- 11. Katrukha AG, Bereznikova AV, Esakova TV, Pettersson K, Lovgren T, Severina ME, et al. Troponin I is released in bloodstream of patients with acute myocardial infarction not in free form but as complex. Clin Chem 1997;43:1379–85.
- Giuliani I, Bertinchant J-P, Granier C, Laprade M, Chocron S, Toubin G, et al. Determination of cardiac troponin I forms in the blood of patients with acute myocardial infarction and patients receiving crystalloid or cold blood cardioplegia. Clin Chem 1999;45:213–22.
- Bodor GS, Oakley AE, Allen PD, Crimmins DL, Ladenson JH, Anderson PAW. Troponin I phosphorylation in the normal and failing adult human heart. Circulation 1997;96:1495–500.
- Ingraham RH, Hodges RS. Effects of Ca<sup>2+</sup> and subunit interaction on surface accessibility of cysteine residues of cardiac troponin. Biochemistry 1988:27:5891–8.
- Dasgupta A, Chow L, Nazareno L, Tso G, Datta P. Analytical performance of a new chemiluminescent troponin I assay using ACS:180 analyzer. [Abstract] Clin Chem 1999;45:A136.
- College of American Pathologists. Chemistry survey 1997: sets C3, C4, C5, C6, C7, and C8. Participant summary. Northfield, IL: College of American Pathologists, 1997.
- Hegner N, Baum H, Eller T, Pohl KH, Messinger M, Mockel M, et al. Multicenter evaluation of OPUS troponin I compared to myoglobin and CK-MB concentrations in cardiac diseases. Clin Lab 1997:43:501–14.
- Bhayana V, Gougoulias T, Gawad Y. A comparison of four serum cardiac troponin I methods [Abstract]. Clin Chem 1997;43:S128.

Glutamic Acid Decarboxylase Antibodies in Screening for Autoimmune Diabetes: Influence of Comorbidity, Age, and Sex on Specificity and Threshold Values, Manou R. Batstra, 1\* Arianne van Driel, 1 Jacob S. Petersen, 2 Cees A. van Donselaar,3,7 Maarten J. van Tol,4 G. Jan Bruining, 1 Diederick E. Grobbee, 5 Thomas Dyrberg, 6 and Henk-Ian Aanstoot<sup>1,8</sup> (Departments of <sup>1</sup> Pediatrics, <sup>3</sup> Neurology, and <sup>5</sup> Epidemiology and Biostatistics, Erasmus University, 3015 GE Rotterdam, The Netherlands; <sup>2</sup> The Hagedorn Research Institute, Gentofte DK2820, Denmark; 4 Department of Pediatrics, Leiden University Medical Center, Leiden 2333 ZA, The Netherlands; <sup>6</sup> Diabetes Immunology, Novo Nordisk A/S, Bagsværd DK2880, Denmark; <sup>7</sup> Hospital St. Clara, Department of Pediatrics, Rotterdam 3078 HT, The Netherlands; 8 IJsselland Hospital, Department of Pediatrics, Capelle a.d. IJssel 2906 ZC, The Netherlands; \* address correspondence to this author at: Department of Immunology, Ee 893, Dr. Molewaterplein 50, 3015 GE Rotterdam, The Netherlands; fax 31-10-4087038, e-mail Batstra@immu.fgg.eur.nl.)

Antibodies against the 65-kDa isoform of glutamic acid decarboxylase ( $GAD_{65}$ ) can be applied as a predictive tool for childhood type-1 diabetes (1-6) and to facilitate the differential diagnosis of diabetes in adults (7-9). However, the sensitivity and specificity of GAD antibody screening have not been fully characterized, and the positive predictive value of screening varies from 20% to 70%, depending on the strategy applied and the population studied (2, 3, 5-7, 9-12). The current study aims to identify factors that may lead to false-positive results in GAD antibody screening.

Previously, it has been demonstrated that the GAD antibody frequency in mixed connective tissue disease and stiff-man syndrome is increased, although not all patients who suffer from these diseases and are positive

for GAD antibodies develop type-1 diabetes (13, 14). GAD is expressed in the islets of Langerhans, neuronal tissue, the ovaries, and the testes (15, 16). Similar to the above-mentioned examples, comorbidity involving these tissues may lead to GAD antibody formation but not to diabetes. Therefore, we compared the prevalence of GAD antibodies in patients with cystic fibrosis, epilepsy, Guillain-Barré syndrome, and premature ovarian failure to the prevalence in an unselected population of 1403 schoolchildren.

In addition, thresholds for positivity for GAD antibodies have generally been defined in children. These thresholds might not be applicable when testing for type-1 diabetes in adults. Therefore, we studied whether GAD antibody concentrations are correlated to age and sex, and whether adjustment of assay thresholds to include these variables may improve screening specificity.

The frequencies of positive results for GAD antibodies and the concentrations of GAD antibodies were established in a population of 1403 schoolchildren, ages 10–12 years, without chronic diseases (17). During a 10-year follow-up, two of these children developed type-1 diabetes [ascertainment >96% (18)].

The influence of comorbidity on GAD antibody concentrations and the frequencies of positive results were studied in four patient populations. The subjects included 394 patients who participated in the Dutch study of epilepsy in childhood (19, 20). These patients were eligible for the current study if the diagnosis was confirmed on the basis of electroencephalograms or therapy and sufficient serum for antibody analysis was available. Sera collected within 2 months after the presenting seizure (mean duration, 0.7 months; n = 228) and at the longest disease duration available of each patient (mean duration, 12.2 months; range, 2-50 months; n = 294) were analyzed separately. The diagnosis and development of diabetes during follow-up (5 years) were recorded from the medical records. Forty-three serum samples from 38 cystic fibrosis patients (collected in 1990–1992) were analyzed for GAD antibodies. In addition, we studied 30 patients with premature ovarian failure and 28 patients with Guillain-Barré syndrome (14 males; age range, 19-64 years). All patient sera were stored at -80 °C.

The influence of age and sex on GAD antibody concentrations and the frequencies of positive results were studied in 1287 individuals from the city of Zoetermeer, who participated in a study of cardiovascular risk factors. Sera were collected in 1976 and stored at −20 °C until testing. The population is described in detail elsewhere (21). The demographic data of the populations are shown in Table 1.

The study protocols were approved by the appropriate medical ethics committees according to the Helsinki Declaration. Informed consent was obtained from all participants or their parents.

Sera were tested for GAD antibodies by radiobinding assay (RBA) (13) or immunoprecipitation and sodium dodecyl sulfate-polyacrylamide gel electrophoresis of the Triton X-100 fraction of [35S]methionine-labeled fetal rat

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Table 1	Demographic dat	a ot nonillations	Tested for GAIL	antinodies

Population	Schoolchildren	General population	Epilepsy, all	Epilepsy, onset	Epilepsy, long duration	Cystic fibrosis	Premature ovarian failure	Guillain- Barré syndrome
n	1403	1287	522	228	294	43	30	28
Age range (mean), years	10–12	6–86 (32)	6–19 (6.42)	0–14 (5.77)	1–19 (6.87)			19–64
Disease duration (mean), months	NA <sup>a</sup>	NA	0–50 (6.9)	0–1.5 (0.23)	2–50 (12)			
Median GAD index	-0.01	$0.04^{b}$	0.01	0.01	0.02	-0.05		
(range)	(-0.05  to  1.53)	(-0.07  to  1.71)	(-0.1  to  1.45)	(-0.1  to  1.45)	(-0.1  to  0.28)	(-0.08  to  0.21)		
Positive RBA (n), %	0.4 (5)	1.0 (13)	0.8 (4)	0.9 (2)	0.6 (2)	2.3 (1)	NT	NT
Positive cIMP, n	5	NT	4	2	2	NT	0	1
cIMP/RBA	RBA/cIMP	RBA	RBA/cIMP	RBA/cIMP	RBA/cIMP	RBA	cIMP	cIMP
<sup>a</sup> NA, not applicable	e; NT, not tested.							

<sup>&</sup>lt;sup>b</sup> P < 0.001 compared with the schoolchildren.

islets (a gift from Dr. T. Dyrberg and H. Richter-Olesen, the Hagedorn Research Institute, Gentofte, Denmark) and evaluated using autoradiography [conventional immunoprecipitation (cIMP)] (22) as indicated in Table 1.

For the RBA, all sera were analyzed in triplicate, and precipitated radioactivity was counted in a microbeta plate reader (EG&G Wallac). Internal reference sera were included in each plate for the epilepsy and cystic fibrosis patients and in each third plate for the schoolchildren and the general population. These internal reference sera were used to calculate an index (GAD index) for the purpose of comparison of experiments (13). The GAD antibody concentration in the positive reference serum was in the linear range of the dilution curve, which allowed the GAD index to be interpreted semiquantitatively. The threshold for positivity was defined as the 99.5th centile of the population of schoolchildren (GAD index >0.21).

The statistical package SPSS for Windows (SPSS) was used for data analysis. The  $\chi^2$ , Mann–Whitney, and Kruskal–Wallis tests were used to analyze differences between groups. Trends within groups were analyzed by the Spearman correlation test.

The GAD antibody frequencies of positive results and concentrations in the six populations studied are shown in Table 1. The positive individuals did not differ from the negative individuals in age or sex distribution in any of the populations studied. The GAD antibody concentrations and frequencies of positive results in the patient populations were not significantly increased compared with the schoolchildren.

In the epilepsy patients, the GAD antibody positive frequency shortly after the presenting seizure did not differ from the frequency at long disease duration, nor was there a correlation between antibody concentrations or positive frequencies and duration of epilepsy. Two children from the epilepsy cohort developed diabetes during follow-up. One was positive for GAD antibodies in a sample taken before diabetes onset (at the presenting seizure) and 7 months after onset of diabetes (13-month epilepsy duration). The other was negative for GAD antibodies in all serum samples

analyzed. Two other patients were positive for GAD antibodies (in only one sample) but did not develop diabetes during follow-up.

One of the cystic fibrosis patients had a GAD index of 0.21, which is just at the defined threshold for positivity. A serum sample from this patient collected 3 months later was negative (GAD index, 0.16). Two out of five school-children who were positive for GAD antibodies developed diabetes during follow-up.

As shown in Fig. 1, there was a slight but statistically significant correlation between age and GAD index (Spearman correlation coefficient, 0.161; P < 0.001). In addition, the GAD index in women was significantly higher than in men (P = 0.009; median GAD index, 0.043 and 0.037, respectively). The correlation between sex and GAD index did not explain the correlation with age and vice versa

To study whether the observed correlation with age affected threshold definition, the general population was split into 10 similarly sized age groups (Fig. 1). The threshold for positivity was adjusted to the 99.5th centile of each age group and compared with the 99.5th centile threshold of the general population (0.68; see Fig. 1) and the threshold established in the schoolchildren. This yielded four additional positive individuals. One of these was negative at the initially applied threshold of 0.21. One individual who was positive at the general population threshold was negative when the age-adjusted thresholds were applied. Adjustment of thresholds for sex did not affect the antibody frequencies.

Strikingly, the GAD index was significantly higher in the general population than in the schoolchildren cohort. This difference remained present when outliers were discarded from both populations. To exclude that this observation was attributable to age effects, we selected all individuals 10–12 years of age from the general population and compared their GAD index to the GAD index in the schoolchildren cohort. The median GAD index in this selection of the general population (0.01) was still significantly higher than in the schoolchildren cohort (-0.04).

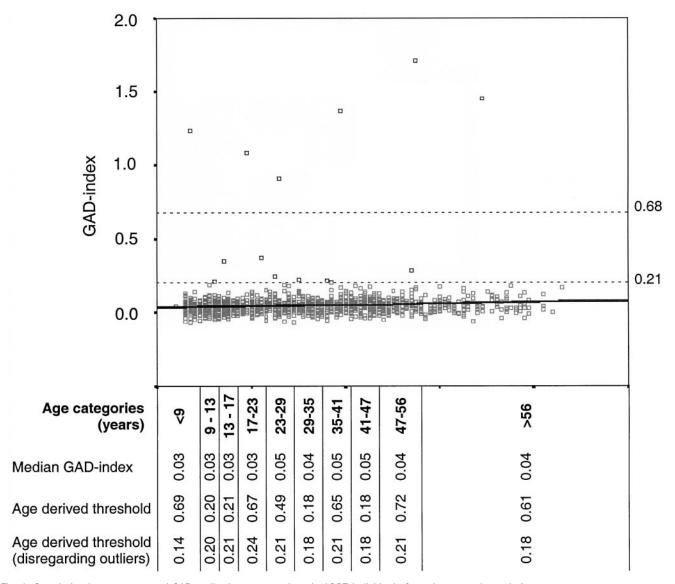


Fig. 1. Correlation between age and GAD antibody concentrations in 1287 individuals from the general population. Spearman correlation coefficient, 0.161; *P* <0.001. *Dotted lines* indicate the applied threshold (0.21) and the population-adjusted threshold (0.68).

None of the diseases involving tissues expressing GAD evaluated in the current study caused a significant increase of GAD antibody concentrations or frequencies of positive results. One might argue that the use of different methods for antibody detection in the patients and reference populations weakens the comparative analysis. However, by evaluation of all epilepsy patients and the schoolchildren who had GAD antibody concentrations higher than the mean + 1 SD in cIMP, we demonstrated that cIMP is at least as sensitive or possibly more sensitive than the RBA (data not shown). In the current study, we hypothesized that the GAD antibody frequency of positive results in neuroendocrine patients was higher because of non-diabetes-associated formation of GAD anti-

bodies. Using the highly sensitive immunoprecipitation technique in the patient populations would only magnify this effect.

One of 28 (3.6%) Guillain-Barré syndrome patients was positive for GAD antibodies. These data clearly suggest that GAD antibodies are not part of the Guillain-Barré syndrome, but additional studies are needed to draw definite conclusions.

We observed a slight, but statistically significant, positive correlation between age and GAD index, but this correlation was not reflected in threshold definition. Exclusion of outliers from the distribution did not alter these perspectives. Although we observed a difference in the mean GAD index between males and females, this did not

affect threshold definition. These results are in concordance with the observations from a previous study in a substantially smaller population (23) and indicate that adjustment of assay thresholds for age or sex is not indicated.

The GAD index in the general population was significantly increased compared with the schoolchildren. We concluded that this difference was attributable to differences in age between the populations. In addition, both populations were tested in one experiment (using one batch of tracer, control sera, and protein A-Sepharose), thus excluding technical variation of the methods used. A noticeable difference between both populations was serum storage. The general population sera were stored at −20 °C for 16 years in tubes with snap caps. When defrosted, some of these sera contained precipitates, which were removed by centrifugation before GAD antibody analysis. The sera of the schoolchildren cohort, on the contrary, were stored at -80 °C for 10 years in tubes with screw caps and had never been defrosted before the GAD antibody analysis. It is possible that the storage conditions of the samples from the general population led to the observed increased antibody concentrations in the general population sera. This observation implies that storage conditions should be monitored carefully to exclude such technical problems in future studies. Because all sera from the general population were stored in one freezer in identical tubes for the same period of time, it is not likely that the storage conditions affected the analysis of age and sex effects in the general population. In addition, positive samples did not differ in macroscopic aspects from negative samples.

This study demonstrates that age- and sex-nondefined populations can be used for definition of reference values for GAD antibodies and that comorbidity involving tissues that express GAD needs not be taken into account when screening for GAD antibodies. Because storage conditions may significantly affect the outcome of GAD antibody tests, these need to be monitored meticulously in collaborative studies.

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## References

- Aanstoot HJ, Kang SM, Kim J, Lindsay LA, Roll U, Knip M, et al. Identification and characterization of glima 38, a glycosylated islet cell membrane antigen, which together with GAD65 and IA2 marks the early phases of autoimmune response in type 1 diabetes. J Clin Investig 1996;97:2772–83.
- Batstra MR, Bruining GJ, Aanstoot HJ. Antibody screening in a population of children. Ann Med 1997;29:453–60.
- 3. Bingley PJ, Bonifacio E, Williams AJ, Genovese S, Bottazzo GF, Gale EA.

- Prediction of IDDM in the general population: strategies based on combinations of autoantibody markers. Diabetes 1997;46:1701–10.
- Wiest-Ladenburger U, Hartmann R, Hartmann U, Berling K, Bohm BO, Richter W. Combined analysis and single-step detection of GAD65 and IA2 autoantibodies in IDDM can replace the histochemical islet cell antibody test. Diabetes 1997;46:565–71.
- Kulmala P, Savola K, Petersen JS, Vahasalo P, Karjalainen J, Lopponen T, et al. Prediction of insulin-dependent diabetes mellitus in siblings of children with diabetes. A population-based study. The Childhood Diabetes in Finland Study Group. J Clin Investig 1998;101:327–36.
- Roll U, Ziegler AG. Combined antibody screening for improved prediction of IDDM-modern strategies. Exp Clin Endocrinol Diabetes 1997;105:1–14.
- Seissler J, de Sonnaville JJ, Morgenthaler NG, Steinbrenner H, Glawe D, Khoo-Morgenthaler UY, et al. Immunological heterogeneity in type I diabetes: presence of distinct autoantibody patterns in patients with acute onset and slowly progressive disease. Diabetologia 1998;41:891–7.
- Hagopian WA, Karlsen AE, Gottsater A, Landin-Olsson M, Grubin CE, Sundkvist G, et al. Quantitative assay using recombinant human islet glutamic acid decarboxylase (GAD65) shows that 64K autoantibody positivity at onset predicts diabetes type. J Clin Investig 1993;91:368-74.
- Zimmet PZ, Tuomi T, Mackay IR, Rowley MJ, Knowles W, Cohen M, et al. Latent autoimmune diabetes mellitus in adults (LADA): the role of antibodies to glutamic acid decarboxylase in diagnosis and prediction of insulin dependency. Diabet Med 1994;11:299–303.
- 10. Borg H, Fernlund P, Sundkvist G. Protein tyrosine phosphatase-like protein IA2-antibodies plus glutamic acid decarboxylase 65 antibodies (GADA) indicates autoimmunity as frequently as islet cell antibodies assay in children with recently diagnosed diabetes mellitus. Clin Chem 1997;43: 2358-63.
- Tuomi T, Groop LC, Zimmet PZ, Rowley MJ, Knowles W, Mackay IR. Antibodies to glutamic acid decarboxylase reveal latent autoimmune diabetes mellitus in adults with a non-insulin-dependent onset of disease. Diabetes 1993;42:359–62.
- Niskanen LK, Tuomi T, Karjalainen J, Groop LC, Uusitupa MIJ. GAD antibodies in NIDDM: ten-year follow-up from the diagnosis. Diabetes Care 1995; 18:1557–65.
- **13.** Petersen JS, Hejnaes KR, Moody A, Karlsen AE, Marshall MO, Hoier-Madsen M, et al. Detection of GAD65 antibodies in diabetes and other autoimmune diseases using a simple radioligand assay. Diabetes 1994;43:459–67.
- 14. Kim J, Namchuk M, Buchawan T, Fu Q, Jaffe M, Shi Y, et al. Higher autoantibody levels and recognition of a linear NH2-terminal epitope in the autoantigen GAD65, distinguish stiff-man syndrome from insulin dependent diabetes mellitus. J Exp Med 1994;180:595–606.
- **15.** Celotti F, Moore GD, Rovescalli AC, Solimena M, Negri-Cesi P, Racagni G. The GABAergic system in the rat oviduct: presence, endocrine regulation and possible localization. Pharmacol Res 1989;21:103–4.
- Erdo SL, Joo F, Wolff JR. Immunohistochemical localization of glutamate decarboxylase in the rat oviduct and ovary: further evidence for non-neural GABA systems. Cell Tissue Res 1989:255:431–4.
- 17. de Man SA, Andre JL, Bachmann H, Grobbee DE, Ibsen KK, Laaser U, et al. Blood pressure in childhood: pooled findings of six European studies. J Hypertens 1991;9:109–14.
- 18. Ruwaard D, Hirasing RA, Reeser HM, van Buuren S, Bakker K, Heine RJ, et al. Increasing incidence of type I diabetes in The Netherlands. The second nationwide study among children under 20 years of age. Diabetes Care 1994;17:599–601.
- 19. Stroink H, Brouwer OF, Arts WF, Geerts AT, Peters AC, van Donselaar CA. The first unprovoked, untreated seizure in childhood: a hospital based study of the accuracy of the diagnosis, rate of recurrence, and long term outcome after recurrence. Dutch study of epilepsy in childhood. J Neurol Neurosurg Psychiatry 1998;64:595–600.
- Carpay H, Arts WFM, Geerts H, Brouwer OF, Peters ACB, Donselaar CAV. Epilepsy in childhood. An audit of clinical practice. Arch Neurol 1998;55: 668-73.
- 21. Hoes AW, Grobbee DE, Valkenburg HA, Lubsen J, Hofman A. Cardiovascular risk and all-cause mortality: a 12 years follow-up study in The Netherlands. Eur J Epidemiol 1993;9:285–92.
- Baekkeskov S, Aanstoot HJ, Christgau S, Reetz A, Solimena M, Cascalho M, et al. Identification of the 64K autoantigen in insulin-dependent diabetes as the GABA-synthesizing enzyme glutamic acid decarboxylase. Nature 1990; 347:151–6.
- 23. Vandewalle CL, Falorni A, Svanholm S, Lernmark A, Pipeleers DG, Gorus FK. High diagnostic sensitivity of glutamate decarboxylase autoantibodies in insulin-dependent diabetes mellitus with clinical onset between age 20 and 40 years. The Belgian Diabetes Registry. J Clin Endocrinol Metab 1995;80:846–51.