

# Mutations in *SPG11* are frequent in autosomal recessive spastic paraplegia with thin *corpus callosum*, cognitive decline and lower motor neuron degeneration

Giovanni Stevanin,<sup>1,2,3,\*</sup> Hamid Azzedine,<sup>1,2,4,\*</sup> Paola Denora,<sup>1,2,5,\*</sup> Amir Boukhris,<sup>1,2,3,6</sup> Meriem Tazir,<sup>7</sup> Alexander Lossos,<sup>8</sup> Alberto Luis Rosa,<sup>9</sup> Israella Lerer,<sup>10</sup> Abdelmadjid Hamri,<sup>11</sup> Paulo Alegria,<sup>12</sup> José Loureiro,<sup>13</sup> Masayoshi Tada,<sup>14</sup> Didier Hannequin,<sup>15,16</sup> Mathieu Anheim,<sup>17</sup> Cyril Goizet,<sup>1,2,18</sup> Victoria Gonzalez-Martinez,<sup>19</sup> Isabelle Le Ber,<sup>1,2</sup> Sylvie Forlani,<sup>1,2</sup> Kiyoshi Iwabuchi,<sup>20</sup> Vardiela Meiner,<sup>10</sup> Goekhan Uyanik,<sup>21</sup> Anne Kjersti Erichsen,<sup>22</sup> Imed Feki,<sup>6</sup> Florence Pasquier,<sup>23</sup> Soreya Belarbi,<sup>7</sup> Vitor T. Cruz,<sup>13</sup> Christel Depienne,<sup>1,2,3</sup> Jeremy Truchetto,<sup>1,2</sup> Guillaume Garrigues,<sup>19</sup> Chantal Tallaksen,<sup>22</sup> Christine Tranchant,<sup>17</sup> Masatoyo Nishizawa,<sup>14</sup> José Vale,<sup>12</sup> Paula Coutinho,<sup>13</sup> Filippo M. Santorelli,<sup>5</sup> Chokri Mhiri,<sup>6</sup> Alexis Brice,<sup>1,2,3</sup> Alexandra Durr<sup>1,2,3</sup> and on behalf of the SPATAX consortium<sup>†</sup>

<sup>1</sup>INSERM, U679, <sup>2</sup>Université Pierre et Marie Curie - Paris 6, UMR S679, <sup>3</sup>APHP, Département de Génétique et Cytogénétique, Groupe Hospitalier Pitié-Salpêtrière, Paris, <sup>4</sup>Centre de Référence de Neurogénétique, CHU d'Angers, France, <sup>5</sup>Unit of Molecular Medicine, IRCCS-Bambino Gesù Children's Hospital, Rome, Italy; <sup>6</sup>Service de Neurologie, Hôpital Habib Bourguiba, Sfax, Tunisia, <sup>7</sup>Service de Neurologie, Hôpital Mustapha, Algiers, Algeria, <sup>8</sup>Department of Neurology, Hadassah-Hebrew University Medical Center, Jerusalem, Israel, <sup>9</sup>Laboratorio de Biología Celular y Molecular, Fundación Allende and Sanatorio Allende, Córdoba, Argentina, <sup>10</sup>Department of Human Genetics, Hadassah-Hebrew University Medical Center, Jerusalem, Israel, <sup>11</sup>Hôpital Benbadis, Constantine, Algeria <sup>12</sup>Serviço Neurologia, Hospital De Egas Moniz, Lisboa, <sup>13</sup>Departamento de Neurologia, Hospital S. Sebastiao, Santa Maria da Feira, Portugal, <sup>14</sup>Department of Neurology, Brain Research Institute, Niigata University, Niigata, Japan, <sup>15</sup>Department of Neurology, Rouen University Hospital and <sup>16</sup>INSERM, U614, Rouen, France, <sup>17</sup>Neurology Department, Hôpital Civil, Strasbourg, France, <sup>18</sup>Service de Génétique, Hôpital Pellegrin, Bordeaux, France, <sup>19</sup>Service de Neurologie, CHU de Montpellier, Hôpital Gui de Chauliac, Montpellier, France, <sup>20</sup>Neurological Clinic of Yokohama, Yamate, Japan, <sup>21</sup>Department of Neurology, University of Regensburg, Germany, <sup>22</sup>Ullevål University Hospital, Oslo, Norway and <sup>23</sup>Centre Hospitalier Régional Universitaire, EA2691, Lille, France

\*The first three authors contributed equally to this work.

†The members of the SPATAX consortium are listed in the Appendix.

Correspondence to: Prof Alexis Brice, INSERM U679, IFR de Neurosciences, Pitié-Salpêtrière Group, 47 Bd de l'Hôpital, 75013 Paris, France

E-mail: brice@ccr.jussieu.fr

**Hereditary spastic paraplegias (HSP) are neurodegenerative diseases mainly characterized by lower limb spasticity associated, in complicated forms, with additional neurological signs. We have analysed a large series of index patients ( $n = 76$ ) with this condition, either from families with an autosomal recessive inheritance ( $n = 43$ ) or isolated patients ( $n = 33$ ), for mutations in the recently identified *SPG11* gene. We found 22 truncating mutations, including the first four splice-site mutations, segregating in seven isolated cases and 13 families. Nineteen mutations were novel. Two recurrent mutations were found in Portuguese and North-African patients indicating founder effects in these populations. The mutation frequency varied according to the phenotype, from 41%, in HSP patients presenting with a thin *corpus callosum* (TCC) visualized by MRI, to 4.5%, in patients with mental impairment without a TCC. Disease onset occurred during the first to the third decade mainly by problems with gait and/or mental retardation. After a mean disease duration of  $14.9 \pm 6.6$  years, the phenotype of 38 *SPG11* patients was severe with 53% of patients wheelchair bound or bedridden. In addition to mental retardation, 80% of the patients showed cognitive decline with executive dysfunction. Interestingly, the phenotype also frequently included lower motor neuron degeneration (81%) with wasting (53%). Slight ocular cerebellar signs were also noted in patients with long disease durations. In addition to a TCC (95%), brain MRI revealed white matter**

alterations (69%) and cortical atrophy (81%), which worsened with disease duration. In conclusion, our study reveals the high frequency of *SPG11* mutations in patients with HSP, a TCC and cognitive impairment, including in isolated patients, and extends the associated phenotype.

**Keywords:** spastic paraplegias; *SPG11*; thin corpus callosum; mental retardation; lower motor neuron degeneration

**Abbreviations:** AR = autosomal recessive; HSP = hereditary spastic paraplegias; IQ = intellectual quotient; LL = lower limbs; MMSE = Mini Mental State Evaluation; UL = upper limbs; SPG = spastic paraplegia gene; TCC = thin corpus callosum; WMA = white matter abnormalities

Received September 28, 2007. Revised November 3, 2007. Accepted November 8, 2007. Advance Access publication December 13, 2007

## Introduction

Hereditary spastic paraplegias (HSP) are neurodegenerative conditions in which the main clinical features are progressive spasticity and weakness of the lower limbs associated with posterior column or bladder involvement (Harding, 1983; Filla *et al.*, 1992; Tallaksen *et al.*, 2001). This phenotype can be complicated by the presence of a wide range of additional neurological and non-neurological signs or symptoms including mental retardation, deafness, cerebellar ataxia, epilepsy, dysarthria, ichthyosis, optic atrophy, peripheral neuropathy, retinitis pigmentosa, cataract, etc. These diseases are inherited in an autosomal dominant, autosomal recessive (AR) or X-linked manner and a wide genetic heterogeneity has been demonstrated with the identification of more than 33 loci and 15 genes (Fink, 2006; Mannan *et al.*, 2006; Valdmanis *et al.*, 2007; Stevanin *et al.*, 2007). The most common forms of autosomal dominant HSP, accounting for about 50% of patients, are caused by mutations in the *SPG4* and *SPG3A* genes that encode for spastin and atlastin, respectively (Hazan *et al.*, 1999; Zhao *et al.*, 2001).

In AR-HSP, observed more frequently than autosomal dominant HSP in inbred populations (Coutinho *et al.*, 1999), the first three genes identified that encode paraplegin (*SPG7*), spartin (*SPG20*) and maspardin (*SPG21*) (Casari *et al.*, 1998; Patel *et al.*, 2002; Simpson *et al.*, 2003), as well as the gene responsible for the related spastic ataxia of Charlevoix Saguenay (Engert *et al.*, 2000), represent only a small proportion of all cases (Fink, 2003). Strikingly, about one-third of AR-HSP index patients have a thin or atrophied *corpus callosum* (ARHSP-TCC) visualized by MRI with different degrees of cognitive deficit (Franca *et al.*, 2007). This form of AR-HSP was initially mapped to chromosome 15q13–15 [*SPG11*, (Martinez *et al.*, 1999)] and accounts for 41–77% of reported ARHSP-TCC families (Shibasaki *et al.*, 2000; Casali *et al.*, 2004; Stevanin *et al.*, 2006). Recently, the *SPG11* gene, also known as *KIAA1840/FLJ21439*, that encodes spatascin, was identified and was mutated in 11 of 12 ARHSP-TCC index patients (Stevanin *et al.*, 2007). Ten different mutations were identified in the 11 families. They were either nonsense mutations, deletions or insertions in the *SPG11* coding sequence, resulting theoretically in an abnormally truncated protein in all cases.

The aims of the present study were to estimate the frequency of *SPG11* mutations in a large series of patients with ARHSP with or without TCC, mental retardation or cognitive impairment, to define the spectrum of the mutations in this gene and to describe the associated phenotypes.

## Materials and Methods

### Subjects

Forty-three kindreds with an autosomal recessive inheritance and 33 isolated cases with no family history of the disease were selected, 22 and 5 of which were consanguineous, respectively. Index patients presented with either (i) spastic paraplegia with a mental retardation or cognitive impairment and TCC visualized at MRI ( $n=26$ ), (ii) spastic paraplegia with TCC without mental retardation and cognitive impairment ( $n=11$ ) or (iii) spastic paraplegia with mental retardation or cognitive impairment without TCC ( $n=22$ ). In addition, 17 index patients presenting with spastic paraplegia and mental retardation or cognitive impairment for which MRI was not available were also analysed. This study has been approved by the local Bioethics committee (approval no 03-12-07 of the Comité Consultatif pour la Protection des Personnes et la Recherche Biomédicale Paris-Necker to Drs A. Durr and A. Brice). Informed written consent has been given by all participating members of the families before blood samples were collected for DNA extraction. All clinical evaluations were performed according to a protocol established by the European and Mediterranean network for spinocerebellar degenerations (SPATAX, coordinator: Dr A. Durr) that included: full medical history and examination, estimation of the age at onset by the patient, presence or absence of additional neurological symptoms/signs, electroneuromyographic (ENMG) studies and brain MRI when possible. IQ or Mini Mental State Evaluations (MMSE) (Folstein *et al.*, 1975) were available for 12 patients, two of whom had detailed neuropsychological evaluations including Standard Progressive Matrices [PM38, (Raven, 1982)] and the Wechsler Memory Scale (Wechsler, 1987). According to the DSM-IV criteria (Diagnosis and statistical Manual of Mental disorders, 2000), mental retardation was considered when the patient had an IQ < 70 before the age of 18 years.

Most patients were French ( $n=37$ ) or North-African ( $n=15$ ), or originated from other European countries ( $n=13$ ), the Middle-East ( $n=6$ ) and elsewhere ( $n=5$ ). Eleven of the 33 sporadic subjects previously tested negative for mutations or

rearrangements in the *SPG4* gene (Depienne *et al.*, 2006, 2007); mutations in the *SPG7* gene were also excluded in a subset ( $n = 20/43$ ) of families (Elleuch *et al.*, 2006).

## Genotyping

Linkage to *SPG11* was investigated in 33 AR-HSP families using the polymorphic markers *D15S781*, *D15S537*, *D15S516* and *D15S659* after DNA amplification by polymerase chain reaction (PCR); the amplicons were sized on an ABI Prism 3730 automated sequencer with GenMapper software (Applied Biosystems, Foster City, CA, USA), as previously described (Stevanin *et al.*, 2006). After haplotype reconstruction, putative linkage was established on the basis of common haplotypes by descent in affected relatives of the same pedigree.

## Mutation detection

The coding sequence and splice site boundaries of the 40 exons of the *SPG11* gene were amplified by PCR and sequenced in both directions as described previously (Stevanin *et al.*, 2007).

Numbering of new mutations/polymorphisms was performed relative to the ATG codon of the first coding exon, as recommended by the Human Genome Variation Society (<http://www.hgvs.org/mutnomen/>). Segregation of the mutations/polymorphisms with the disease was verified by direct sequencing in 64 additional family members whose DNA samples were available. In addition, unrelated healthy subjects were screened to evaluate the frequency of nucleotidic changes: 80 French Caucasians, 31 North-Africans, 103 Palestinians and 48 individuals from Argentina. Synonymous, missense and splice-site variations were systematically evaluated for modifications of exonic splicing enhancers (ESEfinder algorithm available at <http://www.rulai.cshl.edu/cgi-bin/tools/ESE/esefinder.cgi>) or consensus splicing sequences (at [http://rulai.cshl.edu/new\\_alt\\_exon\\_db2/HTML/score.html](http://rulai.cshl.edu/new_alt_exon_db2/HTML/score.html) and [http://www.fruitfly.org/seq\\_tools/splice.html](http://www.fruitfly.org/seq_tools/splice.html)). Multiple alignment with spatacsin orthologs in various species was performed using ClustalW software (<http://www.ebi.ac.uk/clustalw/>) to evaluate conservation of missense variants.

The effect on mRNA splicing of a variant affecting the last codon of exon 15 was analysed by RT-PCR on RNA extracted from the lymphoblasts of patient FSP670-5, as reported elsewhere (Stevanin *et al.*, 2007) using primers cDNA<sup>F</sup>—GCTCTGTGGTGGGATCAACT (exon 14) and cDNA<sup>R</sup>—TGCTTCACTGGCCCTGATTG (exon 18) at an annealing temperature of 60°C, followed by direct sequencing of the PCR product.

## Results

### Linkage studies

We initially analysed the segregation of four microsatellite markers tightly flanking the *SPG11* gene in 33 kindreds, in which at least two affected patients were sampled. Twelve families were excluded because no common haplotypes segregated with the disease in affected relatives. In 21 families (64%), the reconstruction of the haplotypes was compatible or did not exclude linkage to *SPG11*. In these 21 families, the *SPG11* gene was sequenced.

## SPG11 mutation screening

Direct sequencing of the *SPG11* gene was performed in 64 unrelated index patients, including the probands of the 21 putatively linked families mentioned earlier and 43 index patients not analysed by linkage studies. We identified 22 truncating mutations in the index patients of 20 families, 19 were newly identified variants (Table 1). In 14 of these families, the mutations were homozygous. In six kindreds, the patients had two compound heterozygous mutations. The mutations segregated with the disease in the families where this could be tested (Supplementary figure). Unaffected relatives ( $n = 47$ ) never had two mutations in the *SPG11* gene.

Most mutations were nonsense mutations ( $n = 4$ , three new); small deletions ( $n = 13$ , 11 new) or insertions (one new). In addition, we identified four new mutations predicted to affect the splicing of the *KIAA1840* mRNA and that were not found on at least 160 and 62 Caucasian and North-African control chromosomes, respectively. In the Israeli-Arab family FSP670, the homozygous c.2833A>G mutation in the last conserved codon of exon 15, leading to the missense variation p.R945G, was also shown to affect the 5' splice consensus site (score of +2.7 versus +4.9 for the wild-type sequence). This *in silico* prediction was confirmed on mRNA isolated from lymphoblasts of an affected family member (FSP670-5) in which an alternative donor splice site is generated downstream in intron 15 leading to a 65 bp insertion and a premature stop codon (Fig. 1; r.2834 + 1\_2834 + 65ins, p.R945GfsX5). The c.2833A>G mutation was also absent from 103 healthy unrelated Palestinians. In families FSP847 and FSP892, homozygous G>A transitions at positions c.869 + 1 and c.2316 + 1 in intron 4 and intron 12 were predicted to strongly alter the consensus sequence score from +9.8 to −0.9 and from +6.2 to −4.5, respectively. The c.869 + 1G>A mutation, found in family FSP847, was also absent from 48 healthy unrelated Argentineans. The single patient from family FSP830, who carried a heterozygous nonsense mutation in exon 6 (c.1282A>T, p.K428X), also carried an A>G transition at position c.6477 + 4 in intron 34 for which the splice score ([http://rulai.cshl.edu/new\\_alt\\_exon\\_db2/HTML/score.html](http://rulai.cshl.edu/new_alt_exon_db2/HTML/score.html)) was reduced from +9.6 to +6.6. Living cells were not available, however, to confirm the *in silico* predictions of missplicing in families FSP847, FSP892 and FSP830.

The 22 identified mutations were located in—or close to—15 different exons throughout the gene, from exon 1 to exon 37. However, two of these mutations were found in more than one family. Mutation c.6100C>T/p.R2034X was found in four different kindreds from Algeria, Morocco and Tunisia and was previously reported in three North African families (Stevanin *et al.*, 2007). Portions of the haplotypes reconstructed with four closely flanking markers were similar in the four new kindreds and in families previously reported, indicating a common ancestral mutational event (Table 2). The new mutation c.6737\_6740delTTGA/p.I2246\_E2247>S2246fsX was found in two families from

**Table 1** *SPGII* mutations

| Family                           | Inheritance | Consanguinity | Origin         | Exon/intron       | Mutation (s)  |
|----------------------------------|-------------|---------------|----------------|-------------------|---|
| Homozygous non-sense mutations   |             |               |                |                   |   |
| FSP831                           | AR          | yes           | Portugal       | Exon3             | c.529_533delATATT, p.I177SfsX178  |
| SPDI99                           | AR          | yes           | Turkey         | Exon4             | <b>c.704_705delAT, p.H235RfsX246</b>  |
| FSP870                           | AR          | yes           | Tunisia        | Exon4             | c.733_734delAT, p.M245VfsX246   |
| FSP393                           | AR          | yes           | Portugal       | Exon6             | <b>c.I235C&gt;G, p.S412X</b>  |
| FSP838                           | Isolated    | yes           | Saudi Arabia   | Exon30            | <b>c.5769delT, p.S1923RfsX1950</b>  |
| FSP400                           | AR          | yes           | Algeria        | Exon32            | c.6100C>T, p.R2034X   |
| FSP792                           | AR          | yes           | Morocco        | Exon32            | c.6100C>T, p.R2034X   |
| FSP845                           | Isolated    | yes           | Morocco        | Exon32            | c.6100C>T, p.R2034X   |
| FSP881                           | AR          | yes           | Tunisia        | Exon32            | c.6100C>T, p.R2034X   |
| FSP920                           | AR          | yes           | Japan          | Exon36            | <b>c.6737_6740delTTGA, p.I2246SfsX2260</b>  |
| FSP75                            | AR          | no            | Portugal       | Exon37            | <b>c.6832_6833delAG, p.S2278LfsX2338</b>  |
| Homozygous splice site mutations |             |               |                |                   |   |
| FSP847                           | AR          | yes           | Argentina      | Intron4           | <b>c.869 + IG&gt;A, r.?</b>   |
| FSP892                           | Isolated    | no            | Norway         | Intron12          | <b>c.2316 + IG&gt;A, r.?</b>  |
| FSP670                           | AR          | yes           | Isarelian-Arab | Exon15            | <b>c.2833A&gt;G, p.R945GfsX950, r.2834_2835ins2834 + I_2834 + 65</b>                |
| Compound heterozygous mutations  |             |               |                |                   |   |
| FSP830                           | Isolated    | no            | Portugal       | Exon6<br>Intron34 | <b>c.I282A&gt;T, p.K428X</b><br><b>c.6477 + 4A&gt;G, r.? (splicing)</b>             |
| FSP343                           | AR          | no            | Algeria        | Exon7<br>Exon36   | <b>c.I549_I550delCT, p.L517LfsX556</b><br>c.6737_6740delTTGA, p.I2246SfsX2260       |
| FSP522                           | Isolated    | no            | France         | Exon7<br>Exon30   | <b>c.I471_I472delCT, p.L491DfsX556</b><br><b>c.5532_5533delCA, p.S1844SfsX1857</b>  |
| SAL646                           | Isolated    | no            | France         | Exon8<br>Exon36   | <b>c.I668delT, p.F556LfsX577</b><br><b>c.6739_6742delGAGT, p.E2247LfsX2260</b>      |
| FSP683                           | Isolated    | no            | Romania        | Exon10<br>Exon31  | <b>c.I951C&gt;T, p.R651X</b><br><b>c.5989_5992delCTGT, p.LI997MfsX2056</b>          |
| FSP398                           | AR          | no            | Poland-Israel  | Exon25<br>Exon31  | <b>c.4307_4308delAA, p.QI436RfsX1442</b><br><b>c.5986_5987insT, p.CI996LfsX1999</b> |

New mutations are indicated in bold.

Japan (homozygous) and from Algeria (heterozygous) associated with different haplotypes as expected. In addition, the c.529\_533delATATT/p.I177\_I178del>S177fsX mutation (Stevanin *et al.*, 2007), previously found in two Portuguese families, was found in another kindred from the same country (FSP831), associated with the same haplotype (Table 2). In contrast, the c.733\_734delAT/p.M245VfsX mutation, previously found on two different ancestral chromosomes in Italian and French families (Del Bo *et al.*, 2007), was homozygous in patients from a Tunisian kindred associated with different haplotypes, suggesting independent mutational events or a very ancient mutation.

Four exonic nucleotide variants present in patients but also in >2% of control chromosomes from France and North Africa are likely to be polymorphisms: c.808G>A/p.V270I (1.5 versus 2.3% in controls), c.1388T>C/p.F463S (47 versus 51%), c.3420G>A/p.L1140L (2.3 versus 3.3%) and c.7023C>T/p.Y2341Y (1.5 versus 7.1%). The p.F463S and p.L1140L variants have already been described in the NCBI (<http://www.ncbi.nlm.nih.gov>) and Ensembl (<http://www.ensembl.org>) human genome databases, and the silent changes at residues 1140 and 2341 occurred in the presence of homozygous truncating mutations in the *SPG11* gene. None of them were shown to modify splice sites.

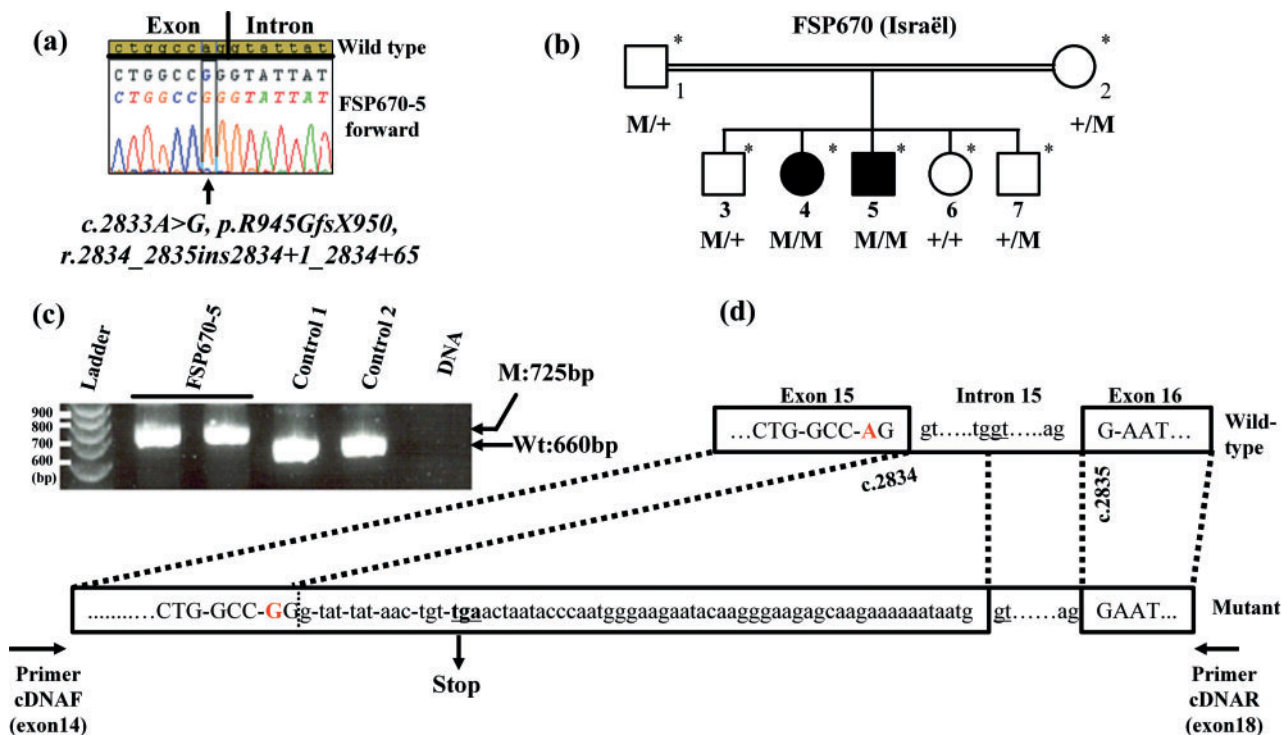
Three additional intronic polymorphisms were detected at positions –139A>G and –141A>C upstream exon 8, and +62C>T downstream exon 37 but were not predicted to cause missplicing of the *SPG11* gene.

### Clinical characteristics of the 20 new *SPGII* families (38 patients)

Six families were North African (two Algerian, two Tunisian and two Moroccan), eight European (four from Portugal, two from France, one from Romania and one from Norway), four Middle-Eastern (two Israeli, one of which of Arabic origin, one Turkish and one Saudi-Arabian) and one each from Argentina and Japan.

Most of the families (65%) had a clear autosomal recessive mode of inheritance, whereas seven index patients (35%), including five without consanguinity, had no family history of neurological disorders.

Age at onset in 37 *SPG11* patients ranged from 2 to 27 with a mean of  $14.0 \pm 5.9$  years. Onset was, in most cases, characterized by gait disorders (30/38, 79%), or less frequently by mental retardation (6/38, 16%), rarely dysarthria and tremor (one each). After a mean disease duration of  $14.9 \pm 6.6$  years (range: 2–35), all patients had a



**Fig. 1** Mutation c.2833A>G in family FSP670 alters *SPGII* mRNA splicing. **(A)** The electropherogram shows the c.2833A>G mutation. **(B)** Pedigree and segregation of the mutation in the family. Square symbols represent men, the circles represent women. The filled symbols are affected individuals. The numbers are an internal reference for each sampled individual. Stars indicate sampled subjects. M = mutation; + = wild type. **(C)** Agarose gel separation of the PCR products generated from *SPGII* cDNA from an affected subject and controls showing abnormal processing of the mRNA. **(D)** Graphical representation of the effect of the mutation. Exons are boxed. An alternative donor splice site in intron 15 is used in the mutant allele leading to a 65 bp insertion and a premature stop codon.

severe clinical picture that included progressive spastic paraplegia (Table 3): most were at least wheelchair bound (20/38, 53%) or needed assistance for walking (6/38, 16%), but 12 could still walk without help (Fig. 2). While only 12% of the patients were confined to a wheelchair or bedridden after <10 years of disease, 60% were in this condition after 18 years of evolution (Fig. 2). Patients were wheelchair bound after a mean disease duration of  $16.5 \pm 5.8$  years (range 9–35,  $n = 20$ ). Lower limb spasticity was severe in 25/37 (67%), associated with severe weakness in 19/37 (51%). Distal or generalized wasting was also noted in 20/38 (53%). Dysarthria was frequently observed ( $n = 16/38$ , 42%). Mental retardation, illustrated by learning difficulties in childhood, was present in 12 patients and confirmed in eight who had a mean IQ of  $58 \pm 9$  (range: 45–69). In addition, in 80% (24/30) of the patients, cognitive decline was evident on examination and worsened with time. MMSE scores were low in 4/5 patients tested (<23/30). Only one patient, who had the shortest disease duration (FSP683-3, 2 years), had no mental retardation and cognitive decline. Two patients underwent detailed neuropsychological evaluation (FSP522-1 and FSP75-21). The non-verbal evaluation of global cognitive efficiency in patient FSP522-1 was normal (PM38 = 46/60), but she had a severe memory impairment (Wechsler Memory Quotient = 72/140) associated with reduced verbal fluency

and an attention deficit indicative of executive dysfunction. A second evaluation, 5 years later, showed deterioration of her cognitive status. Patient FSP75-21, who had mental retardation (IQ = 56), showed a MMSE score of 21/30 at 35 years with psychiatric and cognitive difficulties that included auditory hallucinations and executive dysfunctions.

Cerebellar ocular signs such as abnormal saccadic pursuit and nystagmus were noted in seven patients, most of whom had disease durations >15 years (5/21 versus 2/17). There was *pes cavus* in eight patients, scoliosis in five and other signs were occasionally observed: parkinsonism, orthostatic hypotension, macular excavation or degeneration, strabismus. Four patients, all with disease durations of >18 years, had swallowing difficulties.

Interestingly, ENMG detected lower motor neuron involvement in 13/16 (81%) after a mean disease duration of  $14.4 \pm 4.9$  years (Table 4). In two patients, there was clear anterior horn involvement, while in the others there was axonal neuropathy. Brain MRI showed a TCC (20/21, 95%), with cortical atrophy (17/21, 81%) and associated with diffuse white matter hyperintensities on T2 images (13/19, 69%). The atrophy of the *corpus callosum* was found in all but one patient (FSP400-5, 7-year disease duration), but with variable intensity (Fig. 3). Leucoencephalopathy was periventricular and confluent, and its severity increased

**Table 2** Haplotypes of four close flanking markers segregating with the recurrent mutations in the *SPG11* gene in this study and in previous reports (Stevanin *et al.*, 2007; Del Bo *et al.*, 2007)

| Family  | Stevanin <i>et al.</i> , 2007 |                   |                   |                   |                   | DelBo <i>et al.</i> , 2007 |                |           |                    |                    |
|---------|-------------------------------|-------------------|-------------------|-------------------|-------------------|----------------------------|----------------|-----------|--------------------|--------------------|
|         | Origin                        | FSP400            | FSP446            | FSP881            | FSP792            | FSP845                     | FSP221         | FSP732    | FSP920             | FSP343             |
| D155781 | Algeria                       | 185               | 185               | 185               | 185               | 185                        | 185            | 185       | 183                | 185/187            |
| D155537 | Morocco                       | 180               | 180               | 180               | 176               | 176                        | 176            | 176       | 180                | 184                |
| SPG11   |                               | c.6100C>T         | c.6100C>T         | c.6100C>T         | c.6100C>T         | c.6100C>T                  | c.6100C>T      | c.6100C>T | c.6737.6740delTTGA | c.6737.6740delTTGA |
| D155516 |                               | 191               | 191               | 191               | 191               | 191                        | 191            | 191       | 195                | 195                |
| D155659 |                               | 195               | 179               | 179               | 179               | 179                        | 179            | 175       | 175                | 187                |
| Family  | Stevanin <i>et al.</i> , 2007 |                   |                   |                   |                   | DelBo <i>et al.</i> , 2007 |                |           |                    |                    |
|         | Origin                        | FSP831            | FSP754            | FSP386            | FSP386            | FSP870                     | FSP117         | DelBo     |                    |                    |
| D155781 | Portugal                      | 185               | 185               | 185               | 185               | 187                        | 185            | Italy     |                    |                    |
| D155537 |                               | 172               | 172               | 172               | 172               | 180                        | 172            | France    |                    |                    |
| SPG11   |                               | c.529.533delATATT | c.529.533delATATT | c.529.533delATATT | c.529.533delATATT | c.733.734delAT             | c.733.734delAT | Italy     |                    |                    |
| D155516 |                               | 195               | 195               | 195               | 193               | 195                        | 195            | Italy     |                    |                    |
| D155659 |                               | 203               | 203               | ND                | ND                | 195                        | 179            | Italy     |                    |                    |

ND = not done. Conserved regions are highlighted in grey. Genotypes are indicated in base pairs.

with disease duration. In mild cases, only frontal and occipital periventricular damage was seen (Fig. 3). Finally, visual evoked potentials were abnormal in three out of five patients, indicating an even more diffuse distribution of the lesions.

Discussion

The identification of 22 different truncating mutations (19 new) distributed throughout the *SPG11* gene (Fig. 4) emphasizes the need to analyse the whole gene in clinical practice. Only two of these mutations were found in more than two families in this study and previous studies, suggesting regional founders in these populations (Stevanin *et al.*, 2007): the recurrent mutation c.6100C>T/p.R2034X in families from North Africa where it accounted for 70% of the reported cases (7/10 mutated families); the c.529\_533delATATT/p.I177\_F178>S177fsX mutation in Portuguese families (3/6 mutated families). Conserved haplotypes for flanking microsatellite markers were associated with these mutations.

Most of the 20 new *SPG11* families originated from the Mediterranean basin, but mutations were also found in families from Scandinavia, Japan and South America, indicating a worldwide distribution of this clinico-genetic entity, as previously suggested (Shibasaki *et al.*, 2000; Casali *et al.*, 2004; Winner *et al.*, 2006; Stevanin *et al.*, 2006; Olmez *et al.*, 2006).

When the eleven previously reported cases (Stevanin *et al.*, 2007) from our series are taken into account, *SPG11* mutations are found in ~26% (9/35) of apparently sporadic HSP with TCC patients and ~40% (22/53) of subjects with complex AR-HSP. Interestingly, the frequency varies widely according to the phenotype (Table 5). *SPG11* mutations were found in 59% of patients with TCC and mental impairment collected by our network, a frequency very close to the 41–77% of families from Italy, Japan and Mediterranean countries found in previous linkage analyses (Shibasaki *et al.*, 2000; Casali *et al.*, 2004; Stevanin *et al.*, 2006). Mutations in *SPG11* accounted for only a single family (1/22, 4.5%) of the subgroup of patients with HSP and cognitive impairment without TCC, but patients from this kindred had mild white matter changes and cortical atrophy after a short disease duration of 7 years. *SPG11* is therefore the major identified cause of HSP-TCC and, when taking into account the proportion of 1/3 of HSP-TCC among ARHSP (Franca *et al.*, 2007), *SPG11* is responsible for ~21% of families with ARHSP, making it the most frequent cause of this disease.

The clinical features of the *SPG11* patients studied here were similar to previous reports (Shibasaki *et al.*, 2000; Casali *et al.*, 2004; Winner *et al.*, 2006; Stevanin *et al.*, 2006; Lossos *et al.*, 2006), with a broader range of ages at onset (2–27 years). *SPG11* is severe since patients were wheelchair bound after a mean disease duration of 16.5 ± 5.8 years (range 9–35, *n* = 20) compared to 26.6 ± 15 years (range

**Table 3** Clinical features of 38 patients with mutations in the *SPG11* gene

| Patient (Sex) | Age at onset (years) | Symptoms at onset  | Age at exam (years) | Disease duration (years) | Severity/leg spasticity             | Weakness/wasting               | Pyramidal syndrome [UL/knee/plantar reflexes] | Mental impairment               | Cognitive decline | Other signs or symptoms                                |
|---------------|----------------------|--|---------------------|--------------------------|-------------------------------------|--------------------------------|---|---------------------------------|-------------------|--|
| FSP831-5 (M)  | 6                    | Learning difficulties followed by spastic gait at age 15 | 31                  | 25                       | Bedridden/severe                    | Severe/none                    | ++<br>++<br>↑↑                                | Yes                             | Yes               | Pes cavus, swallowing difficulties                     |
| SPDI99-1 (F)  | 12                   | Tremor at rest and action                                | 28                  | 16                       | Wheelchair at age 22/severe         | Severe/moderate                | ++<br>+<br>↑↑                                 | Yes mild                        | ND                | Parkinsonism with severe akinesia, rigidity and tremor |
| SPDI99-15 (M) | Childhood            | Stiff legs   | 21                  | NA                       | Walking aid at age 20/severe        | Moderate/none                  | ND<br>++ (-at ankles)<br>↑↑                   | Yes (MMSE 21/30)                | Yes               | Parkinsonism with rest tremor, strabism                |
| FSP870-17 (M) | 15                   | Stiff legs   | 25                  | 10                       | Gait possible without help/severe   | Severe/yes                     | N<br>++<br>↑↑                                 | Yes (MMSE<15/30)                | Yes               | Pes cavus, facial dystonia, renal lithiasis            |
| FSP870-20 (F) | 17                   | Weakness legs  | 28                  | 11                       | Wheelchair at age 28/severe         | Severe/none                    | +<br>++<br>↑↑                                 | Yes                             | No                | Dystonia of face and tongue                            |
| FSP870-27 (F) | 14                   | Stiff legs   | 29                  | 15                       | Wheelchair at age 27/severe         | Severe/none                    | +<br>++<br>↑↑                                 | Yes                             | No                | None   |
| FSP870-28 (F) | 20                   | Weakness legs  | 42                  | 22                       | Wheelchair at age 38/severe         | Moderate/none                  | +<br>↑↑<br>+                                  | Yes                             | No                | None   |
| FSP393-11 (F) | 7                    | Learning difficulties                                    | 25                  | 18                       | Gait possible without help/moderate | Mild/none                      | ↑↑<br>++<br>++                                | Yes                             | Yes               | Spastic dysarthria                                     |
| FSP393-12 (M) | 7                    | Learning difficulties                                    | 24                  | 17                       | Gait possible without help/moderate | Mild/none                      | ↑↑<br>++<br>++                                | Yes                             | Yes               | Spastic dysarthria, decreased vibration sense          |
| FSP838-1 (F)  | 13                   | Stiff legs   | 22                  | 9                        | Walking aid/mild                    | Moderate/none                  | ↑↑<br>N<br>++                                 | Learning difficulties at age 15 | ND                | None   |
| FSP400-5 (F)  | 5                    | Leg weakness   | 12                  | 7                        | Gait possible without help/moderate | Moderate/generalized           | ↑↑<br>++<br>++                                | No                              | Yes               | Spastic dysarthria                                     |
| FSP400-10 (M) | 2                    | Delayed walking acquisition, stiff legs                  | 13                  | 11                       | Gait possible without help/moderate | Moderate/none                  | ↑↑<br>++<br>++                                | Yes (learning difficulties)     | Yes               | Spastic dysarthria, scoliosis                          |
| FSP792-4 (M)  | 19                   | Stiff legs   | 32                  | 13                       | Gait possible without help/severe   | Severe/moderate legs mild arms | ↑→<br>N<br>++                                 | Yes (IQ = 45, MMSE = 23/30)     | Yes               | None   |
| FSP792-5 (F)  | 17                   | Stiff legs   | 37                  | 20                       | Walking aid/severe                  | Severe/mild distal             | ↑↑<br>N<br>++<br>↑↑                           | Yes (IQ = 47)                   | Yes               | None   |

|                    |    |                             |    |    |  |                               |                              |                       |                                    |  |
|--------------------|----|-----------------------------|----|----|--|-------------------------------|------------------------------|-----------------------|------------------------------------|--|
| FSP845-7<br>(M)    | 16 | Stiff legs                  | 29 | 13 | Wheelchair/<br>severe                      | Severe/mild                   | N<br>++<br>↑↑                | Yes<br>(MMSE = 15/30) | Yes                                | None   |
| FSP881-143<br>(M)  | 16 | Weakness legs               | 26 | 10 | Wheelchair/<br>severe                      | Severe/mild                   | N<br>++<br>↑↑                | No                    | ND                                 | None   |
| FSP881-144<br>(M)  | 14 | Weakness legs               | 30 | 16 | Wheelchair/<br>severe                      | Severe/severe                 | N<br>++<br>↑↑                | Yes mild              | ND                                 | Pes cavus, scoliosis   |
| FSP881-145<br>(F)  | 16 | Weakness legs               | 26 | 10 | Walking aid/<br>severe                     | Severe/none                   | ++<br>++<br>↑↑               | No                    | ND                                 | None   |
| FSP920-1401<br>(M) | 15 | Stiff legs                  | 26 | 11 | Wheelchair/<br>severe                      | Moderate/<br>generalized mild | +<br>+<br>↑↑                 | Yes (IQ = 63)         | Yes                                | Slow speech, pes<br>cavus, obesity   |
| FSP920-1402<br>(M) | 9  | Stiff legs                  | 18 | 9  | Wheelchair at<br>age 25/na                 | ND/none                       | +<br>+<br>↑↑                 | Yes (IQ = 63)         | Yes                                | Pes cavus  |
| FSP75-21<br>(F)    | 2  | Tip toe walking             | 24 | 22 | Gait possible<br>without help/<br>severe   | Moderate/severe               | ++<br>++<br>↑↑               | Yes (IQ = 52)         | Yes<br>hallucinations<br>at age 25 | Nystagmus,<br>hypertrichosis,<br>dysarthria,<br>orthostatic<br>hypotension<br>Spastic dysarthria |
| FSP75-43<br>(M)    | 7  | Learning<br>difficulties    | 33 | 26 | Wheelchair/<br>severe                      | Severe/generalized            | ++<br>++ (- at ankles)<br>↑↑ | Yes (IQ=64)           | Yes                                |  |
| FSP847-22<br>(M)   | 15 | Unsteadiness,<br>stiff legs | 30 | 15 | Wheelchair/<br>severe                      | Severe/generalized            | ++<br>++<br>↑↑               | Yes                   | ND                                 | Dysarthria,<br>saccadic pursuit,<br>vertical and<br>horizontal gaze<br>limitations, scoliosis    |
| FSP847-23<br>(M)   | 16 | Unsteadiness,<br>stiff legs | 29 | 13 | Wheelchair/<br>severe                      | Severe/generalized            | ++<br>++<br>↑↑               | Yes                   | ND                                 | Dysarthria, saccadic<br>pursuit, vertical<br>and horizontal<br>gaze limitations,<br>scoliosis    |
| FSP847-25<br>(F)   | 15 | Unsteadiness,<br>stiff legs | 22 | 7  | Gait possible<br>without help/<br>moderate | Moderate/mild<br>distal LL    | ++<br>++<br>↑↑               | No                    | ND                                 | Dysarthria, saccadic<br>pursuit  |
| FSP892-3<br>(M)    | 22 | Dysarthria                  | 30 | 8  | Gait possible<br>without<br>help/mild      | Moderate/none                 | N<br>++<br>↑↑                | Yes                   | Yes                                | Spastic dysarthria   |
| FSP670-4<br>(F)    | 12 | Cognitive<br>difficulties   | 31 | 19 | Wheelchair/severe                          | Severe/none                   | +<br>++<br>↑↑                | Yes (IQ = 61)         | Yes                                | High arched palate,<br>hyperpigmented<br>skin  |
| FSP670-5<br>(F)    | 12 | Cognitive<br>difficulties   | 30 | 18 | Wheelchair/<br>Severe                      | Severe/none                   | +<br>++<br>↑↑                | Yes (IQ = 69)         | Yes                                |  |

(continued)

Table 3 Continued

| Patient (Sex)   | Age at onset (years) | Symptoms at onset | Age at exam (years) | Disease duration (years) | Severity/leg spasticity             | Weakness/wasting         | Pyramidal syndrome [UL/knee/plantar reflexes] | Mental impairment           | Cognitive decline                    | Other signs or symptoms  |
|-----------------|----------------------|-------------------|---------------------|--------------------------|-------------------------------------|--------------------------|---|-----------------------------|--------------------------------------|--|
| FSP830-5 (F)    | 27                   | Unsteadiness      | 43                  | 16                       | Wheelchair at age 40/severe         | Severe/Mild and distal   | ++<br>↑↑                                      | No                          | Yes                                  | Macular degeneration   |
| FSP343-I085 (M) | 16                   | Stiff legs        | 33                  | 17                       | Walking aid/severe                  | Moderate/mild UL         | +<br>++<br>↑↑                                 | Yes (learning difficulties) | Yes                                  | Dysarthria, pes cavus, macular excavation                                      |
| FSP343-I08I (F) | 5                    | Leg weakness      | 40                  | 35                       | Wheelchair/moderate                 | Severe/none              | N<br>++<br>↑↑                                 | No                          | No                                   | Meningoencephalitis after measles and sequella (left hemiparesis and epilepsy) |
| FSP343-I084 (M) | 22                   | Weakness legs     | 37                  | 15                       | Walking aid/moderate                | Moderate/none            | +<br>++<br>↑↑                                 | No                          | No                                   | Pes cavus, dysarthria, nystagmus, macular excavation                           |
| FSP522-I (F)    | 19                   | Stiff legs        | 25                  | 6                        | Gait possible without help/moderate | Moderate/none            | +<br>++<br>↑↑                                 | No (MMSE = 26/30)           | Yes (memory and executive functions) | Abnormal fat distribution, orthostatic hypotension                             |
| SAL646-6 (F)    | 15                   | Stiff legs        | 23                  | 8                        | Gait possible without help/moderate | Mild/none                | +<br>++<br>↑↑                                 | Yes                         | Yes                                  | Pes cavus, Spastic dysarthria, Impaired pin-prick sense, deceased at age 36    |
| FSP683-3 (M)    | 18                   | Stiff legs        | 20                  | 2                        | Gait possible without help/moderate | Moderate mild UL/mild LL | +<br>+<br>↑↑                                  | No                          | No                                   | Scoliosis  |
| FSP398-I5 (F)   | 23                   | Gait difficulties | 42                  | 19                       | Wheelchair/severe                   | Moderate/severe          | +<br>++ (- at ankles)<br>NA                   | No                          | Yes                                  | Spastic dysarthria, saccadic pursuit, swallowing difficulties                  |
| FSP398-I7 (F)   | 17                   | Stiff legs        | 38                  | 21                       | Wheelchair at age 26/severe         | Mild/severe              | ++<br>++<br>NA                                | No                          | Yes                                  | Dysarthria, urinary incontinence, swallowing difficulties                      |
| FSP398-I9 (F)   | 15                   | Gait difficulties | 35                  | 20                       | Bedridden/severe                    | Severe/severe            | ++<br>++ (- at ankles)<br>NA                  | No                          | Yes                                  | Strabism, spastic dysarthria, swallowing difficulties, saccadic pursuit        |

M = male; F = female; ND = not done; NA = not assessed; N = normal; UL = upper limbs; LL = lower limbs; IQ = intellectual quotient; MMSE = Mini Mental State Evaluation; + = enhanced; ++ = increased. Left and right plantar reflexes are indicated as: '↑' = extensor, '→' = indifferent.

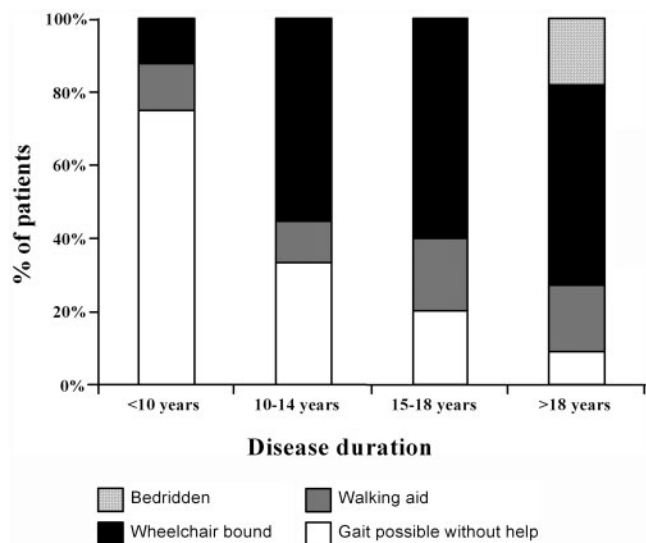


Fig. 2 Severity according to disease duration in SPG11 patients.

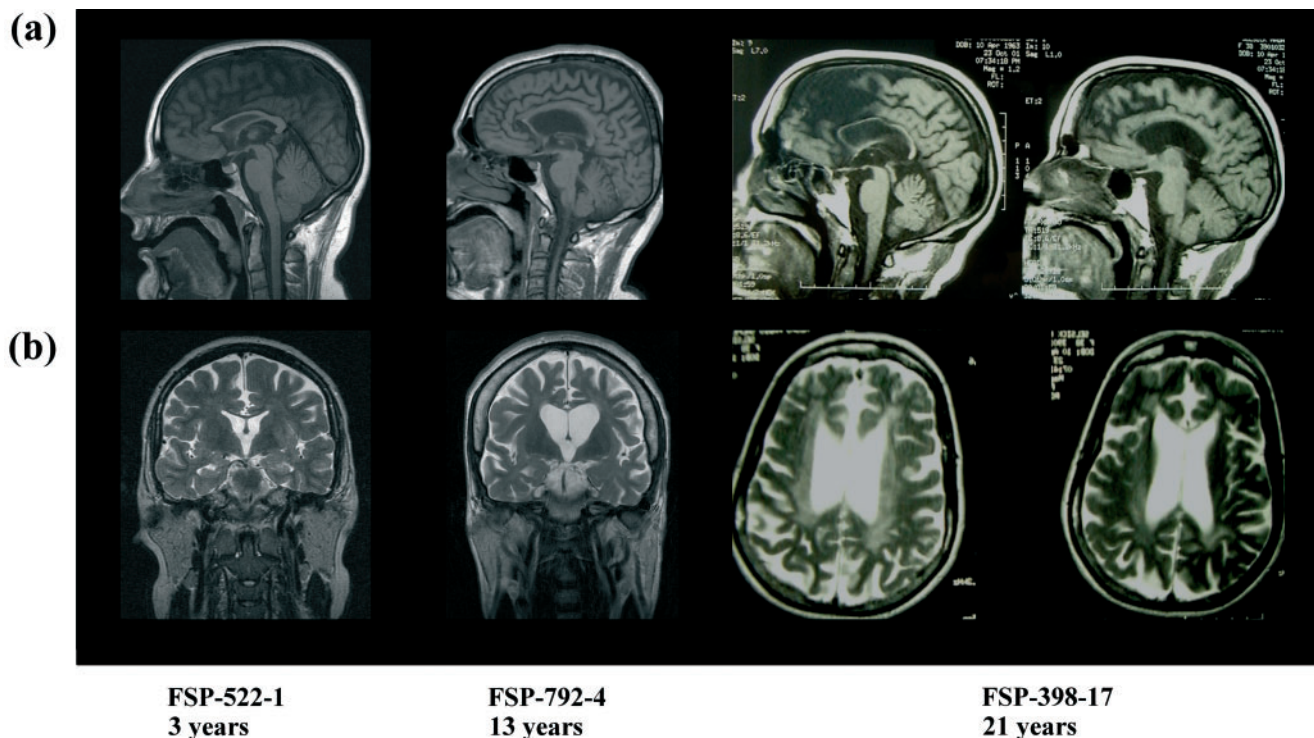
5–49,  $n = 5$ ) in SPG4 patients (Depienne *et al.*, 2006, 2007). Mental retardation or cognitive impairment and atrophy of the *corpus callosum* are the hallmarks of this disorder, but they may be lacking in patients with short disease durations (<10 years). Another frequent sign is axonal neuropathy, sometimes associated with anterior horn signs, observed in 81% of affected subjects. This is indicative of lower motor neuron degeneration and may clinically mimic amyotrophic lateral sclerosis when wasting is marked. White matter abnormalities are also frequently observed on MRI (69%). They start in the periventricular regions close to the frontal and occipital horns and increase in frequency and severity with disease duration, which may lead to a misdiagnosis of leucodystrophy. Finally, cerebellar ocular signs may also occur as the disease progresses.

The phenotype of 21 patients from 15 kindreds with TCC and mental impairment but no mutations in *SPG11* did not differ from SPG11 patients except for an earlier mean age at onset of  $9.6 \pm 13.0$  (range: 6 months to 50 years). Gait instability was the sign at onset in all patients

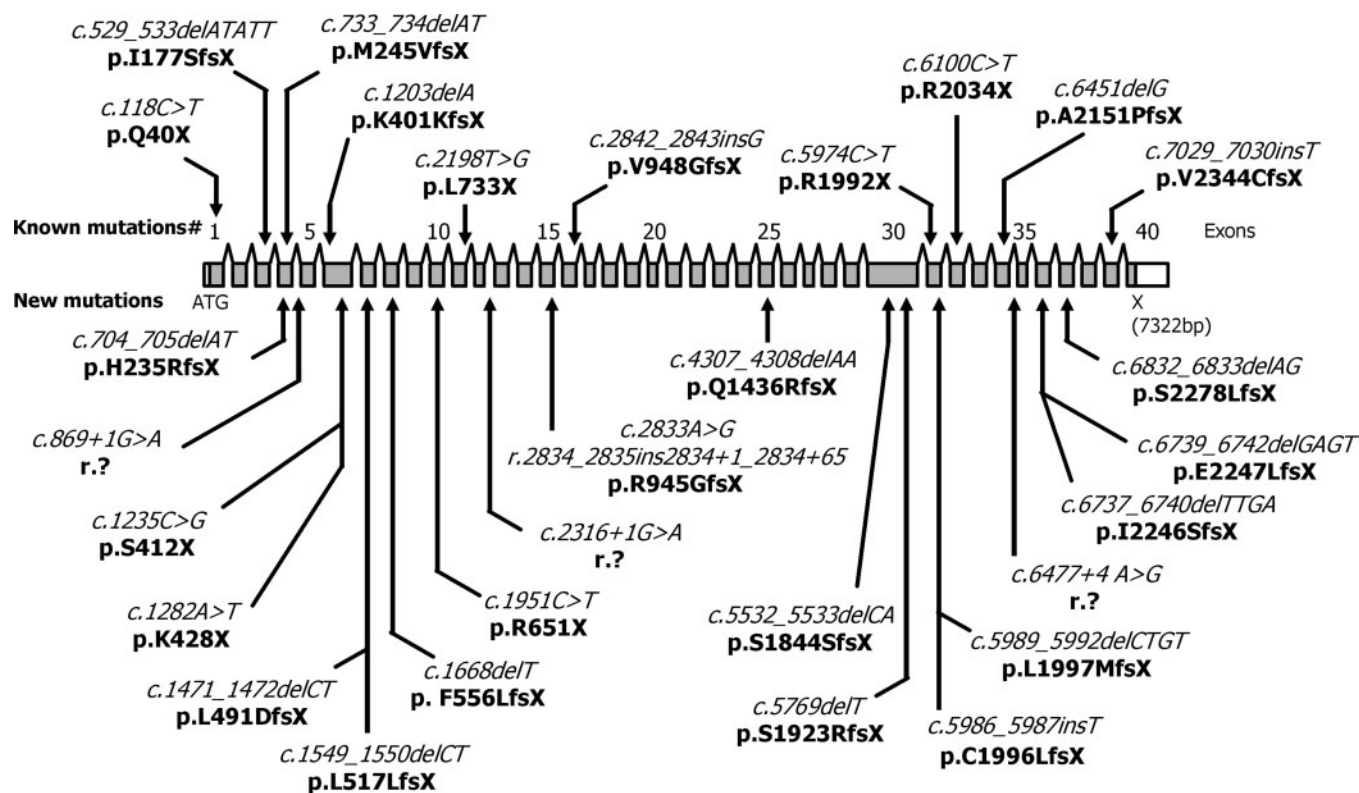
Table 4 Paraclinical investigations in 27 SPG11 patients

| Individual  | Disease duration (years) | Cerebral MRI                         |     |               | ENMG<br>(N neuropathy)                              | Evoked potential<br>(V visual, A auditory, S somatosensory) |
|-------------|--------------------------|--------------------------------------|-----|---------------|---|---|
|             |                          | Cortical atrophy                     | TCC | WMA           |   |   |
| FSP522-1    | 6                        | —                                    | +   | —             | Axonal sensory-motor N                              | A,V Normal  |
| FSP400-5    | 7                        | +                                    | —   | + (mild)      | Normal  | S abnormal, V normal  |
| FSP847-25   | 7                        | +                                    | +   | NA            | ND  | ND  |
| SAL646-1    | 8                        | ND                                   | ND  | ND            | Axonal sensory-motor, neurogenic pattern            | V abnormal, A, S normal                                     |
| FSP892-3    | 8                        | —                                    | +   | —             | Axonal motor N                                      | ND  |
| FSP670-5    | 8                        | + (mild frontal)                     | +   | +             | ND  | ND  |
| FSP838-1    | 9                        | +                                    | +   | + (mild)      | Normal  | ND  |
| FSP920-1402 | 9                        | —                                    | +   | —             | ND  | ND  |
| FSP670-4    | 9                        | ND                                   | ND  | ND            | Axonal sensory-motor N                              | V abnormal  |
| FSP920-1401 | 11                       | —                                    | +   | —             | ND  | ND  |
| FSP870-20   | 11                       | ND                                   | ND  | ND            | Axonal Motor N                                      | ND  |
| FSP792-4    | 13                       | +                                    | +   | +             | Axonal motor N                                      | ND  |
| FSP845-7    | 13                       | +                                    | +   | +             | ND  | ND  |
| FSP847-23   | 13                       | +                                    | +   | NA            | ND  | ND  |
| SPDI99-15   | 15                       | +                                    | +   | +             | Axonal sensory-motor N (biopsy: neurogenic atrophy) | ND  |
| FSP683-3    | 15                       | —                                    | +   | + (posterior) | ND  | ND  |
| SPDI99-1    | 16                       | +                                    | +   | +             | Axonal sensory-motor N                              | ND  |
| FSP830-5    | 16                       | +                                    | +   | +             | Axonal motor N                                      | V abnormal  |
| FSP393-12   | 17                       | ND                                   | ND  | ND            | Anterior horn                                       | ND  |
| FSP343-1085 | 17                       | + (mild)                             | +   | —             | Axonal sensory-motor N                              | ND  |
| FSP881-144  | 19                       | ND                                   | ND  | ND            | Axonal sensory-motor N                              | S normal  |
| FSP792-5    | 20                       | +                                    | +   | +             | ND  | ND  |
| FSP75-21    | 20                       | + (mild, megacisterne posterior)     | +   | —             | ND  | ND  |
| FSP398-17   | 21                       | +                                    | +   | +             | ND  | ND  |
| FSP831-5    | 15                       | +                                    | +   | +             | Normal  | V,A,S normal  |
| FSP75-43    | 26                       | +                                    | +   | + (bifrontal) | Anterior horn involvement                           | ND  |
| FSP343-1081 | 35                       | + (sequellae fronto-parietal lesion) | NA  | NA            | ND  | ND  |

+ = Present; — = absent; ENMG = electroneuromyography; NA = not assessed; ND = not done; WMA = white matter abnormalities; TCC = thin *corpus callosum*.



**Fig. 3** Brain MRI images of three patients with different disease durations. **(A)** T1-weighted sagittal image showing thin *corpus callosum* and global cortical atrophy, **(B)** T2–Fast Spin Echo weighted axial and coronal images showing periventricular hyperintensities of variable severity according to disease duration.



**Fig. 4** Schematic representation of the SPGII gene showing the location of the known (up) and new (down) mutations. # reported by Stevanin et al., 2007 and DelBo et al., 2007.

**Table 5** Frequency of *SPG11* mutations according to the phenotype

| Phenotypes                                 |   | HSP associated with          |                                  |                                     |                                  | Sum         |
|--|---|------------------------------|----------------------------------|-------------------------------------|----------------------------------|-------------|
|  |   | TCC and cognitive impairment | TCC without cognitive impairment | cognitive impairment (MRI not done) | cognitive impairment without TCC |             |
| This study                                 | Number of index cases                             | 26                           | 11                               | 17                                  | 22                               | 76          |
|  | Excluded by linkage analysis/Nb families analyzed | 5/14                         | 0/3                              | 1/6                                 | 6/10                             | 12/33       |
|  | Mutated index cases/Nb sequenced cases            | 11/21                        | 4/11                             | 4/16                                | 1/16                             | 20/64       |
|  | SPG11 frequency                                   | 11/26 (42%)                  | 4/11 (36%)                       | 4/17 (23%)                          | 1/22 (4.5%)                      | 20/76 (20%) |
| This study and Stevanin <i>et al.</i> 2007 |   | 22/37 (59%)                  | 4/12 (33%)                       | 4/17 (23%)                          | 1/22 (4.5%)                      | 31/88 (35%) |

HSP = hereditary spastic paraplegia; TCC = thin corpus callosum

and cerebellar signs were present in 5. Eight of these 15 individuals were sporadic.

In summary, the presence of HSP-TCC is the best single indicator that *SPG11* should be tested in patients with onset in the first to third decade, but the presence of one or more other signs, such as mental retardation and later cognitive deterioration, lower motor neuron involvement and white matter lesions, increases the chance of identifying *SPG11* mutations. Additionally, evidence of white matter abnormalities in the periventricular regions increases even further the probability that *SPG11* is the cause of the disease, rather than other causes of leucodystrophy. HSP mainly affects the corticospinal axons by a dying back mechanism but lesions in *SPG11* are wider, as suggested by the identification of TCC and other white matter abnormalities, signs of lower motor neuron degeneration, cerebellar ataxia and abnormal visual evoked potentials. Further studies are now required to understand the effects of these mutations, all truncating, causing the loss of spatacsin function in upper and lower motor neurons as well as in other regions of the nervous system.

## Supplementary materials

Supplementary materials are available at *Brain* online.

## Acknowledgements

The authors are grateful to the families and to the clinicians who referred patients to us, to Drs Samir Belal, Sebahattin Cirak, Michel Koenig, Clotilde Lagier-Tourenne and Merle Ruberg for their contribution in this study, to Drs Nizar Elleuch, Mohamed Imed Miladi, Catherine Lubetzki, Pilar Mazetti and Frederic Sedel for additional clinical investigations and to Nawal Benammar, Elodie Denis, Estelle Ferdiko and the DNA and cell bank of IFR70 for technical assistance. The study was funded by the Agence Nationale pour la Recherche (France, to A.D. and G.S.), the Verum foundation (Germany, to A.Br), the Hadassah France Fund

for studies in spastic paraplegia (France, to A.L.) and the Groupement d'Intérêt Scientifique – Institut des Maladies Rares (France, A04180DS/A04139DS to G.S.). P.D. and F.M.S. were supported by grants from Fondazione Mariani ONLUS and Telethon Italy (GGP06188).

## References

- Diagnosis and statistical Manual of Mental disorders. Washington DC: American Psychiatric Association, 2000.
- Casali C, Valente EM, Bertini E, Montagna G, Criscuolo C, De Michele G, et al. Clinical and genetic studies in hereditary spastic paraplegia with thin corpus callosum. *Neurology* 2004; 62: 262–8.
- Casari G, De Fusco M, Ciarmatori S, Zeviani M, Mora M, Fernandez P, et al. Spastic paraplegia and OXPHOS impairment caused by mutations in paraplegin, a nuclear-encoded mitochondrial metalloprotease. *Cell* 1998; 93: 973–83.
- Coutinho P, Barros J, Zemmouri R, Guimaraes J, Alves C, Choroa R, et al. Clinical heterogeneity of autosomal recessive spastic paraplegias: analysis of 106 patients in 46 families. *Arch Neurol* 1999; 56: 943–9.
- Del Bo R, Di Fonzo A, Ghezzi S, Locatelli F, Stevanin G, Costa A, et al. SPG11: a consistent clinical phenotype in a family with homozygous Spatacsin truncating mutation. *Neurogenetics* 2007; 8: 301–5.
- Depienne C, Fedirko E, Forlani S, Cazeneuve C, Ribai P, Feki I, et al. Exon deletions of SPG4 are a frequent cause of hereditary spastic paraplegia. *J Med Genet* 2007; 44: 281–4.
- Depienne C, Tallaksen C, Lephay JY, Bricka B, Poeta-Guyon S, Fontaine B, et al. Spastin mutations are frequent in sporadic spastic paraparesis and their spectrum is different from the one observed in familial cases. *J Med Genet* 2006; 43: 259–65.
- Elleuch N, Depienne C, Benomar A, Hernandez AM, Ferrer X, Fontaine B, et al. Mutation analysis of the paraplegin gene (SPG7) in patients with hereditary spastic paraplegia. *Neurology* 2006; 66: 654–9.
- Engert JC, Berube P, Mercier J, Dore C, Lepage P, Ge B, et al. ARSACS, a spastic ataxia common in northeastern Quebec, is caused by mutations in a new gene encoding an 11.5-kb ORF. *Nat Genet* 2000; 24: 120–5.
- Filla A, De MG, Marconi R, Bucci L, Carillo C, Castellano AE, et al. Prevalence of hereditary ataxias and spastic paraplegias in Molise, a region of Italy. *J Neurol* 1992; 239: 351–3.
- Fink JK. Advances in the hereditary spastic paraplegias. *Exp Neurol* 2003; 184 (Suppl 1): S106–10.
- Fink JK. Hereditary spastic paraplegia. *Curr Neurol Neurosci Rep* 2006; 6: 65–76.
- Folstein MF, Folstein SE, McHugh PR. “Mini-mental state”. A practical method for grading the cognitive state of patients for the clinician. *J Psychiatr Res* 1975; 12: 189–98.

- Franca MC Jr., D'Abreu A, Maurer-Morelli CV, Seccolin R, Appenzeller S, Alessio A, et al. Prospective neuroimaging study in hereditary spastic paraplegia with thin corpus callosum. *Mov Disord* 2007; 22: 1556–62.
- Harding AE. Classification of the hereditary ataxias and paraplegias. *Lancet* 1983; 1: 1151–5.
- Hazan J, Fonknechten N, Mavel D, Paternotte C, Samson D, Artiguenave F, et al. Spastin, a new AAA protein, is altered in the most frequent form of autosomal dominant spastic paraplegia. *Nature Genet* 1999; 23: 296–303.
- Lossos A, Stevanin G, Meiner V, Argov Z, Bouslam N, Newman JP, et al. Hereditary spastic paraplegia with thin corpus callosum: reduction of the SPG11 interval and evidence for further genetic heterogeneity. *Arch Neurol* 2006; 63: 756–60.
- Mannan AU, Krawen P, Sauter SM, Boehm J, Chronowska A, Paulus W, et al. ZFYVE27 (SPG33), a novel spastin-binding protein, is mutated in hereditary spastic paraplegia. *Am J Hum Genet* 2006; 79: 351–57.
- Martinez MF, Kobayashi H, Pegoraro E, Galluzzi G, Creel G, Mariani C, et al. Genetic localization of a new locus for recessive familial spastic paraparesis to 15q13-15. *Neurology* 1999; 53: 50–6.
- Olmez A, Uyanik G, Ozgul RK, Gross C, Cirak S, Elibol B, et al. Further Clinical and genetic characterization of SPG11: hereditary spastic paraplegia with thin corpus callosum. *Neuropediatrics* 2006; 37: 59–66.
- Patel H, Cross H, Proukakis C, Hershberger R, Bork P, Ciccarelli FD, et al. SPG20 is mutated in Troyer syndrome, an hereditary spastic paraplegia. *Nature Genet* 2002; 31: 347–8.
- Raven RC. Revised manual for Raven's Progressive Matrices and Vocabulary Scale. UK: Windsor, 1982.
- Shibasaki Y, Tanaka H, Iwabuchi K, Kawasaki S, Kondo H, Uekawa K, et al. Linkage of autosomal recessive hereditary spastic paraplegia with mental impairment and thin corpus callosum to chromosome 15q13-15. *Ann Neurol* 2000; 48: 108–12.
- Simpson MA, Cross H, Proukakis C, Pryde A, Hershberger R, Chatonnet A, et al. Maspardin is mutated in mast syndrome, a complicated form of hereditary spastic paraplegia associated with dementia. *Am J Hum Genet* 2003; 73: 1147–56.
- Stevanin G, Montagna G, Azzedine H, Valente EM, Durr A, Scarano V, et al. Spastic paraplegia with thin corpus callosum: description of 20 new families, refinement of the SPG11 locus, candidate gene analysis and evidence of genetic heterogeneity. *Neurogenetics* 2006; 7: 149–56.
- Stevanin G, Santorelli FM, Azzedine H, Coutinho P, Chomilier J, Denora PS, et al. Mutations in SPG11, encoding spatacin, are a major cause of spastic paraplegia with thin corpus callosum. *Nature Genet* 2007; 39: 366–72.
- Tallaksen CM, Durr A, Brice A. Recent advances in hereditary spastic paraplegia. *Curr Opin Neurol* 2001; 14: 457–63.
- Valdmanis PN, Meijer IA, Reynolds A, Lei A, MacLeod P, Schlesinger D, et al. Mutations in the KIAA0196 gene at the SPG8 locus cause hereditary spastic paraplegia. *Am J Hum Genet* 2007; 80: 152–61.
- Wechsler D. Wechsler Memory Scale-Revised manual. San Antonio, TX: The Psychological Corporation; 1987.
- Winner B, Gross C, Uyanik G, Schulte-Mattler W, Lurding R, Marienhagen J, et al. Thin corpus callosum and amyotrophy in spastic paraplegia-Case report and review of literature. *Clin Neurol Neurosurg* 2006; 692–8.
- Zhao X, Alvarado D, Rainier S, Lemons R, Hedera P, Weber CH, et al. Mutations in a newly identified GTPase gene cause autosomal dominant hereditary spastic paraplegia. *Nature Genet* 2001; 29: 326–31.

## Appendix

Members of the Spastic Paraplegia and ATAXia network (SPATAX): Dr A. Durr (Hôpital Pitié-Salpêtrière, Paris, France), Pr B. Fontaine (Hôpital Pitié-Salpêtrière, Paris, France), Pr J.P. Azulay (Hôpital de la Timone, Marseille, France), Pr A. Benomar (CHU, Rabat, Morocco), Pr E. Bertini (Osp. Bambino Gesù, Roma, Italy), Dr O. Boespflug-Tanguy (Faculté de Médecine, Clermont-Ferrand, France), Pr P. Coutinho (Hospital San Sebastião, Santa Maria da Feira, Portugal), Pr A. Filla (Università Degli Studi Di Napoli Federico II, Napoli, Italy), Pr D. Hannequin (Hôpital Charles Nicolle, Rouen, France), Dr A. Hamri (Hôpital Benbadis, Constantine, Algeria), Pr Michel Koenig (IGBMC, Illkirch, France), Pr P. Labauge (Hôpital Caremeau, Nimes, France), Pr A. Lossos (Hadassah Hebrew University Hospital, Jerusalem, Israel), Pr A. Megarbane (Université Saint-Joseph, Beirut, Lebanon), Pr J.E. Nielsen (The Panum Institute, Copenhagen, Denmark), Pr A.M. Ouvrard Hernandez (CHU, Grenoble, France), Dr E. Reid (Addenbrooke's Hospital, Cambridge, UK), Dr D. Rodriguez (Hôpital St Vincent De Paul, Paris, France) Pr S. Roumani (Dpt of Neurology, Damascus, Syria), Pr M. Salih (University Hospital, Ryadh, Saudi Arabia), Pr J. Sequeiros (University of Porto, Porto, Portugal), Dr C. Tallaksen (Ullevål University Hospital, Oslo, Norway), Pr M. Tazir (CHU Mustapha, Algiers, Algeria), Pr F. Tison (Groupe Hospitalier Sud, Pessac, France), Dr C. Goizet (Hôpital Pellegrin, Bordeaux, France), Dr E.M. Valente (Istituto Di Genetica Medica, Roma, Italy), Pr N. Wood (The National Hospital, London, UK), Dr C. Verny (CHU, Angers, France) and Pr T. Warner (Royal Free and University College Medical School, London, UK).