

Review

# Endothelial aging

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## Abstract

Aging is considered to be the major risk factor for the development of atherosclerosis and, therefore, for coronary artery disease. Apart from age-associated remodeling of the vascular wall, which includes luminal enlargement, intimal and medial thickening, and increased vascular stiffness, endothelial function declines with age. This is most obvious from the attenuation of endothelium-dependent dilator responses, which is a consequence of the alteration in the expression and/or activity of the endothelial NO synthase, upregulation of the inducible NO synthase, and increased formation of reactive oxygen species. Aging is also associated with a reduction in the regenerative capacity of the endothelium and endothelial senescence, which is characterized by an increased rate of endothelial cell apoptosis. © 2005 European Society of Cardiology. Published by Elsevier B.V. All rights reserved.

**Keywords:** Nitric oxide; Vasodilatation; Atherosclerosis; Oxidative stress; Superoxide; Regeneration

## 1. Introduction

The vascular endothelium is situated at the interface between the blood and the vascular wall/tissue and is more than a protective barrier since it possesses anticoagulatory properties and generates a number of autacoids that regulate vascular tone and homeostasis. Nitric oxide (NO) is frequently described as the primary endothelium-derived autacoid [1] and anti-atherosclerotic principle [2]. Apart from its vasodilator property, NO exerts inhibitory effects on leukocyte adhesion [3], thrombocyte aggregation [4] and smooth muscle cell proliferation [5]. Moreover, although the radical NO acts as an antioxidant and terminates lipid peroxidation chain reactions, it also possesses pro-oxidant effects by the formation of peroxynitrite from its reaction with superoxide anions [6].

Early, but reversible manifestations of atherosclerosis, such as fatty streaks, are already found in utero [7], and frequently lead to full blown disease in early adulthood [8]. The slowly progressing atherosclerotic process eventually

results in clinical events such as ischemia and myocardial infarction. Although atherosclerosis is a disease which requires a certain amount of time to become apparent, vascular aging in itself is not synonymous with atherosclerosis. It is however exceedingly difficult to differentiate between the two, especially with the limited data available relating to human subjects.

## 2. Aging and endothelium-dependent vasodilator responses

Endothelial function is usually clinically assessed by determining changes in blood flow or arterial diameter in response to endothelial stimulation, which are estimates of vascular NO bio-availability. The outcome of such tests is not only dependent on endothelial autacoid release (mainly NO versus superoxide anions), but also on smooth muscle function and the extent of neo-intima formation.

Several clinical studies have shown that endothelium-dependent vasodilatation progressively declines with age. This observation has consistently been made in conduit arteries, such as coronary arteries [9], the brachial artery in vivo [10] and the basilar artery ex vivo [11], but also in

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resistance vessels [12,13], and occurs earlier in men than in women [14]. Thus, there is evidence indicating that aging per se leads to an attenuated generation and release or enhanced breakdown of endothelial autacoids. These observations in humans are supported by many studies determining endothelium-dependent vasomotor responses in animals. In particular, in rat conduit [15–17] and resistance vessels [18,19], but also in arteries from pigs [20], rabbits [21] and mice [22], endothelium-dependent vasodilator responses decrease with age, independently of structural changes of the vascular wall. Endothelium-independent relaxations to sodium nitroprusside in contrast are unaffected by aging. One consequence of the aging-associated impairment of endothelial function is an enhanced reactivity to vasoconstrictors [23]. This impairment affects three major endothelium-derived vasodilators, NO, prostacyclin and the endothelium-derived hyperpolarizing factor (EDHF) [24]. For example, in rats the contribution of  $K^+$  channels to endothelium-dependent relaxation progressively declines with age [25], and in aged spontaneously hypertensive rats, EDHF responses are usually completely absent and vasodilator responses are exclusively mediated by NO [26]. In humans, aging is also associated with a decreased urinary excretion of 6-keto-prostaglandin  $F_{1\alpha}$ , the stable metabolite of prostacyclin [27,28], and similar observations have been made in blood vessels from aged male but not female rats [29]. In contrast to the latter studies, an increased generation of prostacyclin was observed in the aorta of aged Wistar rats. This phenomenon was attributed to an age-induced endothelial expression of cyclooxygenase-2, and partially compensated for the age-associated lack of NO-mediated relaxation [30]. Accordingly, the expression of prostaglandin H synthase-1 and prostacyclin synthase was reported to increase with age in the aorta of Wistar–Kyoto rats [31]. The decrease in endothelium-dependent relaxation exhibits heterogeneity throughout the vascular system; with the large conduit vessels, such as the aorta, being most affected [32]. The reason underlying this observation might be that the contribution of the endothelial factors to the overall response varies between vessels as well as the ratio of endothelial cells to smooth muscle cells [32].

### 3. Mechanisms of aging-induced endothelial dysfunction

The mechanism underlying the aging-induced attenuation of endothelium-dependent dilatation is almost impossible to assess in humans, as it necessitates studies in isolated vessels performed on a large cohort of samples. The specimens available from human material; segments of the internal mammary artery from coronary bypass-grafting, coronary artery segments from explants during cardiac transplantation and peripheral arteries from amputation due to angiopathy, are usually heavily affected by the underlying disease. Consequently, more specific time-course studies have to rely on data obtained in animal experi-

ments, traditionally performed in rats. In the rat aorta, the endothelium-dependent relaxation is primarily an indicator of NO bio-availability, which is determined by the rate of NO production, as well as by its scavenging by superoxide anions ( $O_2^-$ ) [33]. Several studies report that endothelial NO synthase (eNOS) expression and NO production decline with age [19,32,34,35], whereas other authors have observed eNOS expression to be increased during aging [36,37]. Several studies also reported an increased vascular formation of  $O_2^-$  [19,22,38] and  $O_2^-$  is known to contribute to impaired relaxation since scavenging of radicals improves endothelium-dependent responses [19]. A concomitant increase in  $O_2^-$  and NO production leads to the generation of peroxynitrite [19,36] and the uncoupling of the eNOS [19]. Some reports also suggest that the cellular antioxidative defense system is attenuated during the aging process. For example, the plasma concentration of superoxide dismutase (SOD), but not the cellular SOD content in rats decreases with age [32]. Although the biological relevance of this observation is uncertain, altered plasma SOD activity is most likely a consequence of the attenuated NO bio-availability, as plasma SOD reflects extracellular SOD (ecSOD) activity in these animals [39] and ecSOD expression is induced by NO [40]. The increased formation of peroxynitrite during aging may also inactivate antioxidative enzymes, and this has been demonstrated for manganese SOD (MnSOD) in mitochondria [36] (Fig. 1).

The mechanism which leads to attenuated eNOS expression in aging is unknown. In vivo, the most important stimulus for the expression of eNOS is the shear stress generated by the flowing blood on the endothelial surface [41], which increases eNOS mRNA expression and stability [42], a fact which may explain why physical training improves eNOS expression in older humans [43,44] and animals [35]. Nevertheless, several other factors/stimuli can increase eNOS expression including estrogens [45], growth factors [46] and hydrogen peroxide [47]. The secretion of many growth factors and hormones declines with age, and this, in particular, has been shown for human growth hormone [48]. Therapy with growth hormone improves endothelial function in aged rats [49] and in humans [50]. The plasma concentration of several steroid hormones also decreases with aging (for review see [51,52]). Of these, estrogens and dehydroepiandrosterone (DHEA) [52] have gained most attention and DHEA has become a widely used “anti-aging” drug. Although hard scientific evidence justifying therapy has not been presented, animal experiments suggest that higher levels of DHEA may protect against the development of atherosclerosis [53–55]. Indeed, treatment with DHEA has been shown to improve endothelium-dependent flow-mediated dilator responses in middle-aged men with hypercholesterolemia [56]. In cultured human endothelial cells, DHEA stimulates NO release [57,58], and, treatment with DHEA increases aortic eNOS expression in ovariectomized Wistar rats via a pathway independent of the sex steroid receptor [59]. Whether, or not, chronic supple-

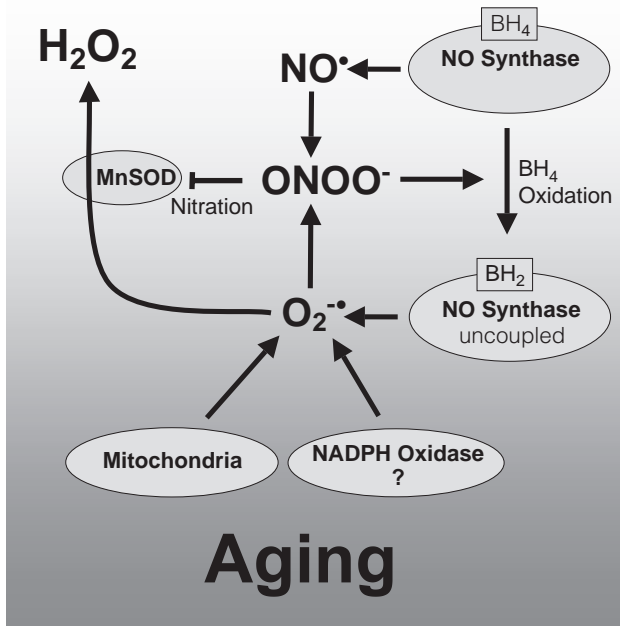


Fig. 1. Potential mechanisms of aging-induced oxidative stress in endothelial cells. Aging leads to mitochondrial dysfunction resulting from cumulative DNA damage. The shortage of enzymes of the respiratory burst promotes mitochondrial formation of superoxide ( $O_2^{\bullet -}$ ), which is usually rapidly detoxified to  $H_2O_2$  by mitochondrial manganese superoxide dismutase (MnSOD). Particularly in endothelial cells, nitric oxide (NO) is present in high concentrations and reacts with  $O_2^{\bullet -}$  to form peroxynitrite ( $ONOO^-$ ).  $ONOO^-$  can inactivate MnSOD by tyrosine nitration and can switch the NO synthase via the oxidation of tetrahydrobiopterin ( $BH_4$ ) from an NO- to an  $O_2^{\bullet -}$ -generating enzyme (NO synthase uncoupling). Both reactions lead to an increase in the concentration of  $O_2^{\bullet -}$ , in terms of a vicious circle, further promoting DNA damage and  $ONOO^-$  formation.

mentation with DHEA is able to prevent aging-induced endothelial dysfunction is still unknown.

Until very recently estrogen supplementation has been advocated as atheroprotective therapy. Each year of delay in the onset of the menopause reduces cardiovascular mortality risk by 2% [60], and epidemiological studies suggest that estrogen replacement in postmenopausal women is associated with a 50% reduction in the incidence of cardiac events [61]. It should however be mentioned that recent large clinical trials using conjugated equine estrogens, which also contain testosterone, and not only  $17\beta$ -estradiol have challenged these observations [62–65]. Estrogens acutely improve the reduced endothelium-dependent dilator response in postmenopausal women [66] and prolonged estrogen therapy has been shown in humans to restore the endothelial function to the premenopausal level [66,67]. In animal experiments, natural  $17\beta$ -estradiol increases eNOS expression and NO production, enhances the generation of prostacyclin and augments the EDHF-mediated relaxation and hyperpolarization (for review see [68,69]).

Despite an attenuated generation of endothelial NO, an excessive constitutive generation of NO by smooth muscle cells might be relevant to endothelial aging. Increased expression of the inducible NO synthase (iNOS) has been

reported in aged rats [19,37]. This enzyme generates high amounts of NO in a calcium-independent manner and is dependent mainly on its level of expression. The latter is in turn determined, at least in part by the activity of the transcription factor  $NF\kappa B$  and by inflammatory cytokines [70]. Indeed, there is evidence linking aging with increased inflammatory burden, since aging is associated with a proinflammatory shift in gene expression, e.g. endothelial expression of interleukin  $1\beta$  and interleukin 6 in aged rats [71]. Oxidative stress and inflammation are the main stimuli determining the activity of  $NF\kappa B$ . The induction of iNOS within the vascular wall would be expected to further promote oxidative stress via two mechanisms; (1) the peroxynitrite formed from the reaction of iNOS-derived NO with  $O_2^{\bullet -}$  is a much stronger oxidizing agent than the two radicals on their own [72]. (2) iNOS, because of its high turnover rate, also generates  $O_2^{\bullet -}$  following uncoupling or the depletion of its substrate L-arginine and subsequent transfer of electrons to  $O_2$  [73]. In this context it is interesting to note that the expression of arginase, which also metabolizes arginine, is increased with age in rabbits [74] and inhibition of arginase in rats improves endothelium-dependent relaxation [75].

#### 4. Endothelial aging and oxidative stress

There is little doubt that aging is associated with increased oxidative stress and oxidative damage [76,77] and the endothelium appears to be an important source of  $O_2^{\bullet -}$  in the vascular wall [38,78–80]. This effect seems to increase with age and leads to an endothelium-dependent attenuation of nitrovasodilator reactivity in aortic and mesenteric rings from rats [23]. Removal of the endothelium, as well as inhibition of the NADPH oxidase and eNO synthase reduce vascular  $O_2^{\bullet -}$  generation in the aorta of aged Wistar–Kyoto rats [38].

There is controversy regarding the enzymatic sources of radical generation. It is widely believed that leakage of  $O_2^{\bullet -}$  and particular of  $H_2O_2$  from mitochondria and mitochondrial dysfunction increase with age. Mitochondria continuously produce  $O_2^{\bullet -}$  and mitochondrial DNA is continuously exposed to oxidative stress during the life span, which in turn results in an ever increasing amount of DNA damage [76,77,81]. The consequence of this is a decrease in the number of mitochondria [82] and impaired expression of mitochondrial proteins as well as the formation of dysfunctional proteins, which leads to cellular energy depletion and further radical formation. Indeed, it has been shown that a senescence-induced lack of the mitochondrial cytochrome *c* oxidase (complex IV) leads to oxidative stress in endothelial cells [83]. In the vasculature, peroxynitrite-dependent inactivation of the mitochondrial MnSOD occurs, which further promotes mitochondrial  $O_2^{\bullet -}$  formation [36] (Fig. 1).

DNA damage, however, is not restricted to the mitochondria and one enzyme family involved in the labeling of damaged DNA—the poly(ADP-ribose) polymerases

(PARP) has been implicated in the aging-induced attenuation of endothelium-dependent relaxation [84,85]. Indeed, inhibition of PARP restored the acetylcholine-induced relaxation in the aorta of aged rats [86].

Several other enzymes are thought to be involved in aging-induced radical formation. As mentioned above, NO synthases can be transformed into radical generating enzymes [87]. In rodents, a role for xanthine oxidase has been suggested [88]. Although direct evidence is lacking, the NADPH oxidase might be a source of radical generation as activation of small GTPases [89] and potentially the NADPH oxidase subunit Nox4 [90] leads to  $O_2^-$  formation and senescence. That the NADPH oxidase may contribute to the aging phenomenon can also be derived from the observation that the stimulation of cultured cells with angiotensin II, a potent inducer and activator of the enzyme in vascular cells, leads to DNA fragmentation [91]. Indeed, treatment with angiotensin converting enzyme (ACE) inhibitors prevents the aging-induced endothelial dysfunction in rats [92]. Hydroxymethylglutaryl-CoA reductase inhibitors (statins), in contrast, had no effect on aging-induced vascular dysfunction, whereas in the rat cyclooxygenase-2 (COX-2) inhibition was just as effective as ACE inhibition [93]. The real contribution of the NADPH oxidase, however, requires further investigation, as the classic, protein kinase C-dependent isoform of this enzyme, is apparently not involved in aging-induced  $O_2^-$  formation [94].

## 5. Aging, endothelial senescence and regeneration

As with most other mammalian cells, the capacity of endothelial cells to divide is limited and ultimately the cells enter a state of irreversible growth arrest, termed senescence [51]. Senescent cells are metabolically active but morphologically altered and express senescence-associated enzymes such as the acidic  $\beta$ -galactosidase (SA- $\beta$ -gal). Under normal conditions endothelial cells rarely divide and exhibit a turnover rate estimated at approximately 3 years. Several processes, such as endothelial injury, wound healing or angiogenesis initiate endothelial proliferation [51], and consequently, the impaired wound healing and angiogenesis that is typically observed in the elderly, has been attributed to endothelial senescence [95]. Nevertheless, it is uncertain, whether or not endothelial cell senescence is an aging-associated or a vascular disease-associated phenomenon. An increased SA- $\beta$ -gal activity has been observed in the endothelial cells within atherosclerotic plaques, but not in human coronary artery explants without signs of atherosclerosis [96]. Another widely used index of senescence is telomere length. Telomeres are critical for chromosomal integrity and during each DNA duplication, approximately 50–200 bp of telomeric DNA fails to replicate. Senescence is reached when the telomeres are shortened below a critical length [97] (for review on telomeres and cardiovascular disease, see [98]). Indeed, telomere length is inversely

correlated to age in endothelial cells in vivo [99–101]. The process of telomere shortening is counteracted by the telomerase reverse transcriptase (TERT). TERT is not expressed in most human somatic cells but in germline cells and most tumor cells, preventing senescence in these lines [97]. Interestingly, cultured endothelial cells possess TERT activity, which potentially delays the onset of senescence [102,103]. Passaging during the cell culture process leads to a decrease in NO formation, which results in a loss of TERT activity. Treatment with NO-donors can however restore TERT activity and prevent the onset of senescence in cultured endothelial cells [102]. Recently, it has been reported that oxidative stress is a central regulator of endothelial nuclear TERT activity. Increased radical production and the subsequent activation of the tyrosine kinase Src leads to the export of TERT from the nucleus [104] (Fig. 2). Indeed, the aging-induced loss in nuclear TERT activity can be prevented by antioxidants and by statins, which also inhibit cellular radical generation [105].

Senescent endothelial cells are more prone to pro-apoptotic stimuli and this is largely a consequence of an

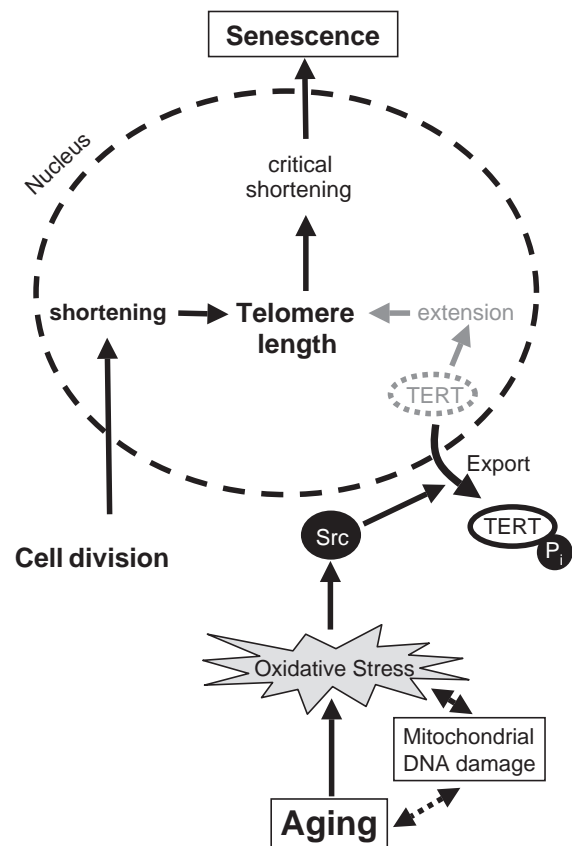


Fig. 2. Model of endothelial senescence in aging. Aging and the accumulation of mitochondrial damage lead to oxidative stress, which activates the tyrosine kinase Src. Src phosphorylates the telomerase reverse transcriptase (TERT) resulting in an export of TERT from the nucleus. Each DNA replication during cell division shortens the telomeres of the chromosomes, a process which is usually compensated by TERT. The aging-induced lack of nuclear TERT activity finally leads to cellular senescence due to telomere shortening.



attenuated NO production [106]. However, passaging of endothelial cells per se renders them susceptible to apoptosis [107] and thus it is uncertain whether the senescence of endothelial cells is a relevant phenomenon in vivo. There is, however, little doubt that endothelial cell apoptosis can occur in vivo. Various stimuli, such as inflammatory cytokines, infection, oxidized lipids and turbulent blood flow seem to promote this process [108]. Indeed, telomere length, as a marker of repetitive cell division, is reduced in endothelial cells localized at sites exposed to high mechanical stress [99–101].

The gap in the endothelial monolayer resulting from endothelial cell injury or apoptosis has to be filled, and this may occur via three different mechanisms: spreading of adjacent endothelial cells, hyperplasia of existing endothelial cells and engraftment of circulating endothelial progenitor cells. Certainly, the contribution of each of these processes to endothelial regeneration is difficult to judge. However, there is evidence that aging is associated with an attenuated capacity of the endothelium to regenerate, which is partially a consequence of an impaired secretion of and/or sensitivity to growth factors [109–111]. Recently, the regeneration of the endothelium by bone marrow-derived circulating progenitor cells has gained particular attention [112]. The number of circulating endothelial progenitor cells (EPCs) decreases with age and is thought to reflect the attenuated mobilization of these cells from the bone marrow [113]. Moreover, EPCs from older subjects have a reduced capacity to engraft. Some studies suggest that the regenerated endothelium is functionally impaired: regenerated endothelium is morphologically different, is dysfunctional [114,115] and exhibits an increased uptake of modified low-density lipoprotein (LDL) and decreased NO production [116]. Aging further impairs endothelial regeneration after injury in rats and this is associated with a decreased expression and phosphorylation of eNOS [116]. Interestingly, arginine supplementation partially restores the “juvenile” phenotype, suggesting that substrate depletion may play a role in this process [86]. The aging-induced loss of eNOS phosphorylation in the aorta from aged rats, in contrast, appears to be a consequence of an attenuated activity of the protein kinase Akt [117].

There is also evidence that angiogenesis is reduced with age. Vascular endothelial growth factor (VEGF)-induced angiogenesis in aged rats [74] and rabbits [95] is attenuated and even angiogenesis-dependent tumor growth is retarded with age [118]. Several reports have demonstrated that wound healing is delayed in aged subjects and this is to some extent attributed to an impaired angiogenic process [110,119]. Apart from the attenuated proliferative and regenerative capacity of the endothelium with age (as discussed above), changes in endothelial gene expression may also contribute to this effect. The inflammation-induced expression of intercellular adhesion molecule 1 (ICAM-1) is retarded and reduced in endothelial cells from aged subjects [119]. In aged mice and in cultured human microvascular

endothelial cells aged by progressive passaging, the expression of the tissue inhibitor of metalloproteinase-2 (TIMP-2) is increased. Enhanced TIMP-2 expression would be expected to correlate with an attenuated capacity of endothelial cells to degrade extracellular matrix, a process required for angiogenesis [120]. Moreover, in mice, the angiogenesis-associated inflammation and matrix deposition as well as the expression of VEGF and transforming growth factor  $\beta$ 1 (TGF  $\beta$ 1) are reduced with age whereas the expression of thrombospondin-2 increases [121,122]. VEGF promotes endothelial proliferation and migration, whereas TGF  $\beta$ 1 is involved in matrix formation. Thrombospondin-2, in contrast, inhibits angiogenesis [123], by reducing endothelial cell migration [123] and proliferation [124].

## 6. Aging and pro-atherosclerotic endothelial phenotype

VEGF is not the only growth factor whose expression is altered by aging. Aging increases the release of endothelin from endothelial cells in man and animals [32,37,125–127]. Endothelin is a potent vascular growth factor and endothelin receptor blockers have been shown to prevent the development of atherosclerosis in ApoE $^{-/-}$  mice [128,129]. Moreover, endothelial overexpression of endothelin in mice results in vascular remodeling and endothelial dysfunction [130]. The vascular reactivity to endothelin is also differentially affected by aging. In the rat aorta and femoral artery, aging impairs the vasoconstrictor response to this peptide [32,131], whereas in rat coronary arteries, it is increased [132,133]. Recently, endothelin has even been suggested to be involved in down-regulation of eNOS in fetal porcine pulmonary artery endothelial cells [134]. The differential expression of endothelin during aging might be a consequence of age-related differences in transcription factor activity particularly that of AP-1, NF $\kappa$ B, CREB, TFIID, CTF and AP-2 but not Sp1 [125,135]. Given the large number of transcription factors altered by aging, it is not surprising that also the expression of a plethora of growth factors and endothelial cell adhesion molecules is altered with aging. In rats, aging leads to an enhanced expression of adhesion molecules in the aorta [136] and in rabbits vascular monocyte adherence is increased with age [137]. A similar process also appears to occur in humans, where the level of serum-soluble adhesion molecules directly correlates with age [138].

## 7. Conclusion

Aging elicits several changes in the vascular endothelium gradually altering its phenotype from an anti- to a pro-atherosclerotic one. Reactive oxygen species and the concomitant oxidative and nitrosative stress may play an important role in the process of endothelial aging, affecting vascular function as well as endothelial gene expression and monolayer integrity.

Although senescence is an attractive concept to account for the attenuated angiogenic and regenerative capacity of endothelial cells with aging, future studies are needed to elucidate whether senescence, as well as alterations in the circulating level of endothelial progenitor cells are of clinical relevance.

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### References

- [1] Radziszewski W, Chopra M, Zembowicz A, Gryglewski R, Ignarro LJ, Chaudhuri G. Nitric oxide donors induce extrusion of cyclic GMP from isolated human blood platelets by a mechanism which may be modulated by prostaglandins. *Int J Cardiol* 1995;51:211–20.
- [2] Fleming I, Busse R. NO: the primary EDRF. *J Mol Cell Cardiol* 1999;31:5–14.
- [3] Kubes P, Suzuki M, Granger DN. Nitric oxide: an endogenous modulator of leukocyte adhesion. *Proc Natl Acad Sci U S A* 1991; 88:4651–5.
- [4] Mellion BT, Ignarro LJ, Ohlstein EH, Pontecorvo EG, Hyman AL, Kadowitz PJ. Evidence for the inhibitory role of guanosine 3', 5'-monophosphate in ADP-induced human platelet aggregation in the presence of nitric oxide and related vasodilators. *Blood* 1981;57: 946–55.
- [5] Garg UC, Hassid A. Nitric oxide-generating vasodilators and 8-bromo-cyclic guanosine monophosphate inhibit mitogenesis and proliferation of cultured rat vascular smooth muscle cells. *J Clin Invest* 1989;83:1774–7.
- [6] Patel RP, Levenon A, Crawford JH, Darley-Usmar VM. Mechanisms of the pro- and anti-oxidant actions of nitric oxide in atherosclerosis. *Cardiovasc Res* 2000;47:465–74.
- [7] Napoli C, D'Armiento FP, Mancini FP, Postiglione A, Witztum JL, Palumbo G, et al. Fatty streak formation occurs in human fetal aortas and is greatly enhanced by maternal hypercholesterolemia. Intimal accumulation of low density lipoprotein and its oxidation precede monocyte recruitment into early atherosclerotic lesions. *J Clin Invest* 1997;100:2680–90.
- [8] Berenson GS, Wattigney WA, Tracy RE, Newman III WP, Srinivasan SR, Webber LS, et al. Atherosclerosis of the aorta and coronary arteries and cardiovascular risk factors in persons aged 6 to 30 years and studied at necropsy (The Bogalusa Heart Study). *Am J Cardiol* 1992;70:851–8.
- [9] Egashira K, Inou T, Hirooka Y, Kai H, Sugimachi M, Suzuki S, et al. Effects of age on endothelium-dependent vasodilation of resistance coronary artery by acetylcholine in humans. *Circulation* 1993;88:77–81.
- [10] Taddei S, Virdis A, Mattei P, Ghiadoni L, Gennari A, Fasolo CB, et al. Aging and endothelial function in normotensive subjects and patients with essential hypertension. *Circulation* 1995;91:1981–7.
- [11] Hatake K, Kakishita E, Wakabayashi I, Sakiyama N, Hishida S. Effect of aging on endothelium-dependent vascular relaxation of isolated human basilar artery to thrombin and bradykinin. *Stroke* 1990; 21:1039–43.
- [12] Singh N, Prasad S, Singer DR, MacAllister RJ. Ageing is associated with impairment of nitric oxide and prostanoid dilator pathways in the human forearm. *Clin Sci (Lond)* 2002;102:595–600.
- [13] Lyons D, Roy S, Patel M, Benjamin N, Swift CG. Impaired nitric oxide-mediated vasodilatation and total body nitric oxide production in healthy old age. *Clin Sci (Lond)* 1997;93:519–25.
- [14] Celermajer DS, Sorensen KE, Spiegelhalter DJ, Georgakopoulos D, Robinson J, Deanfield JE. Aging is associated with endothelial dysfunction in healthy men years before the age-related decline in women. *J Am Coll Cardiol* 1994;24:471–6.
- [15] Hongo K, Nakagomi T, Kassell NF, Sasaki T, Lehman M, Vollmer DG, et al. Effects of aging and hypertension on endothelium-dependent vascular relaxation in rat carotid artery. *Stroke* 1988;19: 892–7.
- [16] Kung CF, Luscher TF. Different mechanisms of endothelial dysfunction with aging and hypertension in rat aorta. *Hypertension* 1995;25:194–200.
- [17] Geary GG, Buchholz JN. Selected contribution: effects of aging on cerebrovascular tone and [Ca<sup>2+</sup>]<sub>i</sub>. *J Appl Physiol* 2003;95:1746–54.
- [18] Muller-Delp JM, Spier SA, Ramsey MW, Delp MD. Aging impairs endothelium-dependent vasodilation in rat skeletal muscle arterioles. *Am J Physiol, Heart Circ Physiol* 2002;283:H1662–72.
- [19] Csiszar A, Ungvari Z, Edwards JG, Kaminski P, Wolin MS, Koller A, et al. Aging-induced phenotypic changes and oxidative stress impair coronary arteriolar function. *Circ Res* 2002;90:1159–66.
- [20] Murohara T, Yasue H, Ohgushi M, Sakaino N, Jougasaki M. Age related attenuation of the endothelium dependent relaxation to noradrenaline in isolated pig coronary arteries. *Cardiovasc Res* 1991;25:1002–9.
- [21] Chinellato A, Pandolfo L, Ragazzi E, Zamboni MR, Froldi G, De Biasi M, et al. Effect of age on rabbit aortic responses to relaxant endothelium-dependent and endothelium-independent agents. *Blood Vessels* 1991;28:358–65.
- [22] Blackwell KA, Sorenson JP, Richardson DM, Smith LA, Suda O, Nath K, et al. Mechanisms of aging-induced impairment of endothelium-dependent relaxations—role of tetrahydrobiopterin. *Am J Physiol, Heart Circ Physiol* 2004;287:2448–53.
- [23] Shirasaki Y, Su C, Lee TJ, Kolm P, Cline Jr WH, Nickols GA. Endothelial modulation of vascular relaxation to nitrovasodilators in aging and hypertension. *J Pharmacol Exp Ther* 1986;239:861–6.
- [24] Busse R, Fleming I. Regulation of endothelium-derived vasoactive autacoid production by hemodynamic forces. *Trends Pharmacol Sci* 2003;24:24–9.
- [25] Mantelli L, Amerini S, Ledda F. Roles of nitric oxide and endothelium-derived hyperpolarizing factor in vasorelaxant effect of acetylcholine as influenced by aging and hypertension. *J Cardiovasc Pharmacol* 1995;25:595–602.
- [26] Bussemaker E, Popp R, Fisslthaler B, Larson CM, Fleming I, Busse R, et al. Aged spontaneously hypertensive rats exhibit a selective loss of EDHF-mediated relaxation in the renal artery. *Hypertension* 2003;42:562–8.
- [27] Gotoh S, Ogihara T, Nakamaru M, Masuo K, Hata T, Kumahara Y. Effect of aging on 6-keto-PGF<sub>1</sub> alpha levels in normotensive and essential hypertensive males. *Jpn Circ J* 1983;47:309–12.
- [28] Hornych A, Forette F, Bariety J, Krief C, Aumont J, Paris M. The influence of age on renal prostaglandin synthesis in man. *Prostaglandins Leukot Essent Fat Acids* 1991;43:191–5.
- [29] Lennon EA, Poyser NL. Effect of age on vascular prostaglandin production in male and female rats. *Prostaglandins Leukot Med* 1986;25:1–15.
- [30] Heymes C, Habib A, Yang D, Mathieu E, Marotte F, Samuel J, et al. Cyclo-oxygenase-1 and-2 contribution to endothelial dysfunction in ageing. *Br J Pharmacol* 2000;131:804–10.
- [31] Numaguchi Y, Harada M, Osanai H, Hayashi K, Toki Y, Okumura K, et al. Altered gene expression of prostacyclin synthase and prostacyclin receptor in the thoracic aorta of spontaneously hypertensive rats. *Cardiovasc Res* 1999;41:682–8.
- [32] Barton M, Cosentino F, Brandes RP, Moreau P, Shaw S, Luscher TF. Anatomic heterogeneity of vascular aging: role of nitric oxide and endothelin. *Hypertension* 1997;30:817–24.

- [33] Cai H, Harrison DG. Endothelial dysfunction in cardiovascular diseases: the role of oxidant stress. *Circ Res* 2000;87:840–4.
- [34] Tschudi MR, Barton M, Bersinger NA, Moreau P, Cosentino F, Noll G, et al. Effect of age on kinetics of nitric oxide release in rat aorta and pulmonary artery. *J Clin Invest* 1996;98:899–905.
- [35] Tanabe T, Maeda S, Miyauchi T, Iemitsu M, Takanashi M, Irukayama-Tomobe Y, et al. Exercise training improves ageing-induced decrease in eNOS expression of the aorta. *Acta Physiol Scand* 2003;178:3–10.
- [36] van der Loo B, Labugger R, Skepper JN, Bachschmid M, Kilo J, Powell JM, et al. Enhanced peroxynitrite formation is associated with vascular aging. *J Exp Med* 2000;192:1731–44.
- [37] Goettsch W, Lattmann T, Amann K, Szibor M, Morawietz H, Munter K, et al. Increased expression of endothelin-1 and inducible nitric oxide synthase isoform II in aging arteries in vivo: implications for atherosclerosis. *Biochem Biophys Res Commun* 2001;280:908–13.
- [38] Hamilton CA, Brosnan MJ, Mcintyre M, Graham D, Dominiczak AF. Superoxide excess in hypertension and aging: a common cause of endothelial dysfunction. *Hypertension* 2001;37:529–34.
- [39] Carlsson LM, Marklund SL, Edlund T. The rat extracellular superoxide dismutase dimer is converted to a tetramer by the exchange of a single amino acid. *Proc Natl Acad Sci U S A* 1996;93:5219–22.
- [40] Fukai T, Siegfried MR, Ushio-Fukai M, Cheng Y, Kojda G, Harrison DG. Regulation of the vascular extracellular superoxide dismutase by nitric oxide and exercise training. *J Clin Invest* 2000;105:1631–9.
- [41] Ranjan V, Xiao Z, Diamond SL. Constitutive NOS expression in cultured endothelial cells is elevated by fluid shear stress. *Am J Physiol* 1995;269:H550–5.
- [42] Davis ME, Cai H, Drummond GR, Harrison DG. Shear stress regulates endothelial nitric oxide synthase expression through c-Src by divergent signaling pathways. *Circ Res* 2001;89:1073–80.
- [43] Taddei S, Galetta F, Virdis A, Ghiadoni L, Salvetti G, Franzoni F, et al. Physical activity prevents age-related impairment in nitric oxide availability in elderly athletes. *Circulation* 2000;101:2896–901.
- [44] Spier SA, Delp MD, Meininger CJ, Donato AJ, Ramsey MW, Muller-Delp JM. Effects of ageing and exercise training on endothelium-dependent vasodilatation and structure of rat skeletal muscle arterioles. *J Physiol* 2004;556:947–58.
- [45] Kleinert H, Wallerath T, Euchenhofer C, Ihrig-Biedert I, Li H, Forstermann U. Estrogens increase transcription of the human endothelial NO synthase gene: analysis of the transcription factors involved. *Hypertension* 1998;31:582–8.
- [46] Bouloumie A, Schini-Kerth VB, Busse R. Vascular endothelial growth factor up-regulates nitric oxide synthase expression in endothelial cells. *Cardiovasc Res* 1999;41:773–80.
- [47] Drummond GR, Cai H, Davis ME, Ramasamy S, Harrison DG. Transcriptional and posttranscriptional regulation of endothelial nitric oxide synthase expression by hydrogen peroxide. *Circ Res* 2000;86:347–54.
- [48] Rosen CJ. Growth hormone and aging. *Endocrine* 2000;12:197–201.
- [49] Ariznavarreta C, Castillo C, Segovia G, Mora F, Azcoitia I, Tresguerres JA. Growth hormone and aging. *Homo* 2003;54:132–41.
- [50] Boger RH, Skamira C, Bode-Boger SM, Brabant G, von zur MA, Frolich JC. Nitric oxide may mediate the hemodynamic effects of recombinant growth hormone in patients with acquired growth hormone deficiency. A double-blind, placebo-controlled study. *J Clin Invest* 1996;98:2706–13.
- [51] Foreman KE, Tang J. Molecular mechanisms of replicative senescence in endothelial cells. *Exp Gerontol* 2003;38:1251–7.
- [52] Lamberts SW, van den Beld AW, van der Lely AJ. The endocrinology of aging. *Science* 1997;278:419–24.
- [53] Arad Y, Badimon JJ, Badimon L, Hembree WC, Ginsberg HN. Dehydroepiandrosterone feeding prevents aortic fatty streak formation and cholesterol accumulation in cholesterol-fed rabbit. *Arteriosclerosis* 1989;9:159–66.
- [54] Gordon GB, Bush DE, Weisman HF. Reduction of atherosclerosis by administration of dehydroepiandrosterone. A study in the hypercholesterolemic New Zealand white rabbit with aortic intimal injury. *J Clin Invest* 1988;82:712–20.
- [55] Hayashi T, Esaki T, Muto E, Kano H, Asai Y, Thakur NK, et al. Dehydroepiandrosterone retards atherosclerosis formation through its conversion to estrogen: the possible role of nitric oxide. *Arterioscler Thromb Vasc Biol* 2000;20:782–92.
- [56] Kawano H, Yasue H, Kitagawa A, Hirai N, Yoshida T, Soejima H, et al. Dehydroepiandrosterone supplementation improves endothelial function and insulin sensitivity in men. *J Clin Endocrinol Metab* 2003;88:3190–5.
- [57] Liu D, Dillon JS. Dehydroepiandrosterone stimulates nitric oxide release in vascular endothelial cells: evidence for a cell surface receptor. *Steroids* 2004;69:279–89.
- [58] Liu D, Dillon JS. Dehydroepiandrosterone activates endothelial cell nitric-oxide synthase by a specific plasma membrane receptor coupled to Galpha(i2,3). *J Biol Chem* 2002;277:21379–88.
- [59] Simoncini T, Mannella P, Fornari L, Varone G, Caruso A, Genazzani AR. Dehydroepiandrosterone modulates endothelial nitric oxide synthesis via direct genomic and nongenomic mechanisms. *Endocrinology* 2003;144:3449–55.
- [60] van der Schouw YT, van der GY, Steyerberg EW, Eijkemans JC, Banga JD. Age at menopause as a risk factor for cardiovascular mortality. *Lancet* 1996;347:714–8.
- [61] Stampfer MJ, Colditz GA, Willett WC, Manson JE, Rosner B, Speizer FE, et al. Postmenopausal estrogen therapy and cardiovascular disease. Ten-year follow-up from the nurses' health study. *N Engl J Med* 1991;325:756–62.
- [62] Anderson GL, Limacher M, Assaf AR, Bassford T, Beresford SA, Black H, et al. Effects of conjugated equine estrogen in postmenopausal women with hysterectomy: the Women's Health Initiative randomized controlled trial. *JAMA* 2004;291:1701–12.
- [63] Barton M. Postmenopausal oestrogen replacement therapy and atherosclerosis: can current compounds provide cardiovascular protection? *Expert Opin Investig Drugs* 2001;10:789–809.
- [64] Manson JE, Martin KA. Clinical practice. Postmenopausal hormone-replacement therapy. *N Engl J Med* 2001;345:34–40.
- [65] Barton M, Dubey RK. Postmenopausal hormone-replacement therapy. *N Engl J Med* 2002;346:63–5.
- [66] Gilligan DM, Badar DM, Panza JA, Quyyumi AA, Cannon III RO. Acute vascular effects of estrogen in postmenopausal women. *Circulation* 1994;90:786–91.
- [67] Majmudar NG, Robson SC, Ford GA. Effects of the menopause, gender, and estrogen replacement therapy on vascular nitric oxide activity. *J Clin Endocrinol Metab* 2000;85:1577–83.
- [68] Orshal JM, Khalil RA. Gender, sex hormones, and vascular tone. *Am J Physiol, Regul Integr Comp Physiol* 2004;286:R233–49.
- [69] Mendelsohn ME. Protective effects of estrogen on the cardiovascular system. *Am J Cardiol* 2002;89:12E–7E.
- [70] Schini-Kerth V, Bara A, Mulsch A, Busse R. Pyrrolidine dithiocarbamate selectively prevents the expression of the inducible nitric oxide synthase in the rat aorta. *Eur J Pharmacol* 1994;265:83–7.
- [71] Csiszar A, Ungvari Z, Koller A, Edwards JG, Kaley G. Aging-induced proinflammatory shift in cytokine expression profile in coronary arteries. *FASEB J* 2003;17:1183–5.
- [72] Xia Y, Zweier JL. Superoxide and peroxynitrite generation from inducible nitric oxide synthase in macrophages. *Proc Natl Acad Sci U S A* 1997;94:6954–8.
- [73] Xia Y, Roman LJ, Masters BS, Zweier JL. Inducible nitric-oxide synthase generates superoxide from the reductase domain. *J Biol Chem* 1998;273:22635–9.
- [74] Sakai Y, Masuda H, Kihara K, Kurosaki E, Yamauchi Y, Azuma H. Involvement of increased arginase activity in impaired cavernous relaxation with aging in the rabbit. *J Urol* 2004;172:369–73.
- [75] Berkowitz DE, White R, Li D, Minhas KM, Cernetich A, Kim S, et al. Arginase reciprocally regulates nitric oxide synthase activity and



- contributes to endothelial dysfunction in aging blood vessels. *Circulation* 2003;108:2000–6.
- [76] Harman D. The free radical theory of aging. *Antioxid Redox Signal* 2003;5:557–61.
- [77] Finkel T, Holbrook NJ. Oxidants, oxidative stress and the biology of ageing. *Nature* 2000;408:239–47.
- [78] Brandes RP, Barton M, Schweitzer G, Phillippens KMH, Mügge A. Endothelial-derived superoxide anion in pig coronary arteries: evidence from lucigenin chemiluminescence and histochemical techniques. *J Physiol (Lond)* 1997;500:331–42.
- [79] Gorlach A, Brandes RP, Nguyen K, Amidi M, Dehghani F, Busse R. A gp91phox containing NADPH oxidase selectively expressed in endothelial cells is a major source of oxygen radical generation in the arterial wall. *Circ Res* 2000;87:26–32.
- [80] Jung O, Schreiber JG, Geiger H, Pedrazzini T, Busse R, Brandes RP. gp91phox-containing NADPH oxidase mediates endothelial dysfunction in renovascular hypertension. *Circulation* 2004;109:1795–1801.
- [81] Vina J, Sastre J, Pallardo F, Borras C. Mitochondrial theory of aging: importance to explain why females live longer than males. *Antioxid Redox Signal* 2003;5:549–56.
- [82] Burns EM, Kruckeberg TW, Comerford LE, Buschmann MT. Thinning of capillary walls and declining numbers of endothelial mitochondria in the cerebral cortex of the aging primate, *Macaca nemestrina*. *J Gerontol* 1979;34:642–50.
- [83] Xin MG, Zhang J, Block ER, Patel JM. Senescence-enhanced oxidative stress is associated with deficiency of mitochondrial cytochrome *c* oxidase in vascular endothelial cells. *Mech Ageing Dev* 2003;124:911–9.
- [84] Pacher P, Mabley JG, Soriano FG, Liaudet L, Komjati K, Szabo C. Endothelial dysfunction in aging animals: the role of poly(ADP-ribose) polymerase activation. *Br J Pharmacol* 2002;135:1347–50.
- [85] Pacher P, Mabley JG, Soriano FG, Liaudet L, Szabo C. Activation of poly(ADP-ribose) polymerase contributes to the endothelial dysfunction associated with hypertension and aging. *Int J Mol Med* 2002;9:659–64.
- [86] Torella D, Leosco D, Indolfi C, Curcio A, Coppola C, Ellison GM, et al. Aging exacerbates negative remodeling and impairs endothelial regeneration after balloon injury. *Am J Physiol, Heart Circ Physiol* 2004;287:2850–60.
- [87] Landmesser U, Dikalov S, Price SR, McCann L, Fukui T, Holland SM, et al. Oxidation of tetrahydrobiopterin leads to uncoupling of endothelial cell nitric oxide synthase in hypertension. *J Clin Invest* 2003;111:1201–9.
- [88] Chung HY, Song SH, Kim HJ, Ikeno Y, Yu BP. Modulation of renal xanthine oxidoreductase in aging: gene expression and reactive oxygen species generation. *J Nutr Health Aging* 1999;3:19–23.
- [89] Deshpande SS, Qi B, Park YC, Irani K. Constitutive activation of rac1 results in mitochondrial oxidative stress and induces premature endothelial cell senescence. *Arterioscler Thromb Vasc Biol* 2003;23:e1–6.
- [90] Geiszt M, Kopp JB, Varnai P, Leto TL. Identification of renox, an NAD(P)H oxidase in kidney. *Proc Natl Acad Sci U S A* 2000;97:8010–4.
- [91] Mazza F, Goodman A, Lombardo G, Vanella A, Abraham NG. Heme oxygenase-1 gene expression attenuates angiotensin II-mediated DNA damage in endothelial cells. *Exp Biol Med (Maywood)* 2003;228:576–83.
- [92] Atkinson J. Effect of aging and chronic angiotensin I converting enzyme inhibition on the endothelial function of the mesenteric arterial bed of the rat. *Am J Cardiol* 1995;76:19E–23E.
- [93] Mukai Y, Shimokawa H, Higashi M, Morikawa K, Matoba T, Hiroki J, et al. Inhibition of renin-angiotensin system ameliorates endothelial dysfunction associated with aging in rats. *Arterioscler Thromb Vasc Biol* 2002;22:1445–50.
- [94] Bachschmid M, van der LB, Schuler K, Labugger R, Thureau S, Eto M, et al. Oxidative stress-associated vascular aging is independent of the protein kinase C/NAD(P)H oxidase pathway. *Arch Gerontol Geriatr* 2004;38:181–90.
- [95] Rivard A, Fabre JE, Silver M, Chen D, Murohara T, Kearney M, et al. Age-dependent impairment of angiogenesis. *Circulation* 1999;99:111–20.
- [96] Minamino T, Miyauchi H, Yoshida T, Ishida Y, Yoshida H, Komuro I. Endothelial cell senescence in human atherosclerosis: role of telomere in endothelial dysfunction. *Circulation* 2002;105:1541–4.
- [97] Buys CH. Telomeres, telomerase, and cancer. *N Engl J Med* 2000;342:1282–3.
- [98] Serrano AL, Andres V. Telomeres and cardiovascular disease: does size matter? *Circ Res* 2004;94:575–84.
- [99] Chang E, Harley CB. Telomere length and replicative aging in human vascular tissues. *Proc Natl Acad Sci U S A* 1995;92:11190–4.
- [100] Aviv H, Khan MY, Skurnick J, Okuda K, Kimura M, Gardner J, et al. Age dependent aneuploidy and telomere length of the human vascular endothelium. *Atherosclerosis* 2001;159:281–7.
- [101] Okuda K, Khan MY, Skurnick J, Kimura M, Aviv H, Aviv A. Telomere attrition of the human abdominal aorta: relationships with age and atherosclerosis. *Atherosclerosis* 2000;152:391–8.
- [102] Vasa M, Breitschopf K, Zeiher AM, Dimmeler S. Nitric oxide activates telomerase and delays endothelial cell senescence. *Circ Res* 2000;87:540–2.
- [103] Murasawa S, Llevadot J, Silver M, Isner JM, Losordo DW, Asahara T. Constitutive human telomerase reverse transcriptase expression enhances regenerative properties of endothelial progenitor cells. *Circulation* 2002;106:1133–9.
- [104] Haendeler J, Hoffmann J, Brandes RP, Zeiher AM, Dimmeler S. Hydrogen peroxide triggers nuclear export of telomerase reverse transcriptase via Src kinase family-dependent phosphorylation of tyrosine 707. *Mol Cell Biol* 2003;23:4598–610.
- [105] Haendeler J, Hoffmann J, Diehl JF, Vasa M, Spyridopoulos I, Zeiher AM, et al. Antioxidants inhibit nuclear export of telomerase reverse transcriptase and delay replicative senescence of endothelial cells. *Circ Res* 2004;94:768–75.
- [106] Matsushita H, Chang E, Glassford AJ, Cooke JP, Chiu CP, Tsao PS. eNOS activity is reduced in senescent human endothelial cells: preservation by hTERT immortalization. *Circ Res* 2001;89:793–8.
- [107] Hoffmann J, Haendeler J, Aicher A, Rossig L, Vasa M, Zeiher AM, et al. Aging enhances the sensitivity of endothelial cells toward apoptotic stimuli: important role of nitric oxide. *Circ Res* 2001;89:709–15.
- [108] Dimmeler S, Haendeler J, Zeiher AM. Regulation of endothelial cell apoptosis in atherothrombosis. *Curr Opin Lipidol* 2002;13:531–6.
- [109] Weinsaft JW, Edelberg JM. Aging-associated changes in vascular activity: a potential link to geriatric cardiovascular disease. *Am J Geriatr Cardiol* 2001;10:348–54.
- [110] Swift ME, Kleinman HK, DiPietro LA. Impaired wound repair and delayed angiogenesis in aged mice. *Lab Invest* 1999;79:1479–87.
- [111] Edelberg JM, Tang L, Hattori K, Lyden D, Rafii S. Young adult bone marrow-derived endothelial precursor cells restore aging-impaired cardiac angiogenic function. *Circ Res* 2002;90:E89–93.
- [112] Walter DH, Dimmeler S. Endothelial progenitor cells: regulation and contribution to adult neovascularization. *Herz* 2002;27:579–88.
- [113] Scheubel RJ, Zorn H, Silber RE, Kuss O, Morawietz H, Holtz J, et al. Age-dependent depression in circulating endothelial progenitor cells in patients undergoing coronary artery bypass grafting. *J Am Coll Cardiol* 2003;42:2073–80.
- [114] Azuma H, Funayama N, Kubota T, Ishikawa M. Regeneration of endothelial cells after balloon denudation of the rabbit carotid artery and changes in responsiveness. *Jpn J Pharmacol* 1990;52:541–52.
- [115] Weidinger FF, McLenachan JM, Cybulsky MI, Gordon JB, Rennke NK, Hollenberg NK, et al. Persistent dysfunction of regenerated endothelium after balloon angioplasty of rabbit iliac artery. *Circulation* 1990;81:1667–79.
- [116] Fournet-Bourguignon MP, Castedo-Delrieu M, Bidouard JP, Leonce S, Saboureaux D, Delescluse I, et al. Phenotypic and functional



- changes in regenerated porcine coronary endothelial cells: increased uptake of modified LDL and reduced production of NO. *Circ Res* 2000;86:854–61.
- [117] Smith AR, Hagen TM. Vascular endothelial dysfunction in aging: loss of Akt-dependent endothelial nitric oxide synthase phosphorylation and partial restoration by (R)-alpha-lipoic acid. *Biochem Soc Trans* 2003;31:1447–9.
- [118] Pili R, Guo Y, Chang J, Nakanishi H, Martin GR, Passaniti A. Altered angiogenesis underlying age-dependent changes in tumor growth. *J Natl Cancer Inst* 1994;86:1303–14.
- [119] Ashcroft GS, Horan MA, Ferguson MW. Aging alters the inflammatory and endothelial cell adhesion molecule profiles during human cutaneous wound healing. *Lab Invest* 1998;78:47–58.
- [120] Koike T, Vernon RB, Gooden MD, Sadoun E, Reed MJ. Inhibited angiogenesis in aging: a role for TIMP-2. *J Gerontol, A Biol Sci Med Sci* 2003;58:B798–805.
- [121] Sadoun E, Reed MJ. Impaired angiogenesis in aging is associated with alterations in vessel density, matrix composition, inflammatory response, and growth factor expression. *J Histochem Cytochem* 2003;51:1119–30.
- [122] Agah A, Kyriakides TR, Letrondo N, Bjorkblom B, Bornstein P. Thrombospondin 2 levels are increased in aged mice: consequences for cutaneous wound healing and angiogenesis. *Matrix Biol* 2004;22:539–47.
- [123] Volpert OV, Tolsma SS, Pellerin S, Feige JJ, Chen H, Mosher DF, et al. Inhibition of angiogenesis by thrombospondin-2. *Biochem Biophys Res Commun* 1995;217:326–32.
- [124] Armstrong LC, Bjorkblom B, Hankenson KD, Siadak AW, Stiles P, Bornstein P. Thrombospondin 2 inhibits microvascular endothelial cell proliferation by a caspase-independent mechanism. *Mol Biol Cell* 2002;13:1893–905.
- [125] Kumazaki T, Wadhwa R, Kaul SC, Mitsui Y. Expression of endothelin, fibronectin, and mortalin as aging and mortality markers. *Exp Gerontol* 1997;32:95–103.
- [126] Kumazaki T. Modulation of gene expression during aging of human vascular endothelial cells. *Hiroshima J Med Sci* 1993;42:97–100.
- [127] White M, Courtemanche M, Stewart DJ, Talajic M, Mikes E, Cernacek P, et al. Age- and gender-related changes in endothelin and catecholamine release, and in autonomic balance in response to head-up tilt. *Clin Sci (Lond)* 1997;93:309–16.
- [128] D'Uscio LV, Barton M, Shaw S, Luscher TF. Endothelin in atherosclerosis: importance of risk factors and therapeutic implications. *J Cardiovasc Pharmacol* 2000;35:S55–9.
- [129] Barton M, Haudenschild CC, D'Uscio LV, Shaw S, Munter K, Luscher TF. Endothelin ETA receptor blockade restores NO-mediated endothelial function and inhibits atherosclerosis in apolipoprotein E-deficient mice. *Proc Natl Acad Sci U S A* 1998;95:14367–72.
- [130] Amiri F, Virdis A, Neves MF, Iglarz M, Seidah NG, Touyz RM, et al. Endothelium-restricted overexpression of human endothelin-1 causes vascular remodeling and endothelial dysfunction. *Circulation* 2004;110:2233–40.
- [131] Dohi Y, Luscher TF. Aging differentially affects direct and indirect actions of endothelin-1 in perfused mesenteric arteries of the rat. *Br J Pharmacol* 1990;100:889–93.
- [132] Tschudi MR, Luscher TF. Age and hypertension differently affect coronary contractions to endothelin-1, serotonin, and angiotensins. *Circulation* 1995;91:2415–22.
- [133] Goodwin AT, Amrani M, Marchbank AJ, Gray CC, Jayakumar J, Yacoub MH. Coronary vasoconstriction to endothelin-1 increases with age before and after ischaemia and reperfusion. *Cardiovasc Res* 1999;41:554–62.
- [134] Wedgwood S, Black SM. Endothelin-1 decreases endothelial NOS expression and activity through eta receptor mediated generation of hydrogen peroxide. *Am J Physiol, Lung Cell Mol Physiol* 2004 [Epub ahead of print].
- [135] Zhou L, Dong J, Yu M, Yin H, She M. Age-dependent increase of NF-kappaB translocation and PDGF-B expression in aortic endothelial cells of hypercholesterolemic rats. *Exp Gerontol* 2003;38:1161–8.
- [136] Li Z, Froehlich J, Galis ZS, Lakatta EG. Increased expression of matrix metalloproteinase-2 in the thickened intima of aged rats. *Hypertension* 1999;33:116–23.
- [137] Orlandi A, Marcellini M, Spagnoli LG. Aging influences development and progression of early aortic atherosclerotic lesions in cholesterol-fed rabbits. *Arterioscler Thromb Vasc Biol* 2000;20:1123–36.
- [138] Morisaki N, Saito I, Tamura K, Tashiro J, Masuda M, Kanzaki T, et al. New indices of ischemic heart disease and aging: studies on the serum levels of soluble intercellular adhesion molecule-1 (ICAM-1) and soluble vascular cell adhesion molecule-1 (VCAM-1) in patients with hypercholesterolemia and ischemic heart disease. *Atherosclerosis* 1997;131:43–8.