Perivascular spaces—MRI marker of inflammatory activity in the brain?

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The Virchow-Robin spaces (VRS), perivascular compartments surrounding small blood vessels as they penetrate the brain parenchyma, are increasingly recognized for their role in leucocyte trafficking as well as for their potential to modulate immune responses. In the present study, we investigated VRS numbers and volumes in different brain regions in 45 multiple sclerosis patients and 30 healthy controls of similar age and gender distribution, applying three different MRI sequence modalities (T2-weighted, T1-weighted and FLAIR). VRS were detected in comparable numbers in both multiple sclerosis patients and healthy individuals, indicating that perivascular compartments present on MRI are not a unique feature of multiple sclerosis. However, multiple sclerosis patients had significantly larger VRS volumes than healthy controls (P = 0.004). This finding was not explained by a significantly lower brain parenchymal fraction (BPF), resulting from a higher degree of atrophy, in the patient cohort. In a multiple linear regression analysis, age had a significant influence on VRS volumes in the control group but not in multiple sclerosis patients (P = 0.023 and P = 0.263, respectively). A subsequent prospective longitudinal substudy with monthly follow-up MRI over a period of up to 12 months in 18 patients revealed a significant increase in VRS volumes and counts accompanying the occurrence of contrast-enhancing lesions (CEL). At time points when blood-brain barrier (BBB) breakdown was indicated by the appearance of CEL, total VRS volumes and counts were significantly higher compared with preceding time points without CEL (P=0.011 and P=0.041, respectively), whereas a decrease thereafter was not statistically significant. Thus, our data points to an association of VRS with CEL as a sign for inflammation rather than with factors such as age, observed in healthy controls, and therefore suggests a role of VRS in inflammatory processes of the brain.

Keywords: perivascular spaces; Virchow-Robin spaces; multiple sclerosis; MRI; neuroinflammation

Abbreviations: BBB = blood-brain barrier; BPF = brain parenchymal fraction; CEL = contrast-enhancing lesion; CNS = central nervous system; CSF = cerbrospinal fluid; EAE = experimental autoimmune encephalomyelitis; EDSS = expanded disability status scale; FLAIR = fluid-attenuated inversion recovery; IFN- β = interferon-beta; VRS = Virchow-Robin spaces

Received April 7, 2008. Revised July 4, 2008. Accepted July 4, 2008. Advance Access publication August I, 2008

Introduction

The perivascular compartment surrounding small cerebral blood vessels (Virchow–Robin spaces, VRS) has been a focus of research for decades, although its role for lymphocyte trafficking into the brain was not appreciated until very recently (Greter *et al.*, 2005; van Horssen *et al.*, 2005; McMahon *et al.*, 2005; Cumurciuc *et al.*, 2006; Bechmann *et al.*, 2007). Owing to the complex topography of the

perivascular spaces, as illustrated in Fig. 1, two steps are considered necessary for cell invasion into the brain parenchyma: transmigration of the endothelium into the perivascular space and penetration through the glia limitans into the brain parenchyma. For the second step, it has been hypothesized that previously activated immune cells require restimulation (Tran *et al.*, 1998). In multiple sclerosis, a chronic inflammatory disease of the CNS, immune cell

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 Table I
 Demographic, clinical and MRI data of multiple sclerosis patients and healthy controls

	Multiple sclerosis patients	Controls
Male/Female	22/23	14/16
Age (years)	39.8 ± 8.2	37.8 ± 11.5
IMT (w/wo IFN-β)	32/I3	NA
EDSS	2.3 ± 1.4	NA
Months since last relapse	16.7 ± 17.9	NA
Disease duration (years)	8.8 ± 6.2	NA
BH count	5.6 ± 8.4	NA
BH volume (mm ³)	556.9 ± 890.5	NA
T2 lesion count	47.9 ± 35.6*	$4.5 \pm 6.0^*$
T2 lesion volume (mm ³)	$5827.9 \pm 5445.2^{\dagger}$	$160.0 \pm 283.0^{\dagger}$
BPF	0.85 ± 0.04 \$	$0.87 \pm 0.02^{\$}$
VRS count	34.4 ± 17.7	$\textbf{30.7} \pm \textbf{22.4}$
VRS volume (mm ³)	280.8 ± 165.1 §	$210.8 \pm 161.5^{\S}$
Ratio VRS volume/BPF	$331.3 \pm 197.8^{\#}$	$\textbf{242.0} \pm \textbf{189.1}^{\#}$

Mean values and SD are presented.

*P < 0.001; †P < 0.001; *P = 0.013; *P = 0.004; #P = 0.003.

IMT = immunomodulatory treatment; BH = black hole;

BPF = brain parenchymal fraction; EDSS = expanded disability status scale; IFN- β = interferon-beta; NA = not applicable.

invasion leads to focal inflammatory and demyelinating lesions as well as axonal and neuronal damage (Frohman et al., 2006; Zipp and Aktas, 2006). Although the exact pathogenesis of multiple sclerosis has not yet been elucidated, the current paradigm states that T cell activation is initiated in the peripheral lymphoid compartment. Once activated, T cells potentially enter the CNS via VRS, where they can be restimulated by antigen-presenting cells (Ransohoff et al., 2003; Greter et al., 2005; McMahon et al., 2005; Becher et al., 2006; Bechmann et al., 2007). Perivascular cuffing (Gareau et al., 2002) and VRS enlargement (Vos et al., 2005) have been observed in multiple sclerosis and its animal model, experimental autoimmune encephalomyelitis (EAE) in post-mortem studies. Perivascular (Achiron and Faibel, 2002; Barkhof, 2004) or perivenular (Ge et al., 2005) spaces detectable by MRI have also been described in vivo in physiological conditions such as the ageing brain as well as in multiple sclerosis. However, it is still a matter of debate as to whether VRS visible on MRI are a physiological feature or may be associated with multiple sclerosis. Therefore, this study aimed to address the following questions by means of a novel, 3D semi-automatic MRI approach: (i) does the presence of VRS on MRI distinguish multiple sclerosis patients from healthy controls?; (ii) do multiple sclerosis patients have more and/or larger VRS than healthy controls? and (iii) do VRS volumes change along with the appearance of contrast-enhancing lesions (CEL) indicating blood-brain barrier (BBB) leakage?

Material and methods

Participants

In this prospective study we enrolled 75 individuals, comprising 45 patients with relapsing–remitting multiple sclerosis, and a

Table 2 Demographic, clinical and MRI data of the I8 multiple sclerosis patients who participated in the longitudinal substudy

Male/Female Age (years) EDSS Months since last relapse Disease duration (years) Number of monthly scans T2 lesion volume (mm ³) (CELneg.0I) T2 lesion volume (mm ³) (CELpos.0I)	10/8 38.8 ± 6.5 1.8 ± 1.1 7.0 ± 3.9 8.3 ± 7.9 7.5 ± 2.6 $4213.4 \pm 3369.6^*$ $4868.5 \pm 3484.3^*$
T2 lesion volume (mm ³) (CELpos ₋ I0)	$4630.7 \pm 3436.5^{\pounds}$
T2 lesion volume (mm ³) (CELneg ₋ I0)	4377.1 ± 3405.3^{t}
VRS volume (mm ³) (CELneg ₀ I)	$151.2 \pm 67.6^{\$}$
VRS volume (mm ³) (CELpos.0I)	$177.3 \pm 71.4^{\$}$
VRS volume (mm ³) (CELpos.I0)	170.8 ± 90.2
VRS volume (mm ³) (CELneg.10)	149.8 ± 69.6
VRS count (CELneg.01)	14.8 ± 6.1 #
VRS count (CELpos.0I)	$18.2 \pm 7.0^{\#}$
VRS count (CELpos.I0)	17.3 ± 9.8
VRS count (CELneg-I0)	14.9 ± 7.0
CEL volume (mm ³) (CELneg.0I)	$8.8 \pm 20.4^{\S}$
CEL volume (mm ³) (CELpos.0I)	$278.4 \pm 140.2^{\S}$
CEL volume (mm ³) (CELpos IO)	$377.7 \pm 522.6^{\$}$
CEL volume (mm ³) (CELneg ₋ I0)	$17.4 \pm 23.8^{\text{\}}$
CEL count (CELneg.0I)	$0.5\pm0.9^{\dagger}$
CEL count (CELpos.0I)	$2.4\pm1.5^{\dagger}$
CEL count (CELpos.I0)	$1.7 \pm 1.1^{\ddagger}$
CEL count (CELneg.I0)	$\textbf{0.5} \pm \textbf{0.5}^{\ddagger}$

Mean values and SD are presented.

 $_{+}^{*}P = 0.001; _{+}^{\pounds}P = 0.041; _{+}^{\$}P = 0.011; _{+}^{\#}P = 0.041; _{+}^{\$}P = 0.001; _{+}^{\$}P < 0.001;$

 $^{\dagger}P = 0.003; \,^{\ddagger}P = 0.003.$

CELneg.0: scan without CEL at shift from 'CEL negative $(0) \rightarrow \text{CEL positive (I)}$ '.

CELpos.01: scan with CEL at shift from 'CEL negative (0) \rightarrow CEL positive (1)'.

CELpos.IO: scan with CEL at shift from 'CEL positive (I) \rightarrow CEL negative (0)'.

CELneg.I0: scan without CEL at shift from 'CEL positive (I) \rightarrow CEL negative (0)'.

control group of 30 healthy volunteers without history of neurological or psychiatric disease (Table 1). The two groups did not differ with respect to age or gender proportion. Patients were screened and enrolled in the outpatient clinic of the Cecilie Vogt Clinic for Neurology in the HKBB, Charité -Universitaetsmedizin Berlin, and met the diagnostic criteria according to McDonald et al. (2001). The study was approved by the Charité Research Ethics Committee, and written informed consent was obtained from all participants according to the Declaration of Helsinki. There was no difference between multiple sclerosis patients and controls with respect to hypertension, migraine or smoking and none of the patients suffered from a relapse or received steroid treatment within 4 weeks prior to an imaging time point. A subgroup of 18 multiple sclerosis patients participated in a longitudinal follow-up study with monthly MRI over a period of up to 12 months (Table 2).

Magnetic resonance imaging

MRI measurements were performed on a 1.5 tesla scanner (Sonata, Siemens Medical Systems, Erlangen, Germany). T₂- and proton density-weighted images were obtained by a triple echo

spin-echo sequence (TR 5780 ms, TE₁ 13 ms, TE₂ 81 ms, TE₃ 121 ms, 3 mm slice thickness and 44 contiguous axial slices). Additionally, we applied a fluid-attenuated inversion recovery (FLAIR) sequence (TIRM, TR 10000 ms, TE 108 ms, TI 2500 ms, 3 mm slice thickness and 44 contiguous axial slices) and a high resolution 3D T₁-weighted sequence (MPRAGE, TR 2110 ms, TE 4.38 ms, TI 1100 ms, flip angle 15°, resolution 1 mm³). Conventional spin-echo T₁-weighted (TR 1060 ms, TE 14 ms, 3 mm slice thickness and 44 contiguous axial slices) images were obtained before and 5 min after injection of 0.1 mmol/kg gadopentate dimeglumine (Gd-DTPA (Magnevist®), Bayer-Schering, Berlin, Germany). A series of axial, coronal and sagittal images was obtained to create a reference scan for subsequent accurate repositioning of patients at follow-up. Blinded MRI data analysis was performed following a semi-automatic procedure, which included an image coregistration (Jenkinson et al., 2002) as well as an inhomogeneity correction routine embedded into the MedX3.4.3 software package (Medical Numerics, Germantown, USA). Time points with total CEL volumes greater than 100 mm³ were considered clearly CEL positive. Bulk white matter lesion load of T2-weighted scans as well as number and volume of hypoand hyperintense lesions on T₁-weighted scans were routinely measured using the MedX3.4.3 software package, as described previously (Wuerfel et al., 2004). Brain parenchymal fraction (BPF) was calculated applying a fully automated software tool (Smith et al., 2002), that yielded stable results in multiple sclerosis (Jasperse et al., 2007).

Establishment of a novel VRS quantification paradigm

VRS are cerebrospinal fluid-filled (CSF) spaces ensheathing small blood vessels as they penetrate the brain parenchyma (Fig. 1A). They are enclosed by endothelial cells towards the blood vessel lumen and by astrocytic endfeet forming the glia limitans on the neuropil side (Fig. 1B and C). VRS were defined on MRI applying

three different sequence modalities after coregistration and inhomogeneity correction of the images, and could thus be differentiated from inflammatory lesions: (i) VRS were visualized as hyperintense regions on T_2 -weighted sequences and as hypointense regions on T_1 -weighted sequences (Fig. 2A and C). VRS within lesions were excluded to avoid partial volume contamination. Depending on the imaging plane, VRS formed punctuate or tubular structures of usually <3 mm in diameter. (ii) Anatomical tracing of blood vessels and VRS was achieved on high-resolution 3D isotropic T_1 -weighted images (MPRAGE), and thus confirmed differentiation from multiple sclerosis lesions

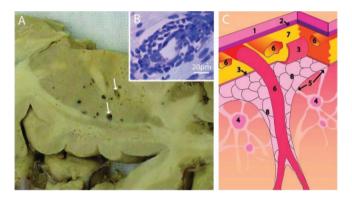


Fig. 1 The topography of VRS is presented in a post-mortem brain section. Arrows highlight VRS surrounding subcortical blood vessels (A). In a highly magnified toluidin blue-stained section (B) a small blood vessel is depicted, surrounded by cells that extravasated through the endothelium into the perivascular space. Very few cells crossed the glia limitans into the neuropil. The illustration (C) shows how the perivascular space is bordered by the outer vascular wall of cerebral vessels (6) and by astrocytic endfeet (5) forming the glia limitans (8) on the neuropil side. Dura mater (I), arachnoidea (2), pia mater (3), astrocytes (4) and the subarachnoidal space (7) are marked.

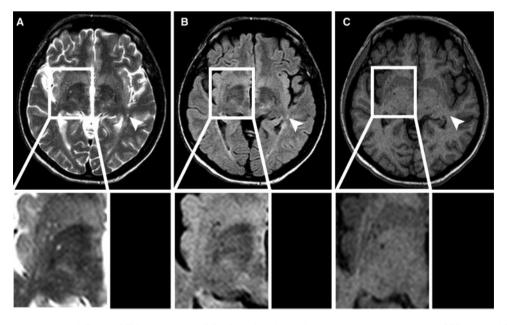


Fig. 2 VRS can be detected on different MRI sequences. VRS of the basal ganglia appear isointense to the CSF on axial T_2 -weighted (A), FLAIR (B) and MPRAGE images (C), and are depicted below at a higher magnification. FLAIR imaging was used to distinguish multiple sclerosis lesions (arrow-head) from VRS; both present as hyperintensities on T_2 -weighted images.

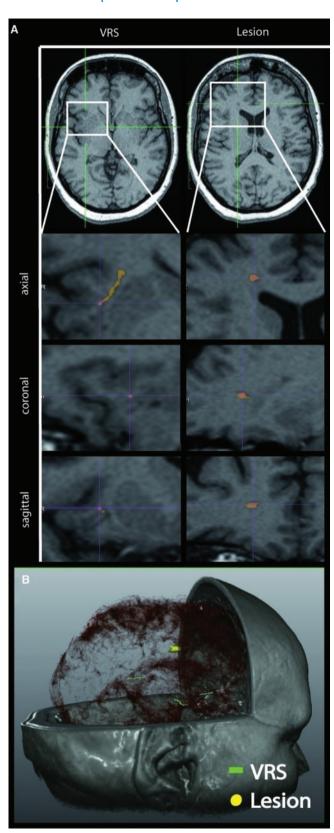


Fig. 3 The 3D volume images (MPRAGE) visualized the tubular structure of blood-vessel ensheathing VRS (A) and facilitated differentiation from small inflammatory lesions (B). Higher magnifications of the three main image planes are presented (A; axial, coronal and sagittal). The difference in the spatial

(Fig. 3A and B). VRS were typically located in conformity with the path of perforating brain vessels, which run perpendicular to the brain surface. (iii) On FLAIR, as in every other pulse sequence applied, VRS appeared isointense to CSF and could be easily differentiated from small inflammatory plaques (Fig. 2B). VRS were identified by two experienced raters (JW, MH) in consensus reading as clearly delineated hyperintense structures on corresponding T2-weighted scans, and both VRS numbers and volumes were quantified in a blinded fashion by applying a raterindependent threshold-based semi-automatic post-processing routine, embedded into the MedX3.4.3 software package following a procedure described elsewhere (Makale et al., 2002). This software tool was further developed for multiple sclerosis lesion quantification, as recently reported (Wuerfel et al., 2004; Paul et al., 2008). For the quantification of very small structures such as VRS (average lesion diameter <3 mm), software parameters including connectivity and threshold values were adjusted and the results of the semi-automatic analysis were tested against manually determined VRS counts and volumes for validation purposes on four participants. As a result of practically identical measurements, comparison of semi-automatic versus manual quantification of VRS counts and volumes yielded an intra-class correlation coefficient of 1. Furthermore, VRS counts and VRS volumes were investigated in 10 randomly selected study participants by one experienced reader (MH) at two different time points to assess the intra-rater reliability of the semi-automatic quantification procedure. The intra-class correlation coefficient was 0.996 (95% CI 0.985-0.999) for VRS counts and 0.998 (95% CI 0.994-1.000) for VRS volumes.

Anatomy and histology

In order to show in principle the topography of VRS, a brain from the Frankfurt Brain Bank was cut in horizontal sections (Fig. 1A). Histological paraffin-sections derived from areas of inflammation were stained with toluidin blue. It has often been argued that VRS are artefacts of fixation, but we have previously demonstrated their functional connection to the subarachnoid space using tracer injections (Bechmann *et al.*, 2001*a*, *b*).

Statistical analysis

Results are expressed as arithmetic mean \pm SD. We applied the Mann–Whitney U-test to assess group differences between the two groups (multiple sclerosis patients versus healthy controls) in age, T2 lesion count, T2 lesion volume, BPF, VRS count, VRS volume and VRS volume/BPF ratio; and to assess group differences between patients with and without immunomodulatory therapy with IFN- β . Correlations between VRS counts and volumes on the one hand and age, EDSS, duration of disease and other MRI parameters on the other were calculated by Spearman's correlation coefficient. The Fisher exact test was used to test for differences regarding gender distribution between the multiple sclerosis and the control group. A multiple linear regression analysis was separately performed for both the multiple sclerosis and the

formation between small ball-shaped multiple sclerosis lesions and tubular VRS is not always obvious depending on the image plane (see sagittal and coronal versus axial slices in A), but can be easily appreciated in a 3D reconstruction (B). The 3D reconstruction was performed with MeVisLab SDK vI.6 (MeVisLab, Bremen, Germany).

healthy control groups to investigate the influence of age and BPF (independent variables) on VRS volumes and on VRS numbers (dependent variables). To assess the degree of agreement between semi-automatic and manual VRS number and volume quantification, the intra-class correlation coefficient was calculated using a one-way random model. Also intra-rater reliability of VRS number and volume quantification at two different time points was calculated by a one-way random model. In the longitudinal observation in 18 patients, Wilcoxon tests were applied to compare VRS volumes, VRS numbers, T2 lesion volumes, CEL volumes and CEL counts between two consecutive time points (months) that were defined by a shift from a CEL-negative scan to a CEL-positive scan ('CEL negative->CEL positive') or from a CEL-positive scan to a CEL-negative scan ('CEL positive→CEL negative'). One patient showed two 'CEL negative

CEL positive' shifts, 12 patients had one 'CEL negative→CEL positive' shift and 5 patients had no 'CEL negative→CEL positive' shift. Four patients showed two 'CEL positive-CEL negative' shifts, and 14 patients experienced only one 'CEL positive

CEL negative' shift. For reasons of clarity, we consistently included only the first 'CEL negative→CEL positive' shift and the first 'CEL positive→CEL negative' shift in our statistical analyses. A two-tailed Pvalue < 0.05 was considered statistically significant. All tests were performed as exploratory data analyses, such that no adjustments for multiple testing were made. The numerical calculations were performed with SPSS Version 13 (SPSS, Inc., Chicago, IL, USA) and SAS Version 9.1 (SAS Institute, Inc., Cary, NC, USA).

Results

VRS are detectable in healthy individuals and in multiple sclerosis patients

VRS were visualized predominantly in two brain regions: (i) the basal ganglia, encompassing lenticulostriate arteries and (ii) the cerebral high convexity along perforating blood vessels. VRS were detectable in the basal ganglia and the high convexity bilaterally in both multiple sclerosis patients and healthy controls, with the exception of two healthy volunteers who did not show subcortical VRS. Thus, the presence of VRS as such did not differentiate multiple sclerosis patients from healthy controls. Moreover, the number of VRS did not differ significantly between the two groups in any of the brain regions investigated (Table 1 and Fig. 4A). Figure 3A gives an example of VRS compared with multiple sclerosis plaques depending on the image plane and impression of their differential appearance in 3D space (Fig. 3B).

Multiple sclerosis patients present with larger VRS volumes compared with healthy controls

In contrast to the similar VRS counts, VRS volumes differed clearly between patients and controls. Multiple sclerosis patients presented with larger VRS bilaterally, resulting in significantly higher total VRS volumes (P=0.004, Table 1 and Fig. 4B). None of the MRI parameters—such as number and volume of T2 lesions, or the number and volume of T1-hypointensities ('black holes')—correlated to VRS numbers and volumes. VRS numbers and volumes also did not correlate with disease duration, EDSS or the time since last relapse. Furthermore, VRS numbers and volumes did not differ between patients with or without immunomodulatory therapy. Interestingly, we found a significant positive correlation between VRS volumes and age only in the control group, i.e. VRS volumes increase with age (r=0.485, P=0.007). This significant correlation remained virtually unchanged when we repeated the calculation without one obvious outlier in the healthy control group (r=0.458, P=0.013). VRS volumes of multiple sclerosis patients do not, however, increase significantly with age (r=0.135, P=0.377). Concerning VRS counts, there was a significant positive correlation with age in the control group (r=0.370, P=0.044), while this did not hold true in the multiple sclerosis group (r = 0.135, P = 0.377).

Multiple sclerosis patients have smaller BPF but reveal no relation between these and VRS

Not surprisingly, and in line with previous reports (Miller et al., 2002), multiple sclerosis patients in our study had

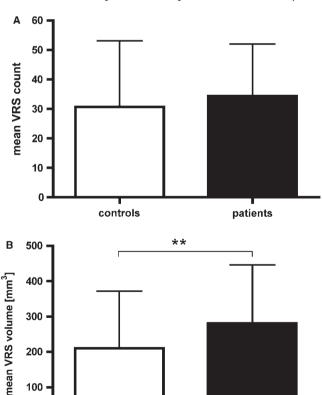


Fig. 4 VRS counts did not differentiate multiple sclerosis patients from healthy controls (A), but VRS volumes were significantly larger in multiple sclerosis patients (**B**; **P = 0.004). Mean values and SD are presented.

patients

controls

100

n

significantly lower BPF, indicating a higher degree of brain atrophy, as compared with healthy controls (P=0.013, Table 1 and Fig. 5). In the control group, there was a significant correlation between age and BPF, i.e. a decrease of BPF with increasing age (r=-0.425, P=0.019), while this did not hold true in the multiple sclerosis group (r=-0.232, P=0.126).

In the entire cohort as well as in both the multiple sclerosis and healthy controls group, there was no correlation between BPF on the one hand and VRS volumes and VRS counts on the other. We further calculated the VRS volume/BPF ratio, which was significantly higher in multiple sclerosis patients compared with controls (P = 0.003, Table 1). The VRS volume/BPF ratio significantly increased with age only in the control group (r = 0.551, P = 0.002, Fig. 6) but not in the multiple sclerosis group (r = 0.178, P = 0.242, Fig. 6). In the multiple sclerosis group, multiple linear regression analyses did not reveal a significant influence of either age or BPF on either VRS volumes (P = 0.263 and P = 0.513, respectively) or on VRS counts (P = 0.376 and P = 0.789, respectively). In the control group, multiple linear regression analysis showed a significant influence of age, but not of BPF, on VRS volumes (P = 0.023 and P = 0.455, respectively) and VRS numbers (P = 0.020 and P = 0.429, respectively).

VRS volumes increase during acute inflammation

Following recent animal studies (Bechmann *et al.*, 2001*a*; Becher *et al.*, 2006), we investigated whether a shift from a CEL-negative to a CEL-positive scan at two consecutive time points indicating BBB breakdown was accompanied by changes in VRS volumes or VRS counts. In the 18 patients who participated in the longitudinal substudy, mean VRS volumes were significantly higher at time points of CEL occurrence compared with the preceding time points

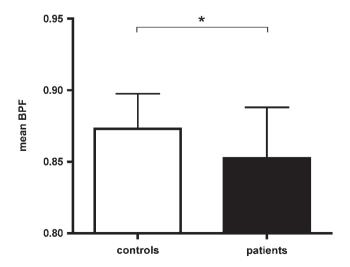


Fig. 5 BPF was significantly lower in multiple sclerosis patients (*P = 0.013). Mean values and SD are presented.

without CEL (shift from 'CEL negative \rightarrow CEL positive', P = 0.011, Table 2 and Fig. 7A, left-hand image). The decrease of VRS volumes upon disappearance of CEL did not reveal statistical significance, however, at CEL-negative time points, VRS volumes were lower compared with the preceding CEL-positive time point (shift from 'CEL positive \rightarrow CEL negative', P = 0.085, Table 2 and Fig. 7A, right-hand image). Also mean VRS numbers were significantly higher at time points of CEL occurrence compared with the preceding time points without CEL (shift from 'CEL negative \rightarrow CEL positive', P = 0.041, Table 2 and Fig. 7B, left-hand image). The decrease of VRS numbers upon disappearance of CEL was not statistically significant, however, at CEL-negative time points, VRS numbers were lower compared with the preceding CEL-positive time point (P = 0.111, Table 2 and Fig. 7B, right-hand image). T2 lesion load also increased when a shift from 'CEL negative -> CEL positive' occurred (P = 0.001, Table 2), and decreased when a shift from 'CEL positive → CEL negative' occurred (P=0.041, Table 2). In Fig. 8, the change in VRS size and number is shown between two time points prior to and during contrast enhancement (Fig. 8A and B, respectively). In none of our patients was meningeal contrast enhancement detected at any time point.

Discussion

In this study, we demonstrate increased size and disease-dependent changes of VRS volumes in multiple sclerosis patients versus healthy controls. In contrast to earlier investigations, we were able to reliably and reproducibly quantify not only subcortical VRS numbers, but also VRS volumes in different anatomical regions. By means of three different imaging modalities, VRS were clearly and

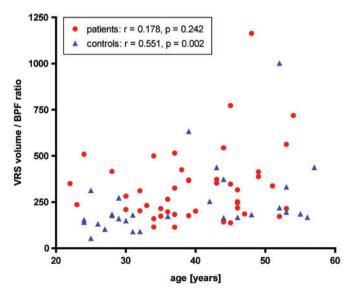


Fig. 6 In healthy individuals, the VRS volume/BPF ratio strongly correlates with age (blue triangles), while this is not the case in multiple sclerosis patients (red circles).

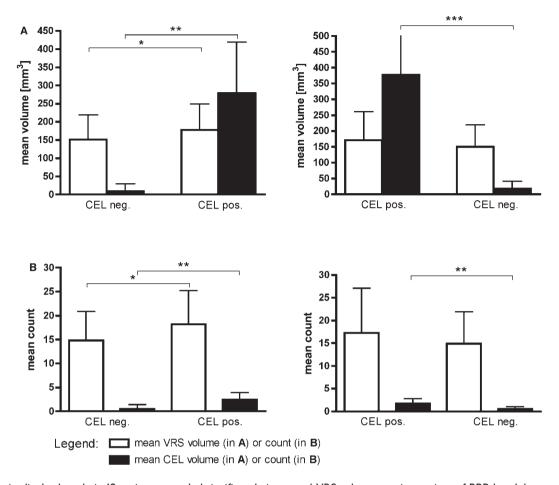


Fig. 7 A longitudinal substudy in 18 patients revealed significantly increased VRS volumes at time points of BBB breakdown with contrast enhancement compared with preceding time points without contrast enhancement ($\bf A$, left-hand image; CEL negative \rightarrow CEL positive, *P=0.011). In parallel with a shift from a CEL-positive time point to a CEL negative time point, VRS volumes did not decrease significantly ($\bf A$, right-hand image; CEL positive \rightarrow CEL negative, P=0.085). VRS counts also significantly increased at time points of BBB breakdown with contrast enhancement compared with preceding time points without contrast enhancement ($\bf B$, left-hand image; CEL negative \rightarrow CEL positive, *P=0.041). In parallel with a shift from a CEL-positive to a CEL-negative time point, VRS numbers did not decrease significantly ($\bf B$, right-hand image; CEL positive \rightarrow CEL negative, P=0.111). Mean values and SD are presented.

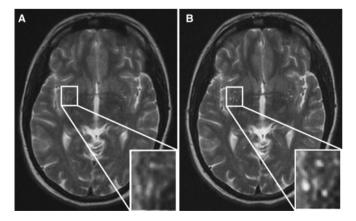


Fig. 8 T₂-weighted images of the same slice position from one study participant at two different time points are shown and magnified (inserts) ($\bf A$: 4 weeks prior to BBB breakdown; $\bf B$: in the presence of CEL) to show the alteration in VRS size and number. A semi-automatic threshold-based software was used to quantify the differences.

systematically distinguished from inflammatory lesions. We showed that (i) in contradiction to previous proposals, the presence of VRS as such is not specific or pathognomonic for multiple sclerosis, but is a physiological phenomenon also detectable in healthy individuals; (ii) VRS are significantly larger in multiple sclerosis patients compared with healthy controls and (iii) VRS volumes and numbers increase when CEL indicating BBB disruption appear on MRI. Despite growing interest in the presumed role of VRS in immune surveillance (Bechmann et al., 2001b; Man et al., 2007; McCandless and Klein, 2007), in vivo imaging of VRS by MRI has been widely neglected until very recently. One study examined VRS in multiple sclerosis patients (Achiron and Faibel, 2002) and suggested these perivascular spaces as a novel neuroradiologic in vivo marker for multiple sclerosis, as a consequence of the finding that a significantly higher proportion of multiple sclerosis patients presented with subcortical VRS compared with controls. In that study, however, and in contrast to

our methods, only two subcortical slices were applied, thus omitting the basal ganglia, and VRS volumes were not compared between groups.

Noteworthy is our finding of significantly higher VRS volumes in multiple sclerosis patients compared with controls despite equal VRS numbers in both groups. One could presume that the higher VRS volumes detected in the multiple sclerosis group might be a result of internal atrophy. In accordance with the literature (Miller et al., 2002), multiple sclerosis patients in our study presented with a higher degree of brain atrophy compared with healthy individuals. However, a significant correlation between VRS volumes, BPF and the VRS volume/BPF ratio on the one hand and age on the other held true only in the control group and not in patients. Moreover, the multiple linear regression analysis revealed a significant influence of age on VRS volumes only in the control group, but not in multiple sclerosis patients. This strongly argues against a mere effect of age or age-related brain volume reduction as the major reason for an increase of VRS volumes in multiple sclerosis patients (Heier et al., 1989; Maclullich et al., 2004). In addition, enlarged VRS in multiple sclerosis patients are not explained by diseaserelated brain atrophy either, as our multiple linear regression analysis did not show a significant influence of brain atrophy on VRS volumes in our patient group. We hypothesized that inflammatory activity leading to infiltration of cells and edema may lead to an enlargement of VRS, resulting in larger VRS volumes and thus also in an increased number of VRS detectable by MRI. Indeed, in our longitudinal observation we found that VRS volumes and numbers significantly increased upon development of BBB disruption, namely contrast enhancement. As the calculated intra-class coefficients showed an excellent intra-rater reliability in our blinded VRS quantification procedure, technical and methodological artefacts are unlikely to influence these changes in VRS volumes and numbers. The fact that there is no significant decrease in VRS volumes and numbers parallel to the disappearance of contrast enhancement suggests that reduction of VRS volumes and counts may lag behind the cessation of contrast enhancement, as we have recently shown for changes in cerebral perfusion at the site of an active lesion (Wuerfel et al., 2004). In sum, despite its exploratory character and thus without making adjustments for multiple testing, our study may indicate that VRS are sites where inflammatory activity is triggered in the CNS.

This assumption is supported by recent experimental data and animal studies. An interaction between immune cells and those of the nervous system is part of a general host defence, and plays an essential role in protecting the CNS from chronic infections and pathogen-induced damage. Therefore, the immune system and the nervous system can no longer be viewed as two compartments sealed off from each other by a tight barrier impeding communication. The CNS is continuously patrolled by

lymphocytes, which—under 'healthy' conditions—do not lead to inflammation or alterations of BBB integrity (Brabb et al., 2000; Hickey, 2001; Ransohoff et al., 2003; Man et al., 2007; Smorodchenko et al., 2007). However, when lymphocytes activated in the context of a local infectious or autoimmune process re-encounter their specific antigens (Hickey and Kimura, 1988), most probably presented within the CNS by perivascular antigen-presenting cells (Hickey and Kimura, 1988; Greter et al., 2005; McMahon et al., 2005; Becher et al., 2006), these lymphocytes may initiate an inflammatory response. This response may then promote BBB disruption and the invasion of leucocytes attracted into CNS parenchyma (Hickey, 1991). These are two key manifestations of early disease stages in multiple sclerosis and its animal model EAE (Kermode et al., 1990), although the primary site of BBB leakage may topographically differ between multiple sclerosis and EAE, as meningeal contrast enhancement is rarely detected in multiple sclerosis, but not uncommon in animal models (Traboulsee and Li, 2006). The perivascular space is potentially one of the most important entry routes of soluble proteins as well as of leucocytes into the CNS (Ransohoff et al., 2003). Within the current concept, VRS directly communicate with the subarachnoid space, and are thus partly filled with CSF. Their outer boundary is formed by a single layer of invaginated pia, and the inner by the tunica adventitia. Histopathologic studies showed that, in addition to dendritic cells and perivascular microglia, T cells are physiologically located within these spaces (Bechmann et al., 2001a; Greter et al., 2005; McMahon et al., 2005), predestining VRS for a key role in immune surveillance (Engelhardt and Ransohoff, 2005). During the generation and maintenance of inflammatory processes (Greter et al., 2005; McMahon et al., 2005; Becher et al., 2006; Bechmann et al., 2007), these spaces may dilate and become more easily visible owing to cell and fluid accumulation. A recent post-mortem study in multiple sclerosis patients lent further weight to this evidence: Vos et al. (2005) demonstrated enlarged VRS containing infiltrated leucocytes associated with diffusely abnormal white matter and focal abnormalities on post-mortem MRI. Furthermore, applying MR microscopy, Gareau et al. (2002) described perivascular cell accumulations during acute inflammation in post-mortem EAE slices. To our knowledge, this is the first longitudinal study to investigate short-term fluctuations in VRS volumes and, more importantly, to correlate these to occurrence and disappearance of CEL. Our findings suggest that enlarged VRS-although not a unique feature of multiple sclerosis-may be involved in neuroinflammatory activity.

Acknowledgements

This work was supported by grants of the German Federal Ministry of Education and Research (BMBF) and of the Institute for Multiple Sclerosis Research Göttingen (IFMS) to F.Z. We wish to thank Alistair Noon for carefully editing the manuscript as an English native speaker.

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