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Endothelial-mesenchymal transition in atherosclerosis

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Abstract

Atherosclerosis is an inflammatory disease resulting in the hardening and thickening of the wall of arteries and the formation of plaques, which comprise immune cells, mesenchymal cells, lipids, and extracellular matrix. The source of mesenchymal cells in the atherosclerotic plaques has been under scrutiny for years. Current endothelial-lineage tracing studies indicate that the endothelium is a source for plaque-associated mesenchymal cells. Endothelial cells can acquire a mesenchymal phenotype through endothelial—mesenchymal transition (EndMT), wherein the expression of endothelial markers and functions is lost and the expression of mesenchymal cell marker and functions acquired. Furthermore, EndMT can result in delamination and migration of endothelial cell-derived mesenchymal cells into the underlying tissue. Here, we review the contribution of EndMT in vascular disease focusing on atherosclerosis and describe the major biochemical and biomechanical signalling pathways in EndMT during atherosclerosis progression. Furthermore, we address how the well-established systemic atherosclerosis risk factors might facilitate EndMT during atherosclerosis.

Keywords

Atherosclerosis • Endothelial function • Endothelial–mesenchymal transition (EndMT) • Transforming growth factor beta (TGF- β) • Mechanotransduction • Regional blood flow • Remodelling • Shear stress

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1. Introduction

Atherosclerosis is a leading cause of death and morbidity. Atherosclerosis is an inflammatory disease resulting in the hardening and thickening of the wall of arteries and the formation of plaques. Atherosclerotic plaques are characterized by the accumulation of immune cells, smooth muscle cells (SMC) and (myo)fibroblasts, lipids, and extracellular matrix (ECM) in the artery wall. Unstable atherosclerotic plaques can rupture resulting in thrombosis and interruption of the blood flow. Despite the systemic nature of atherosclerotic risk factors—which comprise amongst others hypertension, diabetes mellitus (hyperglycaemia), obesity, dyslipidaemia, hyperhomocystinuria, and male gender—atherosclerotic lesions develop preferentially at vessel curvatures, branching points and bifurcations, where the blood flow is disturbed. This suggests the importance of haemodynamic forces in the initiation of the disease.

Endothelial dysfunction plays a critical role in the development of atherosclerosis.^{3,4} The endothelium is the cell layer lining the luminal surface of the blood vessels and, in healthy individuals, is a major regulator

of vascular homeostasis.⁵ The main function of the endothelium is to form a barrier controlling the passage of molecules and cells from the bloodstream to the vessel wall and vice versa. Additionally, the endothelium responds to a series of chemical and biomechanical cues by secreting factors regulating vascular tone, SMC proliferation and migration, immune cell adhesion, thromboresistance, and vessel inflammation.⁶ Most atherosclerotic risk factors can activate the endothelium, resulting in the expression of chemokines and cytokines (e.g. IL1, IL6, IL8, MCP-1) and adhesion molecules (e.g. VCAM-1, ICAM-1, E-selectin) that attract and facilitate immune cell extravasation.⁵ Critical to the activation of endothelial cells (ECs) is the switch from nitric oxide (NO) signalling to reactive oxygen species (ROS) signalling. NO promotes homeostasis and maintains the vascular wall in a guiescent state by the inhibition of proinflammatory cytokine secretion, immune cell extravasation, SMC proliferation, thrombosis and by preventing vascular leakage,³ whereas ROS induces NFkB signalling, the main regulator of inflammation. This state of oxidative stress is a common underlying mechanism for EC dysfunction in response to (biochemical and biomechanical)

pathophysiological stimuli. Upon activation, the endothelium acquires a proinflammatory state and become more permeable, promoting the accumulation of leucocytes and lipids in the intima of the artery, culminating in foam cell formation and the formation of a fatty streak, hallmarks of atherosclerosis development.

EC can acquire myofibroblast-like properties through a specific form of epithelial-to-mesenchymal transition (EMT) known as endothelial-tomesenchymal transition (EndMT). During EndMT, the expression of endothelial markers such as VE-cadherin and CD31 is reduced whereas the expression of mesenchymal markers such as alpha smooth muscle actin (α SMA), N-cadherin, and calponin is acquired. 8,9 EndMT is accompanied by the loss of cell-cell contact and cell polarity, which result in a spindle-shaped morphology and the acquisition of a migratory and invasive phenotype with enhanced ECM production. 10,11 In some instances, EndMT results in delamination and migration of EC-derived mesenchymal cells into the underlying tissue. EndMT was first described in embryonic development during angiogenesis and the formation of the mesenchymal heart cushion, the precursor of the cardiac valves. 12 In adulthood, EndMT is associated with disease including fibrosis in the heart, 13 kidney, 14 dermis, 15 vascular (re-)stenosis, 16 pulmonary arterial hypertension, ¹⁷ and cancer. ¹⁸ Recently, the contribution of EndMT to atherosclerosis development was established. 19-22

In this review, we discuss the contribution of EndMT in vascular disease focusing on atherosclerosis and describe the major biochemical and biomechanical signalling pathways in EndMT during atherosclerosis progression. Furthermore, we address how the well-established atherosclerosis risk factors might facilitate EndMT during atherosclerosis.

2. Endothelial plasticity as origin of mesenchymal cells in atherosclerotic plaques

Plaque formation in atherosclerosis is associated with the accumulation of mesenchymal cells [i.e. vascular SMC, and (myo)fibroblasts] in the intima of the artery. These mesenchymal cells are critical in the progression of atherosclerosis as they secrete proinflammatory molecules and synthetize ECM proteins and metalloproteases which facilitate plaque build-up and regulate plaque stability.²³ The origins of the neointimal mesenchymal cells in the plaque have been under scrutiny for decades but have remained elusive. Under pathological conditions, SMC originating from the media²⁴ and fibroblasts from the adventitia²⁵ migrate, proliferate, and participate to the neointima thickening. Also, bone marrow-derived cells contribute to neointima formation under pathological conditions.²⁶ Recent observations in human, porcine, and mouse plaques suggest a substantial endothelial origin for neointimal mesenchymal cells, which express both endothelial (e.g. PECAM-1, Endocan, and VE-cadherin) and mesenchymal markers [e.g. αSMA, Notch3, Snail, SM22α, fibroblast-associated protein (FAP), FSP1 (S100A4), and Vimentin]. 19-22 Such cells represent the transitioning states of EndMT, where endothelial markers continue to be expressed and new mesenchymal markers have been acquired. Thus, ongoing EndMT results in a panel of phenotypically distinct populations that may exhibit different functional characteristics.

In vivo lineage tracing experiments, employing endothelial-specific Cre-lox systems such as Cdh5-CreERT2 and SCL-CreERT2 in combination with an ApoE^{-/-} or LDLR^{-/-} mouse model of atherosclerosis have been instrumental in understanding the link between EndMT and atherosclerosis.^{21,22} In these mice, EC and their derivatives are irreversibly

labelled with GFP or YFP upon induction with tamoxifen while a high-fat diet induces the formation of atherosclerotic lesions. After 4 months of high-fat diet, approximately 30% of luminal aortic EC are undergoing EndMT as assessed by co-expression of the lineage tracer and the mesenchymal markers Notch3²¹ or FAP²² (*Figure 1*). Half of the endothelial-derived mesenchymal cells completely lost the expression of VE-cadherin, indicating that these cells have undergone a complete EndMT,²² yet the proportion of cells that undergo EndMT and delaminate into the plaque still needs to be defined.

Interestingly, in healthy animals on chow diet, approximately 20% of endothelial-derived cells express the mesenchymal marker FAP.²² These observations suggest that EndMT occurs in normal vascular homeostasis, albeit to a lesser extent than in atherosclerosis. Corroboratively, in healthy arteries, a proportion of EC express both endothelial markers and the EndMT-related transcription factors Snail, Twist1, and Gata4.^{20,27} However, the role of EndMT in the context of physiological vascular turnover warrants further investigation.

Assessing the full extent of EndMT *in vivo* is challenged by (i) the lack of specific fibroblast markers (e.g. vimentin is a marker of activated EC and fibroblasts²⁸), (ii) the potential loss of endothelial marker expression by EC at the end of the transition, and (iii) the heterogeneity of EndMT-derived cell populations.²² Moreover, (iv) the classical endothelial markers (e.g. CD31, Tie-2) are also expressed by monocytes²⁹ which can also express markers of the mesenchyme.³⁰ Caution must thus be taken when double immunofluorescence is used to analyse EndMT as the lack of cell-specific markers can underlie overinterpretation of the data. Thus, the use of a large panel of markers in combination is crucial to assess the full extent of EndMT and to better characterize the contribution of the resultant cells to plaque composition.

In summary, several independent studies have revealed the prevalence of EndMT in atherosclerosis. ^{19–22} Although the contribution of EndMT to disease initiation and progression requires further scrutiny, the extent of EndMT observed in the human plaque strongly correlates with the severity of the disease, implying clinical relevance of the EndMT process. ^{21,22} EndMT may consist of multi-step fate changes, most likely regulated in differential and sequential manners, capable of giving rise to mesenchymal cells in the plaque. Understanding this multi-step process as well as a better characterization of the resultant cells are of great interest in order to control the EC fate and be able to eradicate the emergence of potential detrimental cell populations.

3. Biochemical signals in EndMT

The discovery that EndMT is an active contributor to pathologies such as atherosclerosis and fibrosis has intensified research into the molecular mechanisms that drive EndMT. In this section, we discuss the biochemical signals that induce EndMT, modulate its progression and the currently identified endogenous inhibitors of EndMT.

3.1 Developmental pathways in EndMT: the Gata4–Twist1–Snail pathway

Gata4, Twist1 and Snail1 (Snai1) and Slug (Snai2) are key transcription factors that govern EMT during embryonic development. Notably, Snail, Slug, and Twist1 regulate EndMT during the formation of the endocardial cushions in the atrioventricular canal of the heart. Snail, Slug, and Twist repress endothelial marker gene expression, such as VE-cadherin, PECAM-1, and Claudin-5, and facilitate the expression of mesenchymal genes.

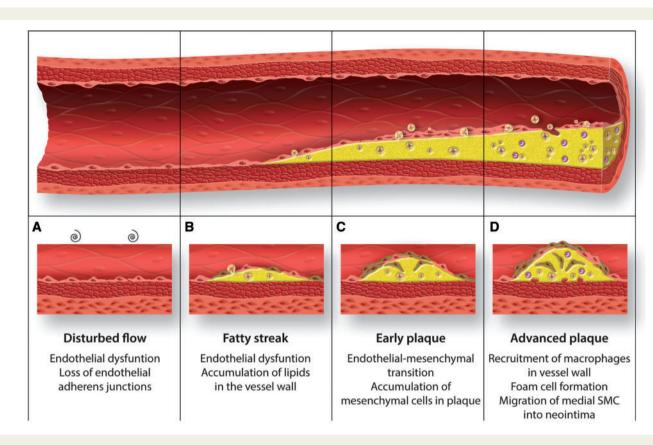


Figure I Potential contribution of EndMT to atherosclerosis progression. At areas of disturbed blood flow, ECs are activated (A), and adherens junctions dissociate resulting in lipid accumulation in the vessel wall (B). Persistent activation of ECs induces EndMT, wherein ECs start to form a multicellular layer and some ECs delaminate and start to migrate into the underlying tissue (C). Recruitment of inflammatory cells, foam cell formation, and adventitial smooth muscle cell proliferation contribute to plaque progression (D).

Snail1 is present in luminal EC overlying human coronary artery plaques, and Gata4, Twist1, and Snail1 are all expressed in luminal EC in atheroprone areas of the mouse aorta. At these sites, Gata4 regulates Twist1 expression, which in turn induces Snail1 in EC. The upstream regulators as well as the downstream targets of this pathway are still unclear, but might include TGF- β , Notch, and Wnt signalling (Figure 2). Endothelial-specific deletion of Gata4 or Twist1 reduces lesion size in atherosclerotic mice, suggesting that the Gata4–Twist1–Snail1 pathway contributes to atherosclerosis. As these TF are critical regulators of EndMT (Figure 2), understanding their regulations and downstream targets in the vascular endothelium is of great interest to understand the molecular basis controlling EndMT in atherosclerosis.

3.2 TGF-β and EndMT

Canonical TGF- β signalling is considered the driving force of EndMT. TGF- β 1 is the prototypic member of a large family of growth factors that includes the activins and bone morphogenetic proteins (BMPs). TGF- β family members mediate their effects by binding to heteromeric receptors consisting of a type I and type II receptors. Upon ligand binding, type I TGF- β receptors induce the phosphorylation of receptor-regulated (R-) SMADs. In the cytosol, activated R-SMADs form heteromeric complexes with SMAD4 and translocate to the nucleus where they regulate gene expression in cooperation with other transcription factors and transcription enhancers. The signal signal is a signal transcription enhancers.

All three TGF- β isoforms (i.e. TGF- β 1, TGF- β 2, and TGF- β 3) can induce EndMT in vitro; 9,38,39 however, TGF- $\beta2$ is essential for EndMT induction during cardiac development. Mice deficient in TGF-β2 signalling do not undergo EndMT during cardiac development, 39,40 whereas a deficiency in TGF- β 3 does not limit cardiac development.⁴¹ Conflicting data exist on the necessity of TGF-\(\beta\)1 for embryonic EndMT. Singleknockout mutant mice lacking TGF-\(\beta\)1 are born without cardiac malformations, 42,43 yet a subset of TGF-β1-/- mice develop multiple cardiac malformations, resulting in embryonic lethality. 42 In atherosclerosis, all TGF- β isoforms are expressed, but with a marked spatial and cellular variability. 44,45 In vitro, TGF-β2 appears to have a higher efficacy for the induction of EndMT;³⁸ however, data exploring the distinct and overlapping functions of the TGF-β isoforms in EndMT induction and progression during adult pathology are lacking. Thus, during atherosclerosis development, EndMT might be induced by multiple TGF- β isoforms and further detailed molecular and quantitative studies are necessary to elucidate this conundrum.

In EC, activin-like kinases (ALK) 1 and ALK5 are the predominant type I TGF- β receptors. ⁴⁶ The binding of TGF- β to ALK1 activates, i.e. phosphorylates, the R-SMADs SMAD1 and SMAD5, which promote vascular homeostasis, EC proliferation, and angiogenesis. ^{47,48} In contrast, binding of TGF- β to ALK5 induces the activation of the R-SMADs SMAD2 and SMAD3, which inhibit EC proliferation and facilitate EndMT. ^{48,49} Interestingly, activation of ALK1 by TGF- β antagonizes the ALK5 activity, ^{46,48} which might explain the pleiotropic responses of the EC to

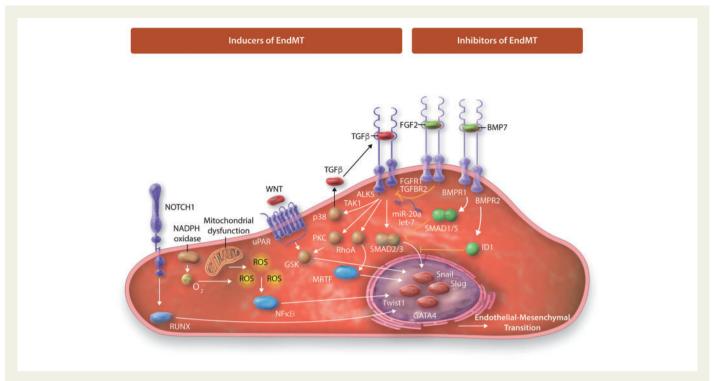


Figure 2 Biochemical signals that induce EndMT. EndMT is driven by the transcription factors Snail, Slug, and Twist1. The expression of the EndMT transcription factors can be induced by multiple upstream signalling cascades. Canonical TGF- β signalling through Smad2/3 directly induces Snail, Slug, and Twist1 expression. Non-canonical TGF- β signalling through RhoA activates the myocardin-related transcription factors (MRTF) that transcriptionally regulate Snail and Slug, whereas TGF- β signalling through TAK1 induces the expression of TGF- β , creating a positive feedback loop for EndMT. Notch signalling, Wnt signalling, and oxidative stress all facilitate in EndMT by either the induction of TGF- β expression or enhancing the nuclear accumulation of transcription factors essential for EndMT.

TGF- β stimulation and suggests a fine balance for TGF- β signalling in vascular homeostasis.

Signalling of TGF- β through ALK5 and the subsequent nuclear translocation and binding of activated SMAD2/3 to SMAD-binding elements (SBE) in the proximal promotors of mesenchymal genes induce their expression. ALK5 activity induces expression of Snail, 32,50 Slug, 49,51 and Twist1, 33 the transcriptional regulators of EndMT (*Figure 2*). Endothelial deficiency in Snail 50 or Slug 31 abrogates EndMT induction by TGF- β , yet overexpression of Snail 50 or Slug 31 alone is not sufficient to induce EndMT, which suggest cooperation between factors in EndMT induction.

Besides canonical TGF- β signalling through Smad2/3, ALK5 activity can activate several non-canonical kinases, such as (i) TGF-β-activated kinase 1 (TAK1),52 whose activity results in the activation of p38 MAPK⁵³ that in turn elevates the endothelial TGF-β expression,⁵⁴ providing a positive-feedback loop for EndMT. Concurrently, pharmacological inhibition of p38 MAPK limits EndMT induced by TGF-β;⁵⁰ (ii) protein kinase C (PKC)- δ and c-Abl induce the phosphorylation of GSK3β, resulting in the increased stability of Snail and thus EndMT.⁵⁵ Besides, PKC- δ activity promotes the activity of protein phosphatase 2a (PP2A),⁵⁶ resulting in the dephosphorylation of occludins and their subsequent loss from the tight junctions.⁵⁷ Indeed, blocking PP2A signalling with neutralizing peptides reduces EndMT;⁵⁸ (iii) RhoA,⁵⁹ which results in the expression of genes encoding cytoskeletal proteins as well as the reorganization of the cytoskeletal structure. ^{59,60} RhoA activation triggers multiple downstream cellular events, including the nuclear accumulation of the myocardin-related transcription factors, 61 which facilitate the

expression of cytoskeletal genes (e.g. transgelin, α SMA, and calponin) and the transcription factor Slug, 60 facilitating EndMT.

TGF- β signalling is more complex than solely receptor-ligand binding and subsequent biochemical signalling. First, all TGF-B isoforms are secreted as inactive precursors containing a C-terminal peptide that renders the TGF-\(\beta\)s inactive. This latency-associated peptide (LAP) needs to be enzymatically removed from TGF- $\!\beta\!$ before signalling can commence. 62 Secondly, within the ECM, multiple matrix-associated proteins such as betaglycan (also known as TGF-β Receptor 3) bind the secreted TGF- β and act as reservoir for the pericellular storage of TGFβ. 63 Thirdly, even when the LAP is removed, within the ECM, the association of TGF- β with the latent TGF- β -binding proteins (LTBP) renders TGF- β inactive, until mechanical forces disrupt the TGF- β -LTBP complex and liberate TGF- β from this inactivation.⁶⁴ Finally, endoglin, a type III TGF- β receptor, directs the actions of TGF- β in ECs by favouring ALK1 signalling and blunting ALK5-SMAD2/3 activation, 65 which suggests that high endoglin expression might inhibit EndMT initiation or progression. Future in-depth molecular investigations are necessary to elucidate the enigma of the contribution of TGF-β-binding proteins on the regulation of EndMT during atherosclerosis development and progression.

Thus, $TGF-\beta$ signalling via ALK5 induces a series of canonical and non-canonical signalling cascades that induce expression of the EndMT transcriptional programme. In coronary artery disease patients, the extend of EndMT (as revealed by Notch3 expression) and the extend of $TGF-\beta$ activation (as revealed by the presence of activated SMAD2) are strongly associated in the luminal endothelium, ²¹ which suggest that endothelial $TGF-\beta$ activity participates to plaque progression by stimulating EndMT.

However, definitive proof for the contribution of endothelial TGF- β signalling in atherosclerosis development is missing.

3.3 Alternative pathways in the induction of EndMT: Notch and Wnt/ β -catenin signalling

Although TGF- β signalling is considered the main driver of EndMT,³⁶ antagonism of TGF- β signalling in EC by, amongst others, TGF- β neutralizing antibodies,⁴⁹ BMPs,¹³ knockdown of SMAD2/3⁴⁹ or microRNA-mediated silencing of TGF- β signalling^{66,67} only partially inhibit EndMT. Likewise, SMAD3 haploinsufficiency^{13,49} or EC-specific SMAD2 deletion⁴⁹ decreases, but does not abolish, EndMT during vein graft remodelling. Therefore, it is conceivable that other signalling mechanisms might induce EndMT.

During development, Notch signalling promotes endocardial EndMT by the activation of a mesenchymal gene expression programme. ^{68,69} Notch can directly activate Twist1 expression ⁷⁰ and facilitates the recruitment of SMAD3 to SMAD-binding elements in the promoters of a variety of mesenchymal genes, ⁷¹ including Slug. ³¹ Additionally, Jagged-1-induced activation of Notch signalling results in the nuclear accumulation of the mesenchymal transcription factor RUNX3 that induces the expression of several mesenchymal genes. ^{72,73}

Wnt/ β -catenin signalling is essential for EndMT during heart valve development. The inhibition of GSK3 β by Wnt/ β -catenin results in the stabilization of Snail and thus increased EndMT; however, Wnt/ β -catenin signalling might also induce EndMT in a direct manner. Indeed, Wnt3a induces the expression of transgelin and other mesenchymal proteins in EC, which can be inhibited by the Wnt-inhibitor DKK1. Contrastingly, signalling through Wnt7a inhibits EndMT. These conflicting data indicate that further studies are necessary to conclusively establish the role of Wnt-signalling in EndMT. Currently, a role for Notch and Wnt signalling in EndMT during atherosclerosis has not been established.

In summary, EndMT can be induced by a variety of signalling mechanisms, both canonical and non-canonical. How these mechanisms cross-communicate, or whether there is a hierarchical order between these mechanisms remains elusive. Yet, the high number of pathways resulting in EndMT suggests that EndMT plays a major role in normal physiology, which might become aberrant or aggravated during disease.

3.4 Modulation of EndMT by disease pathways: hypoxia, inflammation, and oxidative stress

Atherosclerotic plaques are characterized by an increase in inflammatory signalling, hypoxia, and oxidative stress. ²² Hypoxia aggravates EndMT by facilitating the nuclear accumulation of SMAD2/3. ⁸⁰ Hypoxia-inducible factor (HIF)-1 α is the archetype sensory protein for oxygen. HIF-1 α is constitutively expressed in EC, but degraded under normoxic conditions. HIF-1 α degradation is regulated by oxygen sensors that constitutively direct HIF-1 α towards the proteasomal degradation route. During hypoxia, HIF-1 α degradation is inhibited, resulting in its nuclear accumulation. ⁸¹ Interestingly, Snail, Twist1, and ALK5 are transcriptional targets of HIF-1 α , ^{82–84} which might explain the increased sensitivity of EC to TGF- β signalling under hypoxic conditions. Indeed, pharmacological inhibition of HIF-1 α reduces ALK5 expression and abolishes EndMT. ⁸² It must be noted that the contribution of hypoxia to EndMT of luminal ECs is contradictory as atherosclerosis development is not associated with a decrease in pO2. However, the proportion of ECs that delaminate and

migrate into the underlying neointima during EndMT might encounter tissue hypoxia and this hypoxia might aggravate the EndMT process.

Inflammatory signalling synergizes with TGF- β in the induction of EndMT^{38,85} and inflammatory stress exacerbates atherosclerosis progression in mice. Proinflammatory cytokines (e.g. IL-1 β and TNF α) activate the transcription factor NF κ B, resulting in elevated expression of TGF- β 1 and TGF- β 2, ^{38,54} the main inducer of EndMT. Besides the convergence of inflammation with TGF- β signalling, proinflammatory cytokines might also induce EndMT in a TGF- β -independent manner. This inflammation-induced EndMT relies on the induction of Snail by NF κ B (Figure 2), ^{88,89} a major transcriptional regulators of EndMT. The exact pathway of inflammation-induced EndMT has yet to be elucidated and how it converges with or diverges from canonical TGF- β -induced EndMT needs to be further investigated.

TGF- β induces oxidative stress in EC through the induction of mitochondrial dysfunction, ⁵² resulting in the activation of NFkB (*Figure 2*). ⁵² As stated previously, elevated levels of endothelial oxidative stress and NFkB activity increase the expression of TGF- β 1 and TGF- β 2 and consequently EndMT. ^{38,54} Moreover, oxidative stress can activate latent TGF- β in a number of ways; (i) oxidation of the LAP, ^{90,91} and (ii) matrixmetalloproteinase (MMP)-mediated degradation of LAP. ⁹² The reduction of oxidative stress by exogenous antioxidants reduces endothelial oxidative stress and consequently EndMT. ⁹³ Interestingly, MMP-9, a TGF- β activating MMP, facilitates EndMT in kidney fibrosis, indicative of a role for oxidative stress responses in mesenchymal transition. ⁹⁴ Together, these data imply that inflammation, hypoxia, and oxidative stress in the endothelium aggravate EndMT by the induction of canonical TGF- β signalling.

3.5 Endogenous inhibitors of EndMT: fibroblast growth factor signalling and ALK5 antagonism

In contrast to the factors that induce EndMT, endogenous factors that inhibit EndMT have received only limited attention. Fibroblast growth factor (FGF) signalling might be the best characterized endogenous inhibitor of EndMT. FGF signalling in EC induces the expression of several microRNAs that decrease the expression of ALK5 (*Figure 2*). ^{66,67} Moreover, FGF signalling directly antagonizes the ALK5-induced expression of mesenchymal genes in a Ras/MEK1-dependent manner. ⁹⁵ Disruption of FGF signalling by EC-specific deletion of the FGF receptor 1 (FGFR1) ⁹⁶ or FGF Receptor Substrate 2a²¹ aggravates EndMT and the progression of atherosclerosis, ^{21,96} whereas restoration of FGF receptor signalling inhibits EndMT in diabetic nephropathy. ⁹⁷

Endothelial signalling through BMP-7 reduces EndMT (*Figure* 2), albeit through unidentified mechanisms. ^{13,98} Endogenous antagonists of ALK5 signalling, such as ALK1 agonists in EC, might block the induction of EndMT through a variety of mechanisms, namely (i) the induced expression of inhibitory Smads that prevent the phosphorylation of Smad2/3⁹⁹ and thereby EndMT, ¹⁰⁰ and (ii) the ALK1-induced expression of inhibitor of DNA binding (ID) proteins (*Figure* 2). ^{46,48} ID proteins are dominant negative helix—loop—helix proteins that lack a DNA-binding domain. ID proteins heterodimerize with other transcription factors, resulting in the formation of non-functional complexes. ¹⁰¹ Interestingly, EndMT is associated with decreased expression of ID proteins, ¹¹ and restoration of ID protein expression reverts EMT of certain epithelial tumours. ¹⁰² These data suggest a regulatory role for endogenous ALK1 ligands in the regulation of EndMT; however, data are currently unavailable.

In summary, EndMT has emerged as a pathological process that can be induced by multiple signalling cascades (i.e. TGF- β , Notch, and Wnt/ β -catenin). Moreover, the transcriptional regulation of EndMT, primarily regulated by Snail, Slug, and Twist, is facilitated by hypoxia, inflammation, and oxidative stress. Research into endogenous antagonists of the EndMT programme is in its infancy, yet FGF signalling is identified as a potent inhibitor of EndMT and the atherosclerotic process. 21,96

4. Focal risk factors and EndMT in atherosclerosis

Biomechanical forces such as shear stress and (cyclic) wall strain play an important role in vascular development, homeostasis, and the pathology of several cardiovascular conditions including atherosclerosis. 103,104 In this section, we focus on how mechanical forces can influence EndMT and contribute to atherosclerosis.

4.1 EndMT and shear stress in atherosclerosis

EC are extremely sensitive to shear stress, the frictional force exerted by the blood flow oriented tangential to the EC. EC sense shear stress through a large variety of mechanosensory complexes which convert the mechanical stimuli into biochemical signals (reviewed in Ref. 105). Disturbed blood flow, characterized by low and oscillatory shear stress (LOSS) occurring at the branching point and curvatures, induces plaque initiation, whereas high uniform shear stress (HSS) is atheroprotective. 106–108

Snail, Twist1, and Gata4 are expressed in the endothelium at sites of LOSS in healthy murine aorta 20,27 and mesenchymal markers have been detected in the luminal EC exposed to disturbed flow in the porcine abdominal aortic trifurcation. ¹⁹ Concurrently, a combination of signalling pathways favourable to EndMT is present in low shear stress area: FGFR1 expression is decreased in EC by LOSS at atheroprone regions, whereas TGF- β is activated, ²¹ suggesting that LOSS is an activating signal for EndMT. Indeed, the imposition of LOSS on carotid arteries in healthy animals triggers the expression of EndMT transcription factors in EC^{20,27} and aortic constriction-induced LOSS causes co-expression of endothelial and mesenchymal markers. ¹⁹ These data imply that LOSS induces EndMT possibly by regulating the balance between FGF and TGF- β signalling and by activating Gata4—Twist1–Snail pathway.

LOSS induces ROS production and inflammatory signalling, ^{103,109} which both favour EndMT during atherosclerosis (*Figure 3*). LOSS enhances oxidative stress in EC by the production of ROS. ^{52,110} High levels of ROS in EC cause a change in morphology and the induction of EndMT markers with the concurrent down-regulation of endothelial gene expression, ²² suggestive of EndMT induction by LOSS-derived oxidative stress.

LOSS increases vascular inflammation and enhances EC proliferation, apoptosis, and senescence, thereby increasing vascular permeability and contributing to atherosclerosis development. LOSS-induced EndMT may contribute to these alterations of EC function. Moreover, LOSS recruits monocytes to the endothelium and promotes the differentiation of M1-type foam cell which secrete CCL4, IL-1 β , and TNF α . CCL4 can induce EndMT by increasing endogenous TGF- β expression in ECs, 114 suggesting that the interplay between ECs and foam cells is of importance to EndMT induction during atherosclerosis.

EndMT has been linked to an increased vascular inflammation as TAK1, Twist1, and Gata4 promote the endothelial expression of inflammatory molecules (i.e. VCAM-1 and ICAM-1) under LOSS conditions. 27,52 In turn, inflammatory signalling promotes EndMT in an NFκB-dependent pathway, 38,87 thus establishing a positive feedback loop. In addition, Twist1, Gata4, and Snail have all been shown to promote EC proliferation and permeability under LOSS conditions. 20,27

In contrast to the areas of LOSS, EndMT does not occur in HSS regions of the vasculature, suggesting a protective effect of HSS. ^{19–21} HSS inhibits EndMT via activation of the MEK5/ERK5 pathway and KLF4 transcription factors, ^{19,115} which can inhibit Snail expression (*Figure 3*). ¹¹⁶ Interestingly, HSS-induced activity of KLF transcription factors induces the expression of the inhibitory Smad7, ¹¹⁷ thereby directly antagonizing the EndMT programme. Additionally, HSS stabilizes EC junction by controlling VE-cadherin localization at the membrane through Rac/Rho signalling (*Figure 3*). ^{118,119} Loss of adherent junctions is an initiating feature of EndMT.

In summary, it has become apparent that LOSS induces EndMT in atheroprone regions; however, the precise mechanisms linking wall shear stress and the activation of EndMT need further elucidation. On the contrary, HSS is a powerful inhibitor of EndMT via activation of protective ERK5/KLF pathway. Further work is now required to determine at which extend LOSS-induced EndMT contributes to EC dysfunction leading to focal atherosclerosis and to define the underlying mechanisms.

4.2 EndMT and cyclic strain

Cyclic strain (CS) is the blood pressure-derived force causing a repetitive deformation of the artery wall in the circumferential direction perpendicular to the EC as a result of the pulsatile blood flow. The physiological level of CS is around 10% in the human aorta, but this can vary highly during aging 120 (1.5–4%) or when the wall of the aorta becomes stiffer due to diseases such as hypertension (>10%). 121

Mechanical stretch induces the expression of SMC markers in EC. 122,123 Pathological strain induces the expression of TGF- β and BMPs in ECs and drives cardiac valve calcification via EndMT. 124,125 Depending on the strain magnitude, different signalling pathways are activated that can induce EndMT. TGF- β is activated under a relatively low strain (10%), whereas β -catenin/Wnt signalling is increased at high pathological strains (15–20%) (Figure 3). 124 Corroboratively, endothelial TGF- β expression is elevated by hypertension, 126 a condition associated with pathological CS. These data suggest that CS might contribute to the activation of TGF- β signalling during atherosclerosis.

Physiological levels of CS increase the migration and proliferation of EC while inhibiting apoptosis via Pl3-kinase-mediated activation of Akt, 127,128 whereas elevated levels of CS (>15%) induce the expression of metalloproteases (MMPs) in ECs, such as MMP-1, MMP-2, and MMP-9. 128,129 Interestingly, MMP-9 expression by ECs activates the EndMT transcriptional programme through Notch signalling, 94 suggesting that elevated levels of CS in atherosclerosis 130 might aggravate EndMT.

The small GTPases Rac and Rho regulate junctional stability of EC in a CS-dependent manner, ^{131,132} wherein junctional stability is dependent on the membranous localization of VE-cadherin. ¹³³ Physiological CS (5%) favours Rac activation, whereas pathological CS (>15%) suppresses Rac activity and enhances Rho-mediated signalling, ¹³¹ thereby causing the internalization of VE-cadherin and dissociation of endothelial adherens junctions, ¹³⁴ initiating EndMT.

Pathological CS induces mitochondrial ROS release and the subsequent activation of NF κ B signalling. ¹³⁵ The resulting state of oxidative

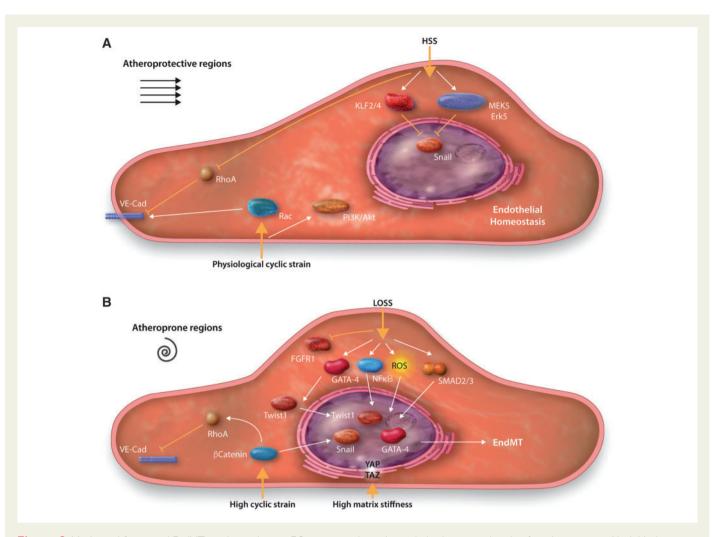


Figure 3 Mechanical forces and EndMT in atherosclerosis. ECs sense mechanical stimuli thanks to a multitude of mechanosensors (thick black arrows; reviewed by Hahn and Schwartz¹⁰⁹). (A) HSS and physiological CS inhibit EndMT. HSS induces MEK5/Erk5 pathway and KLF transcription factors, which inhibit Snail expression. Both HSS and CS control VE-cadherin localization at the cell membrane by regulating the balance between the small GTPases Rac and Rho. Although Erk5 limits Rho activation, physiological strain stimulates Rac, stabilizing VE-cadherin at the cell junction. (B) LOSS, high CS, and matrix stiffness induce EndMT. LOSS inhibits FGFR1 expression which blocks FGF signalling, a potent inhibitor of EndMT. LOSS activates TGF- β and Gata4 and Twist1. LOSS is associated with increased inflammation and oxidative stress. All these pathways culminate in the expression of Snail1 and/or Slug, which trigger EndMT. High CS increases Wnt/ β -catenin activity, which can induce and stabilize Snail1. Increased matrix stiffness promotes nuclear localization of YAP/Taz, which retains activated SMAD2/3 in the nucleus. High CS induces activation of Rho, which causes the translocation of VE-cadherin to the cytoplasm weakening cell–cell junctions, thus facilitation EndMT.

stress and inflammation facilitates EndMT by enhancing the expression of TGF- β and the activation of its signalling through ALK5. ⁵⁴

In summary, EC homeostasis is highly dependent on CS, wherein at physiological levels homeostasis is maintained and at pathological levels multiple EndMT-inducing signalling cascades are activated. Therefore, it is highly likely the interplay between EC and CS contributes to the focal development of atherosclerosis.

4.3 Matrix stiffness and EndMT

Vascular stiffening accompanies several cardiovascular conditions, such as hypertension ¹³⁶ and atherosclerosis. ¹³⁷ Arterial stiffening results from ECM remodelling within the vessel wall, characterized by degradation of elastin in the ECM and deposition and cross-linking of collagen molecules, ¹³⁸ and occurs as a result of aging, diabetes, renal disorders, and atherosclerosis.

EC sense matrix stiffness through focal adhesions. ¹³⁹ Focal adhesions are adhesion plaques, primarily formed by integrins complexes that act as an interface between the actin cytoskeleton and ECM, transmitting mechanical forces across the cell membrane. Matrix stiffness influences a variety of cell functions including proliferation, differentiation, migration, and apoptosis. ^{140–143} Vessel stiffness promotes the adaptation of an atherogenic phenotype in EC, ^{144,145} yet little is known about how arterial stiffness affects ECs within blood vessels where arteriosclerosis initiates. High matrix stiffening promotes the activation of RhoA, ¹⁴⁶ a regulator of Slug-induced EndMT⁶⁰ and nuclear translocation of Twist1. ¹⁴⁷ Matrix stiffness also regulates Yes-associated protein (YAP) and the transcriptional coactivator with PDZ-binding motif (Taz) signal-ling pathway potentially through activation of integrins. ¹⁴⁸

YAP/Taz signalling collaborates with Snail and Slug to induce the mesenchymal transcription factor Runx2. 149 Moreover, increasing matrix

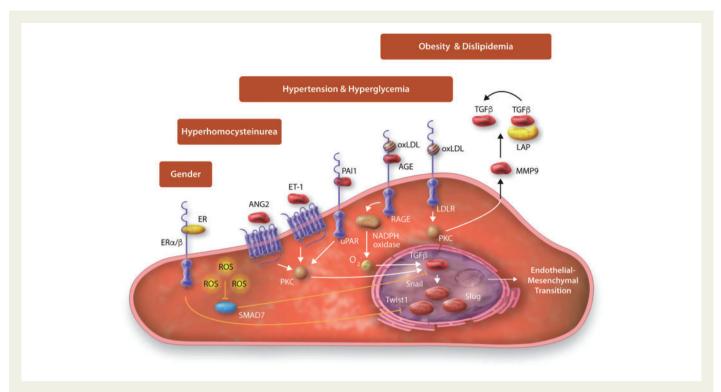


Figure 4 Systemic atherosclerosis risk factors and EndMT. Systemic risk factors for atherosclerosis might facilitate EndMT in a variety of ways. Hypertension and hyperglycaemia increase signalling through angiotensin, ET1 and PAI-1, all resulting in the activation of PKC. Likewise, obesity and dyslipidaemia are associated with increased signalling through PKC. PKC signalling culminates in the expression of TGF-β. TGF-β signalling, either canonical or non-canonical, induces the expression of the EndMT transcription factors Snail, Slug, and Twist1. Additionally, increased activity of RAGE signalling in diabetes, obesity, and dyslipidaemia activates NOX, resulting in oxidative stress-induced TGF-β expression. Hyperhomocystinuria is associated with increased endothelial oxidative stress and the concurrent reduction in SMAD7 expression, an important endogenous inhibitor of TGF-β signalling and thus EndMT. Oestrogen signalling reduces Slug expression in epithelial cells and reduces EndMT, albeit through unidentified mechanisms.

stiffness causes the nuclear translocation of YAP/Taz and promotes TGF- β signalling by the nuclear retention of SMAD2/3 150,151 in epithelial cells and fibroblasts. Corroboratively, endothelial YAP/Taz is increased in atheroprone areas, 152 which coincides with the nuclear accumulation of SMAD2 and EndMT. 21 Together, these data suggest an important role for YAP/Taz signalling in EndMT and atherosclerosis development, resulting in a vicious circle of increasing vessel stiffness and atherosclerosis progression. 153

5. Systemic risk factors and EndMT in atherosclerosis

In the sections above, we have reviewed the biochemical and biomechanical factors that mediate EndMT at atheroprone sites. In the following section, we explore the mechanisms by which systemic atherosclerosis risk factors might induce or facilitate EndMT.

5.1 Hypertension and hyperglycaemia

Hypertension and hyperglycaemia are associated with increased endothelial signalling through endothelin (ET)-1, plasminogen activator inhibitor (PAI)-1, angiotensin (Ang)-II, and NOX, which all enhance TGF- β signalling in ECs (*Figure 4*). ET-1, PAI-1, and Ang-II signalling directly induce the expression of TGF- β in ECs through PKC, ^{154–156} whereas NOX induces endothelial TGF- β expression through the induction of

endothelial oxidative stress. 157,158 In hypertension, endothelial Smad2 activity is increased, 159 indicative of active TGF- β signalling through ALK5. This increase in TGF- β activity associates with an increase in EC expressing mesenchymal marker proteins (e.g. FSP1, α SMA, and Collagen type I), indicative of EndMT. 160,161 Blocking of any of the signalling molecules upstream of TGF- β , i.e. ET-1, 162,163 PAI-1, 164 or Ang-II, 165 drastically reduces fibrogenesis, EndMT, and atherosclerosis progression.

In diabetes, the non-enzymatic reaction between glucose and serum proteins result in increased levels of advanced glycation end products (AGEs) that are biologically active and accumulate in the artery wall. Binding of AGEs to their receptor (RAGE) initiates signalling event that induce or aggravate EndMT in diabetes, 167,168 primarily through the NOX-induced activation of PKC 169 and subsequent increase in TGF- β expression. 170

5.2 Obesity, dyslipidaemia, and EndMT

Obesity and atherosclerosis are characterized by dyslipidaemia, including the increase in serum oxidized low-density lipoprotein (oxLDL) levels. Increased oxLDL signalling might induce EndMT through a variety of cascades all including oxidative stress. 171,172 OxLDL binding to its receptor LDLR induces signalling via PKC which increased endothelial MMP production in an oxidative stress-dependent manner. 173 Elevated levels of PKC signalling and MMP activity might increase endogenous TGF- β production 174 and activation, 92 and thus facilitate EndMT.

Interestingly, oxLDL can also bind and innervate alternative receptors, such as the scavenger receptor LOX-1¹⁷⁵ and RAGE. ¹⁷⁶ OxLDL binding to LOX-1 induces TGF- β expression in arterial EC *in vivo* ¹⁷⁵ and LOX-1 signalling is associated with increased collagen production in myofibroblasts, ¹⁷⁷ suggesting that oxLDL-induced LOX-1 activity might play a role in EndMT. Binding of advanced oxidation protein products, such as oxLDL, to RAGE can induce mesenchymal transition in epithelial cells in a PKC- δ -mediated manner. ¹⁷⁸ Of note, high-density lipoprotein (HDL), an established inhibitor of oxLDL signalling in EC, ¹⁷⁹ abolished EndMT induction by yet unknown mechanisms. ¹⁸⁰

5.3 Hyperhomocystinuria and EndMT

Elevation of plasma homocysteine level is a risk factor for cardiovascular disease, including atherosclerosis. Hyperhomocystinuria induces ER stress and a decrease in the production of the gasotransmitter H_2S . Interestingly, exogenous H_2S supplementation protects against EndMT by the attenuation of ER stress and the resulting TGF- β signalling, potentially through the activation of AMPK signalling, resulting in the expression of inhibitory Smad-7. 183

5.4 Oestrogen receptor signalling and EndMT

Atherosclerosis preferentially develops in male subjects, which might indicate a role for oestrogen signalling in atheroprotection 184 and the inhibition of EndMT. Indeed, in EMT, oestrogen signalling through its receptors $ER\alpha$ and $ER\beta$ maintains the epithelial phenotype by the direct repression of Slug expression and loss of oestrogen signalling induces EMT. 185 In ECs, oestrogen signalling induces endothelial adherens and tight junction formation. 186 These data suggest that oestrogen might be an inhibitor of EndMT, yet, this hypothesis remains to be investigated.

In summary, atherosclerosis risk factors can facilitate EndMT via multiple signalling cascades, all involving oxidative stress-induced expression and the activation of TGF- β signalling. Antagonizing the deranged signalling associated with atherosclerosis risk factors might inhibit EndMT and atherosclerosis progression.

6. Summary and conclusions

Atherosclerosis is a progressive pathological remodelling of the vessel wall, characterized by the accumulation of lipids and intimal mesenchymal cells, resulting in the creation of an atheromatous plaque. It is well-established that endothelial dysfunction plays a pivotal initiating role in

atherosclerosis development.¹⁸⁷ Recent studies indicate another role for the endothelium in atherosclerosis development and progression that links the atherosclerosis initiating events to the subsequent vascular remodelling, i.e. EndMT.^{19–21} EndMT is a process wherein the EC lose the expression of their characteristic marker proteins and cellular functions and acquire the expression of mesenchymal marker proteins and adopt mesenchymal cell functions, such as ECM production and contractile behaviour, thus adding to the pool of mesenchymal cells in the neointima. Here, we have reviewed the major biochemical and biomechanical signalling cascades that induce, modulate, or inhibit EndMT.

It should be emphasized that, for clarity reasons, the biochemical signalling and biomechanical signalling cascades were discussed as separate entities; however, in combination *they* form an intricate signalling network *in vivo*. ^{188,189} It becomes apparent that the native vessel architecture determines the areas at risk for atherosclerosis development through biomechanical signalling, either induced by LOSS or CS. ^{96,153} At these so-called atheroprone areas, biochemical signals originating from the systemic atherosclerosis risk factors (e.g. inflammation, hyperglycaemia, and dyslipidaemia) induce endothelial dysfunction and, if persistent, start to induce EndMT which participates in intimal hyperplasia. ^{19,21} Vascular remodelling, originating from the increase in mesenchymal cells in the intima, results in an increase in vessel stiffness and cyclic wall strain, which might aggravate EndMT creating the vicious cycle that facilitates the progressive nature of atherosclerosis.

Endothelial lineage tracing studies have established the occurrence of EndMT during atherosclerosis. 21,96 Albeit that EndMT is implicated in atherosclerosis progression in humans,²² the extent by which EndMT contributes to the initiation and progression of atherosclerosis needs further elucidation to provide a strong basis for anti-atherosclerosis therapies that specifically target EndMT. Although the endothelialspecific deletion of Erk5, an MAPK that protects against EndMT, ¹⁹ aggravates atherosclerosis development and pharmacological activation of Erk5 diminished atherosclerosis formation, ¹⁹⁰ it is currently unknown whether the modulation of EndMT or the modulation of other antiatherosclerosis genes (i.e. KLF2, KLF4, and eNOS) underlies these observations. Concurrently, pharmacological activation of FGF signalling protects against EndMT-mediated atherosclerosis progression. 21,97 Although these data are encouraging, proof-of-concept studies wherein EndMT is therapeutically targeted in advanced stages of atherosclerosis are missing.

In summary, EndMT contributes to the initiation and progression of atherosclerosis. EndMT is modulated by multiple integrative pathways,

Box 1 Evidence suggesting a link between EndMT and atherosclerosis

- Endothelial-specific lineage tracing experiments showed contribution of endothelial-derived cells to atherosclerotic lesions.
- Expression of EndMT TF (Twist1, Snail, and Gata4) and activity of the potent EndMT inducer TGF- β (revealed by expression of p-Smad2) observed in luminal EC at atheroprone sites in healthy mice.
- TGF- β activity detected in the luminal endothelium overlying the plaque in human.
- Occurrence of EndMT in atherosclerotic plaque in mouse and human as suggested by co-expression of endothelial markers with EndMT TF or mesenchymal markers.
- Correlation between EndMT extent and plaque stability/disease severity in coronary artery disease patient.
- Hypoxia, oxidative stress, and inflammatory cytokines, all present in atherosclerotic plaque, constitute a favourable environment for EndMT
- EC-specific deletion of main EndMT TF, i.e. Twist1 or Gata4 reduces the lesion size in atherosclerotic mice.
- EC-specific disruption of FGF signalling (known inhibitor of EndMT) leads to an increase of atherosclerotic development.

including TGF- β , Wnt/ β -catenin, and Notch, originating from both biochemical and biomechanical stimuli. Therapeutic strategies that reduce EndMT might have potential as anti-atherosclerosis therapies.

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References

- Caro CG, Fitz-Gerald JM, Schroter RC. Arterial wall shear and distribution of early atheroma in man. Nature 1969; 223:1159–1160.
- Zarins CK, Giddens DP, Bharadvaj BK, Sottiurai VS, Mabon RF, Glagov S. Carotid bifurcation atherosclerosis. Quantitative correlation of plaque localization with flow velocity profiles and wall shear stress. *Circ Res* 1983; 53:502–514.
- Mudau M, Genis A, Lochner A, Strijdom H. Endothelial dysfunction: the early predictor of atherosclerosis. Cardiovasc J Afr 2012; 23:222–231.
- Bonetti PO, Lerman LO, Lerman A. Endothelial dysfunction: a marker of atherosclerotic risk. Arterioscler Thromb Vasc Biol 2003; 23:168–175.
- Davignon J, Ganz P. Role of endothelial dysfunction in atherosclerosis. Circulation 2004: 109:III-27-III-32.
- Deanfield JE, Halcox JP, Rabelink TJ. Endothelial function and dysfunction: testing and clinical relevance. Circulation 2007; 115:1285–1295.
- Chang JC, Kou SJ, Lin WT, Liu CS. Regulatory role of mitochondria in oxidative stress and atherosclerosis. World J Cardiol 2010; 2:150–159.
- Frid MG, Kale V, Stenmark K. Mature vascular endothelium can give rise to smooth muscle cells via endothelial-mesenchymal transdifferentiation: in vitro analysis. *Circ* Res 2002: 90:1189–1196.
- Krenning G, Moonen JR, van Luyn MJ, Harmsen MC. Vascular smooth muscle cells for use in vascular tissue engineering obtained by endothelial-to-mesenchymal transdifferentiation (EnMT) on collagen matrices. Biomaterials 2008; 29:3703–3711.
- Kizu A, Medici D, Kalluri R. Endothelial-mesenchymal transition as a novel mechanism for generating myofibroblasts during diabetic nephropathy. Am J Pathol 2009; 175:1371–1373.
- Moonen JR, Krenning G, Brinker MG, Koerts JA, van Luyn MJ, Harmsen MC. Endothelial progenitor cells give rise to pro-angiogenic smooth muscle-like progeny. Cardiovasc Res 2010: 86:506–515.
- Nakajima Y, Yamagishi T, Hokari S, Nakamura H. Mechanisms involved in valvuloseptal endocardial cushion formation in early cardiogenesis: roles of transforming growth factor (TGF)-beta and bone morphogenetic protein (BMP). Anat Rec 2000; 258:119–127.
- Zeisberg EM, Tarnavski O, Zeisberg M, Dorfman AL, McMullen JR, Gustafsson E, Chandraker A, Yuan X, Pu WT, Roberts AB, Neilson EG, Sayegh MH, Izumo S, Kalluri R. Endothelial-to-mesenchymal transition contributes to cardiac fibrosis. Nat Med 2007; 13:952–961.
- Zeisberg EM, Potenta SE, Sugimoto H, Zeisberg M, Kalluri R. Fibroblasts in kidney fibrosis emerge via endothelial-to-mesenchymal transition. J Am Soc Nephrol 2008; 19:2282–2287.
- Manetti M, Romano E, Rosa I, Guiducci S, Bellando-Randone S, De Paulis A, Ibba-Manneschi L, Matucci-Cerinic M. Endothelial-to-mesenchymal transition contributes to endothelial dysfunction and dermal fibrosis in systemic sclerosis. Ann Rheum Dis 2017; 76:924–934.
- Beranek JT. Vascular endothelium-derived cells containing smooth muscle actin are present in restenosis. Lab Invest 1995; 72:771.
- Ranchoux B, Antigny F, Rucker-Martin C, Hautefort A, Pechoux C, Bogaard HJ, Dorfmuller P, Remy S, Lecerf F, Plante S, Chat S, Fadel E, Houssaini A, Anegon I, Adnot S, Simonneau G, Humbert M, Cohen-Kaminsky S, Perros F. Endothelial-to-mesenchymal transition in pulmonary hypertension. *Circulation* 2015; 131: 1006–1018.
- Zeisberg EM, Potenta S, Xie L, Zeisberg M, Kalluri R. Discovery of endothelial to mesenchymal transition as a source for carcinoma-associated fibroblasts. Cancer Res 2007; 67:10123–10128.
- Moonen JA, Lee ES, Schmidt M, Maleszewska M, Koerts JA, Brouwer LA, Van Kooten TG, Van Luyn MJ, Zeebregts CJ, Krenning G, Harmsen MC. Endothelial-tomesenchymal transition contributes to fibro-proliferative vascular disease and is modulated by fluid shear stress. *Cardiovasc Res* 2015; 108:377–386.
- Mahmoud MM, Serbanovic-Canic J, Feng S, Souilhol C, Xing R, Hsiao S, Mammoto A, Chen J, Ariaans M, Francis SE, Van der Heiden K, Ridger V, Evans PC. Shear stress induces endothelial-to-mesenchymal transition via the transcription factor Snail. Sci Rep 2017; 7:3375.

 Chen PY, Qin L, Baeyens N, Li G, Afolabi T, Budatha M, Tellides G, Schwartz MA, Simons M. Endothelial-to-mesenchymal transition drives atherosclerosis progression. J Clin Invest 2015; 125:4514

4528.

- 22. Evrard SM, Lecce L, Michelis KC, Nomura-Kitabayashi A, Pandey G, Purushothaman KR, D'Escamard V, Li JR, Hadri L, Fujitani K, Moreno PR, Benard L, Rimmele P, Cohain A, Mecham B, Randolph GJ, Nabel EG, Hajjar R, Fuster V, Boehm M, Kovacic JC. Endothelial to mesenchymal transition is common in atherosclerotic lesions and is associated with plaque instability. Nat Commun 2016; 7:11853.
- Ross R. Atherosclerosis—an inflammatory disease. N Engl J Med 1999; 340: 115–126.
- Hu YH, Mayr M, Metzler B, Erdel M, Davison F, Xu QB. Both donor and recipient origins of smooth muscle cells in vein graft atherosclerotic lesions. *Circ Res* 2002; 91:E13–E20.
- Shi Y, O'Brien JE, Fard A, Mannion JD, Wang D, Zalewski A. Adventitial myofibroblasts contribute to neointimal formation in injured porcine coronary arteries. Circulation 1996; 94:1655–1664.
- Sata M, Saiura A, Kunisato A, Tojo A, Okada S, Tokuhisa T, Hirai H, Makuuchi M, Hirata Y, Nagai R. Hematopoietic stem cells differentiate into vascular cells that participate in the pathogenesis of atherosclerosis. *Nat Med* 2002; 8:403

 –409.
- 27. Mahmoud MM, Kim HR, Xing R, Hsiao S, Mammoto A, Chen J, Serbanovic-Canic J, Feng S, Bowden NP, Maguire R, Ariaans M, Francis SE, Weinberg PD, van der Heiden K, Jones EA, Chico TJ, Ridger V, Evans PC. TWIST1 integrates endothelial responses to flow in vascular dysfunction and atherosclerosis. *Circ Res* 2016; 119: 450–462.
- Haseloff RF, Krause E, Bigl M, Mikoteit K, Stanimirovic D, Blasig IE. Differential protein expression in brain capillary endothelial cells induced by hypoxia and posthypoxic reoxygenation. *Proteomics* 2006; 6:1803–1809.
- Kim S-J, Kim J-S, Papadopoulos J, Wook Kim S, Maya M, Zhang F, He J, Fan D, Langley R, Fidler IJ. Circulating monocytes expressing CD31: implications for acute and chronic angiogenesis. Am J Pathol 2009; 174:1972–1980.
- Medbury HJ, Tarran SL, Guiffre AK, Williams MM, Lam TH, Vicaretti M, Fletcher JP. Monocytes contribute to the atherosclerotic cap by transformation into fibrocytes. *Int Angiol* 2008; 27:114–123.
- Niessen K, Fu Y, Chang L, Hoodless PA, McFadden D, Karsan A. Slug is a direct Notch target required for initiation of cardiac cushion cellularization. J Cell Biol 2008; 182:315–325.
- Kokudo T, Suzuki Y, Yoshimatsu Y, Yamazaki T, Watabe T, Miyazono K. Snail is required for TGFbeta-induced endothelial-mesenchymal transition of embryonic stem cell-derived endothelial cells. J Cell Sci 2008; 121:3317–3324.
- Chakraborty S, Wirrig EE, Hinton RB, Merrill WH, Spicer DB, Yutzey KE. Twist1
 promotes heart valve cell proliferation and extracellular matrix gene expression
 during development in vivo and is expressed in human diseased aortic valves. Dev
 Biol 2010; 347:167–179.
- 34. Lopez D, Niu G, Huber P, Carter WB. Tumor-induced upregulation of Twist, Snail, and Slug represses the activity of the human VE-cadherin promoter. *Arch Biochem Biophys* 2009; **482**:77–82.
- Aisagbonhi O, Rai M, Ryzhov S, Atria N, Feoktistov I, Hatzopoulos AK. Experimental myocardial infarction triggers canonical Wnt signaling and endothelial-to-mesenchymal transition. Dis Model Mech 2011; 4:469–483.
- van Meeteren LA, ten Dijke P. Regulation of endothelial cell plasticity by TGF-beta. Cell Tissue Res 2012; 347:177–186.
- Feng XH, Derynck R. Specificity and versatility in tgf-beta signaling through Smads. Annu Rev Cell Dev Biol 2005; 21:659–693.
- Maleszewska M, Moonen JR, Huijkman N, van de Sluis B, Krenning G, Harmsen MCIL. 1beta and TGFbeta2 synergistically induce endothelial to mesenchymal transition in an NFkappaB-dependent manner. *Immunobiology* 2013; 218:443–454.
- Camenisch TD, Molin DG, Person A, Runyan RB, Gittenberger-de Groot AC, McDonald JA, Klewer SE. Temporal and distinct TGFbeta ligand requirements during mouse and avian endocardial cushion morphogenesis. Dev Biol 2002; 248: 170–181.
- Azhar M, Runyan RB, Gard C, Sanford LP, Miller ML, Andringa A, Pawlowski S, Rajan S, Doetschman T. Ligand-specific function of transforming growth factor beta in epithelial-mesenchymal transition in heart development. *Dev Dyn* 2009; 238: 431–442.
- Kaartinen V, Voncken JW, Shuler C, Warburton D, Bu D, Heisterkamp N, Groffen J. Abnormal lung development and cleft palate in mice lacking TGF-beta 3 indicates defects of epithelial-mesenchymal interaction. *Nat Genet* 1995; 11:415–421.
- Dickson MC, Martin JS, Cousins FM, Kulkarni AB, Karlsson S, Akhurst RJ. Defective haematopoiesis and vasculogenesis in transforming growth factor-beta 1 knock out mice. Development 1995: 121:1845–1854.
- Diebold RJ, Eis MJ, Yin M, Ormsby I, Boivin GP, Darrow BJ, Saffitz JE, Doetschman T. Early-onset multifocal inflammation in the transforming growth factor beta 1-null mouse is lymphocyte mediated. *Proc Natl Acad Sci U S A* 1995; 92:12215–12219.
- Bahadori L, Milder J, Gold L, Botney M. Active macrophage-associated TGF-beta co-localizes with type I procollagen gene expression in atherosclerotic human pulmonary arteries. Am J Pathol 1995; 146:1140–1149.
- Jeziorska M. Transforming growth factor-betas and CD105 expression in calcification and bone formation in human atherosclerotic lesions. Z Kardiol 2001; 90(Suppl. 3):23–26.

Goumans MJ, Valdimarsdottir G, Itoh S, Lebrin F, Larsson J, Mummery CL, Karlsson S, Ten DP. Activin receptor-like kinase (ALK)1 is an antagonistic mediator of lateral TGFá/ALK5 signaling. *Mol Cell* 2003; 12:817–828.

- 47. Oh SP, Seki T, Goss KA, Imamura T, Yi Y, Donahoe PK, Li L, Miyazono K, ten Dijke P, Kim S, Li E. Activin receptor-like kinase 1 modulates transforming growth factor-β1 signaling in the regulation of angiogenesis. Proc Natl Acad Sci U S A 2000; 97: 2626–2631.
- Goumans MJ, Valdimarsdottir G, Itoh S, Rosendahl A, Sideras P, ten Dijke P. Balancing the activation state of the endothelium via two distinct TGF-β type I receptors. EMBO I 2002;21:1743–1753.
- Cooley BC, Nevado J, Mellad J, Yang D, St Hilaire C, Negro A, Fang F, Chen G, San H, Walts AD, Schwartzbeck RL, Taylor B, Lanzer JD, Wragg A, Elagha A, Beltran LE, Berry C, Feil R, Virmani R, Ladich E, Kovacic JC, Boehm M. TGF-beta signaling mediates endothelial-to-mesenchymal transition (EndMT) during vein graft remodeling. Sci Transl Med 2014: 6:227ra234.
- Medici D, Potenta S, Kalluri R. Transforming growth factor-beta2 promotes Snailmediated endothelial-mesenchymal transition through convergence of Smaddependent and Smad-independent signalling. Biochem J 2011; 437:515–520.
- 51. Romano LA, Runyan RB. Slug is an essential target of TGFbeta2 signaling in the developing chicken heart. Dev Biol 2000: **223**:91–102.
- 52. Lee ES, Boldo LS, Fernandez BO, Feelisch M, Harmsen MC. Suppression of TAK1 pathway by shear stress counteracts the inflammatory endothelial cell phenotype induced by oxidative stress and TGF-beta1. *Sci Rep* 2017; **7**:42487.
- 53. Ninomiya-Tsuji J, Kishimoto K, Hiyama A, Inoue J-I, Cao Z, Matsumoto K. The kinase TAK1 can activate the NIK-I[kappa]B as well as the MAP kinase cascade in the IL-1 signalling pathway. *Nature* 1999; **398**:252–256.
- Montorfano I, Becerra A, Cerro R, Echeverria C, Saez E, Morales MG, Fernandez R, Cabello-Verrugio C, Simon F. Oxidative stress mediates the conversion of endothelial cells into myofibroblasts via a TGF-[beta]1 and TGF-[beta]2-dependent pathway. Lab Invest 2014: 94:1068–1082.
- Li Z, Jimenez SA. Protein kinase Cdelta and c-Abl kinase are required for transforming growth factor beta induction of endothelial-mesenchymal transition in vitro. *Arthritis Rheum* 2011: 63:2473–2483.
- Zhang D, Kanthasamy A, Yang Y, Anantharam V, Kanthasamy A. Protein kinase C delta negatively regulates tyrosine hydroxylase activity and dopamine synthesis by enhancing protein phosphatase-2A activity in dopaminergic neurons. J Neurosci 2007; 27:5349–5362.
- 57. Sheth P, Samak G, Shull JA, Seth A, Rao R. Protein phosphatase 2A plays a role in hydrogen peroxide-induced disruption of tight junctions in Caco-2 cell monolayers. *Biochem J* 2009; **421**:59–70.
- 58. Deng Y, Guo Y, Liu P, Zeng R, Ning Y, Pei G, Li Y, Chen M, Guo S, Li X, Han M, Xu G. Blocking protein phosphatase 2A signaling prevents endothelial-to-mesenchymal transition and renal fibrosis: a peptide-based drug therapy. Sci Rep 2016; 6:19821.
- Mihira H, Suzuki HI, Akatsu Y, Yoshimatsu Y, Igarashi T, Miyazono K, Watabe T. TGF-beta-induced mesenchymal transition of MS-1 endothelial cells requires Smaddependent cooperative activation of Rho signals and MRTF-A. J Biochem 2012; 151: 145–156.
- Bhowmick NA, Ghiassi M, Bakin A, Aakre M, Lundquist CA, Engel ME, Arteaga CL, Moses HL. Transforming growth factor-beta1 mediates epithelial to mesenchymal transdifferentiation through a RhoA-dependent mechanism. *Mol Biol Cell* 2001; 12: 27–36.
- Fang F, Yang Y, Yuan Z, Gao Y, Zhou J, Chen Q, Xu Y. Myocardin-related transcription factor A mediates OxLDL-induced endothelial injury. *Circ Res* 2011; 108: 797–807.
- 62. Khalil N. TGF-beta: from latent to active. Microbes Infect 1999; 1:1255–1263.
- Andres JL, Stanley K, Cheifetz S, Massagué J. Membrane-anchored and soluble forms of betaglycan, a polymorphic proteoglycan that binds transforming growth factorbeta. J Cell Biol 1989; 109:3137–3145.
- 64. Shi M, Zhu J, Wang R, Chen X, Mi L, Walz T, Springer TA. Latent TGF- β structure and activation. *Nature* 2011; **474**:343–349.
- 65. Pece-Barbara N, Vera S, Kathirkamathamby K, Liebner S, Di Guglielmo GM, Dejana E, Wrana JL, Letarte M. Endoglin null endothelial cells proliferate faster and are more responsive to transforming growth factor {beta}1 with higher affinity receptors and an activated Alk1 pathway. | Biol Chem 2005; 280:27800–27808.
- Correia AC, Moonen JR, Brinker MG, Krenning G. FGF2 inhibits endothelialmesenchymal transition through microRNA-20a-mediated repression of canonical TGF-beta signaling. J Cell Sci 2016; 129:569–579.
- 67. Chen PY, Qin L, Barnes C, Charisse K, Yi T, Zhang X, Ali R, Medina PP, Yu J, Slack FJ, Anderson DG, Kotelianski V, Wang F, Tellides G, Simons M. FGF regulates TGF-beta signaling and endothelial-to-mesenchymal transition via control of let-7 miRNA expression. Cell Rep 2012; 2:1684–1696.
- 68. Luna-Zurita L, Prados B, Grego-Bessa J, Luxán G, del Monte G, Benguría A, Adams RH, Pérez-Pomares JM, de la Pompa JL. Integration of a Notch-dependent mesenchymal gene program and Bmp2-driven cell invasiveness regulates murine cardiac valve formation. J Clin Invest 2010; 120:3493–3507.
- Noseda M, McLean G, Niessen K, Chang L, Pollet I, Montpetit R, Shahidi R, Dorovini-Zis K, Li L, Beckstead B. Notch activation results in phenotypic and functional changes consistent with endothelial-to-mesenchymal transformation. Circ Res 2004; 94:910–917.

 Tian Y, Xu Y, Fu Q, Chang M, Wang Y, Shang X, Wan C, Marymont JV, Dong Y. Notch inhibits chondrogenic differentiation of mesenchymal progenitor cells by targeting Twist1. Mol Cell Endocrinol 2015; 403:30–38.

- Fu Y, Chang A, Chang L, Niessen K, Eapen S, Setiadi A, Karsan A. Differential regulation of transforming growth factor β signaling pathways by Notch in human endothelial cells. J Biol Chem 2009; 284:19452–19462.
- Fu Y, Chang AC, Fournier M, Chang L, Niessen K, Karsan A. RUNX3 maintains the mesenchymal phenotype after termination of the Notch signal. J Biol Chem 2011; 286:11803–11813.
- Reichman D, Man L, Park L, Lis R, Gerhardt J, Rosenwaks Z, James D. Notch hyperactivation drives trans-differentiation of hESC-derived endothelium. Stem Cell Res 2016: 17:391–400.
- Hurlstone AF, Haramis AP, Wienholds E, Begthel H, Korving J, Van Eeden F, Cuppen E, Zivkovic D, Plasterk RH, Clevers H. The Wnt/beta-catenin pathway regulates cardiac valve formation. *Nature* 2003: 425:633–637.
- Liebner S, Cattelino A, Gallini R, Rudini N, Iurlaro M, Piccolo S, Dejana E. Beta-catenin is required for endothelial-mesenchymal transformation during heart cushion development in the mouse. J Cell Biol 2004; 166:359–367.
- Thornton TM, Pedraza-Alva G, Deng B, Wood CD, Aronshtam A, Clements JL, Sabio G, Davis RJ, Matthews DE, Doble B, Rincon M. Phosphorylation by p38 MAPK as an alternative pathway for GSK3β inactivation. Science 2008; 320: 667–670.
- Shafer SL, Towler DA. Transcriptional regulation of SM22à by Wnt3a: convergence with TGFá1/Smad signaling at a novel regulatory element. J Mol Cell Cardiol 2009; 46:621–635.
- 78. Li L, Chen L, Zang J, Tang X, Liu Y, Zhang J, Bai L, Yin Q, Lu Y, Cheng J, Fu P, Liu F. C3a and C5a receptor antagonists ameliorate endothelial-myofibroblast transition via the Wnt/β-catenin signaling pathway in diabetic kidney disease. *Metabolism* 2015; 64:597–610.
- 79. Cheng SL, Shao JS, Behrmann A, Krchma K, Towler DA. Dkk1 and MSX2-Wnt7b signaling reciprocally regulate the endothelial-mesenchymal transition in aortic endothelial cells. *Arterioscler Thromb Vasc Biol* 2013; **33**:1679–1689.
- 80. Doerr M, Morrison J, Bergeron L, Coomber BL, Viloria-Petit A. Differential effect of hypoxia on early endothelial–mesenchymal transition response to transforming growth beta isoforms 1 and 2. *Microvasc Res* 2016; **108**:48–63.
- Semenza GL. HIF-1: mediator of physiological and pathophysiological responses to hypoxia. J Appl Physiol 2000; 88:1474–1480.
- 82. Choi SH, Hong ZY, Nam JK, Lee HJ, Jang J, Yoo RJ, Lee YJ, Lee CY, Kim KH, Park S, Ji YH, Lee YS, Cho J, Lee YJ. A hypoxia-induced vascular endothelial-to-mesenchymal transition in development of radiation-induced pulmonary fibrosis. *Clin Cancer Res* 2015; **21**:3716–3726.
- 83. Xu X, Tan X, Tampe B, Sanchez E, Zeisberg M, Zeisberg EM. Snail is a direct target of hypoxia-inducible factor 1alpha (HIF1alpha) in hypoxia-induced endothelial to mesenchymal transition of human coronary endothelial cells. J Biol Chem 2015; 290: 16653–16664.
- Yang MH, Wu MZ, Chiou SH, Chen PM, Chang SY, Liu CJ, Teng SC, Wu KJ. Direct regulation of TWIST by HIF-1alpha promotes metastasis. Nat Cell Biol 2008; 10: 295–305.
- Rieder F, Kessler SP, West GA, Bhilocha S, Motte C, Sadler TM, Gopalan B, Stylianou E, Fiocchi C. Inflammation-induced endothelial-to-mesenchymal transition: a novel mechanism of intestinal fibrosis. Am J Pathol 2011;179:2660.
- 86. Ma KL, Liu J, Ni J, Zhang Y, Lv LL, Tang RN, Ni HF, Ruan XZ, Liu BC. Inflammatory stress exacerbates the progression of cardiac fibrosis in high-fat-fed apolipoprotein E knockout mice via endothelial-mesenchymal transition. *Int J Med Sci* 2013; 10: 420–426.
- Mahler GJ, Farrar EJ, Butcher JT. Inflammatory cytokines promote mesenchymal transformation in embryonic and adult valve endothelial cells. Arterioscler Thromb Vasc Biol 2013; 33:121–130.
- 88. Wu Y, Deng J, Rychahou PG, Qiu S, Evers BM, Zhou BP. Stabilization of Snail by NF-κB is required for inflammation-induced cell migration and invasion. *Cancer Cell* 2009; **15**:416–428.
- Julien S, Puig I, Caretti E, Bonaventure J, Nelles L, van Roy F, Dargemont C, de Herreros AG, Bellacosa A, Larue L. Activation of NF-[kappa]B by Akt upregulates Snail expression and induces epithelium mesenchyme transition. *Oncogene* 2007; 26: 7445–7456.
- Pociask DA, Sime PJ, Brody AR. Asbestos-derived reactive oxygen species activate TGF-beta1. Lab Invest 2004; 84:1013–1023.
- Jobling MF, Mott JD, Finnegan MT, Jurukovski V, Erickson AC, Walian PJ, Taylor SE, Ledbetter S, Lawrence CM, Rifkin DB, Barcellos-Hoff MH. Isoform-specific activation of latent transforming growth factor beta (LTGF-beta) by reactive oxygen species. Radiat Res 2006; 166:839–848.
- Yu Q, Stamenkovic I. Cell surface-localized matrix metalloproteinase-9 proteolytically activates TGF-beta and promotes tumor invasion and angiogenesis. Genes Dev 2000; 14:163–176.
- Ying R, Wang X-Q, Yang Y, Gu Z-J, Mai J-T, Qiu Q, Chen Y-X, Wang J-F. Hydrogen sulfide suppresses endoplasmic reticulum stress-induced endothelial-to-mesenchymal transition through Src pathway. Life Sci 2016; 144:208–217.
- Zhao Y, Qiao X, Wang L, Tan TK, Zhao H, Zhang Y, Zhang J, Rao P, Cao Q, Wang Y, Wang Y, Wang YM, Lee VW, Alexander SI, Harris DC, Zheng G. Matrix

metalloproteinase 9 induces endothelial-mesenchymal transition via Notch activation in human kidney glomerular endothelial cells. *BMC Cell Biol* 2016; **17**:21.

- Ichise T, Yoshida N, Ichise H. FGF2-induced Ras-MAPK signalling maintains lymphatic endothelial cell identity by upregulating endothelial-cell-specific gene expression and suppressing TGFbeta signalling through Smad2. J Cell Sci 2014; 127: 845–857.
- 96. Chen PY, Qin L, Tellides G, Simons M. Fibroblast growth factor receptor 1 is a key inhibitor of TGFbeta signaling in the endothelium. Sci Signal 2014; 7:ra90.
- Nagai T, Kanasaki M, Srivastava SP, Nakamura Y, Ishigaki Y, Kitada M, Shi S, Kanasaki K, Koya D. N-acetyl-seryl-aspartyl-lysyl-proline inhibits diabetes-associated kidney fibrosis and endothelial-mesenchymal transition. *Biomed Res Int* 2014: 2014: 696475.
- Medici D, Kalluri R. Endothelial-mesenchymal transition and its contribution to the emergence of stem cell phenotype. Semin Cancer Biol 2012; 22:379–384.
- Hayashi H, Abdollah S, Qiu Y, Cai J, Xu YY, Grinnell BW, Richardson MA, Topper JN, Gimbrone MA Jr, Wrana JL, Falb D. The MAD-related protein Smad7 associates with the TGFbeta receptor and functions as an antagonist of TGFbeta signaling. *Cell* 1997;89:1165–1173.
- Zhang C, Zhang Y, Zhong B, Luo C-F. SMAD7 prevents heterotopic ossification in a rat Achilles tendon injury model via regulation of endothelial–mesenchymal transition. FEBS J 2016; 283:1275–1285.
- Ling F, Kang B, Sun XH. Id proteins: small molecules, mighty regulators. Curr Top Dev Biol 2014: 110:189–216.
- 102. Stankic M, Pavlovic S, Chin Y, Brogi E, Padua D, Norton L, Massagué J, Benezra R. TGF-β-Id1 signaling opposes Twist1 and promotes metastatic colonization via a mesenchymal-to-epithelial transition. Cell Rep 2013; 5:1228–1242.
- Krenning G, Barauna VG, Krieger JE, Harmsen MC, Moonen JR. Endothelial plasticity: shifting phenotypes through force feedback. Stem Cells Int 2016; 2016:9762959.
- 104. Kwak BR, Back M, Bochaton-Piallat ML, Caligiuri G, Daemen MJ, Davies PF, Hoefer IE, Holvoet P, Jo H, Krams R, Lehoux S, Monaco C, Steffens S, Virmani R, Weber C, Wentzel JJ, Evans PC. Biomechanical factors in atherosclerosis: mechanisms and clinical implications. Eur Heart J 2014; 35:3013–3020.
- Hahn C, Schwartz MA. Mechanotransduction in vascular physiology and atherogenesis. Nat Rev Mol Cell Biol 2009; 10:53–62.
- 106. Dai G, Kaazempur-Mofrad MR, Natarajan S, Zhang Y, Vaughn S, Blackman BR, Kamm RD, Garcia-Cardena G, Gimbrone MA Jr. Distinct endothelial phenotypes evoked by arterial waveforms derived from atherosclerosis-susceptible and -resistant regions of human vasculature. Proc Natl Acad Sci U S A 2004:101:14871–14876.
- Brown AJ, Teng Z, Evans PC, Gillard JH, Samady H, Bennett MR. Role of biomechanical forces in the natural history of coronary atherosclerosis. Nat Rev Cardiol 2016; 13:210–220.
- 108. Warboys CM, Amini N, de Luca A, Evans PC. The role of blood flow in determining the sites of atherosclerotic plaques. F1000 Med Rep 2011; 3:5.
- Bryan MT, Duckles H, Feng S, Hsiao ST, Kim HR, Serbanovic-Canic J, Evans PC.
 Mechanoresponsive networks controlling vascular inflammation. Arterioscler Thromb Vasc Biol 2014: 34:2199–2205.
- 110. De Keulenaer GW, Chappell DC, Ishizaka N, Nerem RM, Alexander RW, Griendling KK. Oscillatory and steady laminar shear stress differentially affect human endothelial redox state: role of a superoxide-producing NADH oxidase. Circ Res 1998; 82:1094–1101.
- 111. Chaudhury H, Zakkar M, Boyle J, Cuhlmann S, van der Heiden K, Luong Le A, Davis J, Platt A, Mason JC, Krams R, Haskard DO, Clark AR, Evans PC. c-Jun N-terminal kinase primes endothelial cells at atheroprone sites for apoptosis. Arterioscler Thromb Vasc Biol 2010; 30:546–553.
- 112. Foteinos G, Hu Y, Xiao Q, Metzler B, Xu Q. Rapid endothelial turnover in atherosclerosis-prone areas coincides with stem cell repair in apolipoprotein E-deficient mice. *Circulation* 2008; **117**:1856–1863.
- 113. Warboys CM, de Luca A, Amini N, Luong L, Duckles H, Hsiao S, White A, Biswas S, Khamis R, Chong CK, Cheung WM, Sherwin SJ, Bennett MR, Gil J, Mason JC, Haskard DO, Evans PC. Disturbed flow promotes endothelial senescence via a p53-dependent pathway. Arterioscler Thromb Vasc Biol 2014; 34:985–995.
- 114. Yang Y, Luo NS, Ying R, Xie Y, Chen JY, Wang XQ, Gu ZJ, Mai JT, Liu WH, Wu MX, Chen ZT, Fang YB, Zhang HF, Zuo ZY, Wang JF, Chen YX. Macrophage-derived foam cells impair endothelial barrier function by inducing endothelial-mesenchymal transition via CCL-4. Int J Mol Med 2017; 40:558–568.
- 115. Young A, Wu W, Sun W, Larman HB, Wang N, Li Y-S, Shyy JY, Chien S, Garcia-Cardena G. Flow activation of AMP-activated protein kinase in vascular endothelium leads to Krüppel-like factor 2 expression. Arterioscler Thromb Vasc Biol 2009; 29: 1902–1908
- 116. Yori JL, Seachrist DD, Johnson E, Lozada KL, Abdul-Karim FW, Chodosh LA, Schiemann WP, Keri RA. Kruppel-like factor 4 inhibits tumorigenic progression and metastasis in a mouse model of breast cancer. Neoplasia 2011; 13:601–610.
- 117. Boon RA, Fledderus JO, Volger OL, van Wanrooij EJ, Pardali E, Weesie F, Kuiper J, Pannekoek H, ten Dijke P, Horrevoets AJ. KLF2 suppresses TGF-beta signaling in endothelium through induction of Smad7 and inhibition of AP-1. Arterioscler Thromb Vasc Biol 2007; 27:532–539.
- Tzima E, del Pozo MA, Shattil SJ, Chien S, Schwartz MA. Activation of integrins in endothelial cells by fluid shear stress mediates Rho-dependent cytoskeletal alignment. EMBO J 2001; 20:4639–4647.

- 119. Yamamoto H, Ehling M, Kato K, Kanai K, van Lessen M, Frye M, Zeuschner D, Nakayama M, Vestweber D, Adams RH. Integrin beta1 controls VE-cadherin localization and blood vessel stability. Nat Commun 2015; 6:6429.
- Morrison TM, Choi G, Zarins CK, Taylor CA. Circumferential and longitudinal cyclic strain of the human thoracic aorta: age-related changes. J Vasc Surg 2009; 49: 1029–1036.
- 121. Bell V, Mitchell WA, Sigurősson S, Westenberg JJM, Gotal JD, Torjesen AA, Aspelund T, Launer LJ, de Roos A, Gudnason V, Harris TB, Mitchell GF. Longitudinal and circumferential strain of the proximal aorta. J Am Heart Assoc 2014; 3:e001536.
- Cevallos M, Riha GM, Wang X, Yang H, Yan S, Li M, Chai H, Yao Q, Chen C. Cyclic strain induces expression of specific smooth muscle cell markers in human endothelial cells. Differentiation 2006: 74:552–561.
- 123. Mai J, Hu Q, Xie Y, Su S, Qiu Q, Yuan W, Yang Y, Song E, Chen Y, Wang J. Dyssynchronous pacing triggers endothelial-mesenchymal transition through heterogeneity of mechanical stretch in a canine model. Circ J 2014; 79:201–209.
- 124. Balachandran K, Sucosky P, Jo H, Yoganathan AP. Elevated cyclic stretch induces aortic valve calcification in a bone morphogenic protein-dependent manner. Am J Pathol 2010; 177:49–57.
- 125. Hjortnaes J, Camci-Unal G, Hutcheson JD, Jung SM, Schoen FJ, Kluin J, Aikawa E, Khademhosseini A. Directing valvular interstitial cell myofibroblast-like differentiation in a hybrid hydrogel platform. Adv Healthc Mater 2015; 4:121–130.
- 126. Baker AB, Ettenson DS, Jonas M, Nugent MA, Iozzo RV, Edelman ER. Endothelial cells provide feedback control for vascular remodeling through a mechanosensitive autocrine TGF-beta signaling pathway. Circ Res 2008; 103:289–297.
- 127. Liu XM, Ensenat D, Wang H, Schafer Al, Durante W. Physiologic cyclic stretch inhibits apoptosis in vascular endothelium. *FEBS Lett* 2003; **541**:52–56.
- 128. Von Offenberg Sweeney N, Cummins PM, Cotter EJ, Fitzpatrick PA, Birney YA, Redmond EM, Cahill PA. Cyclic strain-mediated regulation of vascular endothelial cell migration and tube formation. Biochem Biophys Res Commun 2005; 329:573–582.
- 129. Balachandran K, Sucosky P, Jo H, Yoganathan AP. Elevated cyclic stretch alters matrix remodeling in aortic valve cusps: implications for degenerative aortic valve disease. *Am J Physiol Heart Circ Physiol* 2009; **296**:H756–H764.
- Thubrikar MJ, Robicsek F. Pressure-induced arterial wall stress and atherosclerosis.
 Ann Thorac Surg 1995; 59:1594–1603.
- 131. Birukov KG, Jacobson JR, Flores AA, Ye SQ, Birukova AA, Verin AD, Garcia JG. Magnitude-dependent regulation of pulmonary endothelial cell barrier function by cyclic stretch. Am J Physiol Lung Cell Mol Physiol 2003; 285:L785–L797.
- 132. Shikata Y, Rios A, Kawkitinarong K, DePaola N, Garcia JGN, Birukov KG. Differential effects of shear stress and cyclic stretch on focal adhesion remodeling, site-specific FAK phosphorylation, and small GTPases in human lung endothelial cells. Exp Cell Res 2005; 304:40–49.
- Rolfe BE, Worth NF, World CJ, Campbell JH, Campbell GR. Rho and vascular disease. Atherosclerosis 2005: 183:1–16.
- 134. Aslam M, Gündüz D, Schuler D, Li L, Sharifpanah F, Sedding D, Piper HM, Noll T. Intermedin induces loss of coronary microvascular endothelial barrier via derangement of actin cytoskeleton: role of RhoA and Rac1. Cardiovasc Res 2011; 92: 276–286.
- Ali MH, Pearlstein DP, Mathieu CE, Schumacker PT. Mitochondrial requirement for endothelial responses to cyclic strain: implications for mechanotransduction. Am J Physiol Lung Cell Mol Physiol 2004; 287:L486–L496.
- 136. Mitchell GF. Arterial stiffness and hypertension. Hypertension 2014;64:13-18.
- 137. Fernandes VR, Polak JF, Cheng S, Rosen BD, Carvalho B, Nasir K, McClelland R, Hundley G, Pearson G, O'Leary DH, Bluemke DA, Lima JA. Arterial stiffness is associated with regional ventricular systolic and diastolic dysfunction: the Multi-Ethnic Study of Atherosclerosis. Arterioscler Thromb Vasc Biol 2007; 28:194–201.
- Zieman SJ, Melenovsky V, Kass DA. Mechanisms, pathophysiology, and therapy of arterial stiffness. Arterioscler Thromb Vasc Biol 2005; 25:932–943.
- Discher DE, Janmey P, Wang YL. Tissue cells feel and respond to the stiffness of their substrate. Science 2005; 310:1139–1143.
- 140. Yeh Y-T, Hur SS, Chang J, Wang K-C, Chiu J-J, Li Y-S, Chien S, Leipzig ND. Matrix stiffness regulates endothelial cell proliferation through Septin 9. PLoS One 2012; 7: e46889.
- 141. Du J, Zu Y, Li J, Du S, Xu Y, Zhang L, Jiang L, Wang Z, Chien S, Yang C. Extracellular matrix stiffness dictates Wnt expression through integrin pathway. Sci Rep 2016; 6:20395.
- Fusco S, Panzetta V, Embrione V, Netti PA. Crosstalk between focal adhesions and material mechanical properties governs cell mechanics and functions. Acta Biomater 2015; 23:63–71.
- 143. Lo CM, Wang HB, Dembo M, Wang YL. Cell movement is guided by the rigidity of the substrate. *Biophys J* 2000; **79**:144–152.
- 144. van Bussel BC, Schouten F, Henry RM, Schalkwijk CG, de Boer MR, Ferreira I, Smulders YM, Twisk JW, Stehouwer CD. Endothelial dysfunction and low-grade inflammation are associated with greater arterial stiffness over a 6-year period. Hypertension 2011; 58:588–595.
- 145. Huynh J, Nishimura N, Rana K, Peloquin JM, Califano JP, Montague CR, King MR, Schaffer CB, Reinhart-King CA. Age-related intimal stiffening enhances endothelial permeability and leukocyte transmigration. Sci Transl Med 2011; 3:112ra122.

146. Huang X, Yang N, Fiore VF, Barker TH, Sun Y, Morris SW, Ding Q, Thannickal VJ, Zhou Y. Matrix stiffness-induced myofibroblast differentiation is mediated by intrinsic mechanotransduction. Am J Respir Cell Mol Biol 2012; 47:340–348.

- 147. Wei SC, Fattet L, Tsai JH, Guo Y, Pai VH, Majeski HE, Chen AC, Sah RL, Taylor SS, Engler AJ, Yang J. Matrix stiffness drives epithelial-mesenchymal transition and tumour metastasis through a TWIST1-G3BP2 mechanotransduction pathway. Nat Cell Biol 2015: 17:678–688.
- 148. Dupont S, Morsut L, Aragona M, Enzo E, Giulitti S, Cordenonsi M, Zanconato F, Le Digabel J, Forcato M, Bicciato S, Elvassore N, Piccolo S. Role of YAP/TAZ in mechanotransduction. *Nature* 2011; 474:179–183.
- 149. Tang Y, Feinberg T, Keller ET, Li XY, Weiss SJ. Snail/Slug binding interactions with YAP/TAZ control skeletal stem cell self-renewal and differentiation. Nat Cell Biol 2016; 18:917–929.
- 150. Szeto SG, Narimatsu M, Lu M, He X, Sidiqi AM, Tolosa MF, Chan L, De Freitas K, Bialik JF, Majumder S, Boo S, Hinz B, Dan Q, Advani A, John R, Wrana JL, Kapus A, Yuen DA. YAP/TAZ are mechanoregulators of TGF-beta-Smad signaling and renal fibrogenesis. J Am Soc Nephrol 2016; 27:3117–3128.
- 151. Piersma B, de Rond S, Werker PM, Boo S, Hinz B, van Beuge MM, Bank RA. YAP1 is a driver of myofibroblast differentiation in normal and diseased fibroblasts. *Am J Pathol* 2015; **185**:3326–3337.
- 152. Wang KC, Yeh YT, Nguyen P, Limqueco E, Lopez J, Thorossian S, Guan KL, Li YJ, Chien S. Flow-dependent YAP/TAZ activities regulate endothelial phenotypes and atherosclerosis. *Proc Natl Acad Sci U S A* 2016; **113**:11525–11530.
- 153. Chatzizisis YS, Giannoglou GD. Coronary hemodynamics and atherosclerotic wall stiffness: a vicious cycle. *Med Hypotheses* 2007; **69**:349–355.
- 154. Adiarto S, Heiden S, Vignon-Zellweger N, Nakayama K, Yagi K, Yanagisawa M, Emoto N. ET-1 from endothelial cells is required for complete angiotensin II-induced cardiac fibrosis and hypertrophy. Life Sci 2012; 91:651–657.
- 155. Ren S, Shatadal S, Shen GX. Protein kinase C-β mediates lipoprotein-induced generation of PAI-1 from vascular endothelial cells. Am J Physiol Endocrinol Metab 2000; 278:E656–E662.
- 156. Chua CC, Diglio CA, Siu BB, Chua BHL. Angiotensin II induces TGF-β1 production in rat heart endothelial cells. *Biochim Biophys Acta* 1994; **1223**:141–147.
- 157. Rojas M, Lemtalsi T, Toque H, Xu Z, Fulton D, Caldwell R, Caldwell R. NOX2-induced activation of arginase and diabetes-induced retinal endothelial cell senescence. Antioxidants 2017; 6:43.
- 158. Murdoch CE, Chaubey S, Zeng L, Yu B, Ivetic A, Walker SJ, Vanhoutte D, Heymans S, Grieve DJ, Cave AC, Brewer AC, Zhang M, Shah AM. Endothelial NADPH oxidase-2 promotes interstitial cardiac fibrosis and diastolic dysfunction through proinflammatory effects and endothelial-mesenchymal transition. J Am Coll Cardiol 2014; 63:2734–2741.
- Nakanuma Y, Sato Y, Kiktao A. Pathology and pathogenesis of portal venopathy in idiopathic portal hypertension: hints from systemic sclerosis. *Hepatol Res* 2009; 39: 1023–1031.
- 160. Kitao A, Sato Y, Sawada-Kitamura S, Harada K, Sasaki M, Morikawa H, Shiomi S, Honda M, Matsui O, Nakanuma Y. Endothelial to mesenchymal transition via transforming growth factor-beta1/Smad activation is associated with portal venous stenosis in idiopathic portal hypertension. Am J Pathol 2009;175:616.
- 161. Good RB, Gilbane AJ, Trinder SL, Denton CP, Coghlan G, Abraham DJ, Holmes AM. Endothelial to mesenchymal transition contributes to endothelial dysfunction in pulmonary arterial hypertension. Am J Pathol 2015; 185:1850–1858.
- 162. Wermuth PJ, Li Z, Mendoza FA, Jimenez SA, Trackman PC. Stimulation of transforming growth factor-β1-induced endothelial-to-mesenchymal transition and tissue fibrosis by endothelin-1 (ET-1): a novel profibrotic effect of ET-1. PLoS One 2016; 11:e0161988.
- 163. Widyantoro B, Emoto N, Nakayama K, Anggrahini DW, Adiarto S, Iwasa N, Yagi K, Miyagawa K, Rikitake Y, Suzuki T, Kisanuki YY, Yanagisawa M, Hirata K-I. Endothelial cell-derived endothelin-1 promotes cardiac fibrosis in diabetic hearts through stimulation of endothelial-to-mesenchymal transition. Circulation 2010;121:2407.
- 164. Omori K, Hattori N, Senoo T, Takayama Y, Masuda T, Nakashima T, Iwamoto H, Fujitaka K, Hamada H, Kohno N, Eickelberg O. Inhibition of plasminogen activator inhibitor-1 attenuates transforming growth factor-β-dependent epithelial mesenchymal transition and differentiation of fibroblasts to myofibroblasts. *PLoS One* 2016; 11:e0148969.
- 165. Tang R-N, Lv L-L, Zhang J-D, Dai H-Y, Li Q, Zheng M, Ni J, Ma K-L, Liu B-C. Effects of angiotensin II receptor blocker on myocardial endothelial-to-mesenchymal transition in diabetic rats. Int J Cardiol 2013; 162:92–99.
- 166. Makita Z, Radoff S, Rayfield EJ, Yang Z, Skolnik E, Delaney V, Friedman EA, Cerami A, Vlassara H. Advanced glycosylation end products in patients with diabetic nephropathy. N Engl J Med 1991; 325:836–842.
- 167. Ma J, Liu T, Dong X. Advanced glycation end products of bovine serum albumininduced endothelial-to-mesenchymal transition in cultured human and monkey endothelial cells via protein kinase B signaling cascades. Mol Vis 2010; 16: 2669–2679.
- 168. Sugimoto H, Grahovac G, Zeisberg M, Kalluri R. Renal fibrosis and glomerulosclerosis in a new mouse model of diabetic nephropathy and its regression by bone mor-

- phogenic protein-7 and advanced glycation end product inhibitors. *Diabetes* 2007; **56**:1825–1833
- 169. Thallas-Bonke V, Thorpe SR, Coughlan MT, Fukami K, Yap FYT, Sourris KC, Penfold SA, Bach LA, Cooper ME, Forbes JM. Inhibition of NADPH oxidase prevents advanced glycation end product—mediated damage in diabetic nephropathy through a protein kinase C-α-dependent pathway. Diabetes 2008; 57:460–469.
- 170. Li J, Qu X, Yao J, Caruana G, Ricardo SD, Yamamoto Y, Yamamoto H, Bertram JF. Blockade of endothelial-mesenchymal transition by a Smad3 inhibitor delays the early development of streptozotocin-induced diabetic nephropathy. *Diabetes* 2010; 59:2612–2624.
- 171. Yung L-M, Sánchez-Duffhues G, ten Dijke P, Yu PB. Bone morphogenetic protein 6 and oxidized low-density lipoprotein synergistically recruit osteogenic differentiation in endothelial cells. *Cardiovasc Res* 2015: **108**:278–287.
- 172. Kim M, Choi SH, Jin YB, Lee HJ, Ji YH, Kim J, Lee YS, Lee YJ. The effect of oxidized low-density lipoprotein (ox-LDL) on radiation-induced endothelial-to-mesenchymal transition. *Int J Radiat Biol* 2013; **89**:356–363.
- 173. Kuo M-Y, Ou H-C, Lee W-J, Kuo W-W, Hwang L-L, Song T-Y, Huang C-Y, Chiu T-H, Tsai K-L, Tsai C-S, Sheu WH-H. Ellagic acid inhibits oxidized low-density lipoprotein (OxLDL)-induced metalloproteinase (MMP) expression by modulating the protein kinase C-α/extracellular signal-regulated kinase/peroxisome proliferator-activated receptor γ/nuclear factor-κB (PKC-α/ERK/PPAR-γ/NF-κB) signaling pathway in endothelial cells. *J Agric Food Chem* 2011; **59**:5100–5108.
- 174. Koya D, Jirousek MR, Lin YW, Ishii H, Kuboki K, King GL. Characterization of protein kinase C beta isoform activation on the gene expression of transforming growth factor-beta, extracellular matrix components, and prostanoids in the glomeruli of diabetic rats. *J Clin Invest* 1997; **100**:115–126.
- 175. Chatauret N, Favreau F, Giraud S, Thierry A, Rossard L, Le Pape S, Lerman LO, Hauet T. Diet-induced increase in plasma oxidized LDL promotes early fibrosis in a renal porcine auto-transplantation model. *J Transl Med* 2014; **12**:76-76.
- 176. Guo ZJ, Niu HX, Hou FF, Zhang L, Fu N, Nagai R, Lu X, Chen BH, Shan YX, Tian JW, Nagaraj RH, Xie D, Zhang X. Advanced oxidation protein products activate vascular endothelial cells via a RAGE-mediated signaling pathway. *Antioxid Redox Signal* 2008; 10:1699–1712.
- 177. Hu C, Dandapat A, Sun L, Khan JA, Liu Y, Hermonat PL, Mehta JL. Regulation of TGFβ1-mediated collagen formation by LOX-1: studies based on forced overex-pression of TGFβ1 in wild-type and Lox-1. *J Biol Chem* 2008; **283**:10226–10231.
- 178. Xu X, Sun S, Xie F, Ma J, Tang J, He S, Bai L. Advanced oxidation protein products induce epithelial–mesenchymal transition of intestinal epithelial cells via a PKC δ-mediated, redox-dependent signaling pathway. *Antioxid Redox Signal* 2017; **27**:37–56.
- 179. Emert B, Hasin-Brumshtein Y, Springstead JR, Vakili L, Berliner JA, Lusis AJ. HDL inhibits the effects of oxidized phospholipids on endothelial cell gene expression via multiple mechanisms. J Lipid Res 2014; 55:1678–1692.
- Spillmann F, Miteva K, Pieske B, Tschöpe C, Van Linthout S. High-density lipoproteins reduce endothelial-to-mesenchymal transition. Arterioscler Thromb Vasc Biol 2015; 35:1774–1777.
- 181. Lentz SR. Mechanisms of homocysteine-induced atherothrombosis. J Thromb Haemost 2005; 3:1646–1654.
- Barr LA, Shimizu Y, Lambert JP, Nicholson CK, Calvert JW. Hydrogen sulfide attenuates high fat diet-induced cardiac dysfunction via the suppression of endoplasmic reticulum stress. Nitric Oxide 2015; 46:145–156.
- 183. Dallaglio K, Bruno A, Cantelmo AR, Esposito AI, Ruggiero L, Orecchioni S, Calleri A, Bertolini F, Pfeffer U, Noonan DM, Albini A. Paradoxic effects of metformin on endothelial cells and angiogenesis. *Carcinogenesis* 2014; 35:1055–1066.
- 184. Egan KM, Lawson JA, Fries S, Koller B, Rader DJ, Smyth EM, FitzGerald GA. COX-2-derived prostacyclin confers atheroprotection on female mice. Science 2004; 306: 1954–1957.
- 185. Mak P, Chang C, Pursell B, Mercurio AM. Estrogen receptor β sustains epithelial differentiation by regulating prolyl hydroxylase 2 transcription. *Proc Natl Acad Sci U S A* 2013; **110**:4708–4713.
- Burek M, Arias-Loza PA, Roewer N, Forster CY. Claudin-5 as a novel estrogen target in vascular endothelium. Arterioscler Thromb Vasc Biol 2010; 30:298–304.
- Münzel T, Sinning C, Post F, Warnholtz A, Schulz E. Pathophysiology, diagnosis and prognostic implications of endothelial dysfunction. Ann Med 2008; 40:180–196.
- 188. Amaya R, Pierides A, Tarbell JM, West J. The interaction between fluid wall shear stress and solid circumferential strain affects endothelial gene expression. PLoS One 2015; 10:e0129952.
- 189. Dancu MB, Berardi DE, Vanden Heuvel JP, Tarbell JM. Asynchronous shear stress and circumferential strain reduces endothelial NO synthase and cyclooxygenase-2 but induces endothelin-1 gene expression in endothelial cells. Arterioscler Thromb Vasc Biol 2004; 24:2088–2094.
- 190. Le NT, Heo KS, Takei Y, Lee H, Woo CH, Chang E, McClain C, Hurley C, Wang X, Li F, Xu H, Morrell C, Sullivan MA, Cohen MS, Serafimova IM, Taunton J, Fujiwara K, Abe J. A crucial role for p90RSK-mediated reduction of ERK5 transcriptional activity in endothelial dysfunction and atherosclerosis. *Circulation* 2013; 127: 486–499