# Hyperhomocysteinemia and the Endocrine System: Implications for Atherosclerosis and Thrombosis

VIVIAN FONSECA, SUSAN C. GUBA, AND LOUIS M. FINK

Department of Medicine, Section of Endocrinology, Tulane University Medical School (V.F.), New Orleans, Louisiana 70112; and the Department of Pathology, University of Arkansas for Medical Sciences and the John L. McClellan Memorial Veterans Hospital (S.C.G., L.M.F.), Little Rock, Arkansas

- I. Introduction
- II. Methionine-Homocysteine Metabolism
  - A. Methionine metabolism
  - B. Regulation of remethylation and transsulfuration of H(e)
- III. Nomenclature and Methodology in the Measurement of Plasma H(e)
  - A. Methionine load test
- IV. Determinants of Plasma Homocysteine
  - A. Physiological
  - B. Genetics of hyperhomocysteinemia
  - C. Nutritional
  - D. Hormones and H(e) metabolism
- V. Homocysteine and Diabetes Mellitus (DM)
  - A. Hyperhomocysteinemia, renal failure, and diabetic nephropathy
  - B. Effect of glucose and insulin on H(e) metabolism
- VI. Hyperhomocysteinemia and Cholesterol Metabolism
- VII. Hyperhomocysteinemia in Premature Vascular Disease
  - A. Epidemiological and prospective studies
  - B. Studies in patients with established vascular disease
  - C. Negative studies
  - D. Effect of low plasma H(e) on cardiovascular disease
- VIII. Possible Mechanisms of Accelerated Vascular Disease in Homocysteinemia
  - A. Platelet dysfunction
  - B. Coagulation abnormalities
  - C. Effects on the endothelium
  - D. Effects of hyperhomocysteinemia on the arterial wall
  - E. Coinheritance of factor V Leiden in homocystinuria
- IX. Management of Hyperhomocysteinemia
  - A. Prevention of hyperhomocysteinemia
  - B. Treatment of hyperhomocysteinemia
- X. Conclusion

#### I. Introduction

 $\blacksquare$  OMOCYSTEINE (H(e)) is a nonprotein-forming, thiolcontaining amino acid formed by demethylation of methionine. It is metabolized by remethylation to methionine or by transsulfuration to cysteine. An elevated plasma H(e) level may occur as a result of inherited disorders, which alter enzyme activity in the transsulfuration and remethylation pathways. Alternatively, nutritional deficiencies of essential cofactors or enzyme substrates, including cobalamin (vitamin  $B_{12}$ ), folate, or pyridoxine (vitamin  $B_6$ ), can result in blockade of H(e) metabolic pathways. An elevated plasma H(e) level has recently been established as an independent risk factor for thrombosis and vascular disease (1-11). However, the relationship between hyperhomocysteinemia [HH(e)] and cardiovascular disease remains controversial. Although some prospective studies have confirmed that H(e) is an independent risk factor for cardiovascular disease (3, 6), other studies have not found such a relationship (12, 13). Because of the possibility of reducing plasma H(e) with vitamin therapy, this issue has received considerable public attention. However, no large clinical trials have demonstrated a reduction in cardiovascular risk. Ongoing clinical trials are examining the possibility that vitamin therapy to lower H(e) levels may prevent cardiovascular disease.

Homocystinuria is an inherited disorder characterized by severely elevated plasma H(e). Homocystinuric children are known to develop premature vascular disease involving all major blood vessels (14, 15). McCully (16) first drew attention to a possible link between elevated plasma H(e) and vascular disease, making the seminal observation that extensive arterial thrombosis and atherosclerosis commonly occurs in children with homocystinuria. Boers et al. (17) highlighted the association between accelerated vascular disease and moderate elevation in plasma H(e), without the other manifestations of homocystinuria. Since then, there has been considerable interest in mild HH(e) as a risk factor for coronary artery disease, stroke, and peripheral vascular disease. The understanding of the different etiologies of HH(e) is changing because of the ability to discriminate between the types of mutations present in inherited disorders, the ability to distinguish between homozygous and heterozygous mutations, and recognition of factors that modify H(e) metabo-

A mutation resulting in a thermolabile variant of the meth-

Address reprint requests to: Vivian A. Fonseca, M.D., Tulane University Medical School, Section of Endocrinology, 1430 Tulane Avenue (SL 53), New Orleans, Louisiana 70112 USA. E-mail: vfonseca@mailhost.tcs.tulane.edu

ylene tetrahydrofolate reductase (MTHFR) enzyme is common, occurring in one-third to one-half of alleles and varying slightly with the population studied. In the homozygous state, the thermolabile variant is found in about 8% of the population (1, 2, 18, 19). Mild fasting hyperhomocysteinemia has only been reported in individuals homozygous for this polymorphism (2) and, as discussed below, occurs only with concomitantly low folate levels. Increased cardiovascular risk is not associated with the mutation per se. However, the risk of cardiovascular disease is increased in homozygotes with concomitantly low folate levels. A substantial proportion of patients with cardiovascular disease have post methionine load HH(e), suggesting a possible defect in cystathionine- $\beta$ -synthase (CBS) action. However, mutations of CBS are relatively rare, occurring in only approximately 9% of the population (2).

Thus, the high frequency (25–30%) of postmethionine load HH(e) that occurs in patients with cardiovascular disease again suggests that most cases of HH(e) are due to nongenetic factors. Two possible such factors are nutritional status and hormonal changes. Motulsky (2) highlighted a potential interaction between nutritional and genetic factors, an example of a gene-environment interaction.

The purpose of this review is to update the practicing endocrinologist on methionine-homocysteine metabolism, H(e) measurements, genetics of HH(e), and mechanisms of vascular disease in HH(e). In particular, we will highlight the evidence of interactions between the endocrine system and H(e) metabolism.

#### II. Methionine-Homocysteine Metabolism

## A. Methionine metabolism

Methionine is converted to H(e) through two intermediates: *S*-adenosyl-methionine (SAM) and *S*-adenosyl-homocysteine. The metabolism of H(e) (Fig. 1) occurs either via the transsulfuration pathway or the remethylation pathway (20).

It is likely that the remethylation is active in the fasting state and that transsulfuration is predominant after a methionine load such as a high protein meal (see below). H(e) irreversibly condenses with serine to form cystathionine (reaction 1); this reaction is catalyzed by CBS and is also dependent on pyridoxal-5'-phosphate (the active metabolite of vitamin  $B_6$ ) as a cofactor. Cystathionine is hydrolyzed to cysteine by the enzyme cystathionase (reaction 2) and is also a B<sub>6</sub>-dependent reaction. Alternatively, methionine may be reformed via the remethylation pathway when a methyl group is donated to H(e). In this pathway, 5,10-methylene tetrahydrofolate is converted to N-5-methyl tetrahydrofolate (reaction 3a), in a reaction catalyzed by MTHFR, with riboflavin as a cofactor. *N*-5-methyl tetrahydrofolate then donates a methyl group to H(e) in a reaction catalyzed by 5-methyltetrahydrofolatehomocysteine methyltransferase (methionine synthase) and its cosubstrate B-12 (reaction 3b). Alternatively, the methyl group may be donated by betaine (reaction 4) in a reaction catalyzed by betaine-homocysteine methyltransferase, forming dimethylglycine and methionine. The betaine-homocysteine reaction is neither vitamin B<sub>12</sub> nor folate dependent.

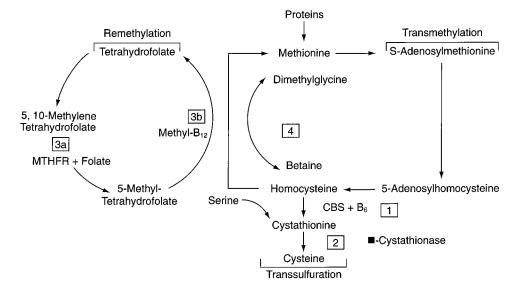
## B. Regulation of remethylation and transsulfuration of H(e)

As described above, H(e) is metabolized by at least two pathways. Selhub and Miller (21) proposed that the partitioning of H(e) between *de novo* methionine synthesis and catabolism through the cystathionine synthesis occurs by coordinate regulation by SAM (21). They have proposed the hypothesis that impairment of one H(e) metabolic pathway must be associated with the impairment of the other metabolic pathway to cause HH(e). However, this mechanism has only been demonstrated in mice. Proof of coordinate regulation in higher mammals is needed.

Dietary and metabolically derived methionine is conjugated by ATP to form SAM. SAM serves primarily as a methyl donor to a variety of acceptors, including guanidinoacetate, nucleic acids, neurotransmitters, phospholipids,

## **Homocysteine Metabolism**

Fig. 1. Summary of the metabolic pathways in homocysteine metabolism. Reaction 1 is catalyzed by choline oxidase; reaction 2, betaine-homocysteine methyltransferase; reaction 3, 5-methyltetrahydrofolate-homocysteine methyltransferase; reaction 4, phosphatidylethanolamine methyltransferase. [Modified from S. Guba et al.: Am J Clin Pathol 106:709–721, 1996 (25). © 1996 by the American Society of Clinical Pathologists. Reprinted with permission.]



and hormones (21). Creatine synthesis accounts for a major portion of SAM consumption. *S*-adenosylhomocysteine is the byproduct of these methyl transfer reactions and is hydrolyzed to form H(e), which then starts a new cycle of methyl group transfer. In one study, red cell SAM levels were found to be low in patients with coronary artery disease (22). However, red cell SAM may not adequately reflect levels in other tissues, such as the liver, which may be metabolically more active (21).

Studies in rodents have demonstrated that SAM is both an allosteric inhibitor of MTHFR (23) and an activator of CBS (24). Selhub and Miller (21) have proposed that the ability of SAM to act as an enzymatic effector provides a means by which remethylation and transsulfuration can be coordinated. When cellular SAM concentration is low, CBS will be suppressed, resulting in increased remethylation of H(e) for methionine synthesis. Conversely, when SAM concentration is high (as occurs after a methionine load), inhibition of methionine synthase is accompanied by diversion of H(e) through the transsulfuration pathway by stimulation of CBS. Figure 2 illustrates the hypothetical regulation of the metabolic pathways by SAM.

Thus, SAM levels may be the key determinant of plasma H(e). Further investigation is needed on the environmental, hormonal, and other factors that affect SAM and its subsequent effects on H(e) metabolism.

# III. Nomenclature and Methodology in the Measurement of Plasma H(e)

We refer the reader to a recent review for details of sample collection (25). Blood should be collected in anticoagulant. EDTA is preferable but heparin or sodium citrate can be used. Plasma H(e) is moderately stable at 22 C and is stable for several weeks at 0-2 C. The plasma can be stored at -20 C; however, repeated thawing should be avoided (26–28). Plasma samples stored for several years have been used in

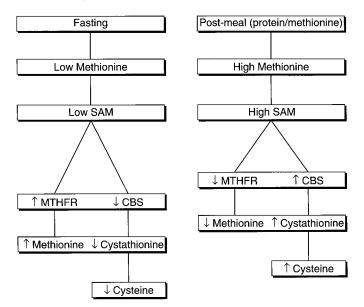


Fig. 2. Regulation of pathways of H(e) metabolism by S-adenosylmethionine (SAM). Activation of either pathway reduces H(e) levels.

some retrospective studies, and the validity of H(e) measurements in such stored plasma samples remains to be determined.

Homocysteine is the reduced (sulfhydryl) form, and homocystine is the oxidized (disulfide) form of the homologs, cysteine and cystine. For the purpose of this review, both forms of "homocyst(e)ine" will be referred to as the H(e) and hyperhomocyst(e)inemia will be referred to as HH(e). There is confusion between the American and European literature in the abbreviations used: H(e) is abbreviated as Hcy or tHcy in the European literature and H(e) in the American literature. It may be important to achieve consensus in nomenclature and use of abbreviations in this field. In patients in whom H(e) levels are normal, about 70-80% of the total H(e) is bound to protein by a disulfide linkage. With elevated H(e) levels, the percentage of H(e) in the sulfhydryl form represents an increasing percentage of the total H(e) concentration and can increase to 10-25% of the total H(e) (26).

The total H(e) is measured as the free thiol, which is obtained by reduction. This is accomplished by treatment with reducing agents such as sodium borohydride, butylphosphine, or monobromobimane (25-28). Methods used to assay H(e) include gas chromatography with mass spectroscopy, HPLC with or without fluorescence detection, and HPLC with electrochemical detection (25, 26). Methods with HPLC coupled to electron capture detectors do not require derivitization (29). There have been new developments for measuring H(e) that will allow more laboratories to measure these metabolites using immunological analyses, including an enzyme-linked immunoassay and an automated fluorescence polarization analyzer (IMX Abbott Diagnostics, Chicago, IL) (30, 31). Quality controls for the standardization of plasma H(e) measurements are not widely available. Coefficients of variation (CV) for intraassay range between 2 and 8% and for interassay the CVs are between 2 and 10%. Studies on a small population for 4 weeks have shown that withinperson variance for a 30-month period showed a high reliability coefficient, but the value was within the accepted range for the most commonly measured chemistry analytes (32).

## A. Methionine load test

The methionine load test (MLT) is essential in the comprehensive assessment of HH(e) because heterozygotes for CBS deficiency have abnormal methionine load test results in the setting of normal fasting H(e) levels (16). Conclusive evidence that an abnormal MLT represents an abnormality of the transulfuration pathway is lacking in humans. However, this hypothesis is supported by the data of Dudman et al. (33) demonstrating decreased CBS activity in most subjects with an abnormal MLT. Bostom et al. (34) have emphasized the importance of the MLT in diagnosing HH(e) in patients with vascular disease. A large proportion of such patients have normal fasting plasma H(e) with an abnormal MLT; and the rate of detection of HH(e) increases significantly when both fasting and postload HH(e) levels are assessed in a study population. The MLT is performed after an overnight fast; blood samples are collected immediately and 2–8 h after a 100 mg/kg methionine load. An abnormal load test results in a peak plasma H(e) level more than 2 sps above normal control levels.

#### IV. Determinants of Plasma Homocysteine

## A. Physiological

Several environmental factors have been found to play a role in determining the presence or absence of HH(e) (35, 36). Lussier-Cacan et al. (36) studied a large number of healthy men and women, excluding individuals with major and common disorders. They determined that gender was a major determinant of fasting plasma H(e) concentration and that women had a 21% lower concentration than men. The gender difference in H(e) concentrations between men and women persist in elderly persons, although postmenopausal women have higher concentrations than premenopausal women. Plasma H(e) concentrations increase with age and remain an independent risk factor for vascular disease in the elderly (37). The marginal folate and other vitamin deficiencies known to be common in the elderly are likely to be contributing factors to HH(e) (38, 39). There are significant negative correlations between plasma H(e) and serum folate and vitamin B<sub>12</sub> concentrations. Plasma H(e) was also highest in individuals in the lowest quartile of serum pyridoxal-5'phosphate, although this active metabolite of vitamin B<sub>6</sub> is more important in determining postmethionine load plasma H(e) than fasting H(e) (36).

Positive correlations have also been found between plasma H(e) and uric acid and creatinine concentrations that may be related to the links between H(e) metabolism with those of creatinine and uric acid (35). Plasma albumin concentration also correlates with plasma H(e) and may reflect an increase in protein-bound H(e). The exact significance of protein binding of H(e) with respect to cardiovascular disease is unknown.

In the Hordaland H(e) study, elevated plasma H(e) was associated with male gender, increasing age, smoking, hypertension, elevated cholesterol, and lack of exercise (40). In a multivariate analysis, Malinow *et al.* (41) demonstrated that systolic blood pressure, plasma uric acid, and hematocrit were predictors of concentrations of plasma H(e) in men who did not have a history of atherosclerotic disease.

It is possible that H(e) in plasma is an "acute phase reactant," rising after vascular injury. Plasma H(e) concentrations rise acutely immediately after a stroke and then decrease over several weeks (42). In contrast, plasma H(e) concentrations tend to be lower immediately after a myocardial infarction (MI) than 6 weeks later (43). The reason for this discrepancy is not clear.

Several other disease states and medications also cause elevations in plasma H(e). The recently described association between HH(e) and diabetes mellitus is described in detail below. These associated factors are outlined in Table 1. The role of genetic mutations, acute events, nutritional status, and hormonal effects are discussed below.

In summary, many nongenetic factors alter plasma H(e) levels either independently or by exacerbating genetic abnormalities in the enzyme.

Table 1. Causes of elevated plasma H(e) levels

Nutritional deficiencies

Folate

Vitamin B<sub>12</sub>

Vitamin B<sub>6</sub>

Medications

Methotrexate

Phenytoin and carbamazepine

Nitrous oxide Theophylline

Metformin

Colestipol and niacin

Disease states

Chronic renal failure

Acute lymphoblastic leukemia

Malignancies

Hypothyroidism

Type 2 diabetes

Type 1 diabetes and nephropathy

Genetic

Transsulfuration abnormalities: cystathionine  $\beta$ -synthase deficiency

Remethylation disorders (e.g., defective vitamin  $B_{12}$  transport or coenzyme synthase, defective methionine synthase)

Mutation in MTHFR

Physiological

Age

#### B. Genetics of hyperhomocysteinemia

Numerous enzyme mutations associated with HH(e) have been described including 17 CBS mutations (reaction 1) and 10 MTHFR point mutations (reaction 3a). One of the mutations for MTHFR, a common polymorphism that is present in one-third to one-half of alleles, results in a thermolabile variant of the MTHFR enzymes (1). The enzyme 5-methyltetrahydrofolate-homocysteine methyltransferase (reaction 3b) (methionine synthase) has been shown to contain one common polymorphism, but no correlation has been found between H(e) levels and genotype. Mutations of the cobalamin coenzyme synthesis enzymes (Cb C, D, E, F, or G) (20) that impair the formation of the cosubstrate methyl-B<sub>12</sub> (reaction 3b) are also rarely involved in HH(e). A comprehensive review of cobalamin coenzyme synthesis enzyme mutations is available elsewhere (44). The characteristics of CBS and MTHFR mutations are summarized in Table 2. Functional mutations are defined as those associated with increased H(e) levels. Whether any of these mutations per se directly increase the risk of arterial or venous occlusive disease remains an area of debate. Most reported studies have not evaluated the genotype of study subjects and have only correlated elevated H(e) levels with risk of vascular occlusive disease. To determine whether genotype is related to the risk of vascular disease, future studies need to correlate genotype, vitamin status (folate, vitamin B<sub>6</sub>, pyridoxal phosphate, vitamin  $B_{12}$ ), H(e) level, and vascular events. These studies may eventually show that genotype does not directly contribute to the risk of vascular disease. Rather, by increasing vitamin requirements, genotype may indirectly affect the risk of vascular disease.

1. Cystathionine  $\beta$ -synthase deficiency. Homozygous CBS mutations are the most common cause of homocystinuria. Mutations of the CBS gene result in an enzyme with decreased

Mutation	Mutation status	Homocysteine levels	Risk of vascular disease
CBS (homo)	Functional	Elevated	Increased
CBS (hetero)	Functional	Normal to elevated	Not increased
G1330A	Functional	$\mathrm{Elevated}^a$	$\mathrm{Unknown}^a$
T833C/68-bp ins	Neutral	Normal	Not increased
MTHFR (TS-homo)	Functional	Elevated	Increased
MTHFR (ts-hetero)	Neutral	Normal	Unknown
MTHFR (C677T, homo)	$Functional^b$	$\mathrm{Normal}^b$	Not increased $^b$
MTHFR (C677T, hetero)	Neutral	Normal	Not increased
Methionine synthase (A2756G transition)	Neutral	Normal	Not increased
Cobalamine coenzyme synthesis enzymes	Functional	Elevated	Increased

Table 2. Relationship of mutations in enzymes in H(e) metabolism to enzyme function, plasma H(e), and risk of vascular disease

affinity for any of its substrates: pyridoxal phosphate, serine, or H(e). Heterozygotes have been found to have variable, but less than 50%, of CBS activity (19). In addition, activity of the mutated CBS enzyme varies as a function of H(e) concentration, possibly due to steric abnormalities in the hybrid normal-mutant molecule. Furthermore, many heterozygotes have normal fasting H(e) levels, and an abnormal MLT does not necessarily imply a heterozygous state. Advances in molecular medicine, however, have now made it easier to assign a heterozygous state to an individual. As a result, CBS genotype is difficult to predict from enzyme activity and emphasizes the need for CBS genotyping in studies correlating CBS mutations with H(e) levels and risk of vascular disease (45).

The CBS gene has been assigned to the subtelomeric region of band 21q22 (46). CBS deficiency is inherited in an autosomal recessive pattern, resulting in homozygous (homocystinuria) and heterozygous (hyperhomocysteinemia) carriers (16). Sequencing of the cDNA for the CBS gene has to date identified 17 mutations (47, 48). Tsai et al. (49) have characterized three of the more common mutations as either a  $G_{919}A$ , a  $T_{833}C$ , or a  $C_{341}T$  transition. The first two mutations account for 50% of affected CBS alleles (50). Different populations demonstrate differing mutation frequencies, with the G<sub>919</sub>A transition occurring in 70% of an Irish cohort while the T<sub>833</sub>C transition occurred in 50% of a Dutch population (1). Tsai et al. (45) have estimated that the heterozygote frequency for a CBS point mutation is 1/20,000 to 1/200,000, and that 30-40% of individuals with premature vascular disease are heterozygous for CBS point mutations.

An additional mutation described by Sebastio *et al.* (51) involves a 68-bp insertion in the coding region of exon 8 of the CBS gene. This insertion mutation, which creates an alternate splice site at the intron 7-exon 8 border, has only been reported in combination with the  $T_{833}C$  missense mutation; the latter is located in *cis* 10 bp upstream of the insertion (46, 49–53). In addition, the insertion sequence contains a premature stop codon; however, Tsai *et al.* (52) reported finding only normal size RNA, implying either that splicing did not introduce the premature stop codon, or that the truncated RNA was not detectable. This insertion results in a benign mutation because the  $T_{833}C$  missense mutation is

eliminated through alternate splicing at the intron 7-exon 8 border (within the insertion there is no substitution of the 833 nucleotide). The prevalence of this mutation varies with the population studied. Tsai *et al.* (52) reported a prevalence of 11.7% in a control population (heterozygotes); the double mutation has been reported to occur in 25.8% of Northern Italians.

Another novel point mutation has been described by Kluijtmans *et al.* (54) in a partially vitamin  $B_6$ -responsive homocystinuric patient. This mutation was a G1330A transition and was unique in that it abrogated CBS responsiveness to SAM. Thus, in contradistinction to the other 17 identified point mutations that result in an altered protein that attenuates the catalytic activity of CBS, this mutation interferes with the regulatory domain of the CBS protein. (54)

2. MTHFR deficiency. MTHFR catalyzes the reduction of 5,10methylenetetrahydrofolate to 5-methyltetrahydrofolate (reaction 3a, Fig. 1). The gene is located on chromosome 1p36.3; 10en different mutations for the MTHFR gene have been identified from isolated cDNA (55). Nine of these mutations result in thermostabile mutations. The two most common of these mutations are a  $C_{559}T$  transition, which converts an arginine codon to a termination codon, and a G<sub>482</sub>A transition, which converts an arginine to a glutamine residue (55). Another mutation in the MTHFR gene, a  $C_{677}$ T transition, results in a thermolabile variant (56-58) of the enzyme. This autosomal recessive mutation creates a Hinfl restriction site and substitutes an alanine for a valine residue in the MTHFR protein (57). The allele frequency of this polymorphism varies slightly with the population studied, but approximately 8% of studied populations are homozygous for this autosomal recessive polymorphism. The homozygous thermolabile MTHFR polymorphism is characterized by an enzyme activity of about 30% of normal. Heat inactivation at 46 C distinguishes the mutant from the normal MTHFR enzyme. The enzyme is considered thermolabile when there is less than 20% residual activity after heating to 46 C. HH(e) does not occur in individuals heterozygous for this polymorphism and appears only to occur in homozygotes who are concomitantly folate deficient (2, 18, 56-62). Some data suggest that

<sup>&</sup>lt;sup>a</sup> Data from a single case report, so cannot generalize conclusions.

<sup>&</sup>lt;sup>b</sup> Data from reported studies that have evaluated both genotype and vitamin status show that in folate-replete patients the risk of coronary artery disease and ischemic stroke is not increased. Some authors have suggested this mutation may increase the folate requirement. For this reason the mutation is listed as functional. In patients with low to low normal folate levels, H(e) levels are increased.

homozygotes for this mutation may actually have a higher folate requirement (59–61).

3. Methionine synthase. Methionine synthase, localized to chromosome 1q42.3–43 (63), catalyzes the transfer of a methyl group from 5-methyltetrahydrofolate to H(e) via the intermediary methyl-B12. Methionine synthase contains one common polymorphism that results in a A2756G transition. This polymorphism does not correlate with H(e) level and does not appear to be a risk factor for vascular occlusive disease, nor neural tube defects (64). Other rare clinical conditions have reported functional methionine synthase deficiency. These have resulted from mutations in cobalamine coenzyme synthesis that has resulted in abnormal methyl B-12 production, a cosubstrate in the methionine synthase reaction (44, 64–69). No discrete mutations of methionine synthase itself that are associated with hyperhomocysteinemia have been described.

4. Does MTHFR polymorphism increase the risk of MI? Data conflict over the risk of vascular occlusive disease and hyperhomocysteinemia in patients with the homozygous thermolabile polymorphism for MTHFR (1). In contrast, no increased risk for vascular occlusive disease or hyperhomocysteinemia is present in heterozygotes for this polymorphism. Kluijtmans et al. (1) screened 60 cardiovascular patients and 111 controls and found that 15% of the cardiovascular patients vs. 5% of controls were homozygous for the thermolabile MTHFR polymorphism (1). This translated into a 3-fold risk of premature cardiovascular disease for the homozygous polymorphism. Kluijtmans and coworkers did not evaluate for folate levels in their study. In contrast, Ma et al. (19) in the Physicians Health Study reported that the thermolabile MTHFR polymorphism in homozygotes was associated with hyperhomocysteinemia only if folate levels were concomitantly low (19). An increased risk for MI, solely on the basis of the homozygous polymorphism, was not found.

In another study by Legnani et al. (18), the presence of the homozygous polymorphism did not increase the risk of thrombosis over control patients (18). Christensen et al. (62) showed that the incidence of the homozygous polymorphism and a normal genotype was not different between patients with coronary artery disease and healthy controls, but patients with the homozygous polymorphism did have higher H(e) levels if the serum folate levels were below the median value. Jacques et al. (61) reported that homozygotes for the polymorphism had H(e) levels 24% greater than those with normal genotype if the serum folate levels were <15.4 nmol/liter. Malinow et al. (59) suggested that homozygotes with the polymorphism may have an increased folate requirement, and Ali et al. (60) reported that serum folate levels are lower in individuals homozygous for the MTHFR polymorphism. The recent large prospective study by Folsom et al. (12) provides additional evidence against the MTHFR mutation being associated with cardiovascular disease (see below for details).

Taken together, these studies suggest that homozygotes may have an increased folate requirement, and that in the presence of normal folate levels homozygotes are not at increased risk for hyperhomocysteinemia or vascular occlusive disease.

#### C. Nutritional

1. Folate. Plasma H(e) is thus a sensitive biomarker of folate deficiency. Lewis et al. (70) demonstrated that in subjects with plasma folate concentrations above 15 nmol/liter the H(e) concentration is on a low, normal plateau. At lower levels of plasma folate, the plasma H(e) concentration increases steadily (38). Rimm et al. (71) reported in a large prospective study of women that the 20% who had the highest consumption of folate (almost all of whom consumed multivitamin tablets) had significantly less cardiovascular disease than the lowest 20% (very few of whom consumed multivitamins). However, there have been no randomized control trials of the effect of folic acid alone on cardiovascular disease.

There is controversy about the exact amount of supplemental folic acid that is required to reduce plasma H(e). Shimakawa *et al.* (72) reported that people who use multivitamin supplements have significantly lower plasma H(e) concentrations than nonusers. The RDA for folate in the United States is 200  $\mu$ g/day. It has been suggested that an intake of 400  $\mu$ g of folic acid above the dietary level will prevent birth defects. Such an increased supplementation will also significantly decrease plasma H(e) concentrations in most of the population (32).

Recently, Malinow *et al.* (73) reported that breakfast cereal fortified with 400  $\mu$ g of folic acid was adequate in lowering plasma H(e) by more than 100  $\mu$ g. Increasing the dose of folate supplementation to 600  $\mu$ g had very little additional effect.

Schorah et al. (74) carried out a randomized double-blind placebo-controlled study in 119 healthy volunteers, whose intake of fortified or supplemental folic acid was low (74). Volunteers were randomized to receive unfortified cereals, or cereals fortified with 200  $\mu$ g of folic acid per portion, with or without other vitamins, for up to 24 weeks. There were no significant changes in plasma H(e) in those eating unfortified cereals. Folic acid fortification of cereals led to significant increases in serum folate (66%), and red cell folate (24%), and a decrease in plasma H(e) (10%) (74). There were no changes in vitamin  $B_{12}$  or cysteine. The H(e) decrease was primarily seen in those who initially had the highest plasma H(e) or the lowest serum folate. Thus, if H(e) is found to be a causative risk factor in occlusive vascular disease, food fortification with physiological levels of folic acid should have a significant impact on the prevalence of the disease in the general population.

de Bree *et al.* (75) evaluated possible inconsistencies between recommended, actual, and desired folate intake in European adult populations. They concluded that in Europe, mean dietary folate intake in adults is 291  $\mu$ g/day (range 197–326) for men and 247  $\mu$ g/day (range 168–320) for women. The recommended intakes vary between 200–300  $\mu$ g/day (men) and 170–300  $\mu$ g/day (women–with higher recommended intakes during pregnancy). The mean dietary folate intake in Europe is in line with recommendations, but the desired dietary intake of more than 350  $\mu$ g/day to pre-

A meta-analysis of randomized trials with folic acid (76) reported that supplementation of the typical Western diet with 0.5–5 mg folic acid along with 0.5 mg vitamin  $B_{12}$  would be expected to reduce plasma H(e) concentrations by one fourth to one third (*e.g.*, from about 12  $\mu$ mol/liter to 8 to 9  $\mu$ mol/liter). Whether such a "population approach" to prevention of HH(e) and associated cardiovascular disease will be effective requires further investigation.

2. Vitamins  $B_6$  and  $B_{12}$ . It is well recognized that pyridoxal-5'-phosphate, the active form of vitamin B<sub>6</sub> pyridoxine, is an important cofactor in the conversion of H(e) to cystathionine. The enzyme involved in this reaction is CBS. Because vitamin B<sub>6</sub> is not involved in remethylation, pyridoxine deficiency will only result in HH(e) when the transsulfuration pathway is activated such as after a methionine load. Thus, the positive MLT that was designed to detect heterozygous CBS defects may result from vitamin B<sub>6</sub> deficiency. Rats fed vitamin B<sub>6</sub>-deficient diets for 4 weeks develop HH(e) after a methionine load (77). However, clinical studies have demonstrated a lack of a relationship between plasma H(e) and vitamin B<sub>6</sub> status. In human subjects made B<sub>6</sub> deficient by diet (78), fasting plasma H(e) levels remain normal. In patients with asthma treated with theophylline (a vitamin B<sub>6</sub> antagonist), plasma H(e) was significantly higher after a methionine load when compared with controls (79). This abnormality was corrected by treatment with pyridoxine supplementation for 6 weeks. Thus, pyridoxine deficiency should be excluded in patients who have HH(e) after a methionine load – a common abnormality in patients with premature cardiovascular disease.

The importance of vitamin  $B_{12}$  in the remethylation of H(e)to methionine is well recognized, and HH(e) is a feature of vitamin B<sub>12</sub> deficiency (19, 39). However, the relationship between vitamin B12 intake/plasma levels and HH(e)related cardiovascular disease is less well defined. In the Framingham study, plasma H(e) exhibited a strong inverse association with plasma folate but a weaker association with plasma vitamin B<sub>12</sub>. Subjects in the lowest decile of plasma B<sub>12</sub> had significantly higher plasma H(e) when compared with those in the highest decile (80). Homocysteine was also inversely associated with intakes of folate and vitamin B<sub>6</sub>, but not vitamin  $B_{12}$ . Many of the case control studies of HH(e) in patients with vascular disease have excluded subjects with vitamin B<sub>12</sub> deficiency. As discussed in the treatment section below, treatment of patients with HH(e) with vitamin B<sub>12</sub> seems to have very little impact on plasma H(e).

## D. Hormones and H(e) metabolism

Studies have been done to clarify the role of hormonal changes in the regulation of H(e) metabolism and in causing HH(e). As indicated above, genetic abnormalities do not fully explain the relatively high prevalence of HH(e) in patients with vascular disease. This is particularly true of post methionine HH(e), which is not often associated with genetic mutations of the CBS gene. Dudman *et al.* (81) suggested that depressed CBS activity in several of their patients was as-

sociated with causes other than abnormal CBS protein. A reasonable alternative explanation for the depressed CBS activity would be that it was metabolically down-regulated. Such down-regulation of CBS as well as other enzymes in H(e) metabolism may be mediated by hormonal changes.

1. Estrogen. There is some data to suggest that estrogen has an effect on H(e) metabolism, although the mechanism involved is not clear. Plasma H(e) in pregnant women decreases to almost half that in nonpregnant women (82, 83), suggesting a possible effect of estrogen, although other factors such as increased vitamin intake obviously play a role.

Estrogen may have an effect on the activity of some of the enzymes in H(e) metabolism (84), although further studies are needed to confirm this effect.

Several investigators have examined the effect of menopausal status on plasma H(e). Wouters et al. (85) measured fasting and postmethionine plasma H(e) concentrations in premenopausal and postmenopausal healthy women without a history of vascular disease. Fasting and postmethionine plasma H(e) was significantly higher in postmenopausal women as compared with premenopausal women. The difference appears to be too large to be explained as an effect of age alone [the increase in plasma H(e) over a decade of life is modest] and is more likely to be related to hormonal status. In premenopausal women, postmethionine plasma H(e) was negatively and significantly correlated to serum  $17\beta$ -estradiol. Andersson et al. (86) reported that levels of fasting and postload plasma H(e) in premenopausal women did not differ from those of men of similar age. In contrast, in postmenopausal women, the level of fasting plasma H(e) was actually lower than that of men of similar age. This was because, in that study, fasting values in men increased with age and was associated with a decrease in serum vitamin  $B_{12}$ , folate, and pyridoxal 5-phosphate.

The rise in H(e) levels after menopause may partly explain the sharp rise in cardiovascular disease that occurs in this age group, and its attenuation by hormone replacement therapy (HRT). However, many cardiovascular risk factors improve with estrogen and further investigation is required to determine whether the lower incidence of vascular disease in premenopausal women and in women taking HRT may be related to the lower concentrations of plasma.

In a prospective study, van der Mooren *et al.* (87) measured fasting serum H(e) during HRT in postmenopausal women. The mean serum H(e) decreased by approximately 11% with a greater decrease (17%) in those women who had a high H(e) level before treatment with very little change in those who had a low level. There are well recognized mechanisms to explain the beneficial effect of HRT on cardiovascular risk, including effects on lipids and fibrinolysis. However, it is possible that a lowering of plasma H(e) also contributes to this benefit.

Oral contraceptive agents, in contrast to HRT, do not affect biochemical folate indices and H(e) concentrations in young women (88).

Tamoxifen, an estrogen antagonist with partial agonist activity, decreased plasma H(e) by a mean of 30% after 9–12 months treatment in postmenopausal women with breast cancer (89). These changes were independent of the tumor

burden. These data, in combination with the effect of estrogen, suggest that there is an estrogen receptor-mediated H(e)-lowering effect. In addition, there may be an indirect effect related to a modest elevation in plasma folate concentrations with tamoxifen (90).

2. Testosterone. Zmuda et al. (91) studied the effect of supraphysiological doses of testosterone on fasting H(e) in normal male weight lifters. Plasma H(e) levels were not significantly altered where testosterone was given alone or together with testolactone. It is likely, therefore, that short-term, high-dose testosterone administration does not affect fasting plasma H(e) levels in normal men. However, the study of Zmuda et al. has limitations in that only fasting plasma H(e) was measured and, therefore, the authors may have underestimated an effect of testosterone on postmethionine H(e).

Since testosterone has a negligible effect on plasma H(e) concentrations, an alternative explanation is needed for the difference in plasma H(e) concentrations between men and women. Another possible explanation for this gender difference is that creatine-creatinine production is directly coupled to *S*-adenosylhomocysteine generation from SAM (89). Lean body mass, muscle mass, amino acid turnover, and creatine-creatinine production tend to be higher in men than in women, all of which may explain in part the higher plasma H(e) in men. Plasma H(e) concentrations correlate directly with serum creatinine concentrations in men and women (90). In fact, Brattstrom *et al.* (92) demonstrated that the gender difference in plasma H(e) disappeared when men and women were matched for serum creatinine concentrations.

In male-to-female transsexuals, plasma H(e) decreased significantly while it increased in female-to-male transsexuals (93). Thus, the difference in sex steroid milieu between men and women may be an important factor in determining the sex difference between men and women in plasma H(e) levels. Interpretation of these data, however, is complex because of the major changes in sex hormone status occurring in transsexuals. In this context, however, it may be relevant that when inbred adult male rats were treated with estrogen, the plasma H(e) concentration fell by approximately 30%, indicating a direct effect of estrogen on lowering plasma H(e) (93). Plasma H(e) concentrations are also lowered in male rats treated with cortisol, estradiol, or a combination of both (94).

3. Thyroid hormones. An elevated plasma H(e) in hypothyroidism has previously been reported (95). In a recent study, Nedrebo et al. (96) demonstrated that plasma H(e) was significantly higher in patients with hypothyroidism (96). In contrast, plasma H(e) in hyperthyroid patients did not differ significantly from that of controls (96). Thus, HH(e) may exacerbate the increased risk of cardiovascular disease in hypothyroidism that is traditionally attributed to lipid changes.

## V. Homocysteine and Diabetes Mellitus (DM)

Macrovascular disease is highly prevalent is patients with type 2 diabetes, has an early onset, and progresses at a more rapid rate than in patients without diabetes. While traditional cardiovascular risk factors, such as hypertension and dyslipidemia, are more common in patients with type 2 diabetes, they do not fully explain this acceleration in macrovascular disease (97). Recent data suggesting that H(e) may be a risk factor for cardiovascular disease in this population are, therefore, important (98–100).

Munshi et al. (98) reported that HH(e) after a methionine load occurred in approximately 40% of patients with type 2 DM who had macrovascular disease, but was normal in patients with insulin-dependent diabetes (type 1 DM). Patients with overt nephropathy had been excluded from that study, and only patients under the age of 60 were studied. In contrast, Araki *et al.* (101) found that fasting plasma H(e) was elevated in some patients with type 2 DM, and this abnormality was corrected with vitamin  $B_{12}$  injections (101). It is possible that these investigators had identified, by chance, patients with concomitant diabetes and B<sub>12</sub> deficiency. In a population-based study of HH(e) and the risk of cardiovascular disease, 631 patients were stratified according to age, sex, and glucose tolerance (100). The authors also investigated the combined effect of HH(e) and DM with regard to cardiovascular disease. Fasting HH(e) was seen in 25.8% of individuals. After adjustment for age, sex, hypertension, hypercholesterolemia, diabetes, and smoking, the odds ratios (ORs; 95% confidence intervals) per 5 µmol/liter increment in H(e) were 1.44 (1.10-1.87) for peripheral arterial, 1.25 (1.03–1.51) for coronary artery, 1.24 (0.97–1.58) for cerebrovascular, and 1.39 (1.15-1.68) for any cardiovascular disease. After stratification by glucose tolerance category and adjustment for the classic risk factors and serum creatinine, the ORs per 5 μmol/liter increment in H(e) for any cardiovascular disease were 1.38 (1.03–1.85) in normal glucose tolerance, 1.55 (1.01–2.38) in impaired glucose tolerance, and 2.33 (1.11–4.90) in non-insulin-dependent diabetes mellitus (P = 0.07 for interaction)(100). Thus, the magnitude of the association between HH(e) and cardiovascular disease is stronger (1.6-fold) for patients with type 2 diabetes than in nondiabetic subjects. Subjects in that study were 50- to 75 yr old, and these findings may relate to an interaction of age, insulin resistance, and diabetes on both H(e) metabolism and cardiovascular disease.

Type 2 DM and HH(e) are both associated with increased lipid peroxidation (oxidative stress) (102, 103). In a study to determine whether the coexistence of elevated H(e) levels stimulate oxidative stress further than that caused by diabetes alone, plasma concentrations of thiobarbituric acid reactive substances (TBARS), an index of lipid peroxidation, were measured in patients with type 2 DM. Plasma TBARS concentrations were elevated in diabetics with vascular disease. The additional presence of hyperhomocysteinemia was not associated with a further increase in plasma TBARS concentrations (104). Thus, diabetes maximally stimulates oxidative stress, and any further acceleration of vascular disease in patients who have coexistent hyperhomocysteinemia is mediated through mechanisms other than lipid peroxidation

Other studies of patients with type 1 DM have confirmed that plasma H(e) levels are normal early in the course of the disease. However, Hultberg *et al.* (105) reported that basal plasma H(e) concentrations were higher in patients who developed nephropathy and had an elevated plasma creat-

inine. These workers recently demonstrated that diabetic patients with the lowest age at onset and poorest metabolic control were most prone to a rapid increase in plasma H(e) (106). They concluded that this increase in plasma H(e) could at least partially be explained by marginal deficiency of blood folate concentrations.

Hofmann *et al.* (107) reported that plasma H(e) levels, both fasting and after a methionine load, were elevated in patients with type 1 DM who had microalbuminuria and were higher still in patients who had overt proteinuria. The patients with type 1 DM and HH(e) also had higher plasma thrombomodulin (TM) levels (indicating endothelial cell damage) and a higher prevalence of late diabetic complications including macrovascular disease.

These workers also reported a significant relationship between plasma H(e) concentrations and urinary albumin excretion rate (Fig. 3), as well as a significant relationship between plasma H(e) and plasma TM (Fig. 4). Thus, HH(e) represents an additional cardiovascular risk factor in patients with microalbuminuria, perhaps contributing to the enhanced risk of cardiovascular disease in this subpopulation of people with diabetes.

Hofmann *et al.* also demonstrated an interaction of H(e) and advanced glycation end products. In human umbilical vein endothelial cells in culture, there was an increased release of TM only when AGE-albumin was added before H(e), indicating a synergistic interaction between advanced glycation end products and H(e), which might contribute to the cardiovascular complications in patients with diabetes (Fig. 5). However, the concentrations of H(e) used in the experiment was more than 100 times that found in plasma in patients with diabetes and HH(e).

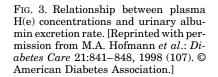
The data of Hofmann *et al.* are limited by the fact that there was a much higher prevalence of hypertension in the hyperhomocysteinemic group than in those with normal H(e). It was, therefore, suggested that these subjects had already developed endothelial damage as a result of their hypertension (108). Nevertheless, the possibility remains that renal MTHFR activity may be impaired even in the early stages of diabetic nephropathy, *i.e.*, microalbuminuria (109).

Three recent reports have confirmed the association between plasma H(e) and albumin excretion rate in patients with DM. Chico *et al.* (110) measured fasting plasma H(e) in

165 diabetic patients and control subjects. Patients with type 2 DM had higher plasma H(e) than controls, whereas patients with type 1 DM did not. Univariate correlations and multiple regression analysis showed albumin excretion rate to be strongly related to plasma H(e) (110). In addition, patients with type 2 DM with hypertension had higher plasma H(e) than patients without hypertension (110). This finding may have been related to the fact that the patients with hypertension had more severe biochemical markers of nephropathy, with a higher albumin excretion rate.

Lanfredini *et al.* (111) recently studied the relationship between homocyst(e)inemia and microalbuminuria in non-insulin-dependent diabetes mellitus (NIDDM) patients. There was a significant correlation between urinary albumin excretion and fasting and postmethionine load plasma H(e) in NIDDM patients. Microalbuminuric NIDDM patients had higher fasting plasma H(e) than normoalbuminuric patients. These investigators also reported that patients with NIDDM and HH(e) had higher diastolic and mean arterial blood pressure than those with a normal plasma H(e). There was a significant correlation between plasma folate and mean arterial pressure (112).

Hoogeven et al. (113) recently reported a relationship between serum H(e) level, protein intake, and risk of microalbuminuria. In a population-based study of 680 subjects stratified according to age, sex, and glucose tolerance, serum total H(e) was positively associated with the presence of microalbuminuria independent of other risk factors including diabetes, hypertension, protein intake, and renal function. For each 5  $\mu$ mol/liter increase in serum H(e), the risk of microalbuminuria being present increased by about 30% (114). The authors suggested that hyperhomocysteinemia may partly explain the link between microalbuminuria and the increased risk of cardiovascular disease. However, it is important to recognize that microalbuminuria is associated with several other cardiovascular risk factors, all of which could contribute to increased cardiovascular disease. Nevertheless, in a recent study, Stehouwer et al. (114) demonstrated that plasma H(e) predicts mortality in patients with NIDDM, with or without albuminuria. Further investigation is required to determine the exact role that HH(e) plays in worsening cardiovascular disease in patients with microalbuminuria and overt proteinuria.



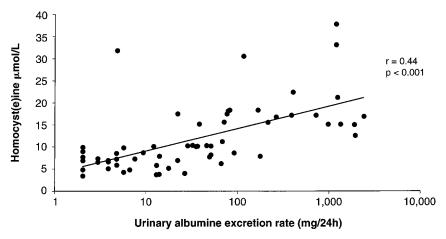


FIG. 4. Relationship between plasma H(e) and plasma TM in patients with type 1 diabetes. [Reprinted with permission from M.A. Hofmann *et al.: Diabetes Care* 21:841–848, 1998 (107). © American Diabetes Association.]

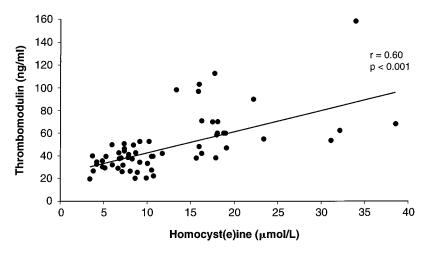
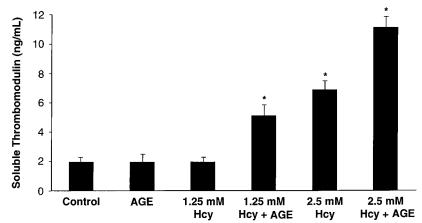


FIG. 5. Effect of the addition of H(e) and HH(e) on human umbilical vein endothelial cells in culture. The release of TM into culture medium has been measured. [Reprinted with permission from M.A. Hofmann et al.: Diabetes Care 21: 841–848, 1998 (107). © American Diabetes Association.]



# A. Hyperhomocysteinemia, renal failure, and diabetic nephropathy

The effect of renal disease on H(e) metabolism has been comprehensively reviewed (115). This subject is of considerable importance to endocrinologists and nephrologists who treat patients with diabetes, as they are at high risk of developing renal failure. Studies to determine whether the treatment of HH(e) in patients with renal insufficiency will reduce morbidity and mortality are thus of considerable importance to physicians treating patients with diabetes.

Recognition of the importance of renal impairment and proteinuria in determining plasma H(e) is vital to the clinician in interpretation of laboratory results. While some of this elevation may be due to decreased clearance of H(e), other mechanisms may also be responsible. Intact kidneys have considerable H(e)-metabolizing capacity (116), impairment of which may be an important determinant of the marked HH(e) frequently observed in end-stage renal disease. High-dose multiple vitamin treatment has been shown to lower plasma H(e) in dialysis patients, although levels remain elevated in many patients (117). Hyperhomocysteinemia has also been demonstrated after successful renal transplantation (118) and may be exacerbated by cyclosporin (119).

## B. Effect of glucose and insulin on H(e) metabolism

It is well recognized that insulin has important effects on protein and amino acid metabolism. However, its effect on H(e) metabolism has not been well studied.

During a hyperinsulinemic euglycemic clamp, the plasma H(e) response to acute hyperinsulinemia was heterogenous (120). Plasma H(e) levels fell by approximately 20% in normal subjects but did not do so in insulin-resistant patients with type 2 DM (Fig. 5) (121). These data suggest that resistance to the effects of insulin on glucose disposal may be associated with resistance to the suppressive effect of insulin on H(e) levels in patients with type 2 DM. Such a resistance to insulin's effect on H(e) may contribute to the increased cardiovascular disease associated with the insulin resistance syndrome and type 2 DM (121).

It is well recognized that insulin decreases plasma methionine, the methionine being incorporated into newly synthesized protein (122). Plasma amino acid concentrations, including methionine, fall significantly after an oral glucose load and the subsequent rise in endogenous plasma insulin (123). However, in patients with diabetes, this fall in amino acids does not occur, indicating a possible resistance to insulin's effect on amino acids in diabetics (123).

The insulin-induced fall in plasma methionine concentra-

tions may be mediated through increased tissue uptake of methionine or metabolism via the transsulfuration pathway resulting in increased levels of H(e). Intracellular and plasma H(e) levels may then rise, particularly if a defect in CBS action exists. Further study is required to determine the effect of insulin on SAM, the key determinant of the relative activity of the transmethylation and transsulfuration pathways (see above).

A recent study has demonstrated an inverse relationship between plasma H(e) and insulin sensitivity in women with preeclampsia (124). However, because of the multiple abnormalities in metabolism and endothelial function present in preeclampsia, it is impossible to determine whether a cause-effect relationship exists between the two variables. Further work is needed in other insulin-resistant states to determine whether insulin resistance causes HH(e).

To examine the effects of hyperinsulinemia induced by a high-fat sucrose (HFS) diet on H(e) metabolism, we measured hepatic mRNA and activity of two key enzymes involved in this metabolic pathway: MTHFR and CBS, in an insulin-resistant rat model (125). Fischer rats made insulin resistant by a HFS diet were examined at 6 months and 2 yr of age and compared with control rats fed a low-fat, complexcarbohydrate (LFCC) diet. At the end of 6 months, the HFSfed rats were heavier than the LFCC rats and had hyperinsulinemia. The plasma H(e) concentrations were elevated in the HFS-fed rats (10.77  $\pm$  0.9 vs. 6.89  $\pm$  0.34  $\mu$ mol/liter; P <0.01). Hepatic CBS mRNA and enzyme activity was significantly lower in the HFS group compared with control. In contrast, hepatic MTHFR enzyme activity and mRNA levels were significantly elevated in the HFS group compared with control. There were significant positive correlations between plasma H(e) and fasting plasma, body weight, and MTHFR activity. There were significant negative correlations between plasma insulin and CBS activity and between CBS and MTHFR activities. The latter inverse relationship supports the hypothesis of Selhub and Miller (21) (Fig. 2) and suggests that insulin's effect on H(e) metabolism may be mediated through SAM. In conclusion, HFS feeding leads to hyperinsulinemia, which may be associated with hyperhomocysteinemia, secondary to down-regulation of CBS message and activity, and may thus contribute to the accelerated macrovascular disease associated with type 2 diabetes. However, we wish to emphasize that the above study was carried out in rodents, and the conclusions may not be applicable to humans.

The effect of drug treatment of diabetes on H(e) metabolism has not been well studied. Evidence suggests that insulin and sulfonylurea treatment do not alter plasma H(e) (101). Hoogeveen *et al.* (126) reported that metformin does not increase plasma H(e). However, in a clinical trial of metformin to assess its lipid-lowering effects in nondiabetic patients with coronary artery disease, there was a moderate but significant increase (7.2% at 12 weeks and 13.8% at 40 weeks) in plasma H(e) with metformin treatment. This was associated with a significant fall in serum  $B_{12}$  levels (127).

In summary, HH(e) is not uncommon in patients with diabetes and may play a role in the accelerated cardiovascular disease in these patients. The mechanism for this increased prevalence of HH(e) is not clear but data suggest a role for insulin in regulation of plasma H(e) levels and that

insulin resistance may lead to HH(e). Tables 3 and 4 summarize current knowledge of the possible interaction between diabetes and HH(e).

## VI. Hyperhomocysteinemia and Cholesterol Metabolism

Evidence is now emerging that the risk for vascular disease from HH(e) is independent of any coexistent abnormalities in lipid metabolism. However, the two risk factors often coexist in the same individual and may have an additive effect.

In 482 patients already at high risk for atherosclerotic vascular disease by virtue of hyperlipidemia, 3.7% had high plasma H(e) (128). In hyperlipidemic patients, the relative risk of atherosclerotic events for the 80th percentile of plasma H(e) was 2.8-fold greater than that seen for the 20th percentile. Furthermore, it was possible to reduce H(e) concentrations in this hyperlipidemic population with vitamins, suggesting a possible therapeutic approach for multiple risk factor intervention. Plasma H(e) has been shown to be associated with a parental history of cardiovascular disease in children with familial hypercholesterolemia (129).

An elevated high-density lipoprotein cholesterol (HDL cholesterol) is well accepted as a protective factor against atherosclerosis. However, a high HDL cholesterol is not necessarily protective in the setting of an elevated plasma H(e) (130).

Considerable *in vitro* data exist to suggest that H(e) may interact with an elevated cholesterol by increasing oxidation of LDL. However, this area remains controversial. It has been suggested that H(e) inhibits glutathione peroxidase activity *in vitro* and leads to a reduction in steady state mRNA levels for the intracellular isoform in endothelial cells (131). Glutathione peroxidase is a member of the antioxidant enzyme family that catalyzes the reduction of lipid peroxides (132).

However, *in vitro* data are not supported by clinical *in vivo* studies, perhaps related to imprecisions in measuring oxidant stress *in vivo*, with current technology. As discussed above in the setting of diabetics with vascular disease, plasma TBARS are not elevated above those levels caused by diabetes alone. These data are supported by two other studies. In one, LDL isolated from two patients with homocystinuria showed a similar extent of copper-catalyzed oxidation as LDL from a group of healthy control subjects (133). Cordoba-Porras *et al.* (134) investigated the existence of oxidized LDL and the susceptibility to oxidation of lipoproteins in six patients with homocystinuria. The proportion of electronegative LDL and concentration of TBARS did not differ between patients and controls (134). Thus, more studies are

TABLE 3. Homocysteine and diabetes

Risk factor for vascular disease
Insulin regulates plasma levels
Elevated in type 2 diabetes
Associated with insulin resistance
Increased in type 1 only with microalbuminuria
Associated with elevated thrombomodulin in type 1 with microalbuminuria
Increased with renal impairment

needed on lipoprotein susceptibility to oxidation in patients with HH(e).

## VII. Hyperhomocysteinemia in Premature Vascular Disease

It is well recognized that children who have homozygous homocystinuria develop premature and severe vascular disease. These patients are prone to sudden death in young adulthood or even in childhood. We will now review the data suggesting that moderately elevated H(e) levels are a risk factor for occlusive arterial disease.

## A. Epidemiological and prospective studies

In the first prospective epidemiological study of H(e) levels as a cardiovascular risk factor, 14,916 male physicians with no prior vascular disease provided plasma samples at baseline and were followed for 5 yr (6). Plasma H(e) levels in 271 physicians who developed a MI were significantly higher than in paired controls. The relative risk for MI in those physicians in the 95th percentile for H(e) levels was 3 times greater than those in the 10th percentile, even after adjustment for other known risk factors for vascular disease (95% confidence interval, 1.3–8.8) (6).

In 1998, Wald *et al.* (3) reported the British United Provident Association study, a prospective nested case-control study of 21,520 men between the ages of 35 and 64. Homocysteine levels were assayed from stored samples in 229 subjects who died of ischemic heart disease, and 1,126 agematched controls. Homocysteine levels were higher in the group that died of ischemic heart disease than in the controls. For men with serum H(e) levels in the highest quartile, the increased risk was 2.9 (after adjustment for other factors) than for men in the lowest quartile (3).

In a prospective, nested case-control study within the British Regional Heart Study cohort, Perry et al. (135) examined the association between serum total H(e) concentration and stroke. Serum was saved from 5,661 men, aged 40-59 yr, randomly selected from general practices. During follow-up of up to 12 yr, there were 141 incident cases of stroke among men with no history of stroke at screening. Serum H(e) was measured in 107 cases and 118 control men (matched for age group and town, without a history of stroke at screening, who did not develop a stroke or MI during follow-up). Serum H(e) concentrations were significantly higher in cases than controls. There was a graded increase in the relative risk of stroke in the second, third, and fourth quarters of the serum H(e) distribution (odds ratios 1.3, 1.9, 2.8; trend, P = 0.005) relative to the first. Adjustment for age group, town, social class, body mass index, hypertensive status, cigarette smoking, forced expiratory volume, packed-cell volume, alcohol intake, diabetes, HDL cholesterol, and serum creatinine did not attenuate the association (135). These findings add to the evidence that H(e) is a strong and independent risk factor for stroke.

#### B. Studies in patients with established vascular disease

Several studies have attempted to establish the prevalence of HH(e) in patients with premature and accelerated vascular

disease. Interest in this field was triggered by the first report by Boers *et al.* (17) who found elevated plasma H(e) after a methionine load in 28% of patients with peripheral vascular and cerebrovascular disease (17). Even after adjustment for other risk factors, plasma H(e) has been found to be significantly higher in patients with peripheral vascular disease compared with healthy individuals (136). Elevations in peak H(e) after a methionine load occur in 28–42% of patients with vascular disease but rarely, if ever, in normal subjects (8, 34, 98). It would appear that the MLT delineates the "at risk" population better than basal levels (34).

In a meta analysis of 27 studies of H(e) in atherosclerotic vascular disease, Boushey *et al.* (7) concluded that elevations of H(e) were an independent risk factor for arteriosclerosis. In addition, approximately 10% of the population's coronary artery disease risk appears attributable to H(e). The odds ratio for development of coronary artery disease from increased plasma H(e) at a level of 5  $\mu$ mol/liter above normal is 1.6 for men and 1.8 for women.

Fermo *et al.* (137) studied patients below the age of 45 with both venous thrombosis and arterial occlusive disease and found moderate HH(e) in 13.1% and 19.2% of patients, respectively. The prevalence of HH(e) was almost twice as high after a methionine load than when based upon fasting levels. Other studies have confirmed the high prevalence of HH(e) in early onset and recurrent venous thrombosis (10).

Compelling recent evidence comes from the multicenter study done in nine European countries (11). In this study, 750 cases of vascular disease were compared with 800 controls of both sexes younger than 60 yr of age. The relative risk for vascular disease in the highest quintile of plasma H(e) was 2.2 when compared with the lower four quintiles. Methionine loading identified an additional 27% of at risk cases. A dose response effect was noted between total plasma H(e) and risk. The risk was similar to and independent of other risk factors including smoking and hyperlipidemia. An elevated plasma H(e) had a multiplicative effect on risk in smokers and subjects with hypertension. Furthermore, subjects taking vitamins appeared to have a substantially lower risk of vascular disease when compared with nonusers of vitamin supplements. This difference was attributed to lower plasma H(e) levels.

A recent study has established the predictive value of plasma H(e) levels in patients with established coronary artery disease (138) in 587 patients with angiographically confirmed coronary artery disease (many of whom underwent bypass surgery and angioplasty). A strong, graded relationship between plasma H(e) levels and overall mortality was found over a 4-yr period. Less than 4% of patients with a plasma H(e) below 9  $\mu$ mol/liter died, as compared with nearly 25% of those with a plasma H(e) greater than 15  $\mu$ mol/liter. The relation of H(e) levels to mortality remains strong after adjustment for other potential confounding variables. It is important to emphasize that plasma H(e) levels are a continuous variable in the population studied, and there is no threshold above which the risk for mortality rises.

In another recent case-controlled study an elevated fasting and postload plasma H(e) showed a positive association with risk of severe coronary atherosclerosis (139). This association

existed over a wide range of plasma H(e), without a clear cut-off point below which there was no increased risk.

Hyperhomocysteinemia appears to have its strongest association with carotid artery disease (140). This finding was also highlighted in a cross-sectional analysis of 1,041 elderly subjects in the Framingham Heart Study. A 2-fold increase in the incidence of carotid disease was seen in patients with the highest plasma H(e) concentrations when compared with those with the lowest concentrations (5). Furthermore, plasma concentrations of folate and pyridoxal-5'-phosphate were also inversely associated with carotid artery stenosis. The Atherosclerosis Risk in Communities Study (ARIC) has reported higher plasma H(e) in subjects with carotid intimalmedia thickening (an early marker of atherosclerosis) when compared with matched controls who did not have such thickening (141). Atherosclerotic disease in the aorta as assessed by transesophageal echocardiography also correlates with plasma H(e) (142).

Bots et al. (143) examined the relationship of H(e) to MI and stroke among older subjects in a nested case-control study of a subset of participants in the Rotterdam Study. One hundred four patients with a MI and 120 with a stroke were compared with 533 control subjects drawn from the study base, who were free of MI and stroke. Nonfasting total H(e) levels were measured. The risk of stroke and MI increased directly with total H(e) (143). The linear coefficient suggested a risk increase by 6-7% for every 1-\mumol/liter increase in total H(e). The odds ratios for subjects in the highest quintile of total H(e) level (>18.6 µmol/liter) compared with those with lower H(e) levels were 2.43 (95% confidence interval, 1.11-5.35) for MI and 2.53 (95% confidence interval, 1.19-5.35) for stroke (143). Associations were more pronounced among those with hypertension (143). The study, based on a relatively short follow-up period, provides evidence that among elderly subjects an elevated H(e) level is associated with an increased risk of cardiovascular disease.

## C. Negative studies

Not all studies have shown that H(e) is an independent risk factor for coronary artery disease. A prospective study in Finland of 7,424 healthy subjects at baseline showed development of stroke in 265 subjects over a 9-yr period (144). The fact that the affected subjects did not have an elevated serum H(e) level has been attributed to the exceptionally low gene frequency predisposing to hyperhomocysteinemia in Finland. However, the study itself did not assess the frequency of mutations in the enzymes concerned, and therefore this explanation must be regarded as speculative.

In 1997 Verhoef *et al.* (145) reported a prospective, nested case-control study that used baseline samples from the Physicians Health Study. After 9 yr of follow-up, subjects with newly diagnosed angina pectoris, without MI, and with subsequent coronary artery bypass graft surgery were studied. Controls had no clinical diagnosis of coronary artery disease. In this study, total H(e) levels and risk of angina pectoris did not correlate (145). These data contrast with other reports from this group, which show that H(e) levels correlate with the extent of coronary occlusive disease (6) and that folate and  $B_6$  levels are inversely related to the risk of MI (146). The

divergent results may be explained because the study subjects from the Physicians Health (Angina) Study are expected to be nutritionally (folate, vitamins  $B_{12}$  and  $B_6$ ) replete (145). Alternatively, H(e) levels are decreased after MI and then increase over several weeks to months (43). Since H(e) levels were measured 2 to 3 months after the cardiac event in the positive studies, the hyperhomocysteinemia may be the result, rather than the cause, of a vascular occlusive event.

In 1997, Evans *et al.* (13) reported the Multiple Risk Factor Intervention Trial (MRFIT). Prospectively obtained, stored serum samples from 712 men (nonfatal MIs or deaths from coronary artery disease) were analyzed for H(e) level. Odds ratios for patients with coronary artery events based upon H(e) levels are quartile 1, 1.00, quartile 2, 1.03, quartile 3, 0.84, and quartile 4, 0.92. Homocysteine was not found to be a risk factor for coronary artery disease in this study (13).

A recent study by Folsom et al. (12) has also questioned the relationship between total H(e) and the risk of coronary artery disease. This prospective case-cohort study consisted of 15,792 men and women ages 45-64, with 232 coronary artery disease cases and 537 controls. Of particular interest, there was no association of coronary heart disease with the thermolabile mutation of the MTHFR gene or with three mutations of the CBS gene (12). These findings in a prospective study add uncertainty to conclusions from other studies that H(e), in general, and the enzyme mutation, in particular, are a major independent risk factor for coronary artery disease. After adjustment for age, plasma H(e) was positively associated with coronary arterial disease in women but not in men (12). Similarly coronary artery disease was negatively associated with plasma folate and vitamin supplementation in women only. After correction for other risk factors, only pyridoxal 5'-phosphate plasma levels were associated with the risk of coronary artery disease (12). The finding that pyridoxal 5'-phosphate levels are important in determining the risk of coronary artery disease may be related to the metabolic mechanisms used by these patients to handle hyperhomocysteinemia. The investigators studied only fasting plasma H(e), and the fact that vitamin B<sub>6</sub> appeared to offer independent protection suggests that high postmethionine load H(e) (determined by vitamin B<sub>6</sub>, as discussed above) may have been a better variable to study as a risk factor for coronary artery disease than fasting H(e).

In summary, data from many studies support the hypothesis that hyperhomocysteinemia is an independent risk factor for coronary artery disease, as well as other arterial occlusive disease. More recently, data from several studies have questioned whether H(e) per se is a risk factor for coronary artery disease. Both positive and negative studies have shown the importance of vitamin levels in study subjects, as well as controlling for other known cardiovascular risk factors, including gender. Indeed, several of the studies indicate that hyperhomocysteinemia is a greater risk factor in women or older subjects in whom vitamin levels may also be lower. The timing of H(e) collection after a vascular occlusive event may also affect the H(e) level. Future studies to clarify the relationship between H(e) and coronary artery disease must therefore be prospective, control for known cardiovascular risk factors, and measure vitamin  $B_6$ ,  $B_{12}$ , and folate levels.

## D. Effect of low plasma H(e) on cardiovascular disease

If an elevated plasma H(e) is associated with an increased risk of coronary heart disease, then a low plasma H(e) should lead to a decreased risk of coronary heart disease. However, information on plasma H(e) concentrations in population groups with low coronary artery disease prevalence is limited and conflicting.

One such population is patients with Down's syndrome, a condition that is associated with a very low prevalence of coronary artery disease (147). The gene for CBS is on chromosome 21, and trisomy 21-associated Down's syndrome is associated with a CBS gene dosage of 150% (148) and significantly lower fasting as well as postmethionine plasma H(e) concentrations (149). However, Brattstrom *et al.* (150) failed to find more effective H(e) metabolism in Down's syndrome patients.

Ubbink *et al.* (151) studied a population of South African black subjects living in a rural area who had a low incidence of coronary artery disease, despite a high prevalence of smoking and in whom plasma H(e) concentrations were significantly lower than those in South African whites. They also reported that the distribution of plasma H(e) concentration frequencies was positively skewed; they suggested that this frequency distribution corresponds to that previously reported in populations prone to coronary artery disease (6, 140). While the postmethionine load H(e) fell after vitamin treatment in white subjects, it did not do so in blacks; indicating relative independence from this nutritional cofactor and the possibility of other cofactors or regulators in H(e) metabolism.

In another study attempting to determine the cause of difference in prevalence of coronary artery disease, plasma H(e) concentrations were found to be higher in people in Ireland compared with those in France (152). The prevalence of coronary artery disease and mortality in Ireland is much higher than that in France. Although there are differences between the two populations in conventional risk factors, these do not account for the large interpopulation difference in coronary artery disease. A higher plasma H(e) in the Irish subjects who suffered an MI when compared with that in the French could explain the different rates of coronary heart disease in the two populations. This study also showed that the risk for MI in both populations was graded across the distribution of plasma H(e) and increased in subjects with the highest plasma H(e)(152).

We have recently studied plasma H(e) concentrations in lean subjects with type 2 diabetes in India and have found that plasma H(e) concentrations are lower than those in normal weight and obese diabetic subjects (S. Das and V. Fonseca, unpublished observations). Coronary artery disease is rare in lean subjects in India despite the presence of DM (152). However, the prevalence of diabetes in Indians is higher than that in Caucasians (153). These patients have been found to be insulin resistant despite modest degrees of obesity, and the incidence of coronary artery disease in Indian immigrants to the United Kingdom has been found to be exceedingly high (154). Thus, it would appear that even modest degrees of obesity are associated with a rise in plasma H(e) concentrations in this population when compared with lean sub-

jects. These findings are compatible with our data on HFS feeding in rats described above.

In summary, the prevalence of vascular disease appears to be lower in populations that have lower levels of plasma H(e). However, a changing phenotype, particularly with an increase in obesity, may be associated with an increase in plasma H(e) and a concomitant increase in cardiovascular risk. Whether these variables are causally related needs to be investigated.

## VIII. Possible Mechanisms Of Accelerated Vascular Disease in Homocysteinemia

Putative mechanisms of atherothrombosis in hyperhomocysteinemia include endothelial cell injury, endothelial dysfunction, increased vascular smooth muscle cell growth, increased platelet adhesiveness, enhanced LDL oxidation and deposition in the arterial wall, and direct activation of the coagulation cascade. Caution should be used in extrapolating the results of *in vitro* studies, as in many of them the concentrations of H(e) used are much higher than those seen in plasma of patients with HH(e). The vascular changes in hyperhomocysteinemia are likely to be multifactorial (Table 4).

#### A. Platelet dysfunction

Platelets from patients with HH(e) have increased adhesiveness which is corrected by pyridoxine (155). Treatment with pyridoxine also restores the decreased platelet survival seen in some patients. Homocysteine alters arachidonic acid metabolism in platelets, with increased release of proaggregatory thromboxane  $A_2$  (156). In rats in whom mild HH(e) has been induced by folate deficiency, an acute methionine load enhances platelet aggregation, thromboxane biosynthesis, and macrophage-derived tissue factor activity (157).

## B. Coagulation abnormalities

Activation of the coagulation cascade by H(e) may also contribute to vascular disease. Homocysteine activates Factor XII and induces arterial endothelial cell Factor V activation (158, 159). In addition, high concentrations of H(e) may inhibit TM (160, 161). Since the binding of thrombin to TM enhances formation of the anticoagulant-activated protein C and inhibits thrombin activation of fibrinogen, a deficiency of TM enhances fibrin formation. All of these events effectively change the balance between procoagulation/anticoagulation and enhance the risk of thrombosis. In patients with the more severe condition of homocystinuria, there is activation and hyperconsumption of Factor VII, Factor X, and consumption of antithrombin III (161-164). In homocystinuria, levels of coagulation factors are reduced. Markers of activation of coagulation, such as F1+2, are elevated and are correctable with treatment (165). H(e) increases tissue factor activity in a dose-dependent fashion, by increasing the rate of synthesis of tissue factor RNA (166).

## C. Effects on the endothelium

Recent investigation has identified the endothelium as a major site of pathological damage caused by HH(e). In par-

TABLE 4. Possible mechanisms contributing to premature atherosclerosis in hyperhomocystinemia

#### Endothelium

Patchy endothelial cell loss

Decreased viability and function

Free radical generation and lipid peroxidation

Induces tissue factor

Increased vWF, thrombomodulin, and PAI-1

Decreased prostacyclin

Decreased DNA synthesis

Decreased nitric oxide

Decreased vascular reactivity

#### **Platelets**

Decreased survival

Increased adhesiveness

Increased aggregation

Increased release of platelet factors that stimulate smooth muscle proliferation, vasoconstriction, and aggregation

Inhibition of cell surface expression of thrombomodulin

Decreased activation of protein C

Decreased activity of antithrombin III, decreased factor VII

Arterial wall

Vascular smooth muscle cell proliferation and hypertrophy Increased intimal-medial thickness

ticular, the interaction between H(e) and nitric oxide (NO) described below has important clinical implications.

Vascular reactivity (determined by assessing the change in brachial artery diameter during reactive hyperemia – an index of flow-mediated, endothelium-dependent, nitric oxide-mediated vasodilatation) is significantly impaired in elderly subjects with HH(e), compared with control subjects (167). In contrast, the vasodilatation after the administration of sublingual nitroglycerin (endothelium-independent) is normal. On linear regression analysis serum H(e) concentrations emerged as the only significant predictor of flow-mediated vasodilatation. These results indicate that the bioavailability of nitric oxide is decreased in human subjects with HH(e) and is compatible with the findings in animals. Celermajer et al. (168) demonstrated that children with homozygous homocystinuria had impaired endothelial function and vascular reactivity. In contrast, endothelial function assessed by similar methodology is preserved in heterozygous adults.

In a primate model, Lentz *et al.* demonstrated that dietinduced HH(e) led to blunted responses of resistant vessels to endothelium-dependent vasodilators and that this effect was accompanied by depressed TM activity and moderately reduced vascular smooth muscle responses to nitroglycerin when vessels were studied *ex vivo* (169). An infusion of H(e) in rats abolishes the endothelium-dependent vasodilation induced by acetylcholine, suggesting that homocysteine's adverse effects are mediated through deficient production of NO (170).

Normal endothelial cells detoxify H(e) by releasing NO, which leads to the formation of *S*-nitroso-homocysteine (171). The formation of S-nitroso-homocysteine attenuates the pathogenicity of H(e) by inhibiting sulfhydryl-dependent generation of free oxygen radicals. This protective action, however, is eventually overcome by chronic exposure of the endothelial cell to HH(e) (103). Homocysteine may also attenuate endothelial production of bioactive NO (103).

The enzymes in H(e) metabolism are present in endothelial cells (172), and H(e) metabolism is active in endothelial cells. In human umbilical vein endothelial cells in culture, H(e) is exported into the culture medium, and this export is decreased by folate in a dose-dependent manner (173).

Endothelial cells from CBS heterozygotes are deficient in CBS and are more susceptible to H(e)-mediated injury (174). Jakubowski has demonstrated that human cells in which H(e) metabolism is deregulated by a mutation in the CBS gene or by the use of antifolate drug produce more H(e) thiolactone than unaffected cells (175). The thiolactone is incorporated into cellular and extracellular proteins and may be more toxic to endothelial cells than H(e).

In response to H(e)-induced toxicity in human endothelial cells, substantial changes in the concentration of intracellular soluble thiols have been observed (176). Large decreases in cellular NAD+ occurred in response to H(e)-induced toxicity, and DNA synthesis was also compromised. Radical scavengers were effective in preventing this H(e) toxicity (176).

Hyperhomocysteinemia after a methionine load in rats is associated with considerable loss of endothelium and degeneration of media cells in the aortic wall. These changes are more pronounced in the spontaneously hypertensive rat than in the normotensive rat (177).

There has been considerable interest in HH(e) causing endothelial damage by increasing free radical production and subsequent lipid peroxidation (178, 179). However, the exact role of free radicals in HH(e)-induced endothelial damage remains unclear.

Anticoagulation and fibrinolysis are also functions of the endothelium that are critical for blood flow. Several studies have been concerned with measuring the endothelialderived proteins important in these processes. Plasma concentrations of proteins secreted by the endothelium such as TM and von Willebrand factor (vWF) are elevated in HH(e) and serve as surrogate markers of endothelial dysfunction (180). Van den Berg et al. assessed endothelial function by measuring plasma concentrations of endothelium-derived proteins such as vWF, TM, and tissue-type plasminogen activator (tPA) (181); vWF and TM were elevated while tPA was normal in patients with HH(e). After treatment with pyridoxine and folic acid, vWF and TM levels decreased and tPA was unchanged. Tissue plasminogen activator inhibitor (PAI-1) antigen has been shown to correlate with total plasma H(e) concentrations and may also be a marker of impaired fibrolytic activity and endothelial function (182). H(e) has also been shown to suppress anticoagulant heparin sulfate expression in cultured porcine aortic endothelial cells and may thus contribute to thrombogenesis (183). Finally, H(e) inhibits cyclooxygenase activity in human endothelial cells, decreasing prostacyclin production (184).

In summary, H(e) metabolism is active in endothelial cells, and endothelial damage with loss of ability to generate adequate NO appears to be the major mediator of vascular disease caused by HH(e). In addition, the endothelial secretion of anticoagulant and fibrinolytic substances is impaired. Depletion of NO results in loss of vasodilatation, generation of reactive oxygen species, inhibition of glutathione peroxidase, proliferation of vascular smooth muscle cells, and suppression of endothelial cell growth. The importance of these

*in vitro* mechanisms *in vivo*, in a setting of more physiological concentrations of H(e), is unclear. Additional *in vivo* studies that utilize physiological concentrations of H(e), demonstrate a H(e)-specific effect, and evaluate reversibility of endothelial damage with vitamin therapy are needed. In particular, these studies must evaluate whether endothelial damage results from or causes elevated plasma H(e) concentrations.

#### D. Effects of hyperhomocysteinemia on the arterial wall

Homocysteine appears to have a growth-promoting effect on the arterial wall through a number of mechanisms. Tsai *et al.* (185) studied the effect of H(e) on the growth of vascular smooth muscle cells and endothelial cells. H(e) causes a 25% increase in DNA synthesis in rat aortic smooth muscle cells. In contrast, in human umbilical vein endothelial cells, H(e) leads to a decrease in DNA synthesis in a dose-dependent manner. These findings suggest that H(e) has a growth-promoting effect on vascular smooth muscle cells along with an inhibitory effect on endothelial cell growth. This combination could lead to atherosclerosis (185). Minipigs fed a methionine-rich casein-based diet develop HH(e). These animals developed aberrations in the elastic lamina with hypertrophy of smooth muscle (186).

Rats fed a high H(e) diet have a stimulation of aortic cyclin-dependent kinase at the transcriptional level, with the possible consequence of proliferation of aortic cells (187). Arterial smooth muscle cells cultured in the presence of H(e) grow to a higher density and produce and accumulate collagen at levels significantly above controls (188). Homocysteine also inhibits growth and p21ras methylation in vascular endothelial cells (189). Homocysteine has also been shown to have a weak mitogenic effect on vascular smooth muscle cells and, in addition, enhances the mitogenic response of platelet-derived growth factor (190).

#### E. Coinheritance of factor V Leiden in homocystinuria

The coinheritance of homocystinuria and factor V Leiden mutation (activated protein C resistance) has been found to have an association with thrombophilia (10). Because only one-third of patients with homocystinuria develop venous or arterial thromboses, a search was made for other contributing factors in patients who developed thrombosis. A mutation in the gene coding for factor V replaces glutamine for arginine at position 506, increasing a patient's risk for thrombosis by altering the first cleavage site involved in the activation of factor V. This suggests that in patients with homocystinemia, the risk of venous or arterial thromboembolic disease may be exacerbated by the presence of other concomitant etiologies of thrombophilia.

## IX. Management of Hyperhomocysteinemia

A number of agents are known to decrease plasma H(e) concentrations in patients with both HH(e) and homocystinuria. Some of these, such as folate and pyridoxine, are inexpensive and safe. However, clinical trials are needed to

demonstrate their efficacy in preventing or halting the progression of vascular disease.

## A. Prevention of hyperhomocysteinemia

The role of folate, B<sub>12</sub>, and pyridoxine in determining plasma H(e) levels in normal subjects has been discussed above. We have also discussed clinical trials of foods fortified with folate in the prevention of HH(e). It is tempting to speculate that such food fortification will be effective in preventing vascular disease. However, outside the setting of a few clinical trials the impact of increased nutritional supplementation with folic acid, which has been recently recommended (191), on H(e) levels in the general population is unknown and needs to be evaluated.

## B. Treatment of hyperhomocysteinemia

There is no consensus on the optimal dosage of vitamins to be used in the treatment of HH(e). Not only are clinical trials necessary to determine whether lowering plasma H(e) will reduce cardiovascular disease, the optimal dose necessary to do so will need to be ascertained. Practicing physicians face a dilemma on prescribing therapy for HH(e) because of media advertising recommending multivitamins and fruit juices, cereals, etc., to reduce cardiovascular disease, when there is very little evidence that such treatments may be effective.

Combination therapy with different vitamins may be necessary to achieve adequate suppression of H(e) levels in many patients. For example, Franken *et al.* treated mild HH(e) patients with vitamin B<sub>6</sub>, 250 mg daily, for 6 weeks after which the post load H(e) concentration fell in 56% of patients (192). Further treatment with the addition of folic acid and/or betaine resulted in normalization of H(e) levels in 95% of the remaining patients. In three patients with homocystinuria, Palareti *et al.* (193) demonstrated not only reduction in H(e) levels but also correction of a number of abnormalities in blood coagulation (193). It is important to determine whether correction of coagulation and other vascular risk factors occurs in patients with mild HH(e) after treatment.

Only half of patients with CBS deficiency respond to pyridoxine. This may be because some nonresponders may have folate deficiency (194) (which can block the response to pyridoxine until folate is replenished) (21) or decreased affinity of the mutant enzyme for the cofactor (195). Van den Berg *et al.* (181) have also demonstrated correction of mild HH(e) in a subset of young patients with cardiovascular disease (below the age of 50). Brattström *et al.* (196) demonstrated that pyridoxine, 240 mg/day, plus folic acid, 10 mg/day, reduced fasting H(e) levels by a mean of 53% and a postmethionine load H(e) by a mean of 39%.

Ubbink and colleagues (197, 198) investigated the roles of three vitamins as determinants of plasma H(e) concentrations in a placebo-controlled study. One hundred individuals with high fasting plasma H(e) were enrolled in the trial, which compared treatment with five different treatments: 1) placebo; 2) folic acid, 0.65 mg; 3) pyridoxine, 10 mg; 4) vitamin B<sub>12</sub>, 0.4 mg; and 5) a combination of all three vitamins.

Plasma samples were assayed 4 and 6 weeks after starting the vitamin therapy. The folic acid lowered plasma H(e) by 42%, but plasma H(e) declined with vitamin  $B_{12}$  by only 15%. Pyridoxine had no significant effect, which is not surprising as the main effect of pyridoxine would be on lowering postmethionine plasma H(e) rather than a fasting H(e) (33, 199). The combination of three vitamins did not differ significantly from folic acid alone (197).

Boers et~al. (200) treated 32 patients with postmethionine load HH(e) with vitamin B<sub>6</sub> 250 mg for 6 weeks. A few patients also received folic acid 5 mg daily in addition (if they were folic deficient); 81% of patients responded with the normalization of postload HH(e). Similarly, Brattström et~al. (196) reported a 26% reduction in postload plasma H(e) between 15 mg daily of vitamin B<sub>6</sub> and further significant reduction of up to 39% when folic acid 10 mg daily was added (196). Dudman et~al. (33) showed similar results with supplementation of 100 mg of vitamin B<sub>6</sub>. There was a further reduction in postload levels using a combination of folic acid, 5 mg daily, and vitamin B<sub>6</sub>, 100 mg daily.

van den Berg *et al.* (201) reported that treatment of patients with HH(e) with vitamin  $B_6$ , 250 mg, plus folic acid, 5 mg daily for 6 weeks, resulted in normalization of the fasting plasma H(e) in 91% and postload plasma H(e) in 92% of patients. Landgren (43) studied the effect of various doses of folic acid on fasting plasma H(e) after myocardial function. Folic acid at 2.5 mg daily and 10 mg daily were equally effective in lowering fasting H(e) levels with a greater reduction seen in patients who had elevated plasma H(e) at the start of the study (43).

Treatment with vitamin  $B_{12}$  does not appear to have a very significant effect on plasma H(e) levels in healthy subjects but may reduce it in some patients with HH(e), particularly those who have low or normal vitamin  $B_{12}$  blood levels (198).

In summary, although no consensus exists it seems likely that folic acid, 0.65 mg daily, may be sufficient to lower mild fasting HH(e) significantly. However, in patients who have a MLT and are found to have an elevated postload plasma HH(e), treatment with at least 100 mg daily of vitamin  $B_6$  and 5 mg of folic acid daily is necessary. The dose of vitamin  $B_6$  may need to be increased to 250 mg daily.

1. Betaine. Betaine is a methyl group donor involved in the metabolism of methionine and has been suggested as a possible treatment for HH(e) (202). Betaine lowers plasma H(e) concentration but raises methionine levels, the significance of which is not clear (203, 204). Betaine has been found to be ineffective in lowering H(e) in patients on hemodialysis (115).

The above treatment strategies may not apply to patients with very advanced aggressive vascular disease or patients with chronic renal failure and/or DM, where other factors may elevate H(e) levels. These patients may require much higher doses of vitamin replacement therapy (115).

# X. Conclusion

Hyperhomocysteinemia is now well established as a risk factor for cardiovascular disease. HH(e) is common in patients with premature and accelerated vascular disease and

is a risk factor for death in patients who have an MI. Genetic abnormalities and nutritional deficiencies explain only a small proportion of the hyperhomocysteinemia associated with vascular disease. Hormonal and metabolic factors such as diabetes, thyroid disease, and estrogen deficiency appear to interact with H(e) metabolism and may thus play a role in the pathogenesis of atherosclerosis seen in patients with these conditions.

Further investigation into the role of hormonal changes in H(e) metabolism may lead to an improvement in our treatment strategies to prevent atherosclerosis. Fortification of food with folate and nutritional supplementation with vitamins lowers plasma H(e). Combination therapy with multiiple vitamins may be nescessary to correct HH(e) in many patients. Clinical trials are needed to determine the optimal doses of vitamins in different clinical settings. Finally, it is important to determine whether such a lowering of plasma H(e) will prevent the onset or progression of cardiovascular disease.

#### References

- 1. Kluijtmans LA, van den Heuvel LP, Boers GH, Frosst P, Stevens EM, van Oost BA, den Heijer M, Trijbels FJ, Rozen R, Blom HJ 1996 Molecular genetic analysis in mild hyperhomocysteinemia: a common mutation in the methylenetetrahydrofolate reductase gene is a genetic risk factor for cardiovascular disease. Am J Hum Genet 58:35–41
- 2. **Motulsky AG** 1996 Nutritional ecogenetics: homocysteine-related arteriosclerotic vascular disease, neural tube defects, and folic acid. Am J Hum Genet 58:17–20
- 3. Wald NJ, Watt HC, Law MR, Weir DG, McPartlin J, Scott JM 1998 Homocysteine and ischemic heart disease: results of a prospective study with implications regarding prevention. Arch Int Med 158: 862–867
- Dalery K, Lussier-Cacan S, Selhub J, Davignon J, Latour Y, Genest Jr J 1995 Homocysteine and coronary artery disease in French Canadian subjects: relation with vitamins B<sub>12</sub>, B<sub>6</sub>, pyridoxal phosphate, and folate. Am J Cardiol 75:1107–1111
- Selhub J, Jacques PR, Bostom AG, D'Agostino RB, Wilson PW, Belanger AJ, O'Leary DH, Wolf PA, Schaefer EJ, Rosenberg IH 1995 Association between plasma homocysteine concentrations and extracranial carotid-artery stenosis. N Engl J Med 332:286–291
- Stampfer MJ, Malinow MR, Willett WC, Newcomer LM, Upson B, Ullmann D, Tishler PV, Hennekens CH 1992 A prospective study of plasma homocyst(e)ine and risk of myocardial infarction in US physicians. JAMA 268:877–881
- 7. **Boushey CJ, Beresford SA, Omenn GS, Motulsky AG** 1995 A quantitative assessment of plasma homocysteine as a risk factor for vascular disease. Probable benefits of increasing folic acid intakes. JAMA 274:1049–1057
- 8. Clarke R, Daly L, Robinson K, Naughten E, Cahalane S, Fowler B, Graham I 1991 Hyperhomocysteinemia: an independent risk factor for vascular disease. N Engl J Med 324:1149–1155
- den Heijer M, Koster T, Blom HJ, Bos GM, Briet E, Reitsma PH 1996 Hyperhomocysteinemia as a risk factor for deep-vein thrombosis. N Engl J Med 334:759–762
- Mandel H, Brenner B, Berant M, Rosenberg N, Lanir N, Jakobs C, Fowler B, Seligsohn U 1996 Coexistence of hereditary homocystinuria and factor V Leiden – effect on thrombosis. N Engl J Med 334:763–768
- 11. **Graham IM, Daly LE, Refsum HM, Robinson K,** 1997 The European Concerted Action Project. Plasma homocysteine as a risk factor for vascular disease. JAMA 277:1775–1781
- 12. Folsom AR, Nieto FJ, McGovern PG, Tsai MY, Malinow MR, Eckfeldt JH, Hess DL, Davis CE 1998 Prospective study of coronary heart disease incidence in relation to fasting total homocysteine, related genetic polymorphisms, and B vitamins: the Athero-

- sclerosis Risk in Communities (ARIC) study. Circulation 98:204–210
- Evans RW, Shaten BJ, Hempel JD, Cutler JA, Kuller LH 1997 Homocyst(e)ine and risk of cardiovascular disease in the Multiple Risk Factor Intervention Trial. Arterioscler Thromb Vasc Biol 17: 1947–1953
- Carson NAJ, Dent CE, Field CMB, Gaull GE 1965 Homocystinuria. Clinical and pathological review of ten cases. J Pediatr 66: 565–583
- Schimke RN, McKusick VA, Huang T, Pollack AD 1965 Homocystinuria. Studies of 20 families with 38 affected members. JAMA 193:711–719
- McCully KS 1969 Vascular pathology of homocysteinemia. Implications for the pathogenesis of arteriosclerosis. Am J Pathol 56: 111–128
- 17. Boers GH, Smals AG, Trijbels FJ, Fowler B, Bakkeren JA, Schoonderwaldt HC, Kleijer WJ, Kloppenborg PW 1985 Heterozygosity for homocystinuria in premature peripheral and cerebral occlusive arterial disease. N Engl J Med 313:709–715
- 18. Legnani C, Palareti G, Grauso F, Sassi S, Grossi G, Piazzi S, Bernardi F, Marchetti G, Ferraresi P, Coccheri S 1997 Hyperhomocyst(e)inemia and a common methylenetetrahydrofolate reductase mutation (Ala<sup>223</sup>Val MTHFR) in patients with inherited thrombophilic coagulation defects. Thromb Vasc Biol 17:2924–2929
- Ma J, Stampfer MJ, Hennekens CH, Frosst P, Selhub J, Horsford J, Malinow R, Willett WC, Rozen R 1996 Methylenetetrahydrofolate reductase polymorphism, plasma folate, homocysteine, and risk of myocardial infarction in US physicians. Circulation 94:2410– 2416
- Mudd SH, Levy HL, Skovby F 1995 Disorders of transsulfuration.
   In: Scriver CR, Beaudet AL, Sly WS, Valle D (eds) The Metabolic Basis of Inherited Disease, ed 7. McGraw-Hill, New York, pp 1279– 1327
- Selhub J, Miller JW 1992 The pathogenesis of homocysteinemia: interruption of the coordinate regulation by S-adenosylmethionine of the remethylation and transsulfuration of homocysteine. Am J Clin Nutr 55:131–138
- 22. Loehrer FMT, Angst CP, Haefeli WE, Jordan PP, Ritz R, Fowler B 1996 Low whole-blood S-adenosylmethionine and correlation between 5-methyltetrahydrofolate and homocysteine in coronary artery disease. Arterioscler Thromb Vasc Biol 16:727–733
- Jencks DA, Matthews RG 1987 Allosteric inhibition of methylenetetra-hydrofolate reductase by adenosylmethionine. J Biol Chem 262:2485–2493
- 24. Finkelstein JD, Lyle WE, Martin JL, Pick AM 1975 Activation of cystathionine synthase by adenosylmethionine and adenosylethionine. Biochem Biophys Res Commun 66:81–87
- Guba S, Fink L, Fonseca V 1996 Hyperhomocysteinemia An emerging and important risk factor for thromboembolic and cardiovascular disease. Am J Clin Pathol 106:709–721
- Ueland PM, Refsum H, Stabler SP, Malinow MR, Andersson A, Allen RH 1993 Total homocysteine in plasma or serum: methods in clinical applications. Clin Chem 39:1764–1779
- Andersson A, Isaksson A, Hultberg B 1992 Homocysteine export from erythrocytes and its implication for plasma sampling. Clin Chem 38:1311–1315
- Fiskerstrand T, Refsum H, Kvalheim G, Ueland PM 1993 Homocysteine and other thiols in plasma and urine: automated determination in sample stability. Clin Chem 39:263–271
- Malinow MR, Sexton G, Averbuch M, Grossman M, Wilson D, Upson B 1990 Homocyst(e)inemia in daily practice: levels in coronary heart disease. Coron Artery Dis 1:215–220
- Shipchandler MT, Moore EG 1995 Rapid, fully automated measurement of plasma homocyst(e)ine with the Abbott Imx<sup>®</sup> Analyser. Clin Chem 41:991–994
- 31. Frantzen F, Faaren AL, Alfheim I, Nordhei AK 1998 Enyzme conversion immunoassay for determining total homocysteine in plasma or serum. Clin Chem 44:311–316
- Garg UC, Zheng ZJ, Folsom AR, Moyer YS, Tsai MY, McGovern P, Eckfeldt JH 1997 Short-term and long-term variability of plasma homocysteine measurement. Clin Chem 43:141–145
- Dudman NP, Wilcken DE, Wang J, Lynch JF, Macey D, Lundberg
   P 1993 Disordered methionine/homocysteine metabolismin pre-

- mature vascular disease. Its occurence, cofactor therapy, and enzymology. Arterioscler Thromb 13:1253–1260
- 34. Bostom AG, Jacques PF, Nadeau MR, Williams RR, Ellison RC, Selhub J 1995 Post-methionine load hyperhomocysteinemia in persons with normal fasting total plasma homocysteine: initial results from the NHLBI Family Heart Study. Atherosclerosis 116:147–151
- Jacobsen DW 1996 Determinants of hyperhomocysteinemia: a matter of nature and nurture. Am J Clin Nutr 64:641–642
- Lussier-Cacan S, Xhignesse M, Piolot A, Selhub J, Davignon J, Genest Jr J 1996 Plasma total homocysteine in healthy subjects: sex-specific relation with biological traits. Am J Clin Nutr 64:587– 593
- 37. von Eckardstein A, Malinow MR, Upson B, Heinrich J, Schulte H, Schonfeld R, Kohler E, Assmann G 1994 Effects of age, lipoproteins, and hemostatic parameters on the role of homocysteinemia as a cardiovascular risk factor in men. Arterioscler Thromb 14:460-464
- 38. **Selhub J, Jacques PF, Wilson PW, Rush D, Rosenberg IH** 1993 Vitamin status and intake as primary determinants of homocysteinemia in an elderly population. JAMA 270:2693–2698
- 39. Joosten E, Pelemans W, Devos P, Lesaffre E, Goossens W, Criel A, Verhaeghe R 1993 Cobalamin absorption and serum homocysteine and methylmalonic acid in elderly subjects with low serum colbalamin. Eur J Haematol 51:25–30
- Nygard O, Vollset SE, Refsum H, Stensvold I, Tverdal A, Nordrehaug JE, Ueland M, Kvale G 1995 Total plasma homocysteine and cardiovascular risk profile. The Hordaland homocysteine study. JAMA 274:1526–1533
- 41. Malinow MR, Levenson J, Giral P, Nieto FJ, Razavian M, Segond P, Simon A 1995 Role of blood pressure, uric acid, and hemorrheological parameters on plasma homocyst(e)ine concentration. Atherosclerosis 114:175–183
- Lindgren A, Brattstrom L, Norrving B, Hultberg B, Andersson A, Johansson BB 1995 Plasma homocysteine in the acute and convalescent phases after stroke. Stroke 26:795–800
- Landgren F, Israelsson B, Lindgren A, Hultberg B, Andersson A, Brattstrom L 1995 Plasma homocysteine in acute myocardial infarction: homocysteine-lowering effect of folic acid. J Intern Med 237:381–388
- 44. **Fenton WA, Rosenberg LE** 1995 Inherited disorders of cobalamin transport and metabolism. In: Scriver CR, Beaudet AL, Sly WS, Valle D (eds) The Metabolic Basis of Inherited Disease, ed. 7. McGraw-Hill, Inc., New York, pp 3129–3149
- 45. Tsai MY, Garg U, Key NS, Hanson NQ, Suh A, Schwichtenberg K 1996 Molecular and biochemical approaches in the identification of heterozygotes for homocystinuria. Atherosclerosis 122:69–77
- 46. **Giusti B, Comeglio P, Attanasio M, Gori AM, Brunelli T, Prisco D, Pepe G, Gensini GF, Abbate R** 1997 Different distribution of the double mutant " $T_{833}$ C/68 bp Insertion" in cystathionine *β*-synthase gene in Northern and Southern Italian populations. Letter to the Editor. Thromb Haemost 78:1293–1303
- 47. Kraus JP, Le K, Swaroop M, Ohura T, Tahara T, Rosenberg LE, Roper MD, Kozich V 1993 Human cystathionine beta-synthase cDNA: sequence, alternate splicing and expression in cultured cells. Hum Mol Genet 2:1633–1638
- 48. Kluijtmans LAJ, Blom HJ, Boers GH van Oost BA, Trijbels FJ, van den Heuvel LP 1995 Two novel missense mutations in the cystathionine β-synthase gene in homocystinuric patients. Hum Genet 96:249–250
- 49. Tsai MY, Hanson NQ, Schwichtenberg K, Garg U 1995 Amplification refractory mutation system to identify mutations in cystathionine  $\beta$ -synthase deficiency. Clin Chem 41:1775–1777
- 50. Tsai MY, Hanson NQ, Bignell MK, Schwichtenberg KA 1996 Simultaneous detection and screening of  $T_{833}C$  and  $G_{919}A$  mutations of the cystathionine  $\beta$ -synthase gene by single-strand conformational polymorphism. Clin Biochem 29:473–477
- 51. Sebastio G, Sperandeo MP, Panico M, de Franchis R, Kraus JP, Andria G 1995 The molecular basis of homocystinuria due to cystathionine β-synthase deficiency in Italian families, and report of four novel mutations. Am J Hum Genet 56:1324–1333
- Tsai MY, Bignell M, Schwichtenberg K, Hanson NQ 1996 High prevalence of a mutation in the cystathionine beta-synthase gene. Am J Hum Genet 59:1262–1267

- 53. Sperandeo MP, Candito M, Sebastio G, Rolland MO, Turc-Carel C, Giudicelli H, Dellamonica P, Andria G 1996 Homocysteine response to methionine challenge in four obligate heterozygotes for homocystinuria and relationship with cystathionine β-synthase mutations. J Inherit Metab Dis 19:351–356
- 54. Kluijtmans LA, Boers GH, Stevens EM, Renier WO, Kraus JP, Trijbels FJ, van den Heuvel LP, Blom HJ 1996 Defective cystathionine β-synthase regulation by S-adenosylmethionine in a partially pyridoxine responsive homocystinuria patient. J Clin Invest 98:285–289
- 55. Goyette P, Sumner JS, Milos R, Duncan AM, Rosenblatt DS, Matthews RG, Rosen R 1994 Human methylenetetrahydrofolate reductase isolation of cDNA mapping and mutation identification. Nat Genet 7:551
- 56. Kang SS, Wong PW, Susmano A, Sora J, Norusis M, Ruggie N 1991 Thermolabile methylenetetrahydrofolate reductase: an inherited risk factor for coronary artery disease. Am J Hum Genet 48: 536–545
- 57. Kang SS, Passen EL, Ruggie N, Wong PW, Sora H 1993 Thermolabile defect of methylenetetrahydrofolate reductase in coronary artery disease. Circulation 88:1463–1469
- 58. Frosst P, Blom HJ, Milos R, Goyette P, Sheppard CA, Matthews RG, Boers GJ, den Heijer M, Kluijtmans LA, van den Heuvel LP 1995 A candidate genetic risk factor for vascular disease: a common mutation in methylenetetrahydrofolate reductase. Nat Genet 10: 111–113
- 59. Malinow MR, Nieto FJ, Kruger WD, Duell PB, Hess DL, Gluckman RA, Block PC, Holzgang CR, Anderson PH, Seltzer D, Upson B, Lin QR 1997 The effects of folic acid supplementation on plasma total homocysteine are modulated by multivitamin use and methylenetetrahydrofolate reductase genotypes. Arterioscler Thromb Vasc Biol 17:1157–1162
- Ali NS, Powell J, Swaminathan R, Markus HS 1997 The relationship between MTHFR genotype, serum homocysteine and folate levels. Biochem Soc Trans 25:386S
- 61. Jacques PF, Bostom AG, Williams RR, Ellison RC, Eckfeldt JH, Rosenberg IH, Selhub J, Rozen R 1996 Relation between folate status, a common mutation in methylenetetrahydrofolate reductase, and plasma homocysteine concentrations. Circulatiaon 93:7–9
- 62. Christensen B, Frosst P, Lussier-Cacan S, Selhub J, Goyette P, Rosenblatt DS, Genest Jr J, Rozen R 1997 Correlation of a common mutation in the methylenetetrahydrofolate reductase gene with plasma homocysteine in patients with premature coronary artery disease. Atheroscler Thromb Vasc Biol 17:569–573
- Chen LH, Liu ML, Hwang HY, Chen LS, Korenberg J, Shane B 1997 Human methionine synthase. CDNA cloning, gene localization, and expression. J Biol Chem 272:3628–3634
- 64. van der Put NM, van der Molen EF, Kluijtmans LA, Heil SG, Trijbels JM, Eskes TK, Van Oppenraaij-Emmerzaal D, Banerjee R, Blom HJ 1997 Sequence analysis of the coding region of human methionine synthase: relevance to hyperhomocysteinaemia in neural-tube defects and vascular disease. QJM 90:511–517
- 65. Hallam LJ, Sawyer M, Clark AC, Van der Weyden MB 1987 Vitamin B12-responsive neonatal megaloblastic anemia and homocystinuria with associated reduced methionine synthase activity. Blood 69:1128–1133
- 66. Harding CO, Arnold G, Barness LA, Wolff JA, Rosenblatt DS 1997 Functional methionine synthase deficiency due to cblG disorder: a report of two patients and a review. Am J Med Genet 71:384–390
- 67. Fowler B, Schutgens RB, Rosenblatt DS, Smit GP, Lindemans J 1997 Folate-responsive homocystinuria and megaloblastic anaemia in a female patients with functional methionine synthase deficiency (cbl E disease). J Inherit Metab Dis 20:731–741
- Rosenblatt DS, Aspler AL, Shevell MI, Pletcher BA, Fenton WA, Seashore MR 1997 Clinical heterogeneity and prognosis in combined methylmalonic aciduria and homocystinuria (cblC). J Inherit Metab Dis 20:528–538
- 69. Gulati S, Baker P, Li YN, Fowler B, Kruger W, Brody LC, Banerjee R 1996 Defects in human methionine synthase in cblG patients. Hum Mol Genet 5:1859–1865
- 70. Lewis CA, Pancharuniti N, Sauberlich HE 1992 Plasma folate

- adequacy as determined by homocysteine level. Ann NY Acad Sci 669:360–362
- 71. Rimm EB, Willett WC, Hu FB, Sampson L, Colditz GA, Manson JE, Hennekens C, Stampfer MJ 1998 Folate and vitamin B6 from diet and supplements in relation to risk of coronary heart disease among women. JAMA 279:359–364
- 72. Shimakawa T, Nieto FJ, Malinow MR, Chambless LE, Schreiner PJ, Szklo M 1997 Vitamin intake: a possible determinant of plasma homocyst(e)ine among middle-aged adults. Ann Epidemiol 7:285–203
- 73. Malinow MR, Duell PB, Hess DL, Anderson PH, Kruger WD, Phillipson BE, Gluckman RA, Block PC, Upson BM 1998 Reduction of plasma homocyst(e)ine levels by breakfast cereal fortified with folic acid in patients with coronary heart disease. N Engl J Med 338:1009–1015
- 74. Schorah CJ, Devitt H, Lucock M, Dowell AC 1998 The responsiveness of plasma homocysteine to small increases in dietary folic acid: a primary care study. Eur J Clin Nutr 52:407–411
- 75. de Bree A, van Dusseldorp M, Brouwer IA, van het Hof KH, Steegers-Theunissen RP 1997 Folate intake in Europe: recommended, actual and desired intake. Eur J Clin Nutr 51:643–660
- 76. Homocysteine Lowering Trialists' Collaboration 1998 Lowering blood homocysteine with folic acid based supplements: meta-analysis of randomised trials. Br Med J 316:894–898
- 77. Miller JW, Nadeau MR, Smith D, Selhub J 1994 Vitamin B-6 deficiency vs. folate deficiency: comparison of responses to methionine loading in rats. Am J Clin Nutr 59:1033–1039
- 78. **Miller JW, Ribaya-Mercado JD, Russell RM** 1991 Total homocysteine in fasting plasma is not a good indicator of B6 deficiency. FASEB J 5:A557 (Abstract)
- 79. Ubbink JB, van der Merwe A, Delport R, Allen RH, Stabler SP, Reizler R, Vermaak WJ 1996 The effect of a subnormal vitamin B-6 status on homocysteine metabolism. J Clin Invest 98:177–184
- 80. Selhub J, Jacques PF, Wilson PW, Rush D, Rosenberg I H 1993
  Vitamin status and intake as primary determinants of homocysteinemia in an elderly population. JAMA 270:2693–2698
  81. Dudman NP, Guo XW, Gordon RB, Dawson PA, Wilcken DE 1996
- 81. Dudman NP, Guo XW, Gordon RB, Dawson PA, Wilcken DE 1996 Human homocysteine catabolism: three major pathways and their relevance to development of arterial occlusive disease. J Nutr 126: 1295S–1300S
- 82. Kang SS, Wong PW, Zhou JM, Cook HY 1986 Total homocyst(e)ine in plasma and amniotic fluid of pregnant women. Metabolism 35:889–891
- 83. Andersson A, Hultberg B, Brattstrom L, Isaksson A 1992 Decreased serum homocysteine in pregnancy. Eur J Clin Chem Biochem 30:377–379
- 84. **Finkelstein JD** 1962 Methionine metabolism in mammals. Effect of age, diet, and hormones on three enzymes of the pathway in rat tissues. Arch Biochem Biophys 122:583–590
- 85. Wouters MG, Moorrees MT, van der Mooren MJ, Blom HJ, Boers GH, Schellekens LA, Thomas Cm Eskes TK 1995 Plasma homocysteine and menopausal status. Eur J Clin Invest 25:801–805
- 86. Ándersson A, Bratistrom L, Israelsson B, Isaksson A, Hamfelt A, Hultberg B 1992 Plasma homocysteine before and after methionine loading with regard to age, gender, and menopausal status. Eur J Clin Invest 22:79–87
- 87. van der Mooren MJ, Wouters MG, Blom HJ, Schellekens LA, Eskes TK, Rolland R 1994 Hormone replacement therapy may reduce high serum homocysteine in postmenopausal women. Eur J Clin Invest 24:733–736
- 88. Green TJ, Houghton LA, Donovan U, Gibson RS, O'Connor DL 1998 Oral contraceptives did not affect biochemical folate indexes and homocysteine concentrations in adolescent females. J Am Diet Assoc 98:49–55
- 89. Anker G, Lonning PE, Ueland PM, Refsum H, Lien EA 1995 Plasma levels of the atherogenic amino acid homocysteine in postmenopausal women with breast cancer treated with tamoxifen. Int J Cancer 60:365–368
- 90. Lien EA, Anker G, Lonning PE, Refsum H, Ueland PM 1997 Effects of hormones on the plasma levels of the atherogenic amino acid homocysteine. Biochem Soc Trans 25:33–35
- 91. Zmuda JM, Bausserman LL, Maceroni D, Thompson PD 1997 The

- effect of supraphysiologic doses of testosterone on fasting total homocysteine levels in normal men. Atherosclerosis 130:199–202
- 92. **Brattstrom L, Lindgren A, Israelsson B, Andersson A, Hultberg B** 1994 Homocysteine and cysteine: determinants of plasma levels in middle-aged and elderly subjects. J Intern Med 236:633–641
- 93. Giltay EJ, Hoogeveen EK, Gooren LJG, Asscheman H, Elbers JMH, Jakobs C, Stehouwer CD, Effects of sex steroids on plasma total homocysteine in male-to-female and female-to-male transsexuals. Program of the 79th Annual Meeting of The Endocrine Society, Minneapolis, MN, 1997 (Abstract P1–326)
- 94. Kim MH, Kim E, Passen EL, Meyer J, Kang SS 1997 Cortisol and estradiol: nongenetic factors for hyperhomocyst(e)inemia. Metabolism 46:247–249
- McCully KS 1996 Homocysteine and vascular disease. Nat Med 2:386–389
- Nedrebo BG, Ericsson UB, Nygard O, Refsum H, Ueland PM, Aakvaag A, Aanderud S, Lien EA 1998 Plasma total homocysteine levels in hyperthyroid and hypothyroid patients. Metabolism 47: 89–93
- Stamler J, Vaccaro O, Neaton JD, Wentworth D 1993 Diabetes, other risk factors, and 12-yr cardiovascular mortality for men screened in the Multiple Risk Factor Intervention Trial. Diabetes Care 16:434–444
- 98. Munshi MN, Stone A, Fink L, Fonseca V 1996 Hyperhomocysteinemia following a methionine load in patients with non-insulindependent diabetes mellitus and macrovascular disease. Metabolism 45:133–135
- Araki A, Sako Y, Ito H 1998 Plasma Homocysteine and its relationship to cardiovascular risk factors in a Japanese population. In: Graham I, Refsum H, Rosenburg I, Ueland PM (eds) Homocysteine Metabolism: From Basic Science to Clinical Medicine. Kluwer Academic Publishers, Boston, pp 205–210
- 100. Hoogeveen EK, Kostense PJ, Beks PJ, Mackaay AJC, Jakobs C, Bouter LM, Heine RJ, Stehouwer CD 1998 Hyperhomocysteinemia is associated with an increased risk of cardiovascular disease, especially in non-insulin-dependent diabetes mellitus: a population-based study. Arterioscler Thromb Vasc Biol 18:133–138
- 101. Araki A, Sako Y, Ito H 1993 Plasma homocysteine concentrations in Japanese patients with non-insulin-dependent diabetes mellitus: effect of parenteral methylcobalamin treatment. Atherosclerosis 103:149–157
- Giugliano D, Ceriello A, Paolisso G 1996 Oxidative stress and diabetic vascular complications. Diabetes Care 19:257–267
- 103. **Welch GN, Loscalzo** Ĵ 1998 Homocysteine and atherothrombosis. N Engl J Med 338:1042–1050
- 104. Fonseca VA, Stone A, Munshi M, Baliga BS, Aljada A, Thusu K, Fink L, Dandona P 1997 Oxidative stress in diabetic macrovascular disease: does homocysteine play a role? South Med J 90:903–906
- 105. Hultberg B, Agardh E, Andersson A, Brattstrom L, Isaksson A, Israelsson B, Agardh CD 1991 Increased levels of plasma homocysteine are associated with nephropathy, but not severe retinopathy in type 1 diabetes mellitus. Scand J Clin Lab Invest 51:277–282
- 106. Hultberg B, Agardh CD, Agardh E, Lovestam-Adrian M 1997 Poor metabolic control, early age at onset, and marginal folate deficiency are associated with increasing levels of plasma homocysteine in insulin-dependent diabetes mellitus. A five-year follow-up study. Scand J Clin Lab Invest 57:595–600
- 107. Hofmann MA, Kohl B, Zumbach MS, Borcea V, Bierhaus A, Henkels M, Amiral J, Schmidt AM, Fiehn W, Ziegler R, Wahl P, Nawroth PP 1998 Hyperhomocyst(e)inemia and endothelial dysfunction in IDDM. Diabetes Care 21:841–848
- Colwell JA 1997 Elevated plasma homocysteine and diabetic vascular disease. Diabetes Care 20:1805–1806
- Fonseca VA, Reynolds T, Fink LM 1998 Hyperhomocysteinemia and microalbuminuria in diabetes. [Letter] Diabetes Care 21:1028
- 110. Chico A, Perez A, Cordoba A, Arcelus R, Carreras G, de Leiva A, Gonzalez-Sastre F, Blanco-Vaca F 1998 Plasma homocysteine is related to albumin excretion rate in patients with diabetes mellitus: a new link between diabetic nephropathy and cardiovascular disease? Diabetologia 41:684–693
- 111. Lanfredini M, Fiorina P, Peca MG, Veronelli A, Mello A, Astorri E, Dall'Aglio P, Craveri A 1998 Fasting and post-methionine load homocyst(e)ine values are correlated with microalbuminuria and

- could contribute to worsening vascular damage in non-insulindependent diabetes mellitus patients. Metabolism 47:915–921
- 112. Fiorina P, Lanfredini M, Montanari A, Peca MG, Veronelli A, Mello A, Astorri E, Craveri A 1998 Plasma homocysteine and folate are related to arterial blood pressure in type 2 diabetes mellitus. Am J Hypertens 11:1100–1107
- 113. Hoogeveen EK, Kostense PJ, Jager A, Heine RJ, Jakobs C, Bouter LM, Donker AJ, Stehouwer CD 1998 Serum homocysteine level and protein intake are related to risk of microalbuminuria: the Hoorn Study. Kidney Int 54:203–209
- 114. **Stehouwer CD, Gall MA, Hougaard P, Jakobs C, Parving HH** 1999 Plasma homocysteine concentration predicts mortality in non-insulin-dependent diabetic patients with and without albuminuria. Kidney Int 55:308–314
- 115. **Bostom AG, Lathrop L** 1997 Hyperhomocysteinemia in end-stage renal disease: prevalence, etiology, and potential relationship to arteriosclerotic outcomes. Kidney Int 52:10–20
- 116. **Bostom A, Brosnan JT, Hall B, Nadeau MR, Selhub J** 1995 Net uptake of plasma homocysteine by the rat kidney *in vivo*. Atherosclerosis 116:59–62
- 117. Bostom AG, Shemin D, Lapane KL, Hume AL, Yoburn D, Nadeau MR, Bendich A, Selhub J, Rosenberg IH 1996 High dose B-vitamin treatment of hyperhomocysteinemia in dialysis patients. Kidney Int 49:147–152
- 118. Massy ZA, Chadefaxu-Vekemans B, Chevalier A, Bader CA, Drueke TB, Legendre C, Lacour B, Kamoun P, Kreis H 1994 Hyperhomocysteinemia: a significant risk factor for cardiovascular disease in renal transplant recipients. Nephrol Dial Transplant 9:1103–1108
- 119. Arnadottir M, Hultberg B, Vladov V, Nilsson-Ehle P, Thysell H 1996 Hyperhomocysteinemia in cyclosporine-treated renal transplant recipients. Transplantation 61:509–512
- 120. Fonseca VA, Mudaliar S, Schmidt B, Fink LM, Kern PA, Henry RR 1998 Plasma homocysteine concentrations are regulated by acute hyperinsulinemia in nondiabetic but not type 2 diabetic subjects. Metabolism 47:686–689
- 121. **Reaven GM** 1988 Banting lecture. Role of insulin resistance in human disease. Diabetes 37:1595–1607
- 122. **Forker LL, Chaikoff IL, Entenman C, Tarver H** 1951 Formation of muscle protein in diabetic dogs studied with S<sup>35</sup> methionine. J Biol Chem 188:37–48
- 123. **Zinneman HH, Nuttall FQ, Goetz FC** 1966 Effect of endogenous insulin on human amino acid metabolism. Diabetes 15:5–8
- 124. Laivuori H, Kaaja R, Turpeinen U, Viinikka L, Ylikorkala O 1999 Plasma homocysteine levels elevated and inversely related to insulin sensitivity in preeclampsia. Obstet Gynecol 93:489–493
- 125. Fonseca V, Dicker-Brown A, Ranganathan S, Barnard RJ, Fink L, Kern PA 1998 Effects of insulin resistance on enzymes of homocysteine metabolism in the rat. Diabetes 47:A113 (Abstract)
- 126. Hoogeveen EK, Kostense PJ, Jakobs C, Bouter LM, Heine RJ, Stehouwer CD 1997 Does metformin increase the serum total homocysteine level in non-insulin-dependent diabetes mellitus? J Intern Med 242:389–394
- 127. Carlsen SM, Folling I, Grill V, Bjerve KS, Schneede J, Refsum H 1997 Metformin increases total serum homocysteine levels in nondiabetic male patients with coronary heart disease. Scand J Clin Lab Invest 57:521–527
- 128. Glueck CJ, Shaw P, Lang JE, Tracy T, Sieve-Smith L, Wang Y 1995 Evidence that homocysteine is an independent risk factor for atherosclerosis in hyperlipidemic patients. Am J Cardiol 75:132–136
- 129. **Tonstad S, Refsum H, Ueland PM** 1997 Association between plasma total homocysteine and parental history of cardiovascular disease in children with familial hypercholesterolemia. Circulation 96:1803–1808
- Superko HR 1997 Elevated high-density lipoprotein cholesterol, not protective in the presence of homocysteinemia. Am J Cardiol 79:705–706
- 131. Upchurch Jr GR, Welch GN, Fabian AJ, Freedman JE, Johnson JL, Keaney Jr JF, Loscalzo J 1997 Homocyst(e)ine decreases bioavailable nitric oxide by a mechanism involving glutathione peroxidase. J Biol Chem 272:17012–17017
- 132. **Loscalzo J** 1996 The oxidant stress of hyperhomocyst(e)inemia. J Clin Invest 98:5–7

- 133. Halvorsen B, Brude I, Drevon CA, Nysom J, Ose L, Christiansen EN, Nenseter MS 1996 Effect of homocysteine on copper ioncatalyzed, azo compound-initiated, and mononuclear cell-mediated oxidative modification of low density lipoprotein. J Lipid Res 37:1591–1600
- 134. Cordoba-Porras A, Sanchez-Quesada JL, Gonzalez-Sastre F, Odonez-Llanos J, Blanco-Vaca F 1996 Susceptibility of plasma low- and high-density lipoproteins to oxidation in patients with severe hyperhomocysteinemia. J Mol Med 74:771–776
- 135. Perry IJ, Refsum H, Morris RW, Ebrahim SB, Ueland PM, Shaper AG 1995 Prospective study of serum total homocysteine concentration and risk of stroke in middle-aged British men. Lancet 346: 1395–1398
- 136. Malinow MR, Kang SS, Taylor LM, Wong PW, Coull B, Inahara T, Mukerjee D, Sexton G, Upson B 1989 Prevalence of hyperhomocyst(e)inemia in patients with peripheral arterial occlusive disease. Circulation 79:1180–1188
- 137. Fermo I, Vigano D'Angelo S, Paroni R, Mazzola G, Calori G, D'Angelo A 1995 Prevalence of moderate hyperhomocysteinemia in patients with early-onset venous and arterial occlusive disease. Ann Intern Med 123:747–753
- 138. Nygard O, Nordrehaug JE, Refsum H, Ueland PM, Farstad M, Vollset SE 1997 Plasma homocysteine levels and mortality in patients with coronary artery disease. N Engl J Med 337:230–236
- 139. Verhoef P, Kok FJ, Kruyssen DA, Schouten EG, Witteman JC, Grobbee DE, Ueland PM, Refsum H 1997 Plasma total homocysteine, B vitamins, and risk of coronary atherosclerosis. Atheroscler Thromb Vasc Biol 17:989–995
- 140. **Aronow WS, Ahn C, Schoenfeld MR** 1997 Association between plasma homocysteine and extracranial carotid arterial disease in older persons. Am J Cardiol 79:1432–1433
- 141. Malinow MR, Nieto FJ, Szklo M, Chambless LE, Bond G 1993 Carotid artery intimal-medial wall thickening and plasma homocyst(e)ine in asymptomatic adults. The Atherosclerosis Risk in Communities Study. Circulation 87:1107–1113
- 142. Konecky N, Malinow MR, Tunick PA, Freedberg RS, Rosenzweig BP, Katz ES, Hess DL, Upson B, Leung B, Perez J, Kronzon I 1997 Correlation between plasma homocyst(e)ine and aortic atherosclerosis. Am Heart J 133:534–540
- 143. Bots ML, Launer LJ, Lindemans J, Hoes AW, Hofman A, Witteman JC, Koudstaal PJ, Grobbee DE 1999 Homocysteine and short-term risk of Myocardial infarction and stroke in the elderly: the Rotterdam Study. Arch Intern Med 159:38–44
- 144. Alfthan G, Pekkanen J, Jauhiainen M, Pitkaniemi J, Karvonen M, Tuomilehto J, Salonen JT, Ehnholm C 1994 Relationship of serum homocysteine and lipoprotein(a) concentrations to atherosclerotic disease in a prospective Finnish population based study. Atherosclerosis 106:9–19
- 145. Verhoef P, Hennekens CH, Allen RH, Stabler SP, Willett WC, Stampfer MJ 1997 Plasma total homocysteine and risk of angina pectoris with subsequent coronary artery bypass surgery. Am J Cardiol 79:799–801
- 146. Verhoef P, Stampfer MJ, Buring JE, Gaziano JM, Allen RH, Stabler SP, Reynolds RD, Kok FJ, Hennekens CH, Willett WC 1996 Homocysteine metabolism and risk of myocardial infarction: relation with vitamins B6, B12, and folate. Am J Epidemiol 143: 845–859
- 147. Murdock JC, Rodger JC, Rao SS, Fletcher CD, Dunnigan MG 1977 Down's syndrome: an atheroma-free model? Br Med J 2:226–228
- 148. Kraus JP, Williamson CL, Firgaira FA, Yang-Feng TL, Munke M, Francke U, Rosenberg LE 1986 Cloning and screening with nanogram amounts of immunopurified mRNAs: cDNA cloning and chromosomal mapping of cystathionine beta-synthase and the beta subunit of propionyl-CoA carboxylase. Proc Natl Acad Sci USA 83:2047–2051
- 149. Chadefaux B, Ceballos I, Hamet M, Coude M, Poissonnier M, Kamoun P, Allard D 1988 Is absence of atheroma in Down syndrome due to decreased homocysteine levels? [Letter] Lancet 2:741
- 150. Brattstrom L, Israelsson B, Tengborn L, Hultberg B 1989 Homocysteine, factor VII and antithrombin III in subjects with different gene dosage for cystathionmine  $\beta$ -synthase. J Inherit Metab Dis 12:475–482
- 151. Ubbink JB, Vermaak WJ, Delport R, van der Merwe A, Becker PJ,

- **Potgieter H** 1995 Effective homocysteine metabolism may protect South African blacks against coronary heart disease. Am J Clin Nutr 62:802–808
- 152. Malinow MR, Ducimetiere P, Luc G, Evans AE, Arveiler D, Cambien F, Upson BM 1996 Plasma homocyst(e)ine levels and graded risk for myocardial infarction: findings in two populations at contrasting risk for coronary heart disease. Atherosclerosis 126:27–34
- 153. **Dhawan J, Bray CL, Warburton R, Ghambhir DS, Morris J** 1994 Insulin resistance, high prevalence of diabetes, and cardiovascular risk in immigrant Asians. Genetic or environmental effect? Br Heart J 72:413–421
- 154. McKeigue PM, Shah B, Marmot MG 1991 Relationship of central obesity and insulin resistance with high diabetes prevalence and cardiovascular risk in South Asians. Lancet 337:382–386
- 155. Harker LA, Ross R, Slichter SJ, Scott CR 1976 Homocystineinduced arteriosclerosis: the role of endothelial cell injury and platelet response in its genesis. J Clin Invest 58:731–741
- 156. Di Minno G, Davi G, Margaglione M, Cirillo F, Grandone E, Ciabattoni G, Catalano I, Strisciuglio P, Andria G, Patrono C 1993 Abnormally high thromboxane biosynthesis in homozygous homocystinuria. Evidence for platelet involvement and probucolsensitive mechanism. J Clin Invest 92:1400–1406
- 157. **Durand P, Lussier-Cacan S, Blache D** 1997 Acute methionine load-induced hyperhomocysteinemia enhances platelet aggregation, thromboxane biosynthesis, and macrophage-derived tissue factor activity in rats. FASEB J 11:1157–1168
- 158. **Ratnoff OD** 1968 Activation of Hageman factor by L-homocystine. Science 162:1007–1009
- 159. **Rodgers GM, Conn MT** 1990 Homocysteine, an atherogenic stimulus, reduces protein C activation by arterial and venous endothelial cells. Blood 75:895–901
- 160. Lentz SR, Sadler JE 1991 Inhibition of thrombomodulin surface expression and protein C activation by the thrombogenic agent homocysteine. J Clin Invest 88:1906–1914
- 161. **Harpel PC, Zhang X, Borth W** 1996 Homocysteine and hemostasis: pathogenetic mechanisms predisposing to thrombosis. J Nutr 126: 1285S–1289S
- 162. Munnich A, Saudubray JM, Dautzenberg MD, Parry P, Ogier H, Girot R, Manigne P, Frezal J 1983 Diet responsive proconvertin (factor VII) deficiency in homocystinuria. J Pediatr 102:730–734
- Giannini MJ, Coleman M, Innerfield I 1975 Antithrombin activity in homocystinuria. [Letter] Lancet 1:1094
- 164. Lieberman ER, Gomperts ED, Shaw KNF, Landing BH, Donnell GN 1993 Homocystinuria: clinical and pathologic review, with emphasis on thrombotic features, including pulmonary artery thrombosis. Perspect Pediatr Pathol 17:125–147
- 165. Schienle HW, Seitz R, Rohner I, Lerch L, Krumpholz B, Krauss G, Fowler B, Baumgartner R, Willenbocker U, Egbring R 1994 Coagulation factors and markers of activation of coagulation in homocystinuria (HOCY): a study in two siblings. Blood Coagul Fibrinolysis 5:873–878
- 166. Fryer RH, Wilson BD, Gubler DB, Fitzgerald LA, Rodgers GM 1993 Homocysteine, a risk factor for premature vascular disease and thrombosis, induces tissue factor activity in endothelial cells. Arterioscler Thromb 13:1327–1333
- 167. Tawakol A, Omland T, Gerhard M, Wu JT, Creager MA 1997 Hyperhomocyst(e)inemia is associated with impaired endothelium-dependent vasodilation in humans. Circulation 95:1119–1121
- 168. Celermajer DS, Sorensen K, Ryalls M, Robinson J, Thomas O, Leonard JV, Deanfield JE 1993 Impaired endothelial function occurs in the systemic arteries of children with homozygous homocystinuria but not in their heterozygous parents. J Am Coll Cardiol 22:854–858
- 169. Lentz SR, Sobey CG, Piegors DJ, Bhopatkar MY, Faraci FM, Malinow MR, Heistad DD 1996 Vascular dysfunction in monkeys with diet-induced hyperhomocyst(e)inemia. J Clin Invest 98:24–29
- 170. Quere I, Hillaire-Buys D, Brunschwig C, Chapal J, Janbon C, Blayac JP, Petit P, Loubatieres-Mariani MM 1997 Effects of homocysteine on acetylcholine- and adenosine-induced vasodilatation on pancreatic vascular bed in rats. Br J Pharmacol 122:351–357
- 171. Stamler JS, Osborne JA, Jaraki O, Rabbani LE, Mullins M, Singel D, Loscalzo J 1993 Adverse vascular effects of homocysteine are

- modulated by endothelium-derived relaxing factor and related oxides of nitrogen. J Clin Invest 91:308–318
- 172. Wang J, Dudman NP, Wilcken DE, Lynch JF 1992 Homocysteine catabolism: levels of 3 enzymes in cultured human vascular endothelium and their relevance to vascular disease. Atherosclerosis 97:97–106
- 173. van der Molen EF, van den Heuvel LP, te Poele-Pothoff MT, Monnens LA, Eskes TK, Blom HJ 1996 The effect of folic acid on the homocysteine metabolism in human umbilical vein endothelial cells (HUVECs). Eur J Clin Invest 26:304–309
- 174. de Groot PG, Willems C, Boers GH, Gonsalves MD, van Aken WG, van Mourik JA 1983 Endothelial cell dysfunction in homocystinuria. Eur J Clin Invest 13:405–410
- 175. Jakubowski H 1997 Metabolism of homocysteine thiolactone in human cell cultures. Possible mechanism for pathological consequences of elevated homocysteine levels. J Biol Chem 272:1935– 1942
- 176. **Blundell G, Jones BG, Rose FA, Tudball N** 1996 Homocysteine mediated endothelial cell toxicity and its amelioration. Atherosclerosis 122:163–172
- 177. Matthias D, Becker CH, Riezler R, Kindling PH 1996 Homocysteine induced arteriosclerosis-like alterations of the aorta in normotensive and hypertensive rats following application of high doses of methionine. Atherosclerosis 122:201–216
- 178. Blom HJ, Kleinveld HA, Boers GH, Demacker PN, Hak-Lemmers HL, Te Poele-Pothoff MT, Trijbels JM 1995 Lipid peroxidation and susceptibility of low-density lipoprotein to *in vitro* oxidation in hyperhomocysteinaemia. Eur J Clin Invest 25:149–154
- 179. **Oszewski ÁJ, McCully KS** 1993 Homocysteine metabolism and the oxidative modification of proteins and lipids. Free Radic Biol Med 14:683–693
- 180. de Jong SC, Stehouwer CD, van den Berg M, Vischer UM, Rauwerda JA, Emeis JJ 1997 Endothelial marker proteins in hyperhomocysteinemia. Thromb Haemost 78:1332–1337
- 181. Van den Berg M, Boers GH, Franken DG, Blom HJ, Van Kamp GJ, Jakobs C, Rauwerda JA, Kluft C, Stehouwert CD 1995 Hyperhomocysteinemia and endothelial dysfunction in young patients with peripheral arterial occlusive disease. Eur J Clin Invest 25:176–181
- 182. Bienvenu T, Ankri A, Chadefaux B, Montalescot G, Kamoun P 1993 Elevated total plasma homocysteine, a risk factor for thrombosis. Relation to coagulation and fibrinolytic parameters. Thromb Res 70:123–129
- 183. Nishinaga M, Ozawa T, Shimada K 1993 Homocysteine, a thrombogenic agent, suppresses anticoagulant heparan sulfate expression in cultured porcine aortic endothelial cells. J Clin Invest 92: 1381–1386
- 184. **Quere I, Habib A, Tobelem G, Maclouf J** 1995 Inhibition of cyclooxygenase activity in human endothelial cells by homocysteine. Adv Prostaglandin Thromboxane Leukot Res 23:397–399
- 185. Tsai JC, Perrella MA, Yoshizumi M, Hsieh CM, Haber E, Schlegel R, Lee ME 1994 Promotion of vascular smooth muscle cell growth by homocysteine: a link to atherosclerosis. Proc Natl Acad Sci USA 91:6369–6373
- 186. Rolland PH, Friggi A, Barlatier A, Piquet P, Latrille V, Faye MM, Guillou J, Charpiot P, Bodard H, Ghiringhelli O 1995 Hyperhomocysteinemia-induced vascular damage in the minipig. Captopril-hydrochlorothiazide combination prevents elastic alterations. Circulation 91:1161–1174
- 187. Lubec B, Labudova O, Hoeger H, Muehl A, Fang-Kircher S, Marx M, Mosgoeller W, Gialamas J 1996 Homocysteine increases cyclindependent kinase in aortic rat tissue. Circulation 94:2620–2625

- 188. Majors A, Ehrhart LA, Pezacka EH 1997 Homocysteine as a risk factor for vascular disease. Enhanced collagen production and accumulation by smooth muscle cells. Arterioscler Thromb Vasc Biol 17:2074–2081
- 189. Wang H, Yoshizumi M, Lai K, Tsai JC, Perrella MA, Haber E, Lee ME 1997 Inhibition of growth and p21ras methylation in vascular endothelial cells by homocysteine but not cysteine. J Biol Chem 272:25380–25385
- 190. **Nishio E, Watanabe Y** 1997 Homocysteine as a modulator of platelet-derived growth factor action in vascular smooth muscle cells: a possible role for hydrogen peroxide. Br J Pharmacol 122:269 –274
- 191. Department of Health and Human Services 1996 Food standards: amendment of standards of identity for enriched grain products to require addition of folic acid. In: Federal Register Rules and Regulations, vol 61:8781–8807
- 192. Franken DG, Boers GH, Blom HJ, Trijbels FJ, Kloppenborg PW 1994 Treatment of mild hyperhomocysteinemia in vascular disease patients. Arterioscler Thromb 14:465–470
- 193. Palareti G, Salardi S, Piazzi S, Legnani C, Poggi M, Grauso F, Caniato A, Coccheri S, Cacciari E 1986 Blood coagulation changes in homocystinuria: effects of pyridoxine and other specific therapy. J Pediatr 109:1001–1006
- 194. **Barber GW, Spaeth GL** 1967 Pyridoxine therapy in homocystinuria. Lancet 1:337
- 195. Lipsom MH, Kraus J, Rosenberg LD 1980 Affinity of cystathionine β-synthase for pyridoxal 5'-phosphate in cultured cells. A mechanism for pyridoxine-responsive homocystinuria. J Clin Invest 66: 188–193
- 196. Brattström L, Israelsson B, Norrving B, Bergqvist D, Thorne J, Hultberg B, Hamfelt A 1990 Impaired homocysteine metabolism in early-onset cerebral and peripheral occlusive arterial disease. Effects of pyridoxine and folic acid treatment. Atherosclerosis 81: 51–60
- 197. Ubbink JB, Vermaak WJ, van der Merwe A, Becker PJ, Delport R, Potgieter HC 1994 Vitamin requirements for the treatment of hyperhomocysteinemia in humans. J Nutr 124:1927–1933
- 198. **Ubbink JB** 1998 Vitamin status and hyperhomocysteinemia in a healthy population. In: Graham I, Refsum H, Rosenburg I, Ueland PM (eds) Homocysteine Metabolism: From Basic Science to Clinical Medicine. Kluwer Academic Publishers, Boston, pp 93–98
- 199. Miller JW, Ribaya-Mercado JD, Russell RM, Shepard DC, Morrow FD, Cochary EF, Sadowski JA, Gershoff SN, Selhub J 1992 Effect of vitamin B-6 deficiency on fasting plasma homocysteine concentrations. Am J Clin Nutr 55:1154–1160
- 200. Boers GHJ, van den Berg M, Franken DG 1997 Treatment of mild hyperhomocysteinemia. In: Graham I, Refsum H, Rosenburg I, Ueland PM (eds) Homocysteine Metabolism: From Basic Science to Clinical Medicine. Kluwer Academic Publishers, Boston, pp 111– 116
- 201. van den Berg M, Franken DG, Boers GH, Blom HJ, Jakobs C, Stehouwer CD, Rauwerda JA 1994 Combined vitamin B6 plus folic acid therapy in young patients with arteriosclerosis and hyperhomocysteinemia. J Vasc Surg 20:933–940
- 202. Anonymous 1997 Betaine for homocystinuria. Med Lett 39:12
- Smolin LA, Benevenga NJ, Berlow S 1981 The use of betaine for the treatment of homocystinuria. J Pediatr 99:467–472
- 204. Wilcken DE, Wilcken B, Dudman NP, Tyrrell PA 1983 Homocystinuia the effect of betaine in the treatment of patients not responsive to pyridoxine. N Engl J Med 309:448–453