yang, Li & Wang, 1991; Hong, 1992, 1993; Hedberg, 1997), although fruit characters have mostly been investigated locally for a limited number of taxa and not at the level of entire tribes. In order to increase our knowledge of fruit morphology and anatomy in *Polygonum* s.l. and critically assess the classification of Ronse Decraene & Akeroyd (1988), we studied a representative number of species of tribes Persicarieae and Polygoneae.

General description of fruit in Polygonaceae

Fruits of Polygonaceae are invariably indehiscent achenes or nuts formed by three carpels (pyramidal or triangular: Figs 21, 36, 55, 60, 61, 70) or two carpels (lenticular fruits: Fig. 16). Rarely, there are four carpels, which is a constant feature for the genus *Calligonum*. The number of styles or stigmatic lobes corresponds with the number of carpels. The fruit is enclosed by the persistent perianth, which may play an important role in the dispersal mechanisms. For example, the tepal lobes can become accrescent and covered with hooks in *Rumex* and *Emex*. In *Persicaria* sect. *Tovara* tepals are undifferentiated but dispersal occurs through the hooked persistent

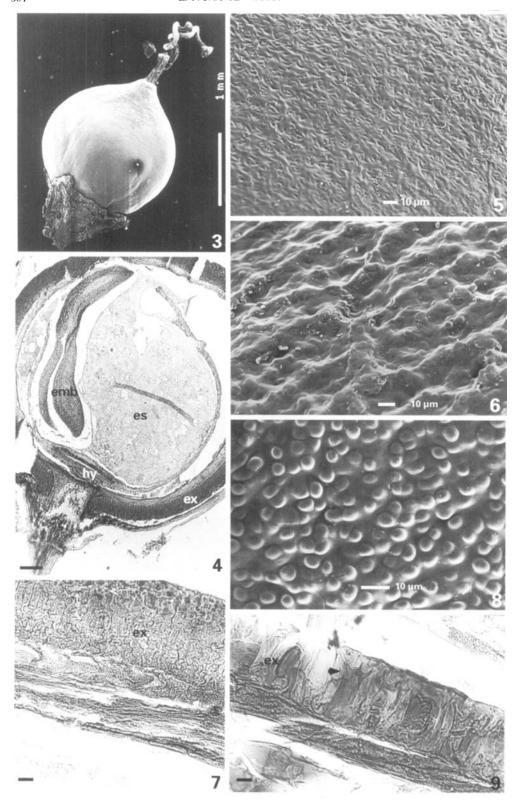


Figures 1, 2. Fig. 1. Schematic LS fruit of Polygonaceae. Fig. 2. Details of transverse section (TS) and longitudinal section (LS) of pericarp. Different shapes are shown for exocarp cells in TS and LS. However, both sections are similar for a given species.

styles (Fig. 16). In *Persicaria lapathifolia* (sect. *Persicaria*) the vasculature of the disintegrating tepals enclosing the fruit functions as an agent of dispersal, developing as anchor burrs (Wisskirchen, 1991).

The morphological configuration of the fruit (an achene) is relatively simple and basically similar in all Polygonaceae (Fig. 1; e.g. Dammer, 1893; Graham & Wood, 1965; Roth, 1977; Brandbyge, 1993). The three or two carpels enclose an orthotropous ovule arising from the apical meristem of the flower. Vautier (1949) studied the vascular arrangement of several taxa using a clearing treatment for whole flowers. The vasculature in the ovary is represented by three dorsal traces running into the styles; they are located in the mesocarp and may be flanked with marginal traces. The single placental trace (rarely three, as in Fagopyrum) is well-developed. The vasculature is sometimes interrupted (especially in younger stages) by a non-lignified zone at the level of the hypostase (Figs 1, 4). There has been discussion in the past about the origin of the free-central placentation (e.g. Laubengayer, 1937; Vautier, 1949). While the ovule develops as a seed, the carpel wall differentiates as the pericarp (Figs 1, 2).

The most conspicuous feature of the pericarp is the exocarp or epidermis, which is sclerified in most cases (Figs 1, 2). The young fruit consists of a well developed exocarp (epidermis), several layers of parenchymatous mesocarp cells (sometimes subdivided in an outer and inner portion, depending on the inclusion of cell contents) and an endocarp. During maturation the inner layers of the fruit wall (mesocarp and endocarp) become mostly flattened or destroyed. Only the exocarp increases in size and becomes heavily sclerified. Thickenings most commonly occur on the anticlinal (radial) walls, but not on the inner tangential (periclinal) walls. As thickening of the anticlinal walls increases from the inner tangential wall to the outer, the lumen often takes a triangular to trapezoid shape in section. The thickening is often U-shaped or horseshoe-shaped (cf. Roth, 1977). Very often small canals branch



from the triangular lumen into the surrounding sclerenchymatous tissue. Due to space constraints the anticlinal walls can become variously convoluted or bent in cross- or longitudinal sections (Fig. 2). As convolutions of the anticlinal walls may also occur tangentially this gives the cells a star- or puzzle piece-shape in surface view or in paradermal section, or the cells appear polygonal if no convolutions occur. Shifts in shape may occur within the cells from the periphery to the mesocarp (cf. Lonay, 1922). The outer surface of the epidermis represents interesting systematic characters, as emphasized by Barthlott (1981, 1990). Barthlott distinguished between primary cell sculpturing (including cell shape, cell boundaries and curvature of outer periclinal wall), secondary sculpturing (including microscopic sculpturing of an often cuticular nature), and tertiary sculpturing (epicuticular wax secretions). These characteristics apply equally for the fruit surfaces of Polygonaceae, which can be highly variable. However, in the past the descriptive terminology has often been vague, ill-defined and sometimes technically incorrect (cf. Wolf & McNeill, 1986). SEM studies reveal smooth, pitted surfaces, surfaces with a reticulate thickening, or surfaces covered with tubercles (Fig. 2). Wolf & McNeill (1986) recognized four surface patterns: smooth, roughened, papillose and striate-papillose. The achene can sometimes be supplied with spines and wings (e.g. Fagopyrum giraldii, Oxygonum, Pteropyrum).

Seeds are small and the testa is reduced to the outer integument with little specialization. The seed develops an outer layer of rectangular cells (exotesta), surrounding a mealy or horny endosperm and embryo. The embryo is mostly straight with incumbent cotyledons and is positioned excentrically in one corner of the seed. The outer layer of the endosperm is differentiated as a nutritional layer (Neubauer, 1971), which is a rare feature in dicotyledons. Seed and fruit development are little known in Polygonaceae (Corner, 1976), except for detailed studies by Neubauer (1971) on *Persicaria pennsylvanica* and Lonay (1922) on *Polygonum aviculare*. Corner (1976) was uncertain whether the seeds are truly exotestal or merely undeveloped because they are enclosed in indehiscent syncarpous fruits.

MATERIAL AND METHODS

The study of fruits was conducted mostly on material taken from herbarium specimens on loan from the following herbaria (abbreviations according to Holmgren, Holmgren & Barnett, 1990): BOL, BM, BR, E, GB, K, LD, LV, S, and UPS, but also partly from pickled material. A list of all species and specimens investigated is provided in the Appendix.

Figures 3–9. Fruits of *Persicaria* sect. *Persicaria* (Figs 3–8) and sect. *Rubrivena* (Fig. 9). Figs 3, 5, 6, 8: SEM. Figs 4, 7, 9: LM. Fig. 3. Lateral view of mature fruit of *P. attenuata* (Mohoro 588) with twisted styles and globular fruit. Scale bar = 1 mm. Fig. 4. *P. viscosa* (Ryding 322). LS mature fruit; note thick pericarp. Scale bar = 100 μ m. Fig. 5. *P. attenuata* ssp. *pulchra* (Mohoro 588). Detail of fruit outer surface with pits arranged in star-like pattern. Scale bar = 10 μ m. Fig. 6. *P. hydropiper* ssp. *microcarpum* (H. Smith 4778). Detail of fruit outer surface with irregular ridges on anticlinal cell walls. Scale bar = 10 μ m. Fig. 7. *P. acris* (Rosas 362). LS pericarp. Scale bar = 10 μ m. Fig. 8. *P. orientalis* (Brumbach 6921). Detail of fruit outer surface with tubercles more or less set on anticlinal cell walls. Scale bar = 10 μ m. Fig. 9. *P. wallichii* (Ronse Decr. 15Le). LS pericarp. Arrow points to dendritic branching of cell lumen. Scale bar = 10 μ m. Abbreviations: emb = embryo; es = endosperm; ex = exocarp: hy = hypostrase.

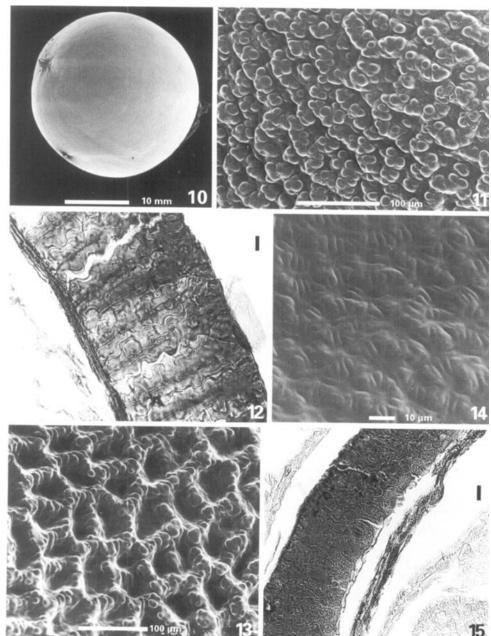
The following fruit characters were investigated:

- (1) External structure. This includes fruit shape (correlated with carpel number), presence versus absence of a beak (e.g. in Aconogonon campanulatum: Hong, 1992) on which the style is inserted, presence of one or more styles which are free or fused at the base, and whether fruits are stipitate versus non-stipitate (the achene can have a stalk or be sessile).
- (2) Perianth structure (accrescent versus non accrescent). For example, in species of Fagopyrum the perianth persists as small basally inserted appendages, while the inner perianth whorl becomes strongly accrescent in Rumex.
- (3) Fruit surface patterns. We follow the terminology of Barthlott (1981, 1990). Cell shape (primary sculpturing) is hardly visible externally, as cell boundaries are often unclear. Cells are most often isodiametric, rarely elongated (Fig. 28). Anticlinal walls are either straight or undulating, giving a puzzle piece-shaped pattern to the surface. At one extreme the anticlinal walls are not visible externally and the whole fruit surface is smooth (Figs 3, 10, 54, 60); in other cases the walls are collapsed and present as small pits lying radially around the cell (Figs 5, 17), as a continuous ridge or folds (Figs 6, 22, 24, 28, 30, 43, 51, 68, 78), or high walls giving a reticulate pattern to the surface (Figs 13, 46). Papillae (with a diameter below 10 µm: Figs 8, 11, 13, 14, 71) or strongly developed tubercles (with a diameter exceeding 10 µm: Figs 37, 50, 62) may be superimposed on the anticlinal walls, in some cases delimiting the boundaries of the cells when no ridge is visible (Fig. 37). In other instances the tubercles appear set in radial lines (Fig. 62; striate-papillose according to Wolf & McNeill, 1986). Papillae or tubercles are not necessarily linked with convoluted anticlinal walls, and their number may be variable. Ridges are not necessarily connected with the cell boundaries (Fig. 49). The outer periclinal wall of the cell is either flat (Figs 32, 46), or convex (Fig. 35), although this may be related to the collapsing of the wall. In Oxygonum short trichomes are present (Fig. 74).

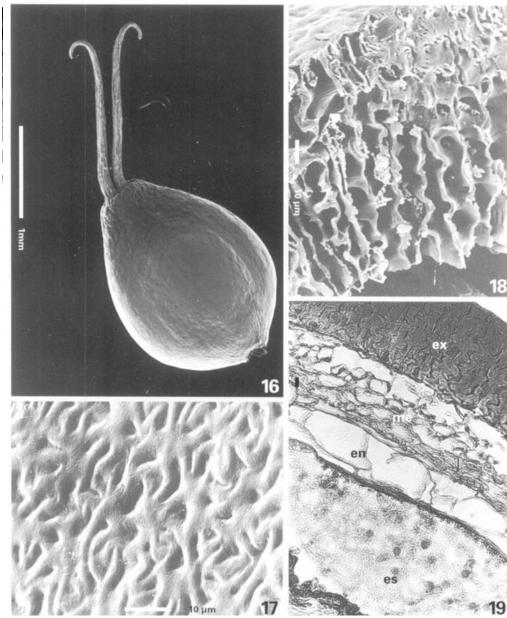
The secondary sculpturing of the cell wall consists in some cases of longitudinal cuticular striae, which may be weakly or strongly developed (Figs 54, 59). The limits between micropapillae (which are part of the secondary sculpture according to Barthlott), papillae, and tubercles are a matter of degree. Epicuticular wax secretions are only occasionally present (e.g. *Polygonum undulatum*, *P. minimum*).

(4) Anatomical structure of pericarp. In the simplest case we found one layer of cells aligned in parallel, the exocarp, resembling a palisade. These were normally stone cells, with a variable distribution of pit channels. Mesocarp and endocarp were often torn apart and were therefore mostly not included in the observations. Characters of testa and embryo were also not studied here.

Exocarp cell shape (in longitudinal or cross section) was square (isodiametric), shortly rectangular, or elongated-rectangular. Anticlinal walls were either straight (resembling 'corrugated sheets': Figs 20, 23, 25, 26, 29, 48, 54, 58, 64, 66, 69, 72), or undulating (resembling 'egg cartons': Figs 7, 12, 15, 18, 19, 38, 44, 50, 57, 65, 76, 78). Cell lumen size depends on cell wall thickening, which may be weak (limited to the outer tangential and radial walls, leading to a broad lumen: Figs 9, 42, 48, 52, 56), trapezoid or triangular (narrowing towards the periphery: Figs 29, 41, 47, 54, 63, 73), completely enclosed by thickenings (also with thickenings of the inner tangential wall: Figs 79, 80), or narrow with or without connection to the periphery (Figs 12, 19, 57, 64, 65, 66, 69, 72, 76, 77). In addition the lumen may show branches



Figures 10–15. Fruits of *Persicaria* sect. *Echinocaulon* (10–12) and sect. *Cephalophilon* (13–15). Figs 10, 11, 13, 14: SEM. Figs 12, 15: LM. Fig. 10. *P. perfoliata* (H. Smith 10000). Apical view of globular fruit. Scale bar = 10 mm. Fig. 11. *P. sieboldii* (Ohwi 2569). Surface detail with tubercled anticlinal cell walls. Scale bar = 10 μm. Fig. 12. *P. hastato-sagittata* (Kanai 10383). LS exocarp. Scale bar = 100 μm. Fig. 13. *P. alata* (Hedberg s.n.). Fruit surface; polygons surrounded by tubercled wall. Scale bar = 100 μm. Fig. 14. *P. runcinata* (Ludlow & Sherriff 8250). Detail of fruit with shallow papillae on anticlinal cell walls. Scale bar = 10 μm. Fig. 15. *P. capitata* (Polunin *et al.* 5248). TS fruit. Scale bar = 10 μm.



Figures 16–19. Fruits of *Persicaria* sect. *Tovara*. Figs 16–18: SEM. Fig. 19: LM. Fig. 16. Mature fruit of *P. neofiliformis* (Ohwi 331); fruit lenticular with long hooked styles. Scale bar=1 mm. Figs 17–18. *P. virginiana* (Ronse Decr. 214 Lb). Fig. 17. Surface, with pits set in a star-like pattern. Scale bar=10 µm. Fig. 18. TS pericarp showing cells with convoluted anticlinal walls and 'swiss cheese'-holes. Scale bar=10 µm. Fig. 19. *P. virginiana* (McDonald s.n.). LS fruit. Scale bar=10 µm.

or canals situated at the outer limits of the lumen and branching dichotomously or trichotomously (Figs 9, 23, 25, 26, 40, 41, 63, 64, 66, 73), or as dendritic ramifications along the anticlinal sides of the lumen (as a result of the convolutions of the wall: Fig. 65). As seen in paradermal (tangential) section the shape of the palisade may

change at different levels of the cell. Near the outer surface the cells often have undulating anticlinal walls (as discerned in surface view); towards the mesocarp the same cell may be polygonal with straight anticlinal walls. However, in cross or longitudinal sections the anticlinal walls may appear straight, although undulating in surface view (e.g. *Aconogonon, Bistorta, Fallopia, Koenigia*). One must distinguish between the undulations of the anticlinal walls seen in cross or longitudinal sections from those seen in paradermal section, confirming the existence of two different patterns.

For examination of fruits, well-developed, mature flowers were selected. Material was examined with the dissecting microscope, light microscope (LM) and scanning electron microscope (SEM). Most material from herbarium specimens was directly mounted onto aluminum stubs, coated and photographed as outlined below. A number of fruits was also treated with aerosol-OT (dioctyl sodium sulfosuccinate) following Erbar (1995) in order to compare dried fruits with rehydrated fruits. Sectioning of fruits was problematical, as the material was too hard to cut or became overstained. A solution to this was soaking the fruits in warm 10% KOH for a couple of hours, and the use of Stockwell's bleach (Schmid, 1977) to remove extra tannins. Material was subsequently treated with aerosol-OT and stored in 70% ethanol before sectioning.

Fruits collected from pickled material were prepared for the SEM as follows. Samples were dehydrated using formaldehyde-dimethylacetal (Gerstberger & Leins, 1978). Critical point drying was carried out using a CP-Drier (CPD 030, Balzers). The dried samples and those taken from the herbarium specimens were mounted on aluminium stubs and coated with c. 30 mm layer of Au or Au/Pd before being studied under the SEM (JEOL T-20, JEOL-5200 and JEOL-6200), operating at 15 kV, in Seoul, Reading and Leuven respectively.

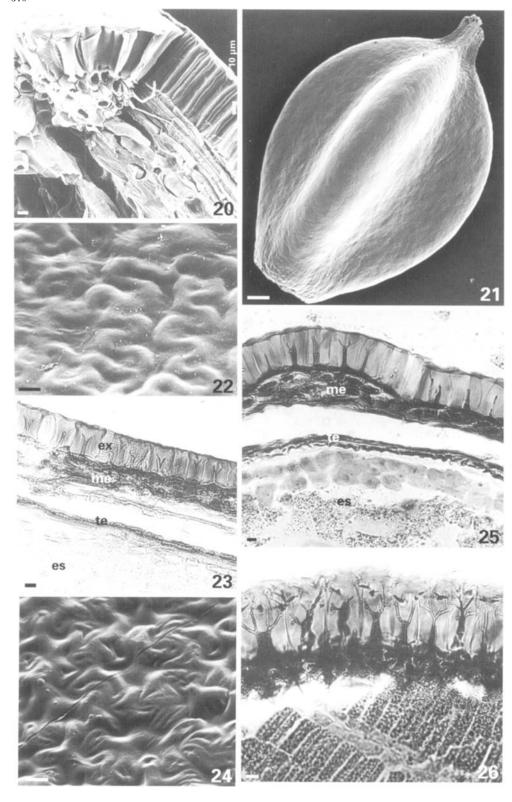
RESULTS

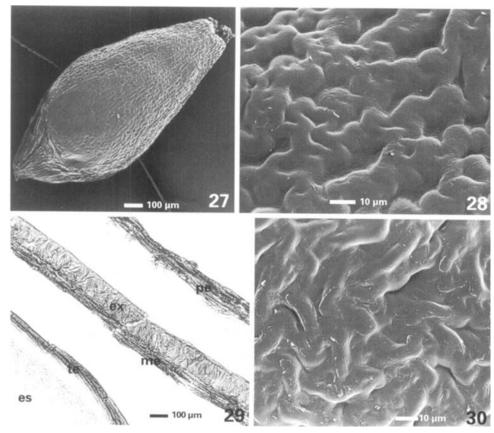
Tribe Persicarieae

Genus Persicaria Mill. [including sects. Persicaria (Mill.) Gross, Echinocaulon (Meisn.) H. Gross, Cephalophilon (Meisn.) H. Gross, Rubrivena (M. Král) S.-P. Hong, Tovara (Adans.) H. Gross] (Figs 3–19)

In all taxa of *Persicaria* investigated except *P. wallichii* (Fig. 9), the epidermal cells are considerably higher than wide in cross section, with sinuate anticlinal walls (Figs 7, 12, 15, 18, 19). The thickness of the exocarp varies between 50 and 120 μ m. The surface is rarely smooth, often pitted along the cell walls or with anticlinal walls covered by papillae or tubercles. The lumen of the cells is very narrow, running straight towards the periphery and is without branching.

Globular fruits are found in section *Persicaria (Persicaria attenuata:* Fig. 3, *P. viscosa:* Fig. 4, *P. strigosa)*, or contrary to other sections the fruit is often elongated with a short beak (e.g. *P. acris*), bicarpellate and with almost no stipe, embedded in the perianth bases (Fig. 4). The fruit surface pattern is almost smooth (eventually with small pits set radially around the cells: *P. attenuata:* Fig. 5), with a variable number of papillae set on anticlinal ridges (*P. viscosum*), in *P. hydropiper* with the ridges following a longitudinal pattern (Fig. 6), or it consists of numerous tubercles delimiting





Figures 27–30. Fruits of *Koenigia*. All scale bars = 10 μm, except Fig. 27 = 100 μm. Figs 27, 28, 30: SEM. Fig. 29: LM. Fig. 27–28. *K. islandica* (Horsen 601). Fig. 27. Lateral view of mature fruit. Fig. 28. Surface detail; elongated cells with folded anticlinal cell walls. Fig. 29. *K. nepalensis* (Einarsson *et al.* 3394). LS fruit wall. Fig. 30. *K. nummularifolia* (Stainton 728). Surface detail. Abbreviations: es = endosperm; ex = exocarp; me = mesocarp; pe = perianth; te = testa.

individual cells (*P. bungeana*, *P. orientalis*: Fig. 8). The exocarp cells are narrowly rectangular (70–150 μ m thick), with undulating anticlinal walls and narrow unbranched lumina (Fig. 7).

Fruits of sect. Echinocaulon are broadly triangular (e.g. Persicaria sieboldii, P. hastato-sagittata), rounded to almost globular (Persicaria perfoliata: Fig. 10), shortly stipitate, with a narrow beak. Tepals are not accrescent and shrivel at fruiting. Externally the surface consists of randomly grouped tubercles separated by small pits or deeper

Figures 20–26. Fruits of Aconogonon (Persicaria sect. Aconogonon) and Bistorta (Persicaria sect. Bistorta). All scale bars = 10 µm, except Fig. 21 = 100 µm. Figs 20–22, 24: SEM. Figs 23, 25, 26: LM. Fig. 20. Section through pericarp of A. molle var. rude (Ronse Decr. 642 Le). Fig. 21. Lateral view of fruit of B. purpusella (Ludlow et al. 21115). Fig. 22. A. molle var. rude (Ronse Decr. 642 Le). Surface detail with folded puzzle piece-like cell walls. Fig. 23. B. griffithii (Ludlow et al. 13097). LS fruit. Fig. 24. B. macrophylla (H. Smith 10932). Surface detail. Fig. 25. A. hooken (Smith 4172). LS fruit. Fig. 26. A. molle var. rude (Hooker & Thompson s.n.). Section of fruit. Abbreviations: es=endosperm; ex=exocarp; me=mesocarp; te=testa.

invaginations. The number of tubercles ranges from a few large papillae arranged on the anticlinal wall (*P. hastato-sagittata*) to a high number of papillae grouped in polygons around depressions (*P. sieboldii*: Fig. 11). Anatomically one finds similar exocarp cells as in previous section (Fig. 12).

In section Cephalophilon one finds identical rectangular cells with undulating anticlinal walls (Fig. 15). The external fruit surface is constructed along the same pattern as in sect. Echinocaulon and consists of depressions (the periclinal wall of the cells) bordered by ridges covered with tubercles (P. alatum: Fig. 13, with a reticulate pattern, P. nepalensis). In P. runcinata the papillae are weakly developed in a position similar to the tubercles in other species (Fig. 14). Tepals are not accrescent but are larger than in section Echinocaulon. Fruits are more elongated and rounded in outline with a short stipe.

Section *Tovara* has bicarpellate lenticular fruits without beak and topped by two long styles with curved apex (Fig. 16). There is almost no stipe and the fruit surface is almost smooth, consisting of scattered small pits (in *P. virginiana*: Fig. 17) or shallow depressions in rows (in *P. neofiliformis*: Fig. 16), representing the collapsed anticlinal walls. In cross-section one finds a narrowly elongated exocarp with undulating tangential walls (Figs 18, 19). The mesocarp is loose and in some cases an endocarp of large cells is visible (mostly torn apart). The outer layer of the seed consists of darkly staining cells (Fig. 19). With the SEM one can observe holes in the undulating anticlinal walls which are connections between the lumina of adjacent cells.

Section Rubrivena was erected by Hong (1993) to include two polymorphic species *P. wallichii* Greuter & Burdet and *P. pinetorum* (Hemsley) H. Gross. Ronse Decraene & Akeroyd (1988) included *P. wallichii* in their section Aconogonon (as Persicaria polystachya (Wall. ex Meisn.) H. Gross). The exocarp of *P. wallichii* consists of rectangular cells with straight anticlinal walls and broad ellipsoidal lumen, branching dichotomously towards the outer tangential wall (Fig. 9). External fruit morphology and fruit surface are illustrated by Hong (1993).

Aconogonon (Figs 20, 22, 25, 26)

Data have been partly obtained from Hong (1989, 1992). In Aconogonon exocarp thickness varies between 30 and 65 μ m. Fruits are trigonous, often with prominent ribs and almost no beak (except in A. campanulatum). Styles are short and topped with a cap-like stigma. The achenes are included, slightly exserted to well exserted above the persistent tepals.

Fruit surfaces have cells with undulating anticlinal walls, and are smooth (A. coriarium), sometimes slightly rough with weak depressions (A. hookeri, A. campanulatum, A. angustifolium), or with weak undulating ridges following the cell wall (A. molle: Fig. 22); there is no secondary sculpturing and rarely dispersed epicuticular wax secretions (A. hookeri). The pericarp is uniform in LM view with an exocarp of rectangular to square cells with straight anticlinal walls (Figs 20, 25, 26) and cylindrical lumina often branching dichotomously towards the outer periclinal wall. Dichotomous branching may be repeated (Fig. 26).

Bistorta (Figs 21, 23, 24)

Fruits are similar to those of *Aconogonon* with well developed ribs, but often more elongated (Fig. 21). There is virtually no stipe but the beak is well developed. Styles are short and topped with a cap-like stigma. The achenes are included, slightly

exserted to well exserted above the persistent tepals. Fruit surfaces are smooth with tesselated cell outline, sometimes slightly pitted (*B. perpusella*), rough with weak folds (*B. griffithii*), or with undulating ridges along the anticlinal walls (*B. macrophylla*: Fig. 24). The exocarp consists of narrowly rectangular cells with straight anticlinal walls, and with a narrow lumen branching dichotomously towards the periphery (Fig. 23).

Koenigia (Figs 27–30)

Compared with *Persicaria* the exocarp anatomy of *Koenigia* is less elaborate and consists of one layer of more or less isodiametric cells in cross section (see also Hedberg, 1997). The cells show weak anticlinal thickenings delimiting a trapezoid lumen (Fig. 29). The mesocarp is flattened. Fruits are either elongated or globular with a short beak; there is a short stipe present (Fig. 27). The epidermal surface is either nearly smooth with inconspicuous ridges (*K. nepalensis*) or convolute with few to many folds on the anticlinal walls of the cells (*K. nummularifolia*, *K. islandica*: Figs 28, 30). In *K. islandica* the convolutions run in longitudinal rows and suggest an elongated shape of the cells (Figs 27, 28). Hedberg (1997) described the surface of the cells as smooth; otherwise his description corresponds with our data.

Fagopyrum (Figs 31–35)

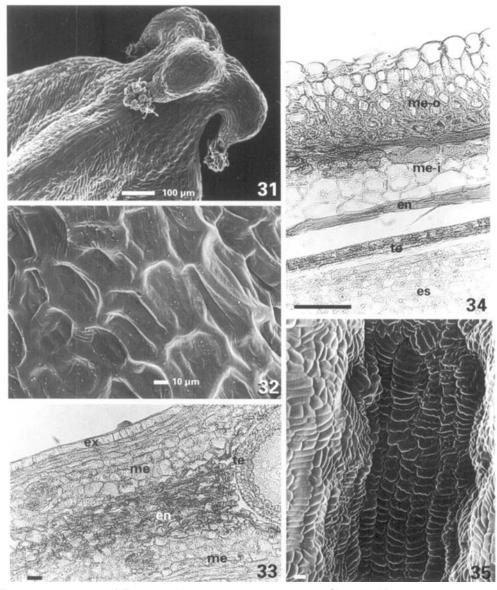
Mature fruits of Fagopyrum are readily recognizable by their massive triangular body with well developed ribs and alternating depressions, a non-accrescent perianth which persists at the base of the fruit, and the short tripartite style split to the base with reflexed, persistent style branches and small stigmas (Fig. 31). Fruit surface patterning is non-differentiated and thin-walled, and consists of rectangular cells which are collapsed in dry state (Figs 32, 35). Cell size may differ between the ribs and the intervening regions (Fig. 35). In section the pericarp consists of several layers (Figs 33, 34). The exocarp (epidermis) is thin-walled and isodiametric to rectangular in shape. The mesocarp consists of several cell layers: an outer mesocarp has small cylindrical cells occasionally becoming thickened towards the inner layers (Fig. 34), a transitional zone consisting of tannin-filled cells, and an inner mesocarp of parenchymatous cells containing starch. In other cases no thickenings are found (Fig. 33). There are several vascular bundles distributed in the mesocarp. The endocarp consists of larger parenchymatous cells disintegrating on the internal side of the fruit (Figs 33, 34).

Tribe Polygoneae

Polygonum L. s.s. [including sect. Polygonum, Pseudomollia, Duravia, Tephis, and Polygonella] (Figs 36-59)

A detailed description of fruit structure and anatomy is given in Ronse Decraene, Hong & Smets (in press).

Fruits of sect. *Polygonum* are variously included or exserted from the perianth. The shape of the fruit is ellipsoidal-pyriform, often unequally triangular with one side broader than the other two, with a short stipe and long beak (Fig. 36). Styles are basally fused with capitate stigmas. Several species of sect. *Polygonum* and some of sect. *Duravia* have a distinctively tubercled surface pattern. Tubercles may be equidistant and sparsely covering the fruit (e.g. *P. bellardi*, *P. equisetiforme*), crowded



Figures 31–35. Fruits of Fagopyrum. All scale bars = 100 μm, except Fig. 32 = 10 μm. Figs 31, 32, 35: SEM. Figs 33, 34: LM. Fig. 31. F. esculentum (Reekmans s.n.). Detail of mature fruit; three hooked styles with terminal stigmas clustered with pollen. Fig. 32. F. cymosum (H. Smith 13315). Detail of collapsed cells on fruit surface. Fig. 33. F. esculentum (Ronse Decr. 30 Le). TS fruit; scattered bundles in mesocarp, crushed endocarp and absence of inner space between opposite wall-layers. Figs 34, 35. F. tataricum (Ronse Decr. 1335). Fig. 34. LS fruit and outer surface of fruit. Note the mesocarp with thickened walls. Fig. 35. Fruit surface with detail of interrib region. Abbreviations: en = endocarp; ex = exocarp; me = mesocarp (inner and outer); te = testa.

on the anticlinal ridges (e.g. *Polygomum achoreum*, *P. erectum*: Fig. 37), or in longitudinal rows without connection to the anticlinal walls of the cells (e.g. *P. arenastrum*, *P. aviculare*: Fig. 36). Tubercles may be associated with pits, but never with superficial

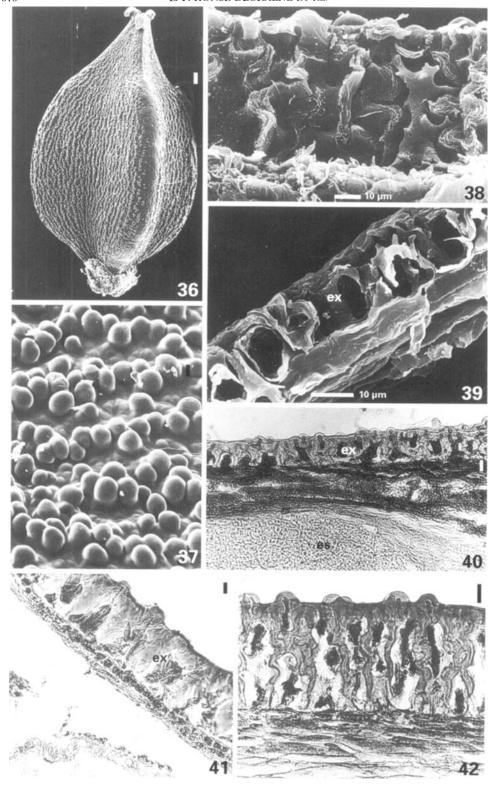
striae. In other species the fruit surface is smooth (e.g. *P. maritimum*) or the upper half of the fruit is tubercled and the lower half smooth (e.g. *P. oxyspermum*, *P. arenastrum*). In the exocarp of sect. *Polygonum* two kinds of cell structure were observed. In several species the anticlinal walls are strongly convolute with numerous folds and gaps in the walls (Figs 38, 44). The lumen of the cells is broadly rectangular with dendricular branching along the sides (e.g. *P. ramosissimum*, *P. tubulosum*, *P. equisetiforme*, *P. bellardi*). The exocarp is usually thick (above 60 µm). The second cell type consists of smaller, isodiametric or shortly rectangular cells with a broad lumen, triangular to trapezoid in shape with the smallest side towards the periphery. The lumen is often dichotomously or trichotomously branching towards the periphery (Figs 39, 40; e.g. *P. oxyspermum* var. *raii*, *P. arenastrum*, *P. aviculare*). The exocarp is thinner (less than 40 µm). Some species have exocarps that are intermediate between both types (e.g. *P. bellardi*: Fig. 41), depending on the origin of the material.

P. sect. Tephis (with two species P. afromontanum and P. undulatum) is distinctive in its lenticular cells with long styles and capitate stigmas. The fruit surface is almost smooth or the outer periclinal walls are curved with slightly undulating anticlinal walls filled with epicuticular secretions (Fig. 43). In section, the exocarp consists of long rectangular cells (about 70 μm thick) with undulating anticlinal walls and a narrow lumen (Figs 44, 45).

The fruit surface of *P. molliaeforme* (sect. *Pseudomollia*) is distinctive with a roughly reticulate surface formed by the straight anticlinal walls (Fig. 46) along the wings lining the beak of the fruit; the reticulation is more compact and is covered with tubercles in some places (see also Ronse Decraene, Hong & Smets, in press). In section the exocarp consists of square cells with straight anticlinal walls and tangential lumina, similar to species of *Polygonella* and sect. *Duravia* (Fig. 47).

P. sect. Duravia shows the same variability as sect. Polygonum. Smooth fruit surfaces are found in P. minimum and P. paronychia. The fruit surface is almost smooth with low ridges (P. bolanderi) or with pits (P. shastense, P. californicum) along the anticlinal walls depending on the material, and there is an occasional superficial cuticular striation present. The fruit surface of P. polygaloides s.l. (including P. kellogii, P. confertiflorum) is the most distinctive and shows a reticulation of longitudinally running ridges bridged at regular intervals (Fig. 49). These ridges do not necessarily correspond with the anticlinal cell walls. P. douglasii and P. tenue are polymorphic in having some materials with a smooth surface and others with tubercles more or less following the anticlinal walls (Figs 50, 51, 53). The exocarp is most often narrow with isodiametric cells, straight anticlinal walls, and a broadly trapezoid lumen (Figs 48, 52). In a few species (e.g. P. douglasii, P. tenue: Figs 50, 53), the exocarp resembles that of sect. Polygonum in being rectangular with undulating anticlinal walls.

In *Polygonella* the outer surface of the fruit is most often smooth (*P. americana*, *P. gracilis*, *P. polygama*: Figs 54, 55, 59), or with weak depressions along the anticlinal walls (*P. articulata*). Longitudinal cuticular striae are often present (Figs 54, 59), but not in *P. fimbriata* or *P. gracilis*. *P. fimbriata* has a combination of a close packing of tubercles in the upper part of the fruit (without relation to the anticlinal walls), and a roughly pitted surface in the lower part (see also Ronse Decraene, Hong & Smets, in press). The shape of the fruit is narrowly triangular with prominent ribs, a long beak and short stipe. The short styles with globular stigmas drop early. Most species have a narrow exocarp (40 µm thick) of square cells with straight anticlinal walls and limited thickening (Figs 54, 56, 58). The only exception with rectangular cells, a narrow dendritically branching lumen, and undulating anticlinal walls is *P. americana* (Fig. 57).



Fallopia (Figs 60–66)

Three to four sections are recognized (see Holub, 1970; Haraldson, 1978; Ronse Decraene & Akeroyd, 1988). Fruits are triquetrous with strong equal ribs, a beak is virtually absent or very short (except for sect. *Reynoutria*: Fig. 60) and styles are short and fused with capitate stigmas (Fig. 61; except for sect. *Reynoutria* with free styles and fimbriate stigmas). Both sect. *Reynoutria* and sect. *Sarmentosae* possess winged, enlarged outer tepals surrounding the fruit.

The fruit surface is either smooth (e.g. *F. cilinodis*, *F. sachalinensis*, *F. multiflorum*), with unequal ridges following the anticlinal cell walls (*F. japonica*), or covered with tubercles without link to the anticlinal walls (e.g. *F. convolvulus*, *F. dumetorum*, *F. aubertii*: Figs 60–62). Tubercles may be distributed without apparent order (e.g. *F. dumetorum*) or in regular sequences or lines (*F. convolvulus*: Fig. 62, *F. scandens*, *F. aubertii*). The exocarp is often represented by rectangular cells with straight anticlinal walls and a narrow lumen dichotomously or trichotomously branching towards the periphery (Figs 64, 66). There is a gradation towards cells with undulating anticlinal walls. In *Fallopia convolvulus* we found a dendritically branching lumen (probably the result of the convoluting radial walls: compare with Roth, 1977: fig. 12b) and anticlinal walls were slightly undulating (Fig. 65). Section *Reynoutria* differs in the more isodiametric exocarp cells with broad trapezoid lumen (Fig. 63) reminiscent of those found in some species of sect. *Polygonum*.

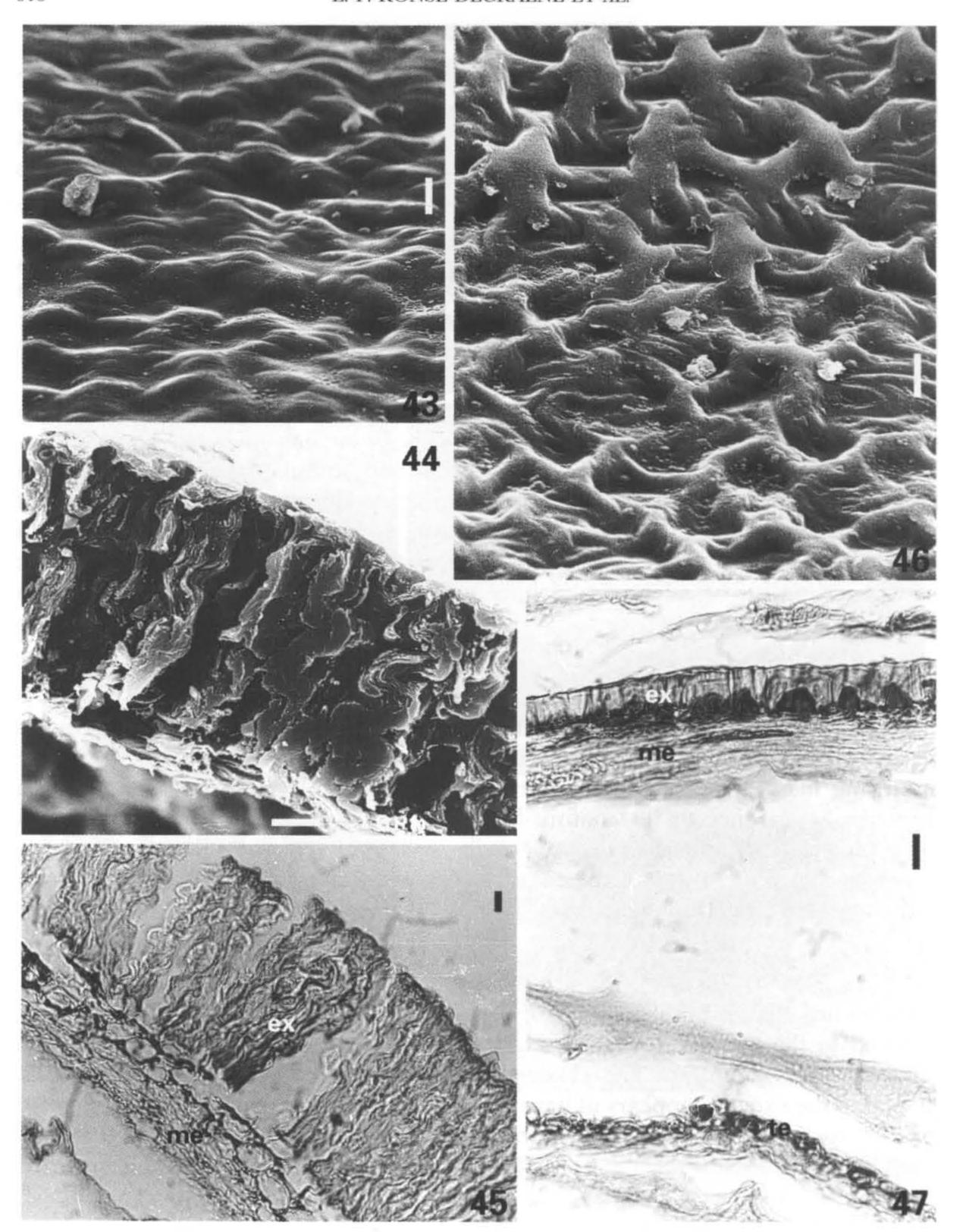
Calligonum (Figs 67–69)

Fruits are tetramerous, with a peculiar dispersal mechanism. Perianth segments are deflexed in the fruit and the achene surface is covered with strong dendritically branching hairs or cristae, arising along the ribs of the young fruit and giving a shrublike appearance to the mature fruit (Fig. 67). In between the ribs, the fruit surface consists of smooth polygonal cells (collapsed: Fig. 68). The exocarp (*C. eriopodum*: Fig. 69) consists of rectangular cells with narrow lumina dichotomously branching and straight anticlinal walls.

Atraphaxis (Figs 70–73)

Fruits are 'hazelnut'-like (Fig. 70: A. suaedifolia) or lenticular (e.g. A. spinosa), enclosed in the accrescent inner perianth parts. The fruit surface is covered with small papillae. (A. buxifolia: Fig. 71) or pits (A. frutescens) set along the anticlinal walls. In section the exocarp appears to have rectangular cells with narrow (e.g. A. pyrifolia: Fig. 72) to broad lumina (A. billardieri: Fig. 73), dichotomously or trichotomously branching towards the periphery. In the latter species the shape of the lumen is trapezoidal.

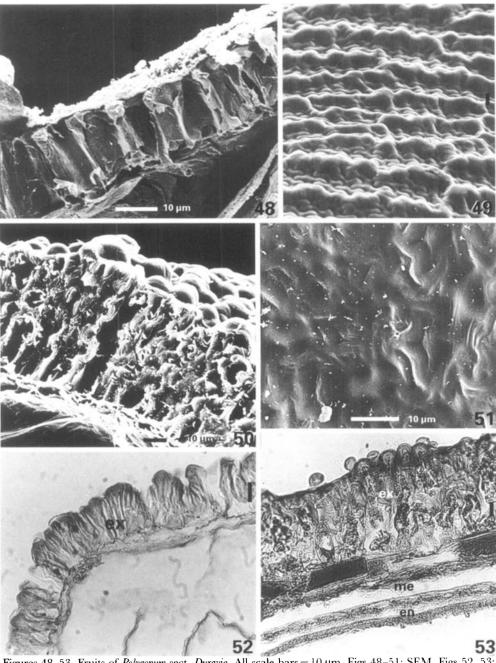
Figures 36–42. Fruits of *Polygonum* sect. *Polygonum*. All scale bars = 10 μm, except Fig. 36 = 100 μm. Figs 36–39: SEM. Figs 40–42: LM. Fig. 36. *P. aviculare* var. *aequale* (Ronse Decr. 25 Lb). Lateral view of mature fruit; tubercles arranged in longitudinal rows. Fig. 37. *P. erectum* (Bebb s.n.). Surface detail. Fig. 38. *P. ramosissimum* (Puissant s.n.). Section through exocarp. Note strong convolution and holes in anticlinal walls. Fig. 39. *P. arenastrum* (Ronse Decr. 1157). Section through exocarp. Note nearly isodiametric cells and straight anticlinal walls. Fig. 40. *P. oxyspermum* var. *raii* (Corbière 5306). LS fruit wall. Fig. 41. *P. bellardi* (Heldreich 877). LS pericarp. Fig. 42. *P. ramosissimum* (Puissant s.n.). LS pericarp; note surface tubercles and convoluted anticlinal walls. Abbreviations: es = endosperm; ex = exocarp.



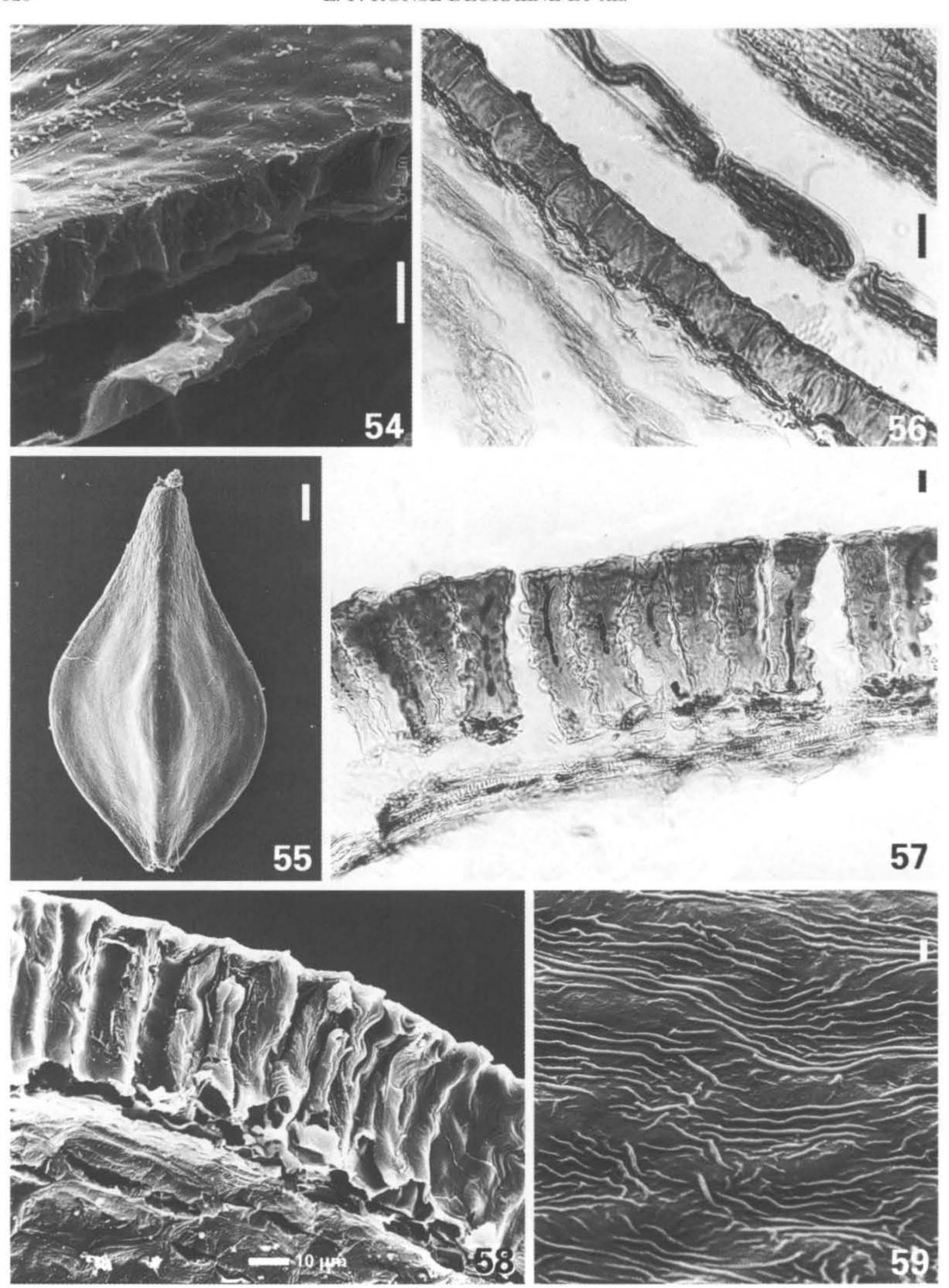
Figures 43–47. Fruits of *Polygonum* sect. *Tephis* (43–45) and sect. *Pseudomollia* (46–47). Scale bars = 10 μm. Figs 43, 44, 46: SEM. Figs 45, 47: LM. 43. *P. afromontanum* (Hedberg 4320) Surface detail. Figs 44–45. *P. undulatum* (Fourcale 5962). Fig. 44. LS exocarp. Fig. 45. LS pericarp. Figs 46–47. *P. molliaeforme* (Kotschy 778). Fig. 46. Detail of fruit surface. Fig. 47. LS fruit. Abbreviations: ex = exocarp; me = mesocarp.

Oxygonum (Figs 74-77)

Achenes are tightly enclosed within the fused perianth and receptacle. Several fruits of Oxygonum appear cross-like in longisection (Fig. 75). This is caused by the



Figures 48–53. Fruits of *Polygonum* sect. *Duravia*. All scale bars = 10 µm. Figs 48–51: SEM. Figs 52, 53: LM. Fig. 48. *P. shastense* (C.F. Sonne s.n.). Section through pericarp. Note large lumen and erect anticlinal walls. Fig. 49. *P. polygaloides* (Nelson & Nelson 6463). Surface detail; ridges run between puzzle piece-like cells in longitudinal rows. Fig. 50. *P. douglasii* (Howell s.n.). Section through pericarp; note tubercles on surface of fruit and strong convolutions. Fig. 51. *P. douglasii* var. *latifolium* (Howell s.n.). Detail of fruit surface. Fig. 52. *P. polygaloides* (Howell s.n.). Section through fruit wall. Fig. 53. *P. tenue* (Puissant s.n.). Section through fruit wall. Abbreviations: en = endocarp; ex = exocarp; me = mesocarp.



Figures 54–59. Fruits of *Polygonella*. All scale bars = 10 μm, except Fig. 55 = 100 μm. Figs 54, 55, 58, 59: SEM. Figs 56, 57: LM. Figs 54–56. *P. polygama*. 54. Detail of fruit surface and exocarp (Curtiss 2433) with surface striations. Fig. 55. Lateral view of mature fruit (Curtiss s.n.). Fig. 56. LS fruit wall; thick exocarp with straight anticlinal walls (Curtiss s.n.). Fig. 57. *P. americana* (Meisner 1845). LS fruit wall with convoluted anticlinal walls. Figs 58–59. *P. articulata* (Bodin s.n.). Fig. 58. Section through pericarp seen from inside of fruit. Note slightly convoluted anticlinal walls. Fig. 59. Surface detail with cuticular striations.

presence of spines or wings in the middle of the fruit. Fruits have a trigonous shape and are elongated-pyriform. The fruit surface appears fimbriate to wrinkled and is covered with short trichomes (e.g. O. stuhlmannii, O. delagoense: Fig. 74) which may resemble spines (O. sinuatum). The exocarp consists of narrowly rectangular cells with undulating anticlinal walls. The lumen is narrow without dichotomous branching (Figs 76, 77).

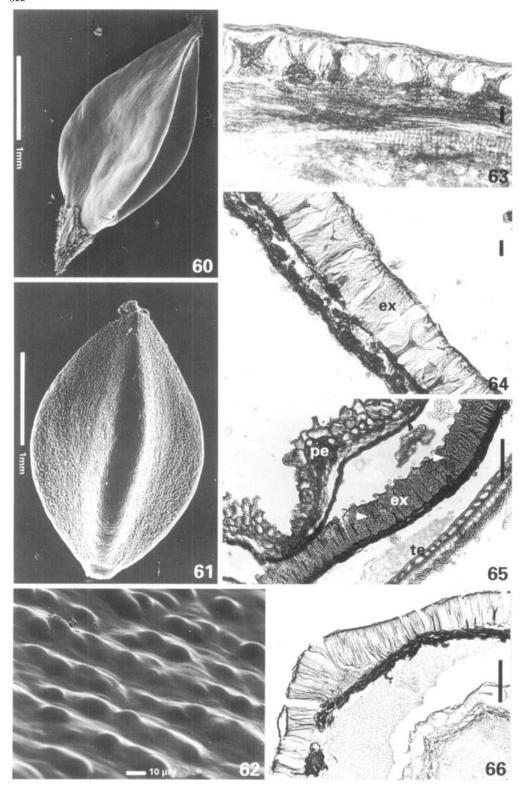
Pteropyrum (Figs 78–80)

Fruits are surrounded by two strongly winged outer tepals and smaller inner tepals. The fruit is beaked and has three narrow wings covered by the folds of the outer tepals (Fig. 80). The wings are biparted by an invagination in the middle with appendages running alongside the fruit. The fruit surface has cells surrounded by ridges sparsely covered with papillae (Fig. 78: *P. noëanum*) or is almost smooth (*P. aucheri*). The exocarp is distinctive, consisting of square stone-cells with central lumina (Figs 79, 80).

DISCUSSION

Table 1 lists the different fruit morphological and anatomical characters of the genera presented here. The anatomy of the outer epidermal cells (the exocarp) has more systematic value than any other fruit character in Persicarieae and Polygoneae. It can be used consistently to delimit genera. However, no clear-cut fruit characters were found to delimit tribes Persicarieae and Polygoneae. This lack of distinctive features may be related to the common *Bauplan* of the fruits in Polygonaceae.

Neubauer (1971) argued that development of the cell wall thickenings differs between Polygonum aviculare, as described by Lonay (1922), and Persicaria pennsylvanica. He described the former species as having anticlinal walls with smooth, flat outer surfaces and convoluted inner surfaces (formed by localized internal thickenings), contrasting with the entire anticlinal walls becoming convoluted in the latter species. Indeed, cell wall thickening of *Polygonum aviculare* is different, but not in the manner described by Neubauer. Lonay (1922) clearly showed (backed by illustrations) that the thickening of P. aviculare is different, stating "Les parois latérales sont aussi très épaissies; cependant ces épaississements vont en diminuant vers l'intérieur, tantôt progressivement, et alors la cavité cellulaire est conique, tantôt brusquement à partir du quart interne de ces parois, auquel cas la cavité cellulaire a la forme d'une cloche évasée vers ses bords" [There is also considerable thickening in the lateral walls; however, this diminishes towards the centre of the fruit, either progressively, resulting in a conical cavity, or abruptly about three quarters of the way through the cell wall, resulting in a splayed, bell-shaped cavity.] (Lonay, 1922: 15). This shows that at least in P. aviculare, anticlinal cell wall thickening is different from that of P. pennsylvanica and represents a different character state, as we describe in this paper. We found that the structure of the convoluted anticlinal fruit wall (the 'egg-case' type) is basically similar in all investigated species (e.g. Polygonum, Polygonella, Persicaria, Oxygonum) and is obviously the result of tangential pressures during fruit development. The exocarp with straight anticlinal walls (the 'corrugated sheet' type) appears to be more basic, although wall thickening is variable. Seen from the surface and in paradermal section, exocarp cell shape often changes from the periphery to the



mesocarp. In most genera the cells are puzzle piece- or star-shaped on the surface and become polygonal towards the centre of the fruit. We observed this transition in several species and it was beautifully illustrated by Lonay (1922) for *P. aviculare*. Anticlinal cells which are straight over their whole surface are restricted to *Fagopyrum*, *Calligonum* and *Polygonum* sect. *Pseudomollia*.

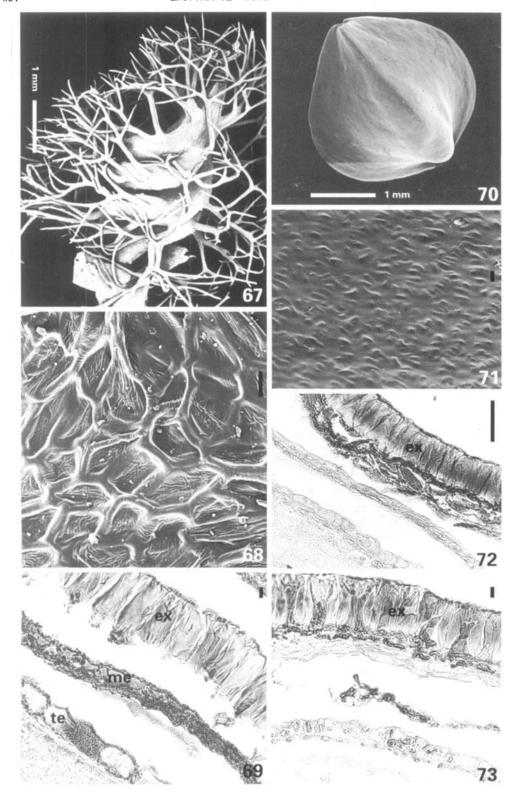
Primary sculpturing of fruit wall cells often includes ridges or folds following the outline of the anticlinal walls. In other instances the walls are collapsed and pits are visible. The ridges are often covered with papillae or tubercles, following the outline of the walls, even when the ridge is not visible, or running independently of the anticlinal walls in longitudinal lines. The association of tubercles with presence versus absence of convolutions in cross or longitudinal sections is incidental.

On the basis of fruit anatomy and the external cell surface, sections Tovara, Cephalophilon and Echinocaulon cannot be distinguished from sect. Persicaria. All share a similar thick exocarp of narrowly rectangular cells with undulating anticlinal walls. The lumen is narrow, eventually with short dendritic branches, but it does not show any dichotomous splitting towards the periphery. Differences are expressed in the thickness of the exocarp, being relatively thin in sections Echinocaulon and Cephalophilon, but thick in sections Persicaria and Tovara (Table 1). The thickest exocarp occurs in Persicaria orientalis with an upper limit of 150 µm. A frequently occurring surface pattern consists of a reticulate arrangement of ridges covered with tubercles running along the anticlinal walls of the cells, and there is a gradual transition from an almost smooth surface (e.g. P. runcinata) occasionally with pits, to weakly developed papillae and finally strongly developed tubercles. Section Persicana is even more variable with a different extent of coverage with tubercles. Given the shared pattern of tepal epidermal cells (rectangular cells with straight to undulating anticlinal walls, and smooth surface or with longitudinal ridges: Hong, Ronse Decraene & Smets, 1998), the classification of Ronse Decraene & Akeroyd (1988) is supported. Cephalophilon and Echinocaulon cannot be distinguished from sect. Persicaria using the external morphology on fruit sections. However, section Tovara is readily recognizable macromorphologically by its dimerous fruits with long hooked styles.

P. wallichii (sect. Rubrivena) does not correspond with Persicaria in its fruit anatomy, but appears similar to Aconogonon. Section Rubrivena differs in pollen (granulated muri) and inflorescence structure from Aconogonon and from the other sections of Persicaria (Hong, 1993). However, exocarp morphology lends support to the inclusion of section Rubrivena in Aconogonon. This is also supported by floral characters (e.g. Ronse Decraene & Akeroyd, 1988).

Aconogonon differs anatomically from *Persicaria* in the almost isodiametric to short rectangular exocarp cells with lumina branching towards the outer tangential walls. Fruit anatomy and fruit surface pattern are almost identical to those of *Bistorta*. Fruit

Figures 60–66. Fruits of Fallopia sect. Reynoutria (60, 63), sect. Fallopia (61, 62, 65, 66) and sect. Sarmentosae (64). Figs 60–62: SEM. Figs 63–66: LM. Fig. 60. F. sachalinensis (Anonym). Lateral view of mature fruit. Scale bar = 1 mm. Fig. 61. F. dumetorum (Ronse Decr. 1309). Lateral view of mature fruit. Scale bar = 1 mm. Fig. 62. F. convolvulus (H. Smith 4372). Surface detail; note rows of tubercles. Scale bar = 10 mm. Fig. 63. F. japonica (Clément Aigret s.n.). Section through fruit wall. Scale bar = 10 mm. Fig. 64. F. multiflorum (Hers 813). Section through pericarp. Scale bar = 10 mm. Fig. 65. F. convolvulus (Kalheber 78–640). LS fruit wall; note dendritic branching in lumen (arrows). Scale bar = 100 μ m. Fig. 66. F. cilinodis (Graxton 9631). TS fruit. Scale bar = 100 μ m. Abbreviations; ex = exocarp; pe = perianth; te = testa.

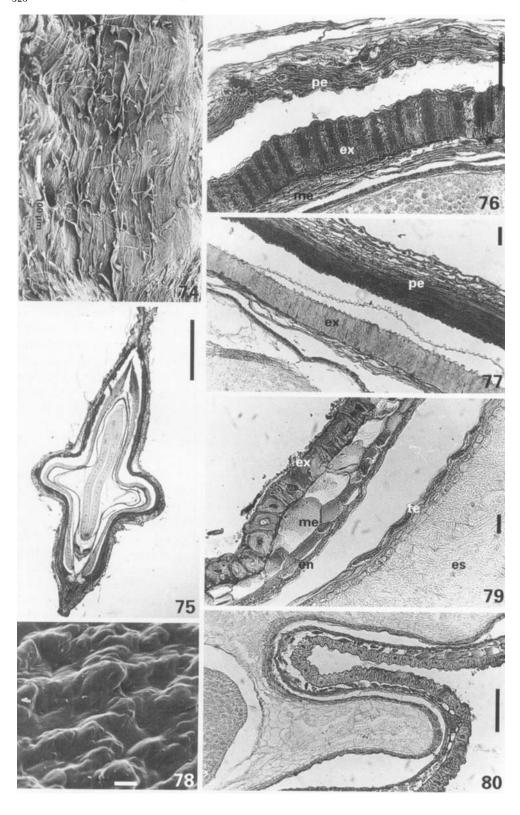


anatomy supports other evidence (see e.g. Haraldson, 1978; Ronse Decraene & Akeroyd, 1988) in assigning a close relationship between *Bistorta* and *Aconogonon*. They share the same pollen morphology (tricolpate with smooth exine covered with microspinules: see also Jaretzky, 1925), fusion of nectaries with stamen bases into a disc (Ronse Decraene & Smets, 1991), constant octomerous androecium, comparable inflorescence and sharing of anthraquinones (Jaretzky, 1925), and embryos with accumbent cotyledons. However, tepal epidermal cell structure is not much different from that of *Persicaria*. In our opinion the two genera show sufficient similarities to be grouped into a single genus as sister to *Koenigia* and *Persicaria*. *Koenigia* differs in fruit anatomy in the isodiametric cells with thinner thickening (see also Hedberg, 1997), but the external epidermal surface is strongly similar.

A similar exocarp structure to that of Aconogonon has been found in Knorringia (but see Hong, 1989). Hong (1989) erected Knorringia (formerly known as Polygonum sibiricum) as a genus separate from Persicarieae in tribe Coccolobeae on the basis of differential characters. He described the pericarp as distinctive and two-layered, with a lumen narrowing triangularly in cross section. However, in the material we studied (Table 1) the one-layered exocarp has straight anticlinal walls; the lumen is cylindrical, only branching at the surface. From the illustrations in Hong (1989) the external surface of the fruit appears rather smooth with shallow depressions. The achenes are included to slightly exserted above the tepals. Unfortunately little information is available on the structure of the flower (e.g. nectarics, tepal epidermis). Knorringia shares the dendritic venation of the tepals with tribe Polygoneae, and closely resembles the genus Fallopia in pollen morphology, exocarp structure, chemical substances (presence of anthraquinones) and chromosome number. Style and stigma shape are reminiscent of sect. Reynoutria. Haraldson (1978) placed Fallopia in the Coccolobeae.

There is strong overlap between some species of sect. Duravia and sect. Polygonum on the one hand, and between sect. Duravia and Polygonella on the other. Similar smooth fruit surfaces (eventually with pits aligning the anticlinal cell walls) are found in Polygonum sect. Duravia (e.g. P. californicum, P. minimum), and sect. Polygonum (e.g. P. maritimum). In some cases superficial cuticular striations are superimposed on the smooth surfaces as in some species of Polygonella and sect. Duravia. Smooth or rough surfaces with irregular ridges lining the anticlinal walls were found in sect. Tephis, sect. Pseudomollia and sect. Duravia (e.g. P. douglasii). In some cases fruits with smooth-pitted surfaces and with tubercles occur side by side in the same species (e.g. P. arenastrum, P. douglasii). More or less similar cell anatomy was observed in a few species of sect. Duravia (e.g. P. douglasii, P. tenue), sect. Tephis (although the convolution is more important and it lacks the dendritic branching), and exceptionally in Polygonella (e.g. P. americana). Almost straight anticlinal walls (less than 40 µm wide) with narrow lumina occur in most other species of sect. Duravia (e.g. P. coarctatum, P.

Figures 67–73. Fruit of Calligonum (67–69) and Atraphaxis (70–73). Figs 67–68. C. microcarpum (Sintenis 2202). Figs 67, 68, 70, 71: SEM. Figs 69, 72, 73: LM. Fig. 67. Part of fruit with branch-like extensions. Scale bar = 1 mm. Fig. 68. Surface detail between branches. Scale bar = 100 μ m. Fig. 69. C. eriopodum (Michelson s.n.). LS fruit wall. Scale bar = 10 μ m. Fig. 70. A. suaedifolia (Wendelbo & Assadi 28026), lateral view of fruit. Scale bar = 1 mm. Fig. 71. A. frutescens (Alm s.n.). Detail of fruit surface. Scale bar = 10 μ m. Fig. 72. A. prifolia (Regel s.n.). LS fruit. Scale bar = 10 μ m. Fig. 73. A. billardieri (Heldreich 617). LS fruit wall. Scale bar = 10 μ m. Abbreviations: ex = exocarp: me = mesocarp: te = testa.



shastense, P. californicum), most Polygonella (e.g. P. articulata, P. polygama), and in sect. Pseudomollia. The lumen appears narrow-rectangular, x-shaped to triangular with the broad base on the inner tangential wall. Dendritic canals were not observed. Ronse Decraene, Hong & Smets (in press) could not find straightforward differences between Polygonella and Polygonum s.s. in fruit anatomy, flower structure, pollen and micromophological details. The identity of Duravia is not clear with some species closely related to Polygonella, and others to sect. Polygonum. The tribe is probably artificial.

A similar exocarp morphology is shared by some species of Fallopia, Calligonum and Atraphaxis. Fallopia is polymorphic with even more variation of cell shapes than in Polygonum sect. Polygonum. The distinctness of sect. Reynoutria is best supported by fruit anatomy, but sect. Sarmentosae and Fallopia have a strongly overlapping pattern, both in outer morphology and cell structure.

Fagopyrum is isolated in its fruit morphology and anatomy. Several characteristics link the genus to Rheum, such as the reflexed styles with central aperture, smooth and unthickened fruit surface, small undifferentiated perianth lobes in fruit, pollen and anther morphology, and tepal epidermis. However, Rheum differs in being trimerous and has a fused disc-like nectary, while there are free lobes in Fagopyrum. Ronse Decraene & Smets (1991) suggested that Fagopyrum occupies a basal position in the tribe Persicarieae, being primitive in many respects and forming a link with a Rheum-like ancestor. The suggestion of Marek (1958) to place Fagopyrum in a separate subfamily makes sense, but we are more inclined to assign it to its own tribe Fagopyreae. More study of related tribes as well as its sectional taxonomy are necessary to understand the relationships of this enigmatic genus.

Relationships between different pericarp types

We have constructed a hypothetical scheme showing the relationships between different pericarp types (Fig. 81). An exocarp with trapezoidal to triangular lumina, relatively square cells and straight anticlinal walls can be considered as basal (Fig. 81C). Evidence supporting this assumption is the fact that such exocarp is found in the neighbouring tribe Rheae (e.g. Rumex, Oxyria, Rheum: Marek, 1954). Increased thickening of the exocarp leads to rectangular cells with straight anticlinal walls and dichotomous branching (e.g. Bistorta, Aconogonon, Calligonum, Atraphaxis, Fallopia: Fig. 81F). Due to constraints of development the cells often become undulating; this has occurred independently in several genera, with the two conditions often occurring side by side (e.g. Polygonella, Polygonum, Fallopia, Oxygonum: Fig. 81A,B). A limited thickening linked with isodiametric exocarp cells is found in Koenigia, Polygonum sect.

Figures 74–80. Fruits of Oxygonum (74–77) and Pteropyrum (78–80). Figs 74, 78: SEM. Figs 75–77, 79, 80: LM. Fig. 74. O. delagoense (Richards 14666). Surface detail with fimbriate tissue and trichomes. Scale bar = $100 \, \mu m$. Fig. 75. O. stuhlmannii (Mhoro & Backeus 2236). LS entire fruit with cross-like median extensions of the fruit and coherent perianth. Scale bar = $10 \, \mu m$. Fig. 76. O. dregeanum var. strictum (Napper 1054). Section of fruit wall. Scale bar = $100 \, \mu m$. Fig. 77. O. sinuatum (Friis et al. 2696). Section of fruit wall. Scale bar = $100 \, \mu m$. Fig. 78. P. noëanum (Horsain & Horsain 53). Detail of fruit surface. Scale bar = $10 \, \mu m$. Fig. 79. P. aucheri (Rechinger 27441): TS fruit. Scale bar = $10 \, \mu m$. Fig. 80. P. gracile (Strauss s.n.). TS fruit with strongly ribbed structure. Scale bar = $100 \, \mu m$. Abbreviations: en = endocarp; es = endosperm; ex = exocarp; me = mesocarp; te = testa.

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Table 1. Fruit morphological and anatomical characters of tribes Persicarieae and Polygoneae (Polygonaceae). Mostly from personal observations, also literature	data from Dammer (1893), Hong (1989, 1992, 1993), Li (1952), Lonay (1922), Lousley & Kent (1981), Neubauer (1971), Marck (1954), Mitchell & Dean (1978),	Dance Dance & Court I'm man (1020)

Genus	Styles	Stigma	Fruit shape	Fruit surface	Tepals	Exocarp cell shape in cross-and longitudinal section	Exocarp thickness	Anticlinal walls of exocarp	Presence of dendritic branching
.lconogonon	free and short, rarely long, occ. heterostylous	capitate	triquetrous, strongly angular, not beaked	cells puzzle piece-shaped, almost smooth, with pits or inconspicuous folds along the anticlinal walls	achene included to exserted	square to rectangular cells, narrow lumen	25 40 µm	straight	dichotomous
Atraphaxis	short, free or basally fused	capitate	triquetrous, lenticular, subangular	smooth, or with pits or papillae on the anticlinal walls	achene included by accrescence of inner tepals	rectangular, lumen narrow	40-55 µm	straight	dichotomous or trichotomous
Bistorta	free, long, deciduous	capitate	triquetrous	cells puzzle piece-shaped, almost smooth, or with folds along the anticlinal walls	achene included to exserted	rectangular with narrow lumen	about 50 μm	straight	dichotomous, with 2nd order branching
Calligonum	very short to long, basally fused or free	capitate	tri- to often tetraquetrous, subangular	polygonal cells, forming reticulate pattern by collapse of cell wall; ribs covered with dendritic branching hairs	achene exserted	rectangular, lumen broad	55–60 µm	straight	dichotomous
Fagofyrum	free, reflexed, persistent, heterostylous	small, capitate, converging towards a central aperture	sharply iriquetrous, beaked, angles inflated- rounded with deep grooves, or subangular	aggregation of rectangular cells, 15–60 × 15–20 μm in LS or TS, polygonal in surface view, smooth and smaller in grooves	achene exserted, perianth typically persistent at the base of the fruit	unthickened, rectangular	40–50 µm	straight to slightly undulating	absent

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TABLE	

Crema Styles Sigma Fruit damps Fruit surface Tepak Exocrap cell shape in control of c										
short to capitate displayers cells parale piece-dapped achone spanter luncath, and achone surgate, nor slightly scattered or in rows, or surged or sippate state of in rows, or surged or subject or suppated in the state of the subject or suppated in the subject of suppated in related. Very short capitate displayers state of the subject of many subject or suppated in the subject of many subject or suppated in the subject of suppared in the suppared in the suppared in the subject of suppared in the suppared in the subject of suppared in the suppared in the suppared in the subject of suppared in the subject of suppared in the subject of suppared in the suppar	Genus	Styles	Stigma	Fruit shape	Fruit surface	Tepals	Exocarp cell shape in cross-and longitudinal section	Exocarp thickness	Anticlinal walls of exocarp	Presence of dendritic branching
short, in finith are triquetrous, subought a subject to angular, anticinal walks and included, subject to angular, and the subject to angular subject to angular, and the subject to angular, subject to angular, subject to angular, subject to subject to more or longitudinal rows outer transfer to a subject to more or longitudinal rows outer transfer to a subject to subject to a s	Vallopia ect. Fallopia		capitate	triquetrous, angular, not or slightly beaked, not stipitate	cells puzzle piece-shaped with tubercles, randomly scattered or in rows, or smooth to ribbed surface	achene included. outer tepals winged or not winged	square, lumen broadly triangular	20 25 µm	straight	dichotomous to trichotomous
winged short capitate triquetrous, with tubercles forming archene rectangular, Inner narrow subangular incrupted ridges in the feast angular shortly shortly backed not braked shortly another collaboration. Fire, very capitate subangular cells puzzle piece-shaped, achene half square, with straight absent triquetrous, capitate with ± slightly folded included influence absent capitate with ± slightly folded included influence shaped richer short capitate with ± slightly folded included influence shaped richer short capitate with ± slightly folded included influence shaped richer short capitate richer short capitate richer short capitate with ± slightly folded included influence shaped richer short capitate of triquetrous, with longitudinal ridges and force short fixed styles often with trichomes and shortly folded included influence shortly the shaped richer short capitate of triquetrous. A three short capitate of triquetrous with strong or wings. A three shortly shortly shortly included inc	'allopia ect. Revnoutria	short, basally fused	fimbriate	triquetrous, subangular to angular, beaked, not stipitate	smooth surface or with scattered tubercles on anticlinal walls	achene included, outer tepals winged	rectangular, narrow lumen	30-20 µm	straight	dichotomous, with 2nd order branching
firee, very capitate subangular, cells puzzle piece-shaped, achene half short to absent triquetrous, folds along anticlinal walls and narrow absent triquetrous, folds along anticlinal walls and narrow triquetrous, with ± slightly folded included in lumen based tribs, ovoid tribs, ovoid tribs, and three short capitate tripquetrous, with longitudinal ridges and fixed styles three spines or wings, elongated with strong tribs are tripquetrous.	'allopia ect. iamentosae	very short	capitate	triquetrous, subangular to more or less angular, shortly beaked, not stipitate	with tubercles forming interrupted ridges in longitudinal rows	achene included, outer tepals winged	rectangular, lumen narrow	50 75 µm	occ. with undulations, or straight	dendritic or dichotomous branching
deciduous, (oblong)- triquetrous apparently smooth to achene rectangular, with narrow 100 µm straight to fused at capitate with ± slightly folded included in humen prominent ribs, owid prominent ribs, owid trichomes and crebsed by narrowly rectangular. 40 80 µm undulating three short capitate triquetrous, with longitudinal ridges and crebsed by narrowly rectangular. 40 80 µm undulating fused styles of wings, and and serious with strong beak.	mănəo	free, very short to absent	capitate	subangular, not beaked, triquetrous, lenticular	cells puzzle piece-shaped, sometimes elongated with folds along anticlinal walls	achene half included	square, with straight anticlinal walls and narrow lumen, x-shaped	20-40 µm	straight	absent
three short capitate triquetrous, with longitudinal ridges and enclosed by narrowly rectangular. 40 80 µm undulating basally fused styles three spines and and perianth elongated with strong beak	ับดราชานูนั้น	deciduous, fused at base	(oblong)- capitate	triquetrous with ± prominent ribs, ovoid	apparently smooth to slightly folded	achene included in persistent tepals	rectangular, with narrow lumen	100 µm	straight to slightly convoluted	dichotomous
	нашобох	three short basally fused styles	capitate	triquetrous, often with three spines or wings, elongated with strong beak	with longitudinal ridges and trichomes	enclosed by floral tube and perianth	narrowly rectangular, lumen slitike	40 80 µm	undulating	absem

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Presence of dendritic branching	absent	absent	dichotomous	dichotomous	absent	absent
Anticlinal walls of exocarp	strongly undulating	strongly undulating	strongly undulating	straight	undulating	straight or undulating
Exocarp	100-120 µm	70–150 µm	about 75 µm	about 75 μm	70–80 µm	25–70 µm
Exocarp cell shape in cross-and longitudinal section	long rectangular, lumen narrow	narrowly rectangular, with (mostly) regularly undulating anticlinal walls, lumen slitlike	rectangular with broad ellipsoid lumen	broadly rectangular, lumen broadly trapezoidal	narrowly to broadly rectangular with regularly undulating anticlinal walls, lumen slitlike	rectangular to isodiametric, lumen broad to narrow
Tepals	achene included	achene included	achene included	achene included	achene included	included to slightly exserted
Fruit surface	with tubercles on ridges along anticlinal walls forming a reticulate structure, or smooth	with tubercles on ridges along anticlinal walls forming a reticulate structure, or almost smooth	cells with anticlinal walls collapsed as pits or as irregular ridges with scattered tubercles or papillae	almost smooth (P. wallichii) to strongly folded (P. pinetorum)	puzzle piece-shaped cells; anticlinal walls collapsed as pits, or with pits aligned in longitudinal rows	puzzle piece-shaped cells, with folds or tubercles along anticlinal walls, or almost smooth with longitudinal cuticular striae
Fruit shape	triquetrous, v subangular, d beaked, d ovate to s	triquetrous, subangular, beaked	triquetrous to lenicular, beaked, ovate to rounded, subangular	triquetrous, ovoid, ovoid- orbicular, not beaked, not stipitate	lenticular, not beaked, not stipitate	triangular, not beaked to conspicuously so, slightly stipitate
Stigma	capitate	capitate	capitate	capitate	minute	small, capitate
Styles	free or fused above or up to middle, deciduous	fused above the middle, deciduous	basally fused, long, deciduous	deciduous, free, long	free and persistent, hooked at tip	short to long, basally fused, erect
Genus	Persicaria sect. Echinocaulon	Persicaria fused abov sect. the middle Cephalophilon deciduous	Persicaria sect. Persicaria	Persicana sect. Rubrivena	Persicaria sect. Tovara	Polygonum sect. Duravia

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Genus	Styles	Stigma	Fruit shape	Fruit surface	Tepals	Exocarp cell shape in cross-and longitudinal section	Exocarp thickness	Anticlinal walls of exocarp	Presence of dendritic branching
Polygonum sect. Polygonella	erect, short to very short	small, capitate	narrow, elongated, subangular, beaked	surface smooth, covered with longitudinal cuticular striae, rarely with tubercles	achene surrounded by three inner accrescent tepals, or exserted	rectangular or isodiametric, lumen broad to slitlike	25–75 µm	straight or undulating	absent
Polygonum sect. Polygonum	short to very short, free or basally fused	capitate	trigonous, beaked, subangular to angular, often with combinations of subequal convex and concave sides	surface smooth, with tubercles covering ridges of anticlinal walls, in longitudinal rows, or randomly distributed	perianth not accrescent, fruit variously inserted or exserted	rectangular to isodiametric, lumen variously narrow, triangular or trapezoid	25–95 µm	undulating	dichotomous, trichotomous, dendritic, or absent
Polygonum sect. Pseudomollia	erect, short, basally fused	capitate	triangular, angles winged in upper part	polygonal cells, collapsed with broad anticlinal wall segments, tubercles in the angles	included to slightly exserted	isodiametric, lumen triangular	about 16 μm	straight	absent
Polygonum sect. Tephis	erect, long, basally fused	capitate	lenticular, rounded	cells puzzle piece-shaped with anticlinal walls collapsed or with irregular ridges	achene included	rectangular, lumen narrow	50–70 µm	strongly undulating	absent
Pteropyrum	basally converging, well developed	capitate	triquetrous, producing three wings or spines in alternation, in the middle interrupted	cells with anticlinal walls ridge-like covered with scattered tubercles	achene exserted	square, lumen large, central	25–30 μm	straight	absent

Table 1—continued

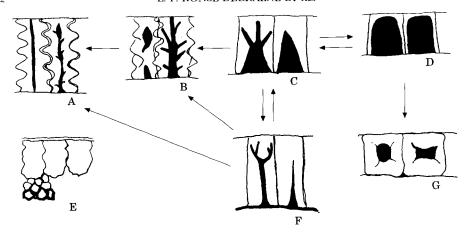


Figure 81. Schematic presentation of hypothetical evolutionary trends in exocarp anatomy. A. Persicaria sect. Persicaria, P. sect. Echinocaulon, P. sect. Cephalophilon, P. sect. Tovara, Oxygonum, Polygonum sect. Polygonum partly, P. sect. Duravia partly, P. sect. Tephis, Polygonella partly. B. Polygonum sect. Polygonum partly, Polygonella partly, Fallopia partly. C. Polygonum sect. Polygonum partly, P. sect. Pseudomollia, Persicaria sect. Rubrivena, Atraphaxis, Fallopia partly, Aconogonon partly, D. Koenigia, Polygonella partly, Polygonum sect. Duravia partly. E. Fagopyrum. F. Fallopia partly, Aconogonon partly, Bistoria, Calligonum, Atraphaxis. G. Pteropyrum.

Duravia, P. sect. Pseudomollia, and Polygonella (Fig. 81D). Pteropyrum (Fig. 81G) is distinctive in that the thickening also occupies the inner tangential walls. Fagopyrum (Fig. 81E) cannot be related to any of the previous types. Persicaria (except for sect. Rubrivena which does not belong here) is distinct in the association of narrow rectangular cells with undulating anticlinal walls and the absence of branching in the lumen. Also if there are papillae or tubercles, these are associated with ridges formed on the anticlinal cell walls. No distinction can be made between different sections of Persicaria on the basis of fruit anatomy. In Polygonum and Fallopia the presence of undulating anticlinal walls is often accompanied by dichotomous to dendritic branching of the lumen.

There is no correlation between anatomy and outer surface structure. Fruit anatomical characters appear more constant within separate sections, contrary to fruit surface characters, which may be strongly variable within genera. The value of external surface patterns is limited and must be sought at subgeneric level, as several conditions coexist in one genus, and even in a single species. However, there are recurring patterns, such as smooth surfaces with striae in *Polygonella* and *Polygonum* sect. *Duravia*, the reticulate pattern in the polymorphic *Polygonum polygaloides*, the presence of tubercles in sect. *Polygonum*, and the folded anticlinal walls of the epidermal cells shared by *Koenigia*, *Bistorta*, and *Aconogonon*.

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REFERENCES

- **Barthlott W. 1981.** Epidermal and seed surface characters of plants: systematic applicability and some evolutionary aspects. *Nordic Journal of Botany* 1: 345–355.
- **Barthlott W. 1990.** Scanning electron microscopy of the epidermal surface of plants. In: Claugher D, ed. *Scanning electron microscopy in taxonomy and functional morphology*. Systematics Association Special Volume 41. Oxford: Clarendon Press, 69–83.
- Brandbyge J. 1993. Polygonaceae. In: Kubitzki K, Rohweder J, Bittrich V, eds. The families and genera of vascular plants. vol. 2. Berlin: Springer-Verlag, 531–544.
- Corner EJH. 1976. The seeds of dicotyledons, 2 vols. Cambridge: Cambridge University Press.
- **Dammer U. 1893.** Polygonaceac. In: Engler A, Prantl K, eds. *Die natürlichen Pflanzenfamilien 3, 1a.* Leipzig: Engelmann, 1–36.
- Erbar C. 1995. On the floral development of *Sphenoclea zeylanica* (Sphenocleaceae, Campanulales) SEM-investigations on herbarium material. *Botanische Jahrbücher für Systematik* 117: 469–483.
- Gerstberger P, Leins P. 1978. Rasterclektronenmikroskopische Untersuchungen an Blütenknospen von *Physalis philadelphica* (Solanaceae). Anwendung einer neuen Präparationsmethode. *Berichte der Deutschen Botanischen Gesellschaft* 91: 381–387.
- Graham SA, Wood CE. 1965. The genera of Polygonaceae in the South Eastern United States. Journal of the Arnold Arboretum 46: 91-121.
- Haraldson K. 1978. Anatomy and taxonomy in Polygonaccae subfam. Polygonoideae Meissn. emend. Jaretzky. Symbolae Botanicae Upsalienses 22: 1–95.
- **Hedberg O. 1997.** The genus *Koenigia L.* emend. Hedberg (Polygonaceae). *Botanical Journal of the Linnean Society* **124:** 295–330.
- Holmgren PK, Holmgren NH, Barnett LC. 1990. Index Herbariorum. Part I: The herbaria of the world, 8th ed. New York: New York Botanical Garden.
- Holub J. 1970. Fallopia Adans. 1763 instead of Bilderdykia Dum. 1827. Folia Geobotanica e Phytotaxonomica, Praha 6: 171–177.
- Hong S-P. 1989. Knorringia (= Aconogonon sect. Knorringia), a new genus in the Polygonaccae. Nordic Journal of Botany 9: 343-357.
- **Hong S-P. 1992.** Taxonomy of the genus *Aconogonon* (Polygonaceae) in Himalaya and adjacent regions. *Symbolae Botanicae Upsalienses* **30:** 1–118.
- Hong S-P. 1993. Reconsideration of the generic status of Rubrivena (Polygonaceae, Persicaricae). Plant Systematics and Evolution 186: 95-122.
- Hong S-P, Ronse Decraene LP, Smets EF. 1998. Systematic significance of tepal surface morphology in tribes Persicarieae and Polygoneae (Polygonaceae). *Botanical Journal of the Linnean Society* 127: 91-116.
- Jaretzky R, 1925. Beiträge zur Systematik der Polygonaccae unter Berücksichtigung des Oxymethylanthrachinon-Vorkommens. Feddes Repertorium 22: 49–83.
- Kanai H, Takeno S, Taniguchi S. 1983. An approach to a logical identification of Japanese plants, 3: a census of achenes of the genus *Polygonum* (Polygonaceae). *Bulletin of the National Science Museum of Tokyo* Ser. B 9: 51–74[in]apanese with English summary].
- **Laubengayer RA**, 1937. Studies in the anatomy and morphology of the polygonaceous flower. *American Journal of Botany* 24: 329–343.
- Li HL. 1952. The genus Tovara (Polygonaceae). Rhodora 54: 19–25.

- Lonay H. 1922. Génèse et anatomie des péricarpes et des spermodermes chez les Polygonacées. Mémoires de la Société Royale des Sciences de Liège, Sér. 3, Tome 2, Part 2, 6: 1-109.
- Lousley JE, Kent DH. 1981. Docks and knotweeds of the British Isles. London: Botanical Society of the British Isles.
- Marek S. 1954. Morphological and anatomical features of the fruits of genera *Polygonum L., Rumex* L. and keys for their determination. *Monographiae Botanicae* 2: 77–161. [in Polish]
- Marek S. 1958. European genera of Polygonaceae in the light of anatomical and morphological investigations on their fruits and seeds. *Monographiae Botanicae* 6: 57–79. [in Polish]
- Mitchell RS, Dean JK. 1978. Polygonaceae (Buckwheat family) of New York State. Albany: University of the State of New York, Education Department.
- Neubauer BF. 1971. The development of the achene of *Polygonum pensylvanicum*: embryo, endosperm, and pericarp. *American Journal of Botany* 58: 655-664.
- Ronse Decraene LP, Akeroyd J. 1988. Generic limits in *Polygonum* and related genera (Polygonaceae) on the basis of floral characters. *Botanical Journal of the Linnean Society* 98: 321–371.
- Ronse Decraene LP, Hong S-P, Smets E. (in press). What is the taxonomic status of *Polygonella?* Evidence of floral morphology. *Annals of the Missouri Botanical Garden*.
- Ronse Decraene LP, Smets E. 1991. The floral nectaries of *Polygonum* s.l. and related genera (Persicarieae and Polygoneae): Position, morphological nature and semophylesis. *Flora* 185: 165–185.
- Roth I. 1977. Fruits of angiosperms. Encyclopedia of plant anatomy X1. Berlin: Borntraeger.
- Schmid R. 1977. Stockwell's bleach, an effective remover of tannin from plant tissues. Botanische Jahrbücher für Systematik 98: 278–287.
- Steward AN. 1930. The Polygonaceae of Eastern Asia. Contributions from the Gray Herbarium of Harvard University 5: 1-129.
- Vautier S. 1949. La vascularisation florale chez les Polygonacées. Candollea 12: 219-343.
- Wisskirchen R. 1991. Zur Biologie und Variabilität von Polygonum lapathifolium L. Flora 185: 267–295.
 Wolf SJ, McNeill J. 1986. Synopsis and achene morphology of Polygonum section Polygonum (Polygonaceae) in Canada. Rhodora 88: 457–479.
- Yang J, Li Y, Wang J. 1991. A primary study on the microstructural features of achene surfaces and its taxonomic importance to *Polygonum*. *Acta Bot. Boreal.—Occident. Sin* 11: 1–7 [Chinese, with English summary].

APPENDIX: ORIGIN OF MATERIAL INVESTIGATED

Aconogonon (Meisn.) Reichenb.

A. alpinum (Allioni) Schur: India, Kashmir, Iragbel; Clarke 292108 (K). A. campanulatum (Hook.f.) Hara var. campanulatum S.-P. Hong: China, Tibet, Tin To Shan; McLaren 8313 (E). A. coriarium (Grigorjev) Soják var. coriarium: Kazakhstan, Kaskelen; Roldugin & Tzagolova 5363 (S). A. hookeri (Meisn.) Hara: China, Szechuan, Tsipula; Smith 4172 (UPS). A. molle (D.Don) Hara var. rude (Meisner) Hara: India, Mt. Khasia; Hooker & Thompson s.n. (BM); originally cultivated in Kew Bot. Gard. Kew, 267–72. 02550 B.L. & M., Ronse Decr. (spirit) 642 Le (LV). A. tortuosum (D.Don) Hara var. tibetanum (Meisn.) S.-P. Hong: India, Kashmir, Lamayuru; Koelz 2748 (E).

Atraphaxis L.

A. billardieri Jaub. & Spach: Syria, Hymetti, Heldreich 617 (LV). A. frutescens (L.) Koch: cultivated in UPS, Alm s.n. (UPS). A. pyrifolia Bunge: Turkestan, Kokschar, Regel s.n. (UPS). A. suaedifolia Jaub. & Spach: Iran, Azarbayejan, Miaveh to Zanjan, Wendelbo & Assadi 28026 (GB).

Bistorta MILL.

B. amplexicaulis (D.Don) Greene: Nepal, Chiong, Dhwoj 108 (UPS). B. griffithii (Hook.) Grierson: SE Tibet, Showa Dzong, Ludlow et al. 13097 (UPS). B. macrophylla (D.Don) Soják: China, Sikang, Kangting, H. Smith 10932 (UPS). B. major Gray: Hercegovina, Murbeck s.n. (UPS). B. perpusella (Hook.) Greene: Bhutan, Shingh, Ludlow et al. 21115 (UPS). B. suffulta (Maxim.) Greene ex H. Gross: Japan, Honshu, Nagano Pref., Yahara et al. 6998 (UPS).

Calligonum L.

C. eriopodum Bunge: Russia, Transcasparian, Michelson s.n. (GB). C. litvinowii Drobov: Uzbekistan, Kokangom, Prjachin s.n. (GB). C. microcarpum Borszcz.: Russia, Transcaucasia, Aschabad, Sintenis 2202 (UPS).

Fagopyrum MILL.

E cymosum Meisn.: China, Sikang, Kangtung, Tachinlu, Vaszeko, H. Smith 13315 (UPS); China, Yunnan, Tong-Achouan, Maire s.n. (G). E esculentum Moench.: China, Shansi, Chieh-Hsiuh Distr., Sung-Lin-Miao, Smith 7666 (UPS); USA, Pennsylvania, Monroe Co., Delaware Water Gap, Brumbach 6902 (K); Burundi, Bururi Prov., Botare, Reekmans s.n. (UPS); cult. Reading Botanic Garden, Ronse Decr. (spirit) 30 Le (LV). E tataricum (L.) Gaertn.: China, Fiouping-Route, Qutae, Yang Tao-Li (9vi 1925), Anonymous 649 (UPS); Belgium, cultivated Nodebais, seeds from Bot. Gard. Göttingen 1385, Ronse Decr. 1335 (spirit 626Ln) (LV).

Fallopia Adans.

Sect. Sarmentosae (I. Grintz.) Holub

E aubertii (L. Henry) Holub: China, Szchuan, Wen-Chuan-Hsien, H. Smith 2316 (UPS). F. baldshuanica (Regel) Holub: Turkestan, cult. in Uppsala, Lundquist s.n. (UPS). F. multiflorum (Maxim.) Haraldson: China, Hunan, Changmau, Herb. J. Hers 813 (BR).

Sect. Reynoutria (Houtt.) Ronse Decr.

E japonica (Houtt.) Ronse Decr.: Japan, Hondo, Yamamoto, Togashi 1264 (UPS); ex Hort. Mus. Paris, Herb. L. de Bullemont 206 (LV); Herb. Clément Aigret s.n. (LV); Belgium, cult. Bruges, Ronse Decr. 1150 (spirit 11 Lb) (LV). E sachalinensis (F. Schmidt) Ronse Decr.: Japan, Sachalin, Anonym. S.n. (LV); FAA, Leg. J. Bailey, Cirencester.

Sect. Fallopia (Adans.) Holub

E. cilinodis (Michx.) Holub: Canada, Ontario, Black Bay, Graxton 9631 (UPS); New York, N. of N. Hudson, off Rd 9, Ronse Decr. 1342 (spirit 646 La) (LV). F. convolvulus (L.) A. Löve: Germany, Hessen, Kalheber 78–640 (UPS); China, Szechuan, Matang, H. Smith 4372 (UPS). F. dumetorum (L.) Holub: Austria, Vienna, Leopoldsberg, Ronse Decr. (spirit) 447 Lo (LV); Belgium, Oud-Heverlee, Ronse Decr. 1309 (spirit 541 L1) (LV). F. scandens (L.) Holub: China, SE Tibet, Mönyul Prov., Nyam Jang Chu, Ludlow et al. 7059 (UPS).

Knorringia (Czuk.) S.-P. Hong (Aconogonon sect. Knorringia (Czuk.) Soják) K. sibirica (Laxm.) Tzvcl. Ssp. sibirica: China, Hopeh, Peitaino Ho, Hopkinson 1620 (S).

Koenigia L.

K. islandica L.: Greenland, E. of Sukkertoppen, Horsen 601 (UPS). K. filicaulis (Wallich ex Meisn.) Hedb.: Nepal Rambrong, Stainton et al. 6014 (UPS). K. nepalensis D. Don: Nepal, Jangla, Einarsson et al. 3394 (UPS). K. nummularifolia Meisn.: Nepal, Arun Valley, W. of Num, Stainton 728 (UPS).

Oxygonum Burch.

O. alatum Burch.: Zimbabwe, Chipinga, Goodier 861 (BR). O. atriplicifolium (Meisn.) Martel: Kenya, East. Prov., S. of Mtiti Andei, Strid 2405 (UPS); Kenya, East. Prov., Machakos Distr. Near Embakasi Station, Haraldson 401 (UPS). O. delagoense Kuntze: Nyamaland, Francis Town, Richards 14666 (UPS); Rhodesia, Aborecorn, Fries 1258 (UPS). O. dregeanum Meisn. var. strictum (C.H. Wright) R. Graham: Zimbabwe, Sipolilo. Nyamunyeche Estate, Nyariti 588 (UPS); Tanzania, Mbizi Forrest, Ufipa Distr. Napper 1054 (UPS). O. sinuatum (Meisn.) Dammer: Kenya, Coast. Prov. Taxa Distr., 8 km NE of Voz. Zumer 56 (UPS); Ethiopia, Genale Gorge, Friis et al. 2696 (UPS). O. stuhlmanii Dammer: Kenya, Eastern Prov., S. of Mtizo Andei, Jonsson 576 (UPS); Tanzania, Moshi Distr., Nyumbaya, Mhoro & Backeus 2236 (UPS).

Persicaria MILL.

Sect. Persicaria Mill.

P. acris (H.B. & K.) Gomez de la Maza: Mexico, Veracruz, N. of Alzacan, Rosas 362 (UPS). P. attenuata (R. Br.) Soják ssp. attenuata [= P. javanicum De Bruyn]: Thailand, Kanchanaburi Prov. Ryding 688 (UPS); ssp. pulchra (Blumc) K.L. Wilson: Tanzania, Iringa Distr. Kidatu, Mohoro 588 (UPS). P. bungeana

(Turcz.) Nakai: China, Chili Prov., Cho-kou-tien, Bohlin 268 (UPS). P. hydropiper (L.) Opiz ssp. microcarpum Denser: China, Szechuan, Hsu-tsing, H. Smith 4778 (UPS). P. decipiens (R. Br.) K.L. Wilson [=P. salicifolia (Brociss. Ex Willd.) Assenov]: Malawi, Southern Region, Blantyre, Brummitt & Williams 9856 (UPS). P. orientalis (L.) Spach: U.K.? Reading, Penna, Brumbach 6921 (UPS). P. pubescens Blume: Nepal, Jumla, Einarsson et al. 3961 (UPS). P. strigosa (R.Br.) H. Gross: Tanzania, Iringa Distr., Mufindi, Bidgood & Loveth 4 (UPS). P. tinctoria (Aiton) H. Gross): Japan, cultivated, Togasi 1518 (UPS). P. viscosa (Buch.-Ham. ex D. Don) H. Gross: Nepal, Bagmati, Kathmandu, Ryding 332 (UPS).

Sect. Echinocaulon (Meisn.) H. Gross

P. hastato-sagittata (Nakai) Ohki: Japan, Numanotaira, Nikko, Kanai 10383 (UPS). P. perfoliata (L.) H. Gross: China, Szechuan, Iping, H. Smith 10000 (UPS). P. sagittata (L.) H. Gross: Nepal, Jumla, Einarsson et al. 3915 (UPS). P. sieboldii (Meisn.) Ohki: Japan, Hondo, Koshigaya in Musashi (1950), Ohwi 110 (K); Korea, Daitaku, Ohwi 2569 (UPS). P. thunbergii (Sieb. & Zucc.) H. Gross ex Nakai: Japan, Hondo, Musashi, Ohwi 170 (UPS).

Sect. Cephalophilon (Meisn.) H. Gross

P. alata (Buch.-Ham.) Nakai: Kenya, Aberdare Range, cult. in UPS, (Hedberg s.n. (UPS). P. capitata (Hamilt. Ex D. Don) H. Gross: Nepal, Muga Karnali Valley, Polunin et al. 5248 (UPS). P. chinensis (L.) H. Gross: Nepal, Chin Hills, Venning 88 (K). P. nepalensis (Meisn.) H. Gross: Nepal, Muga Khola, Polunin et al. 3017 (UPS). P. nuncinata (Buch.-Ham. ex D.Don) Roxb. var. sinuata: India, Kashmir, Kel, Ludlow & Sherriff 8250 (UPS).

Sect. Tovara (Adans.) H. Gross

P. neofiliformis (Nakai) Ohki: Japan, Hondo, Koshigaya, Ohwi 331 (UPS). P. virginiana (L.) Gaertn.: Belgium, seed from Bot. Garden of Bordeaux, cult. Bruges, Ronse Decr. (spirit) 214 Lb (LV); USA, Illinois, Peoria, McDonald s.n. (UPS).

Sect. Rubrivena (M. Král) S.-P. Hong

P. wallichii Greuter & Burdet: U.K., cult. Reading Bot. Gard., Ronse Decr. (spirit) 15 Le (LV).

Polygonum L.

Sect. Polygonum L.

P. achoreum Blake: USA, Wyoming, Crook Co., Sundance, Porter & Porter 8384 (UPS). P. arenastrum Boreau: U.K., Devon, Prawle Point, E. Prawle, Ronse Decr. 1157 (spirit 405 Le) (LV). P. aviculare L.: Belgium, Bruges, ssp. aequale; Ronse Decr. (spirit) 25 Lb (LV). P. bellardi All. Greece, Thessalia, Pharsalum, Heldreich 877, ex Herb. Bullemont 8369 (LV). P. erectum L.: USA, Illinois, Fountaindale, Bebb s.n. (UPS). P. equisetiforme Sibth. & Sm.: Tunisia, Jerba, near hotel Toumana, Ronse Decr. 975 (spirit 309 Lt) (LV). P. maritimum L.: France, Bouches-du-Rhône, Camargue, étang d'Iland, Nordin 102 (UPS). P. oxyspermum Meyer & Bunge ex Ledeb. subsp. raii (Bab.) D.A. Webb & Chater: France, Manche, Vauville et Denneville, L. Corbière 5306, ex Herb. Bullemont 8443 (LV). P. ramosissimum Michx., USA., Kansas, St. Mary's, P.A. Puissant s.n. (LV). P. tubulosum Boiss.: India, Kashmir, Sind Valley, Ludlow & Sherriff 7948 (UPS).

Sect. Tephis (Adans.) Meisn. Emend. Haraldson

P. afromontanum Greenway, Ethiopia, S. Bale, Ririka, Hedberg 377 (UPS); Kenya, Nanyuki Distr., Mt. Kenya, Hedberg 4320 (UPS). P. undulatum (L.) Bergius: South Africa, Humansdorp Distr., 2.6 miles from Assegaai Bosch, Fourcale 5962 (BOL).

Sect. Pseudomollia Boiss.

P. molliaeforme Boiss.: Iran, Kuh-Daena, Kotschy 778 (UPS); Iran. Denawand, Bornmüller 8181 (K). Sect. Duravia S. Wats.

P. bolanderi Brew. ex A. Gray: USA, California: Napa Hills, C.G. Pringle s.n. ex Herb. Wibbe 7829 (LV). P. californicum Meisn.: USA, California, Eal River, Jepson s.n. ex Herb. Wibbe 7830 (LV). P. douglasii E. Greene: USA, Oregon, Siskiyou Mountains, as var. latifolium E. Greene. Howell s.n. ex Herb Wibbe 7842 (LV); Placer Co., Sierra Nevada Mts., A.M. Carpenter s.n. ex Herb. Wibbe 7840 (LV); Wyoming, Southern Carbon Co., W slope of Sierra Madre, Porter & Porter 9722 (UPS); Oregon, base of Stein's mountain, Howell s.n. ex herb. Wibbe 7841 (LV). USA. Montana, as P. douglasii montanum Small, Rydberg & Bessey 5364 (K). ssp. spenulariiforme (Meisn. ex Small) Hickman [=P. coarctatum Dougl. ex Meisn.]: USA, Oregon, Grant's Pass, Howell s.n. ex Herb. Wibbe 7881 (LV); Lyal 1858 (K). P. minimum S. Wats.: USA, Washington, Cascades, Sterens Pass, Sandberg & Leiberg 799 (UPS). P. paronychia Cham. & Schlecht.: USA, California, San Francisco, Golden Gate Heights, Rose 47103 (UPS). P. polygaloides Meisn.: USA, Oregon, Howell s.n. ex Herb. Wibbe 7881 (LV); USA, Wyoming, Uinta Co., Snake River, Nelson & Nelson 6463 (UPS). ssp. kelloggii (E. Greene) Hickman [=P. kelloggii Greene]; USA. Wyoming, Johnson Co., Big Horn Range, W. of Buffalo, Circle Park, Porter & Porter 7555 (UPS). ssp. confertiflorum (Nutt. ex

Piper) Hickman [= P. confertiflorum Nutt. ex Piper]: USA, California, Modoc Co., 9 miles NW of Camby, Bacigalupi & Smith 5982 (UPS). P. shastense Brewer: USA, California, E. North Peak, Nigsen s.n. (UPS); USA, California?, Placer Co., A.M. Carpenter s.n. ex Herb. Wibbe 7880 (LV); USA, California, N. slopes above Coldsham, Sierra Nevada, C.F. Sonne s.n. ex Herb. Wibbe 7879 (LV). P. tenue Michx.: USA, Massachussets, Milten Blue Hills. Forbers 1346 (UPS); Baltic Co. P.A. Puissant (LV). USA.

Polygonella Michx.

P. americana (Fisch. et Mey.) Small: USA, Texas, Cat. Springs, Fischer 10 (UPS); Alabama, Blackwarrior Creek, Blount County, Meisner 1845 (BR), P. articulata (L.) Meisn.: USA, Massachussets, Oakes? s.n. (UPS); New Jersey, Woodbury, Henry S. Conard s.n. (LV); New Jersey, Middlesex Co., South Amboy, Bodin s.n. (UPS). P. fimbriata. (Ell.) Horton: USA, Florida, Tampa, Blanton 6825 (S). P. gracilis Meisn.: USA, Mississipi, Sandy barrens, Pass Christian, A.B. Langlois 177, ex Herb. Wibbe 7817 (LV); Florida, Orange Co., Killarney, O. Vestulund s.n. (UPS). P. polygama (Vent.) Engelm. & Gray [= Polygonella parvifolia Michx.]: USA, N. Carolina, Brunswick Co., Orton Plantation Garden, Godfrey 1046 (K): California, Curtiss s.n. (UPS): Florida, Indian River. Curtiss 2433 (LV).

Pteropyrum JAUB. & SPACH

P. aucheri Jaub. & Spach: Pakistan, Baluchistan, Kalat, Rechinger 27441 (LD). P. gracile Boiss.: Iran, Snetaunabad?, Strauss s.n. (LD). P. noëanum Boiss. Ex Meisn.: Iraq, Koisanjak, Horsain & Horsain 53 (UPS).