

REVIEW ARTICLE

Verrucomicrobial methanotrophs: ecophysiology of metabolically versatile acidophiles

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One sentence summary: This review discusses the metabolic versatility of verrucomicrobial methanotrophs regarding the acidic volcanic ecosystems they thrive in and a comparison is made with the canonical proteobacterial methanotrophs.

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ABSTRACT

Methanotrophs are an important group of microorganisms that counteract methane emissions to the atmosphere. Methane-oxidising bacteria of the Alpha- and Gammaproteobacteria have been studied for over a century, while methanotrophs of the phylum Verrucomicrobia are a more recent discovery. Verrucomicrobial methanotrophs are extremophiles that live in very acidic geothermal ecosystems. Currently, more than a dozen strains have been isolated, belonging to the genera *Methylacidiphilum* and *Methylacidimicrobium*. Initially, these methanotrophs were thought to be metabolically confined. However, genomic analyses and physiological and biochemical experiments over the past years revealed that verrucomicrobial methanotrophs, as well as proteobacterial methanotrophs, are much more metabolically versatile than previously assumed. Several inorganic gases and other molecules present in acidic geothermal ecosystems can be utilised, such as methane, hydrogen gas, carbon dioxide, ammonium, nitrogen gas and perhaps also hydrogen sulfide. Verrucomicrobial methanotrophs could therefore represent key players in multiple volcanic nutrient cycles and in the mitigation of greenhouse gas emissions from geothermal ecosystems. Here, we summarise the current knowledge on verrucomicrobial methanotrophs with respect to their metabolic versatility and discuss the factors that determine their diversity in their natural environment. In addition, key metabolic, morphological and ecological characteristics of verrucomicrobial and proteobacterial methanotrophs are reviewed.

Keywords: verrucomicrobial methanotrophs; geothermal ecosystems; methane; proteobacterial methanotrophs; hydrogen gas; metabolism; comparative genomic analysis

INTRODUCTION

The atmospheric concentration of methane (CH₄) has been increasing rapidly over the past quarter millennium due to

anthropogenic activities (Etheridge *et al.* 1998; Turner, Frankenberg and Kort 2019). Currently, the amount of methane emitted from sources exceeds the amount of methane taken up by sinks, resulting in an imbalanced carbon cycle (Saunio *et al.* 2020).

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Consequently, the present atmospheric methane concentration (1.86 ppmv) is 2.6 times higher than the preindustrial concentration (Etheridge et al. 1998; Cai et al. 2016; Etminan et al. 2016; Saunio et al. 2020). Since methane is a powerful greenhouse gas, atmospheric methane is now estimated to contribute approximately 16% to global warming (Saunio et al. 2016). Moreover, an increase in global temperature can induce additional release of methane to the atmosphere, e.g. through permafrost thawing, causing positive climate feedback that results in an acceleration of climate change (Dean et al. 2018). Hence, it is important to understand the microbial sources and sinks of methane (Stein 2020).

Each year, 548 to 678 Tg (10^{12} g) CH_4 is emitted from various sources into the atmosphere, of which 33%–54% and 46%–67% are from natural and anthropogenic origin, respectively (Kirschke et al. 2013; Dean et al. 2018). The majority of these sources contain methanogenic archaea that catalyse the final step of the biological degradation of organic matter by producing methane (Balch et al. 1979; Conrad 2020). The largest sources of methane are wetlands, which are anoxic, water-logged soils in which methanogenic archaea are present and active (Angel, Claus and Conrad 2012; Bridgham et al. 2013; Evans et al. 2019). Other natural sources of biogenic methane are aquatic systems (e.g. lakes), marine systems (especially coastal sediments), termites and wild animals (Jensen 1996; Reeburgh 2007; Bastviken et al. 2008; Brune 2010; Dean et al. 2018; Evans et al. 2019). Additional microbial processes that are implicated in methane production include conversion of methylphosphonates by Thaumarchaeota in the ocean and by cyanobacteria in freshwater and terrestrial ecosystems (Metcalfe et al. 2012; Bižić et al. 2020). Natural sources in which methane is not produced microbially but through abiotic processes include wildfires and methane hydrates (located inside ocean floors and permafrost) (Buffett 2000; Vasileva and Moiseenko 2013; Dean et al. 2018). In addition, various geothermal systems such as fumaroles, mud volcanoes and hydrothermal vents emit 40 to 60 Tg CH_4 annually (Etiope 2009). Methane released from these geothermal systems either has a thermogenic origin (e.g. formed through the thermogenic breakdown of organic matter over millions of years), an abiogenic origin (e.g. formed through the reduction of CO_2 at high temperature in the Earth's crust) or even a biogenic origin, produced by methanogens in the deep subsurface (Sherwood Lollar, Lacrampe-Couloume and Slater 2006; Tassi et al. 2012; Stolper et al. 2015). As is the case for several natural sources, various man-made systems emit significant amounts of methane produced by methanogenic archaea. These sources include livestock (e.g. ruminants), landfills and rice paddies (Themelis and Ulloa 2007; Carlson et al. 2017; Dean et al. 2018; Chang et al. 2019). Other anthropogenic, non-biological sources are comprised of fossil fuels (through mining, combustion and industry) and biofuel/biomass (through combustion) (Hao and Ward 1993; Heede 2014; Hanaki and Portugal-Pereira 2018). The predominant sink conversion of methane occurs chemically in the troposphere by hydroxyl radicals ($\cdot\text{OH}$) and to a lesser extent by chlorine and oxygen radicals (Allan, Struthers and Lowe 2007; Rigby et al. 2017). In various ecosystems, methane is produced in deeper anoxic layers that are covered by more oxidised sediments or water columns. In oxidised zones such as those present in wetlands, and notably at the oxic-anoxic interface, methanotrophic prokaryotes consume a major part of methane as energy and carbon source, prior to emission to the atmosphere (Brune, Frenzel and Cypionka 2000; Conrad 2009). These microorganisms therefore act as a biofilter for emissions of this potent greenhouse gas (La et al. 2018). In addition, specialised high-affinity

methanotrophs present in soil seem to oxidise methane from the atmosphere, although only present at a very low concentration (Holmes et al. 1999; Cai et al. 2016; Tveit et al. 2019). Considering the role methanotrophs play in mitigating methane emissions, they are an important topic of study in understanding and counteracting global warming.

METHANOTROPHIC MICROORGANISMS

Aerobic methanotrophs conserve energy by oxidising CH_4 with O_2 to CO_2 . The first aerobic methanotrophs were discovered and isolated more than a century ago (Kaserer 1905; Söhngen 1906). In the remainder of the twentieth century, numerous novel isolates were described, all belonging to the Alpha- and Gammaproteobacteria (Whittenbury, Phillips and Wilkinson 1970; Hanson and Hanson 1996). Methanotrophy is not restricted to oxic environments, but can take place anaerobically as well. Already decades ago, methane consumption in anoxic, sulfate-rich marine sediments was observed, but the microorganisms responsible for this process remained elusive (Barnes and Goldberg 1976; Reeburgh 1976; Iversen and Jørgensen 1985). At the beginning of the current century, a marine consortium of methane-oxidising archaea and sulfate-reducing bacteria was discovered, mediating sulfate-dependent methane oxidation (Boetius et al. 2000). This finding was followed by the discovery of a consortium of *Methylomirabilis* bacteria and *Methanoperedens* archaea, capable of coupling anaerobic methane oxidation to denitrification of nitrite and nitrate (Raghoebarsing et al. 2006). Hereafter it was shown that both the bacterium and the archaea were capable of methane oxidation independently. *Methanoperedens* archaea are able to couple methane oxidation to the reduction of nitrate (Haroon et al. 2013), iron (Beal, House and Orphan 2009; Ettwig et al. 2016; Cai et al. 2018) and manganese (Beal, House and Orphan 2009; Ettwig et al. 2016; Leu et al. 2020), whereas *Methylomirabilis* bacteria (of the candidate phylum NC10) couple methane oxidation to the reduction of nitrite (Ettwig et al. 2008, 2010; He et al. 2016; Versantvoort et al. 2018). Remarkably, *Methylomirabilis* bacteria possess the complete aerobic methane oxidation pathway and are postulated to produce oxygen internally by reducing nitrite to nitric oxide, which is subsequently dismutated into O_2 and N_2 (Ettwig et al. 2010, 2012). Most aerobic and anaerobic methanotrophs live in environments with moderate temperature and circum-neutral pH (Bowman et al. 1993). Exceptions are several aerobic proteobacterial methanotrophs of the genera *Methylocapsa*, *Methylocella*, *Methylocystis* and *Methylosinus* (Dedysh et al. 1998; Dunfield and Dedysh 2010; Kip et al. 2011). These bacteria are moderate acidophiles growing in acidic peat environments with a pH as low as 4.2 and are frequently found as intracellular symbionts of *Sphagnum* mosses (Raghoebarsing et al. 2005; Kostka, Weston and Glass 2016; Kox et al. 2018). In addition, methanotrophs of the gammaproteobacterial genus *Methylothermus* are thermophiles, growing in hot springs at temperatures up to 72°C (Bodrossy et al. 1999; Hirayama et al. 2011; Houghton et al. 2019). In 2005, methane oxidation was observed in volcanic soils of the Solfatara volcano (Campi Flegrei, Pozzuoli, Italy) that are characterized by a high temperature (50 to 95°C) and an extreme acidity (pH 1.0) (Castaldi and Tedesco 2005). Two years later, thermoacidophilic methanotrophs were isolated from hot and acidic volcanic ecosystems at the Solfatara volcano (near Naples, Italy), in the Uzon Caldera (Kamchatka, Russia) and in Hell's gate (Tikitere, New Zealand) (Dunfield et al. 2007; Pol et al. 2007; Islam et al. 2008). Surprisingly, phylogenetic analyses revealed that these microbes belong to the phylum Verrucomicrobia, refuting

the dogma that all aerobic methanotrophs are part of the phylum Proteobacteria.

METHANOTROPHS OF THE PHYLUM VERRUCOMICROBIA

The first isolated verrucomicrobial methanotrophs were classified in the novel genus *Methylacidiphilum* (Op den Camp et al. 2009). These methanotrophs were observed to have temperature optima of 50 to 60°C (Dunfield et al. 2007; Pol et al. 2007; Islam et al. 2008). Currently, five *Methylacidiphilum* strains have been isolated that differ enough, based on DNA-DNA hybridisation and on the analysis of 16S rRNA and housekeeping genes, to be classified as separate species (Erikstad et al. 2019): *Methylacidiphilum fumariolicum* SolV, *Methylacidiphilum infernorum* V4, *Methylacidiphilum kamchatkense* Kam1, *Methylacidiphilum* sp. Yel and *Methylacidiphilum* sp. Phi (Table 1). The geothermal ecosystems they were isolated from have an extremely low pH, primarily as a result of the biogenic formation of sulfuric acid (H₂SO₄) from the oxidation of hydrogen sulfide (H₂S) (Schoen and Rye 1970; Quatrini and Johnson 2018). Accordingly, the pH optima of all isolated *Methylacidiphilum* strains range from pH 2.0 to 3.0. Still, the pH range in which these strains can grow is much broader (Table 1). *Methylacidiphilum fumariolicum* SolV was shown to grow up to pH 6 when it was slowly adapted to higher pH (Mohammadi et al. 2017b). Moreover, this strain was shown to grow even below pH 1 (Pol et al. 2007) (Table 1). Through current culture-independent molecular methods, clones related to the isolated strains were found at geothermal sites across the globe, of which several could be novel species (Fig. 1).

Since all isolated *Methylacidiphilum* strains were found in hot and acidic geothermal habitats, the question arose whether verrucomicrobial methanotrophs could have a more widespread distribution in habitats with different physicochemical parameters (Dunfield et al. 2007). Therefore, Sharp et al. (2014) conducted pyrosequencing of 16S rRNA genes on samples derived from geothermal habitats, acidic peat bogs and fens with a temperature range of 6.3 to 81.6°C and a pH range of 1.8 to 8.6. Surprisingly, 16S rRNA gene sequences of verrucomicrobial methanotrophs were detected in the full range of 22.5 to 81.6°C, but only below pH 5.0 (Sharp et al. 2014). From a sediment sample of 22°C and pH 2.6, the novel verrucomicrobial strain LP2A was enriched and isolated when incubated with methane as the sole energy source (Sharp et al. 2014). Interestingly, the 16S rRNA sequence of this newly isolated strain was only 90.6% identical to that of *Methylacidiphilum infernorum* V4, isolated from hot and acidic soil sediment (Dunfield et al. 2007). The novel mesophilic strain LP2A was subsequently placed in the novel genus *Methylacidimicrobium* within the phylum Verrucomicrobia (Van Teeseling et al. 2014). That same year, three additional mesophilic strains within this genus were isolated from geothermal soil at the Solfataria in Italy: *Methylacidimicrobium tartarophylax* 4AC, *Methylacidimicrobium fagopyrum* 3C and *Methylacidimicrobium cyclopophantes* 3B (Van Teeseling et al. 2014) (Fig. 1). These acidophiles have pH growth optima of 1.0 to 3.0, while strain 4AC can even grow at pH 0.5 (Van Teeseling et al. 2014) (Table 1). The major difference between *Methylacidiphilum* and *Methylacidimicrobium* strains was thought to be the growth temperature. *Methylacidimicrobium* strains were reported to be mesophiles, with temperature growth optima of 30 to 44°C and the apparent inability to grow at temperatures

above 49°C (Sharp et al. 2014; Van Teeseling et al. 2014). However, recently, *Methylacidimicrobium thermophilum* AP8 was isolated from Pantelleria island (Italy) with a temperature optimum of 50°C (Picone 2020), questioning the clear temperature-based division between *Methylacidiphilum* and *Methylacidimicrobium* strains.

Apart from the strains isolated from geothermal soils in Italy, Russia and New-Zealand, 16S rRNA gene sequences belonging to strains of the *Methylacidimicrobium* genus were detected in hot springs and geothermal soil in other parts of the world (Fig. 1). In addition, a metagenome-assembled genome (MAG) representing a novel verrucomicrobial methanotroph genus and species, Ca. 'Methylacidithermus pantelleriae' PQ17, was obtained through sampling of geothermal soil on Pantelleria island (Picone et al. 2020a). Interestingly, verrucomicrobial methanotrophs do not seem to be restricted to geothermal habitats (Pagaling, Yang and Yan 2014). Clones related to *Methylacidimicrobium* strains were abundantly present in biofilms in the crown of corroded concrete sewage pipes on Hawaii, which suggests that verrucomicrobial methanotrophs have adapted to man-made systems (Pagaling, Yang and Yan 2014) (Fig. 1). These sewage pipes are characterised by a low pH and the presence of methane and sulfur compounds (e.g. H₂S), which could explain the abundance of verrucomicrobial methanotrophs in this environment. Sequencing of samples from other acidic, methane-rich environments could reveal whether verrucomicrobial methanotrophs are present in other non-geothermal habitats.

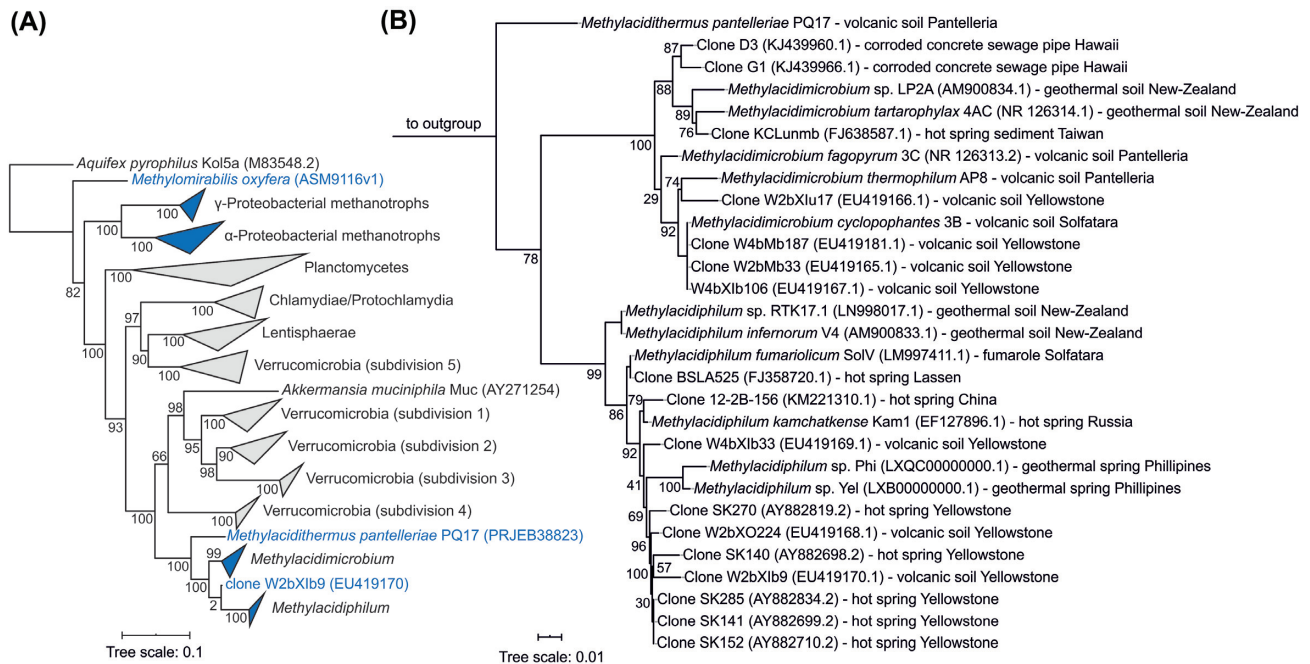
Besides the importance of verrucomicrobial methanotrophs in the environment, they could also be relevant in biotechnology and industry, as they are capable of producing valuable molecules such as methanol, glycogen, polyhydroxybutyric acid, vitamins, compatible solutes and (thermostable) enzymes (Kalyuzhnaya, Puri and Lidstrom 2015; Strong, Xie and Clarke 2015; Bodelier et al. 2019). Although still in its infancy, notable examples of the biotechnological potential of verrucomicrobial methanotrophs are the recently shown improved methanol production (63% mole methanol produced per mole methane) by *Methylacidiphilum fumariolicum* SolV (Hogendoorn 2020; Hogendoorn et al. 2020) and the purification of a thermostable, high-affinity [NiFe] hydrogenase from the same bacterium (Schmitz et al. 2020a).

KEY SIMILARITIES AND DIFFERENCES BETWEEN VERRUCOMICROBIAL AND PROTEOBACTERIAL METHANOTROPHS

Proteobacterial methanotrophs can be found in diverse oxic environments in which methane is present, such as wetlands, soil, peat lands, marine sediments, landfills and rice paddies (Bodelier et al. 2019). On the contrary, verrucomicrobial methanotrophs are almost exclusively found in acidic (pH < 3.5) geothermal systems. Whether proteobacterial methanotrophs are present in such acidic habitats is unknown. However, phylogenetic analyses of *pmoA* genes present in acidic geothermal soil and mud pots show clustering with proteobacterial *pmoA* gene sequences (Pol et al. 2007; Gagliano et al. 2014). This finding suggests that verrucomicrobial and proteobacterial methanotrophs could share habitats, which could ultimately be proven by the isolation of a proteobacterial methanotroph from an acidic geothermal ecosystem where *Methylacidiphilum* or *Methylacidimicrobium* were isolated from. Although mostly living in vastly different habitats, they share their preference for the

Table 1. Summary of the pH and temperature optima, isolation location and verified growth substrates of the isolated verrucomicrobial methanotrophs discussed in this review.

Strain	pH _{optimum} (and range)	T _{optimum} (°C) (and range)	Verified growth substrates	Isolation location	References
<i>Methylacidiphilum fumarolicum</i> SolV	2 (0.8–6.0)	55 (40–65)	CH ₄ , H ₂ , methanol, propane, ethane	Acidic thermal mudpot, Solfatara, Italy	Pol et al. 2007; Mohammadi et al. 2017a; Picone et al. 2020b
<i>Methylacidiphilum inferorum</i> V4	2–2.5 (1.0–6.0)	60 (40–60)	CH ₄ , methanol	Acidic thermal soil, Tikitere, New Zealand	Dunfield et al. 2007; Hou et al. 2008
<i>Methylacidiphilum kamchatkense</i> Kam1	2–2.5 (2.0–5.0)	55 (37–60)	CH ₄ , methanol	Acidic thermal spring, Kamchatka, Russia	Islam et al. 2008
<i>Methylacidiphilum</i> sp. Phi	3	55–65	CH ₄	Acidic hot spring, Makiling, The Philippines	Erikstad et al. 2019
<i>Methylacidiphilum</i> sp. Yel	2.8	50	CH ₄	Acidic hot spring, Yellowstone, USA	Erikstad et al. 2019
<i>Methylacidimicrobium tartarophylax</i> 4AC	1–3 (0.5–5.5)	38 (?–43)	CH ₄ , H ₂ , methanol	Acidic soil, Solfatara, Italy	Van Teeseling et al. 2014; Mohammadi et al. 2019
<i>Methylacidimicrobium cyclophantes</i> 3B	1.5–3 (0.6–5.5)	44 (?–49)	CH ₄ , methanol	Acidic soil, Solfatara, Italy	Van Teeseling et al. 2014
<i>Methylacidimicrobium fagopyrum</i> 3C	1.5–3 (0.6–5.5)	35 (?–39)	CH ₄ , methanol	Acidic soil, Solfatara, Italy	Van Teeseling et al. 2014
<i>Methylacidimicrobium</i> sp. LP2A	3.1 (1.0–5.2)	30 (17–37)	CH ₄	Acidic mud pool, Reporoa, New Zealand	Sharp et al. 2014
<i>Methylacidimicrobium thermophilum</i> AP8	3–5 (1.5–5.5)	50 (30–55)	CH ₄ , H ₂	Acidic geothermal soil, Pantelleria island, Italy	Picone 2020

**Figure 1.** (A) Overview 16S rRNA gene phylogenetic tree of Verrucomicrobia, Lentisphaerae, Chlamydiae, Planctomycetes and methanotrophic Alpha- and Gammaproteobacteria. Blue shading indicates species that are methanotrophic. Accession numbers are indicated between brackets. 16S rRNA gene alignment was constructed using SINA v1.2.11 (Pruesse, Peplies and Glöckner 2012) and a tree that was constructed using FastTree v2.1 (Price, Dehal and Arkin 2010) with 1000 bootstraps and substitution model GTR-gamma. The tree was visualised using ITOL v5.5.1 (Letunic and Bork 2007). Bootstrap values are indicated as a proportion of 1000 re-samplings ranging from 1 to 100. (B) Detailed 16S rRNA gene tree of the verrucomicrobial methanotrophs. 16S rRNA gene alignment of verrucomicrobial methanotrophs together with an outgroup of Planctomycetes constructed using SINA v1.2.11 (Pruesse, Peplies and Glöckner 2012) and a tree that was constructed using FastTree v2.1 (Price, Dehal and Arkin 2010) with 1000 bootstraps and substitution model GTR-gamma. The tree was visualised using ITOL v5.5.1 (Letunic and Bork 2007). Bootstrap values are indicated as a proportion of 1000 re-samplings ranging from 1 to 100. Isolation location and accession number are indicated after each strain.

greenhouse gas methane. Besides this metabolic trait, several noteworthy differences exist between the two groups. Classically, an important property for the classification of proteobacterial methanotrophs is the type of intracytoplasmic membrane (ICM) structures they have in electron micrographs, either visible as membrane pairs or vesicular discs (Davies and Whittenbury 1970). In verrucomicrobial methanotrophs, ICM structures are mostly absent, although membrane stacks have been found in *Methylacidimicrobium fagopyrum* 3C and also in some cells of *Methylacidiphilum inferorum* V4 tubular membrane structures were observed (Dunfield et al. 2007; Van Teeseling et al. 2014). Another interesting difference between verrucomicrobial and proteobacterial methanotrophs are the phospholipid fatty acids (PLFAs) of their membranes (Bodelier et al. 2009; Op den Camp et al. 2009). Membranes of verrucomicrobial methanotrophs are almost exclusively made up of saturated fatty acids, whereas membranes of proteobacterial methanotrophs are mainly composed of unsaturated fatty acids (Bodelier et al. 2009; Op den Camp et al. 2009; Erikstad et al. 2019). Verrucomicrobial methanotrophs probably require a saturated membrane to minimise proton permeability in an extremely acidic environment (Siliakus, van der Oost and Kengen 2017).

Remarkably, the genome sizes of verrucomicrobial and proteobacterial methanotrophs differ significantly. The former typically have a genome size of 2.2 to 2.5 Mbp (Hou et al. 2008; Anvar et al. 2014; Kruse et al. 2019; Cremers et al. 2020) while the latter have larger genomes of 3.3 to 5.1 Mbp (Ward et al. 2004; Chen et al. 2010; Stein et al. 2010; Boden et al. 2011; Svenning et al. 2011; Vuilleumier et al. 2012). The larger genomes of proteobacterial methanotrophs could render them better adapted to more diverse environments in comparison with verrucomicrobial methanotrophs (Cobo-Simón and Tamames 2017). Interestingly, genome comparison of eleven verrucomicrobial methanotrophs reveals 317 core gene clusters that are shared by all these verrucomicrobial strains, whereas the same analysis of eleven proteobacterial methanotrophs reveals 74 core gene clusters shared by all these proteobacterial strains (Fig. 2). The smaller amount of core gene clusters shared by proteobacterial methanotrophs could be explained by the fact that these microbes are found in very distinct habitats under various circumstances. The verrucomicrobial methanotrophs, on the other hand, are adapted to a specific niche. The composition of Clusters of Orthologous Groups of proteins (COGS) of the core genomes of verrucomicrobial and proteobacterial methanotrophs show a similar distribution (Fig. 2). A notable example is the relatively large percentage unique gene clusters involved in cell wall/membrane/envelope biogenesis (COG group M) found in verrucomicrobial methanotrophs. This might be explained by the harsh environment in which these microbes thrive, which could require additional machinery to cope with acid stress.

Although verrucomicrobial methanotrophs have smaller genomes, genomic analyses suggest verrucomicrobial methanotrophs are metabolically quite versatile microorganisms. In this review, we discuss the physiological and biochemical knowledge that has been obtained on the key metabolic pathways of verrucomicrobial methanotrophs since their discovery 13 years ago. As such, a connection between the metabolic potential in the genome and experimental observations is made. Metabolic genes investigated in this review involved in the oxidation of methane, ammonia, hydrogen and sulfur compounds, nitrogen assimilation, the respiratory chain and the synthesis of tetrahydrofolate and menaquinone can be found in Table S1 (Supporting Information).

METABOLIC VERSATILITY OF VERRUCOMICROBIAL METHANOTROPHS

Initially, the first isolated strains appeared to be obligate methanotrophs, growing only on methane and methanol (CH₃OH) (Op den Camp et al. 2009). In fact, all described *Methylacidiphilum* and *Methylacidimicrobium* strains were isolated on methane (Dunfield et al. 2007; Pol et al. 2007; Islam et al. 2008; Sharp et al. 2014; Erikstad et al. 2019). However, after more than a decade of additional experimental research, we now know that verrucomicrobial methanotrophs are actually metabolically quite versatile microorganisms, able to metabolise a variety of compounds present in volcanic ecosystems, which together could influence methanotrophy.

Carbon and nitrogen assimilation

Similar to any other microorganism, methanotrophs need to assimilate carbon and nitrogen to generate biomass (Levicán et al. 2008). Methanotrophs use either formaldehyde (CH₂O) or CO₂ as a carbon source to generate biomass (Hanson and Hanson 1996). In general, gammaproteobacterial methanotrophs assimilate formaldehyde via the ribulose monophosphate pathway, whereas alphaproteobacterial methanotrophs employ the serine pathway, in which both formaldehyde and CO₂ are used as a carbon source (Chistoserdova, Kalyuzhnaya and Lidstrom 2009). Verrucomicrobial methanotrophs lack a few essential genes for the complete serine cycle (i.e. hydroxypyruvate reductase and malyl-CoA lyase) or ribulose monophosphate pathway (i.e. 3-hexulose-6-phosphate synthase and 6-phospho-3-hexuloisomerase), but possess genes encoding the complete set of enzymes of the Calvin-Benson-Bassham (CBB) cycle (Khadem et al. 2011; Van Teeseling et al. 2014). In this cycle, the enzyme ribulose-1,5-bisphosphate carboxylase/oxygenase (RuBisCO) is involved and CO₂ is used as the sole carbon source. Several proteobacterial methanotrophs also harbour genes involved in the CBB cycle, although activity of this cycle has not yet been experimentally validated in these bacteria (Stein, Roy and Dunfield 2012; Khmelenina et al. 2018). Through ¹³C stable isotope measurements, *Methylacidiphilum fumarolicum* SolV and three *Methylacidimicrobium* strains were shown to indeed exclusively use CO₂ as carbon source, not methane-derived formaldehyde (Khadem et al. 2011; Van Teeseling et al. 2014). The autotrophic CO₂ fixation via the CBB cycle was also experimentally shown for the denitrifying methanotroph *Ca. 'Methylomirabilis oxyfera'* (Rasigraf et al. 2014). In geothermal habitats, high concentrations of ammonium can be present (Khadem et al. 2010; Holloway et al. 2011; Mohammadi et al. 2017b). Although methanotrophs cannot use ammonium as an energy source, this reduced form of fixed nitrogen is the preferred inorganic nitrogen source as it can be directly assimilated (Wang et al. 2016). Indeed, all known verrucomicrobial methanotrophs carry a gene encoding a highly-conserved ammonium transporter. Central molecules in nitrogen assimilation are glutamate and glutamine, from which other amino acids and purines and pyrimidines can be synthesised (Prusiner and Stadtman 1973; Zalkin and Smith 1998). Genes encoding glutamate dehydrogenase, glutamine synthetase and glutamate synthase are found in all isolated verrucomicrobial methanotrophs. The utilisation of these three enzymes for nitrogen assimilation has been validated experimentally in several proteobacterial methanotrophs (Murrell and Dalton 1983a; Lees, Owens and Murrell 1991).

Although ammonium is often present in the geothermal environment, exponentially growing microorganisms can

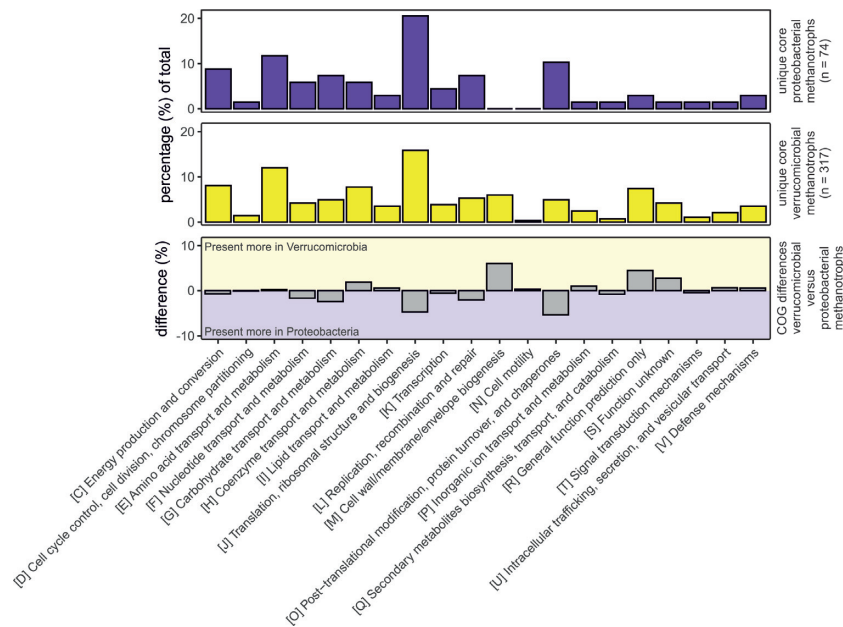


Figure 2. Bar graphs comparing the composition of Clusters of Orthologous Groups of proteins (COGs) of the unique core genomes of the verrucomicrobial methanotrophs and the proteobacterial methanotrophs. Core genomes were calculated by using usearch (Edgar 2010) with a cut-off value of 0.5 to group all genes into gene clusters. Clusters present in all genomes of the verrucomicrobial methanotrophs, as well as absent in all genomes of the proteobacterial methanotrophs, are defined as the unique core genome of verrucomicrobial methanotrophs. Clusters present in all genomes of the proteobacterial methanotrophs, as well as absent in all genomes of the verrucomicrobial methanotrophs, are defined as the unique core genome of proteobacterial methanotrophs. Analysis performed with 22 genomes in total, of which 11 verrucomicrobial methanotrophs: *Methylacidiphilum fumariolicum* SolV, *Methylacidiphilum inferorum* V4, *Methylacidiphilum kamchatkense* Kam1, *Methylacidiphilum* sp. Phi, *Methylacidiphilum* sp. Yel, *Methylacidimicrobium cyclophantes* 3B, *Methylacidimicrobium fagopyrum* 3C, *Methylacidimicrobium tartarophylax* 4A, *Methylacidimicrobium thermophilum* AP8, *Methylacidimicrobium* sp. LP2A and *Methylacidithermus pantelleriae* PQ17, and 11 proteobacterial methanotrophs: *Methylobacterium extorquens* AM1, *Methylocaldum szegediense* O-12, *Methylocapsa acidiphila* B2, *Methylocella silvestris* BLZ, *Methylococcus capsulatus* Bath, *Methylocystis rosea* SV97T, *Methyloferula stellata* AR4, *Methylomarinum vadi* IT4, *Methylomicrobium alcaliphilum* 20Z, *Methylomonas denitrificans* FJG1 and *Methylorinus trichosporium* OB3B.

rapidly deplete this source of nitrogen from their direct surroundings (Van Heeswijk, Westerhoff and Boogerd 2013). If insufficient ammonium is present in the environment, many microorganisms have the possibility to take up nitrate or nitrite from the environment and invest energy to produce ammonium (Moreno-Vivián et al. 1999). All isolated *Methylacidiphilum* strains possess a gene encoding a transporter that could transport nitrate and/or nitrite across the membrane. In addition, they harbour a gene encoding a cytoplasmic NAD(P)H-dependent nitrate reductase that reduces nitrate to nitrite and a cytoplasmic NAD(P)H-dependent nitrite reductase that catalyses the 6-electron reduction to ammonium. The assimilatory reduction of nitrate to ammonium is catalysed at the expense of four NAD(P)H molecules and is therefore only used when ammonium in the environment is scarce. In several *Methylacidimicrobium* strains, the pathway for nitrate assimilation is unclear, or could be absent.

All isolated verrucomicrobial methanotrophs possess the *nifHDK* operon, indicating their ability to fix N_2 from the atmosphere. *Methylacidiphilum fumariolicum* SolV and *Methylacidiphilum kamchatkense* Kam1 were experimentally shown to use N_2 as nitrogen source (Islam et al. 2008; Khadem et al. 2010). Diazotrophy has also been shown for several proteobacterial methanotrophs (Murrell and Dalton 1983b; Khmelenina et al. 2018). Although the ability to obtain nitrogen from the atmosphere provides a major advantage in an environment devoid of fixed nitrogen molecules, the fixation of nitrogen gas is very energy-demanding, requiring 16 ATP molecules per N_2 molecule fixed (Dixon and Kahn 2004). As a result, the maximum specific growth rate of *Methylacidiphilum fumariolicum* SolV with N_2 as nitrogen source is 2.8 times slower than when supplied with

ammonium (Khadem et al. 2010). The absence of fixed nitrogen in the environment might therefore lead to a reduction in growth of verrucomicrobial methanotrophs and therefore to increased methane emissions.

Oxidation of methane and other hydrocarbons

Significant amounts of methane are emitted from mud pools, hot springs and fumaroles in terrestrial geothermal systems (Etiope and Klusman 2002; Castaldi and Tedesco 2005; Kvenvolden and Rogers 2005). Typically, geothermal gas consists of 0.01 to 1% (v/v) methane, although much higher concentrations have been detected (Giggenbach 1995; Etiope and Klusman 2002; D'Alessandro et al. 2009; Dunfield and Dedysh 2010). Under suitable conditions, verrucomicrobial methanotrophs could represent a significant filter against emissions of methane to the atmosphere in these environments (Op den Camp et al. 2009; Venturi et al. 2019). Studying in which circumstances verrucomicrobial methanotrophs consume more methane than in others is worth investigating to elucidate which factors enhance or inhibit methane oxidation.

The first step in methane oxidation is the conversion of methane to methanol (CH_3OH), which can be catalysed by two genetically unrelated enzymes: the membrane-bound copper-containing particulate methane monooxygenase (pMMO) and the cytosolic iron-containing methane monooxygenase (sMMO) (Ross and Rosenzweig 2017). Several proteobacterial methanotrophs harbour genes encoding both the soluble and the particulate methane monooxygenase (Semrau, DiSpirito and Yoon 2010). However, none of the verrucomicrobial methanotrophs carry a gene encoding sMMO. *Methylacidiphilum* strains possess

three complete but distinct *pmoCAB* operons and therefore in all probability oxidise methane to methanol via pMMO (Hou et al. 2008; Anvar et al. 2014; Van Teeseling et al. 2014; Kruse et al. 2019) (Figs 3 and 4). As exception, *Methylacidiphilum* sp. Yel only carries a single complete *pmoCAB* operon most closely related to the *pmoCAB3* of the other *Methylacidiphilum* strains. All pMMOs characterised thus far show an $(\alpha\beta\gamma)_3$ oligomeric state, consisting of the two membrane-spanning subunits PmoC and PmoA and a predominantly periplasmic subunit PmoB, which is tethered to the membrane by two transmembrane helices (Lieberman and Rosenzweig 2005; Ross and Rosenzweig 2017). The enzyme activates O_2 to break the C-H bond of methane, producing methanol and water, via a yet unresolved mechanism (Ross and Rosenzweig 2017; Ross et al. 2019) (Fig. 4). Interestingly, similarities between two *pmo* operon structures of proteobacterial and verrucomicrobial methanotrophs indicate evolution from a common ancestor (Op den Camp et al. 2009; Van Teeseling et al. 2014). Besides, the most distant *pmoCAB3* operon seems to be obtained through horizontal gene transfer from an unknown microorganism (Dunfield et al. 2007). In fact, a large variety of key metabolic genes of verrucomicrobial methanotrophs are thought to be derived via horizontal gene transfer, especially from Proteobacteria (Sharp et al. 2013).

In *Methylacidiphilum fumariolicum* SolV, the oxygen concentration seems to regulate the expression of the *pmoCAB1* and *pmoCAB2* operons (Khadem et al. 2012). Notably, the *pmoCAB3* operon is barely expressed in *Methylacidiphilum fumariolicum* SolV when grown on methane (Khadem et al. 2012). Recently, *Methylacidiphilum fumariolicum* SolV was shown to grow on ethane and propane but not on butane (Picone et al. 2020b). *pmoCAB3* could be specifically tuned towards the oxidation of alkanes, enabling *Methylacidiphilum fumariolicum* SolV to utilise gaseous hydrocarbons other than methane (Picone et al. 2020b). However, *Methylacidiphilum* sp. Yel only carries *pmoCAB3* and is able to grow on methane, suggesting the enzyme encoded by this operon is at least also involved in the oxidation of methane (Erikstad et al. 2019). In contrast to *Methylacidiphilum* strains, *Methylacidimicrobium* strains and Ca. 'Methylacidithermus pantelleriae' PQ17 possess a single *pmoCAB* operon, related to the closely related *pmoCAB1* and *pmoCAB2* types of *Methylacidiphilum* species. As exception, *Methylacidimicrobium thermophilum* AP8 and *Methylacidimicrobium* sp. LP2A possess two almost identical operons (Sharp et al. 2014).

For a long time, the well-studied calcium-dependent methanol dehydrogenase (MDH) MxaFI was thought to be the key enzyme for methanol oxidation in both methanotrophs and methylotrophs (Anthony 2004). This periplasmic enzyme contains a pyrroloquinoline quinone (PQQ) prosthetic group at the catalytic centre and converts methanol to formaldehyde (CH_2O) (Anthony 2004; Keltjens et al. 2014). The alphaproteobacterial methylotroph *Methylobacterium extorquens* AM1 carries genes encoding MxaFI and an MDH homolog named XoxF (Nakagawa et al. 2012). A $\Delta mxaF$ mutant strain was unable to grow on methanol when supplied with calcium, but growth on methanol was restored when cells were supplied with the lanthanide lanthanum (La^{3+}) in the medium. Similarly, *Methylacidiphilum fumariolicum* SolV is only able to grow on methane when lanthanide-containing mud pot water (from the environment where the strain was isolated from) is added to the growth medium, or when the mineral medium contains micromolar concentrations of one or more lanthanides (Pol et al. 2014). These lanthanides are essential for verrucomicrobial methanotrophs when using methanol as an energy source, because they do not possess genes encoding

the canonical calcium-dependent MxaFI, but a gene encoding the novel lanthanide-dependent MDH (XoxF) (Pol et al. 2014) (Fig. 3). Remarkably, the biological relevance of these rare earth elements was previously deemed unthinkable (Lim and Franklin 2004). Genomic analyses surprisingly revealed *xoxF* to be widespread in methylotrophs and methanotrophs, and is classified into five different clades (Chistoserdova 2011; Keltjens et al. 2014; Pol et al. 2014; Wu et al. 2015; Versantvoort et al. 2018; Kato et al. 2020). In fact, XoxF-type MDHs are actually much more abundant in nature than the MxaFI-type (Keltjens et al. 2014, Chistoserdova and Kalyuzhnaya 2018). The MxaF protein (large subunit) seems to have descended from a XoxF prototype (Keltjens et al. 2014) and all methylotrophs that harbour genes encoding MxaFI also possess a gene encoding the XoxF-type MDH (Chistoserdova and Kalyuzhnaya 2018). Dissolved lanthanide concentrations in most environments are very low (nM range) and its limitation may have driven the evolutionary adaptation of the proteobacterial methylotrophs towards an MDH variant that depends on calcium, present in excess. The discovery of lanthanide-dependent XoxF-type MDHs has led to a completely new field of research, including the differential regulation in bacteria containing both types of MDHs (Zheng et al. 2018; Daumann 2019; Good et al. 2019; Picone and Op den Camp 2019; Good et al. 2020; Yanpirat et al. 2020). As example, the alphaproteobacterium *Methylobacterium extorquens* AM1 carries genes encoding both MDH types and was shown to upregulate *xoxF* expression already at a lanthanide concentration of 2.5 nM and downregulate *mxaf* expression at a lanthanide concentration of 25 nM or above (Vu et al. 2016; Daumann 2019).

Electrons yielded from methanol oxidation by XoxF are first transferred to a dedicated cytochrome partner (Zheng et al. 2018; Versantvoort et al. 2019) homologous to cytochrome c_L , the electron acceptor of MxaFI-type MDH (Anthony 1992) (Fig. 4). In *Methylacidiphilum* strains, this cytochrome is a fusion protein of the cytochrome c protein XoxG and the periplasmic substrate binding protein XoxJ, but they also harbour a separate *xoxJ* gene. In *Methylacidimicrobium* strains, XoxG and XoxJ are encoded by two separate genes. *Methylacidiphilum* strains possess only one XoxF enzyme, whereas *Methylacidimicrobium* strains carry genes encoding for a copy of both the XoxF1 and XoxF2 clade that are less than 50% identical, which suggests that they could be expressed differentially under different conditions (Chistoserdova 2011; Keltjens et al. 2014; Picone and Op den Camp 2019). In addition, *Methylacidimicrobium* sp. LP2A harbours two genes encoding XoxF1 that are 96% identical to each other, presumably as a result of a relatively recent gene duplication (Van Teeseling et al. 2014; Op den Camp et al. 2018).

Similar to MxaFI-type MDH, XoxF-type MDH also contains a PQQ cofactor, and its overall structure is conserved in comparison with the MxaFI-type MDH (Keltjens et al. 2014). From *Methylacidiphilum fumariolicum* SolV, cultivated on lanthanide-containing mud pot water, XoxF was crystallised as a homodimer with a cerium ion coordinating with the PQQ cofactor (PDB: 4MAE; Pol et al. 2014). In addition, XoxF-type MDHs with a europium or lanthanum ion in the active site were crystallised (Deng, Ro and Rosenzweig 2018; Jahn et al. 2018; Good et al. 2020). The lanthanide coordinating with the PQQ cofactor of XoxF seems to render this enzyme catalytically more efficient compared to MxaF, since lanthanides are stronger Lewis acids than calcium, facilitating the hydride transfer (Keltjens et al. 2014; Daumann 2019). MxaFI-type MDHs produce formaldehyde from methanol oxidation, but XoxF was shown to produce formate (HCOOH) *in vitro* (Pol et al. 2014). Moreover, XoxF

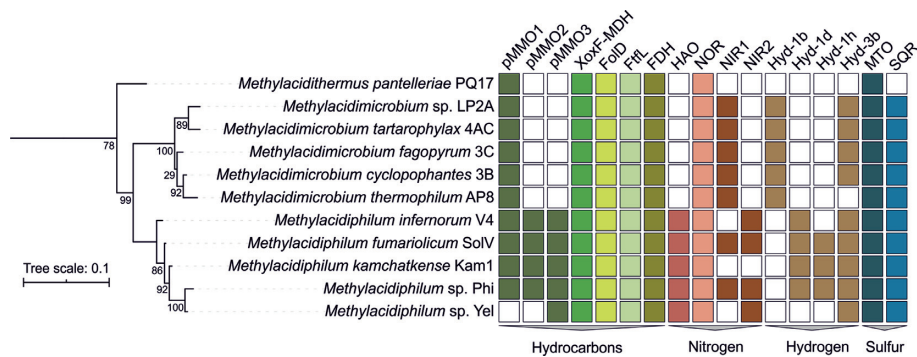


Figure 3. 16S rRNA gene phylogenetic tree of verrucomicrobial methanotrophs and the presence or absence of genes involved in the oxidation of hydrocarbons, ammonia, hydrogen and sulfur compounds. Planctomycetes were used as an outgroup. Sequences were aligned using SINA v1.2.11 (Pruesse, Peplies and Glöckner 2012) and a tree was constructed using FastTree v2.1 (Price, Dehal and Arkin 2010) with 1000 bootstraps and substitution model GRT-GAMMA. The tree was visualised using ITOL v5.5.1 (Letunic and Bork 2007). Bootstrap values are indicated as a proportion of 1000 re-samplings ranging from 1 to 100. Presence of genes of interest was examined using BLASTp and of each hit the amino acid sequence was manually inspected for domains and identity. The result table was transformed to a binary ITOL dataset and both the 16S tree and the table were visualised using ITOL v5.5.1. pMMO: particulate methane monooxygenase; XoxF-MDH: lanthanide-dependent methanol dehydrogenase; FoD: methylenetetrahydrofolate dehydrogenase/methenyltetrahydrofolate cyclohydrolase; FtlL: formate-tetrahydrofolate ligase; FDH: formate dehydrogenase; HAO: hydroxylamine oxidoreductase; NOR: nitric oxide reductase; NIR: nitrite reductase; Hyd: type of [NiFe] hydrogenase (based on Søndergaard, Pedersen and Greening 2016); MTO: methanethiol oxidase; SQR: sulfide:quinone oxidoreductase.

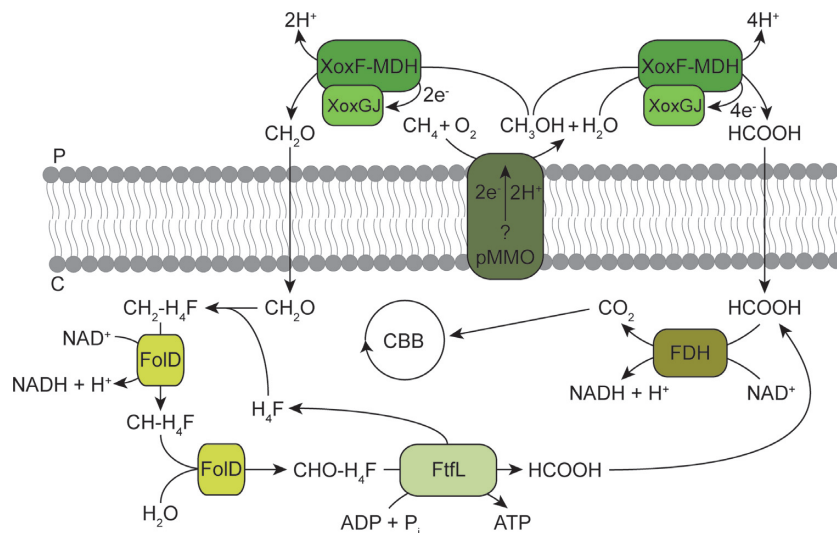


Figure 4. Possible pathways for methane oxidation in verrucomicrobial methanotrophs. pMMO oxidises methane to methanol (CH_3OH), while an unknown electron donor is oxidised. The lanthanide-dependent XoxF methanol dehydrogenase (MDH) could subsequently oxidise methanol to either formate (HCOOH) or formaldehyde (CH_2O), while donating electrons to its redox partner XoxGj. If formate is produced it could diffuse into the cytoplasm and be converted to CO_2 by the NAD^+ -dependent formate dehydrogenase (FDH). CO_2 is fixed into biomass via the Calvin-Benson-Bassham (CBB) cycle. Alternatively, if formaldehyde is produced, it could bind to tetrahydrofolate (H_4F) spontaneously or enzymatically by an unidentified enzyme, to form methylene-tetrahydrofolate ($\text{CH}_2\text{-H}_4\text{F}$). The enzyme FoD converts $\text{CH}_2\text{-H}_4\text{F}$ to methenyl-tetrahydrofolate ($\text{CH-H}_4\text{F}$), which is subsequently converted to formyl-tetrahydrofolate ($\text{CHO-H}_4\text{F}$). This product is then converted to H_4F and formate, while producing ATP. P: periplasm; C: cytoplasm.

from *Methylococcus fumariolicum* SolV can oxidise formaldehyde with high affinity and MxaFI-type MDHs in general are also able to oxidise formaldehyde (Keltjens et al. 2014). However, recently the XoxF5 of the alphaproteobacterium *Methylobacterium extorquens* AM1 was shown to oxidise methanol to formaldehyde *in vivo*, not to formate (Good et al. 2019). Currently, it is unresolved how formaldehyde would be oxidised further to formate in verrucomicrobial methanotrophs (Chistoserdova 2011). Formaldehyde could be oxidised directly to formate by formaldehyde dehydrogenase (Chistoserdova, Kalyuzhnaia and Lidstrom 2009), but verrucomicrobial methanotrophs do not possess a gene encoding this enzyme. Alternatively, methanotrophs can make use of the tetrahydromethanopterin (H_4MPT) or the tetrahydrofolate (H_4F) pathway for formaldehyde oxidation, as found in proteobacterial methanotrophs (Vorholt 2002;

Marx, van Dien and Lidstrom 2005; Chistoserdova 2011). H_4F is synthesised by bacteria *de novo* (Birmingham and Derrick 2002). Indeed, genes encoding all enzymes involved in tetrahydrofolate biosynthesis are present in the genomes of the verrucomicrobial methanotrophs. Formaldehyde produced by periplasmic XoxF could diffuse into the cytoplasm and bind spontaneously to H_4F to form methylene- H_4F (Kallen and Jencks 1966). Whether this condensation is of sufficient high rate for bacterial growth *in vivo* is under debate (Crowther, Kosály and Lidstrom 2008; He et al. 2020). In the proteobacterium *Methylobacterium extorquens* AM1, the formaldehyde-activating enzyme actually catalyses this reaction at high rate (Vorholt et al. 2000). Whereas this enzyme is present in proteobacterial methanotrophs, it is absent in verrucomicrobial methanotrophs, suggesting another enzyme should be involved in the condensation of formaldehyde and

H₄F. All verrucomicrobial methanotrophs carry a gene encoding the bifunctional enzyme FolD (Fig. 3), which is known to convert methylene-H₄F to methenyl-H₄F, and produce NAD(P)H, similar to the methylene-H₄F dehydrogenase (Pawelek and MacKenzie 1998; Hou et al. 2008; Chistoserdova 2011; Eadsforth, Maluf and Hunter 2012). Subsequently, FolD acts as a methenyl-H₄F cyclohydrolase, converting methenyl-H₄F to formyl-H₄F (Fig. 4). Ultimately, the enzyme formate-tetrahydroformate ligase FtfL, found in all verrucomicrobial methanotrophs, could convert formyl-H₄F to tetrahydrofolate and formate, while producing one ATP molecule (Marx, Laukel and Vorholt 2003). If this pathway oxidises formaldehyde instead of XoxF, fewer electrons are donated to the cytochrome c protein XoxGJ, which is postulated to donate its electrons to the terminal oxidase to fuel respiration (Versantvoort et al. 2019). Nevertheless, the production of formaldehyde instead of formate by XoxF seems logical since formaldehyde oxidation via the H₄F pathway provides valuable reducing equivalents in the form of NAD(P)H and ATP. Finally, in the last step of aerobic methane oxidation in verrucomicrobial methanotrophs, formate is oxidised to CO₂ (Pol et al. 2014). All verrucomicrobial methanotrophs possess one complete *fdsDABG* operon, encoding a cytosolic formate dehydrogenase that is thought to oxidise formate, while producing NADH needed for the generation of biomass (Op den Camp et al. 2018). CO₂ can ultimately be assimilated via the CBB cycle. The key enzyme RuBisCO found in verrucomicrobial methanotrophs forms a novel cluster within the form I RuBisCO phylogenetic tree and is most closely related to form IC RuBisCOs found in various Proteobacteria (Khadem et al. 2011).

Coping with toxic nitrogen compounds from ammonia oxidation

In the geothermal habitat where *Methylacidiphilum fumariolicum* SolV was isolated from, high concentrations (1–28 mM) of ammonium (NH₄⁺) are present (Khadem et al. 2010; Mohammadi et al. 2017b). Methane and ammonia (NH₃) are highly-reduced molecules with structural similarities. Nitrifying microorganisms make a living from the oxidation of ammonia, initiated via the oxidation of ammonia to hydroxylamine (NH₂OH), catalysed by ammonia monooxygenase (AMO) (Klotz and Stein 2008). AMO is a membrane-bound copper-dependent enzyme homologous to pMMO of methanotrophs (Tavormina et al. 2011). Consequently, many nitrifiers can oxidise methane and many methanotrophs can oxidise ammonia (Bédard and Knowles 1989). Methanotrophs use ammonium as nitrogen source; however, ammonia in the periplasm can also fortuitously be oxidised by pMMO to hydroxylamine, which is a toxic compound (Nyerges and Stein 2009). The affinity of pMMO for ammonia decreases with increasing CH₄ concentrations, which indicates competitive substrate inhibition (Nyerges, Han and Stein 2010). Hydroxylamine could impede various cellular processes by damaging proteins or DNA and strongly inhibit MDHs (Kaplan and Ciotti 1953; Duine and Frank 1980; Versantvoort et al. 2020). Therefore, verrucomicrobial and proteobacterial methanotrophs have developed mechanisms to balance the assimilation of ammonium and the detoxification of deleterious nitrogen compounds.

In order to detoxify hydroxylamine, many methanotrophs possess a gene encoding a hydroxylamine oxidoreductase (HAO), oxidising hydroxylamine to nitric oxide (NO) (Fig. 5) (Caranto and Lancaster 2017; Versantvoort et al. 2020). All known *Methylacidiphilum* strains carry a gene encoding an HAO (Fig. 3), as do many proteobacterial methanotrophs (Campbell et al.

2011; Stein and Klotz 2011; Versantvoort et al. 2020). In contrast, HAO is not found in any of the *Methylacidimicrobium* strains and as such, these strains might be more sensitive to ammonium (Stein 2018). Alternatively, *Methylacidimicrobium* strains could have developed a different strategy to deal with ammonia that does not require an HAO, analogous to the gammaproteobacterial methanotroph *Methylomonas methanica*. This methanotroph also lacks an HAO-like protein (Campbell et al. 2011) and was unable to oxidise ammonia to nitrite, although its methane oxidation rate was inhibited by ammonia (Nyerges and Stein 2009). This could indicate that its pMMO is unable to oxidise ammonia to hydroxylamine or the formed hydroxylamine is converted to a product by an unknown enzyme other HAO. In either case, an HAO-like protein is not necessary to deal with ammonia toxicity. In nitrifiers, electrons yielded from the oxidation of hydroxylamine are transferred to the quinone pool for energy conservation via the hydroxylamine:ubiquinone reductase module (Klotz and Stein 2008). However, methanotrophs are thought to lack this module. Still, per ammonia molecule oxidised, four electrons are yielded when ammonia is oxidised to nitrite (Fig. 5). Since two electrons are needed to activate O₂ for the oxidation of ammonia to hydroxylamine by pMMO, the remaining two electrons might be used at the terminal oxidase for energy conservation (Stein and Klotz 2011).

Nitric oxide produced by HAO is a toxic compound. However, when ammonia is oxidised by *Methylacidiphilum fumariolicum* SolV under aerobic conditions, nitrite is observed as the main end product (Mohammadi et al. 2017b), similar to aerobic ammonia oxidisers and various proteobacterial methanotrophs (Nyerges, Han and Stein 2010; Campbell et al. 2011; Lehtovirta-Morley 2018). These cases necessitate an additional enzyme that oxidises the NO produced by HAO to nitrite. The nature of this nitric oxide-oxidising enzyme in methanotrophs and ammonia-oxidisers is unknown, although it is postulated that the copper-dependent nitrite reductase NirK could function in the opposite direction to oxidise NO to nitrite (Caranto and Lancaster 2017). In addition, NO can rapidly react with oxygen resulting in the formation of nitrite and nitrate in aqueous solutions (Udert, Larsen and Gujer 2005; Hughes 2008) and therefore the production of nitrite observed for *Methylacidiphilum fumariolicum* SolV could also be non-enzymatic (Fig. 5). Under anoxic conditions, nitrite reduction rates in *Methylacidiphilum fumariolicum* SolV are higher compared with rates under oxic conditions (Mohammadi et al. 2017b). As such, the alternative electron acceptor nitrite could be converted to NO by a nitrite reductase, of which one or two orthologues are present in most verrucomicrobial methanotrophs (Fig. 3). Nitric oxide can subsequently be reduced to N₂O, as was shown for the gammaproteobacterial methanotrophs *Methylomicrobium album* and *Methylomonas denitrificans* and for *Methylacidiphilum fumariolicum* SolV (Nyerges, Han and Stein 2010; Kits, Klotz and Stein 2015; Mohammadi et al. 2017b). The membrane-bound nitric oxide reductase NorCB reduces NO to N₂O and is found in all known verrucomicrobial methanotrophs (Figs 3 and 5). It is therefore conceivable that verrucomicrobial methanotrophs detoxify nitric oxide to inert N₂O under anoxic or oxygen-limited conditions (Acton and Baggs 2011; Bodelier and Steenbergh 2014). In addition, several proteobacterial methanotrophs were shown to produce N₂O without the initial formation of nitrite (Hoefman, van der Ha and Boon 2014). Since N₂O is strong greenhouse gas with a significant role in global warming, methanotrophs might not only have a mitigating effect on climate change by oxidising methane, but also aggravate this change by producing N₂O (Acton and Baggs

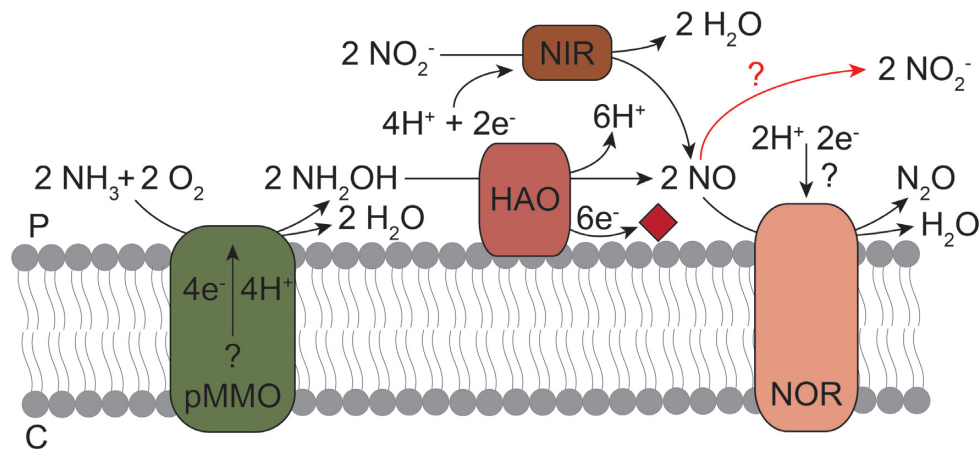


Figure 5. Ammonia (NH_3) oxidation by pMMO and fate of the reaction products. pMMO fortuitously oxidises ammonia to hydroxylamine. Subsequently, hydroxylamine oxidoreductase (HAO) oxidises hydroxylamine to nitric oxide, donating electrons to an unknown cytochrome c protein (indicated by the red diamond). Nitric oxide can be reduced to inert N_2O by the nitric oxide reductase (NOR), using an unknown electron donor. Alternatively, nitric oxide is converted chemically to nitrite in the presence of oxygen, or by an unknown enzyme (red arrow). Under anaerobic conditions, nitrite could be utilised as alternative electron acceptor. The nitrite reductase (NIR) could then reduce nitrite to nitric oxide, which is subsequently reduced to N_2O . P: periplasm; C: cytoplasm.

2011; Versantvoort et al. 2020). Altogether, the accidental oxidation of ammonia by pMMO results in the formation of various toxic compounds, for which *Methylacidiphilum* strains typically appear to have multiple detoxification pathways.

Oxidation of hydrogen gas

Multiple proteobacterial methanotrophs carry hydrogenase genes and were shown to consume H_2 (De Bont 1976; Csáki et al. 2001; Kelly, Anthony and Murrell 2005). These methanotrophs were proposed to use these hydrogenases to produce reducing equivalents to drive the energy-demanding conversion of methane to methanol by pMMO (Hanczár et al. 2002). Besides, various proteobacterial methanotrophs possess genes encoding both a hydrogenase and ribulose-1,5-biphosphate carboxylase/oxygenase (RuBisCO), suggesting autotrophic growth on H_2 is possible (Mohammadi et al. 2019). However, autotrophic growth on H_2 in liquid media without methane has never been observed for proteobacterial methanotrophs (Taylor, Dalton and Dow 1981; Baxter et al. 2002).

Hydrogen gas (H_2) is typically emitted in geothermal habitats in high concentrations (often > 1% v/v) and is a potential alternative energy source for various microorganisms (Aragno 1992; Chiodini et al. 2001; Carere et al. 2017; Mohammadi et al. 2019). Indeed, *Methylacidiphilum fumariolicum* SolV was experimentally shown to make a living as autotrophic hydrogenotroph in the absence of methane (Mohammadi et al. 2017a). Since methane emissions can heavily fluctuate in geothermal habitats, the ability to grow on another emitted, energy-rich gas is highly advantageous. *Methylacidiphilum* sp. RTK17.1 (an isolate almost identical to *Methylacidiphilum infernorum* V4) was shown to grow as a mixotroph on H_2 and CH_4 , a trait hypothesised to have driven niche expansion, which could explain the dominance of verrucomicrobial methanotrophs in geothermal habitats (Carere et al. 2017; Mohammadi et al. 2019). Recently, this mixotrophic lifestyle on H_2 and CH_4 was also demonstrated by the alphaproteobacterium *Methylcystis* sp. strain SC2 (Hakobyan, Zhu and Glatter 2020).

Hydrogenases are a very diverse group of enzymes that convert H_2 into two protons and two electrons, or vice versa (Lubitz et al. 2014). Three different metal compositions are known in the active site: [NiFe], [FeFe] and [Fe]. Genome comparisons of

several verrucomicrobial methanotrophs revealed that all these microbes carry genes encoding one or more different [NiFe] hydrogenases, except for *Ca. Methylacidithermus pantelleriae*' PQ17 (Fig. 3) (Mohammadi et al. 2019). It has to be noted that the genome of strain PQ17 was retrieved from a metagenome, not from an isolate. It might therefore still possess several genes that are not directly found through genetic analyses, although the genome is more than 98% complete and shows little contamination (Picone 2020). All isolated *Methylacidiphilum* strains carry genes encoding the group 1d [NiFe] hydrogenase, except for *Methylacidiphilum* sp. Yel. This membrane-bound enzyme has a relatively high O_2 tolerance compared to other hydrogenases and is involved in the aerobic respiration of H_2 (Greening et al. 2015). Both *Methylacidiphilum* sp. RTK17.1 and *Methylacidiphilum fumariolicum* SolV were experimentally shown to grow on H_2 using this hydrogenase under various oxygen concentrations (Carere et al. 2017; Mohammadi et al. 2017a).

All of the *Methylacidimicrobium* strains and none of the *Methylacidiphilum* strains possess genes encoding the oxygen-sensitive group 1b [NiFe] hydrogenase (Fig. 3) (Mohammadi et al. 2019). *Methylacidimicrobium tartarophylax* 4AC was shown to grow as an autotroph on H_2 using this enzyme under microoxic conditions (Mohammadi et al. 2019). The group 1b [NiFe] hydrogenase, typically found in Proteobacteria that perform anaerobic respiration (Greening et al. 2015), therefore enables *Methylacidimicrobium* strains to respire H_2 aerobically. In addition, all *Methylacidimicrobium* and *Methylacidiphilum* strains, except for *Methylacidimicrobium thermophilum* AP8, carry genes encoding a group 3b [NiFe] hydrogenase. This heterotetrameric cytosolic enzyme is known to couple the oxidation of NADPH to the production of H_2 during fermentation (Berney et al. 2014). However, in the verrucomicrobial methanotrophs it is hypothesised to oxidise H_2 and produce NADH for CO_2 fixation (Carere et al. 2017). This proposed catalysis was demonstrated by the group 3b [NiFe] hydrogenase of *Hydrogenobacter thermophilus* (Yoon et al. 1996).

Most hydrogen-oxidising microorganisms live in habitats with relatively high H_2 concentrations, such as animal guts and leguminous soils (Conrad and Seiler 1979; Pester and Brune 2007). In addition, several soil-inhabiting Actinobacteria were shown to consume H_2 present in the atmosphere (Constant, Poissant and Villemur 2008; Constant et al. 2010; Constant et al. 2011). Although the atmospheric H_2 concentration is very low

(0.53 ppmv H₂), soil systems are the largest sink of atmospheric H₂, consuming 75 Tg H₂ annually (Novelli et al. 1999). These H₂ scavengers possess genes encoding a putative high-affinity [NiFe] hydrogenase, of which the oxygen-tolerant group 1h [NiFe] hydrogenase seems to be dominant. Remarkably, this group 1h [NiFe] hydrogenase is also found in the genomes of *Methylacidiphilum fumariolicum* SolV, *Methylacidiphilum kamchatkense* Kam1 and *Methylacidiphilum* sp. Phi (Fig. 3) (Mohammadi et al. 2017a; Kruse et al. 2019). In the related *Methylacidiphilum infernorum* V4, this gene may have been lost over time due to the movement of transposable elements in the genome (Kruse et al. 2019). *Methylacidiphilum fumariolicum* SolV was shown to express both the group 1d and group 1h [NiFe] hydrogenase to grow on H₂ (Mohammadi et al. 2017a). At dissolved oxygen concentrations between 0.2 and 1.5%, only the group 1h [NiFe] hydrogenase supported growth, up to a growth rate of 0.03 h⁻¹, which is almost 40% of the growth rate on methane (Mohammadi et al. 2017a). The enzyme has a remarkable tolerance towards O₂, suggested to be the result of the unique coordination of the proximal [4Fe4S] clusters by three cysteines and an aspartate residue, instead of the usual coordination by four cysteine residues (Schäfer, Friedrich and Lenz 2013; Schäfer et al. 2016). The group 1h [NiFe] hydrogenase was purified from *Methylacidiphilum fumariolicum* SolV and kinetic experiments revealed an unusually high affinity for H₂, enabling this methanotroph to oxidise atmospheric H₂ (Schmitz et al. 2020a). It is hypothesised that this hydrogenase in particular could aid verrucomicrobial methanotrophs to thrive in geothermal systems (Carere et al. 2017; Schmitz et al. 2020a).

Oxidation of sulfur compounds

A variety of inorganic sulfur compounds such as hydrogen sulfide (H₂S) and sulfur dioxide (SO₂) are present in or released from geothermal systems (Spiro, Jacob and Logan 1992; Vasiliakos et al. 2005). Additionally, organic sulfur compounds such as methanethiol (CH₃SH) could be present, but data on its presence in terrestrial volcanic ecosystems are lacking. In seafloor hydrothermal systems, methanethiol is formed abiotically from CO₂, H₂S and H₂, derived from organic matter below the ocean floor (Rogers and Schulte 2012; Reeves et al. 2014). It is conceivable that in terrestrial mud volcanoes with comparable conditions, thermogenic production of methanethiol could occur as well. This compound could then be used by methylotrophs such as the verrucomicrobial methanotrophs as carbon and sulfur source, or even as energy source, as observed in the methylotrophic alphaproteobacterium *Hyphomicrobium* sp. VS (Pol et al. 1994). Studying (organic) sulfur compounds and how microorganisms cope with these compounds is important, as several of these compounds have profound effects on the environment, causing acid precipitation and cloud formation (Lomans, Pol and Op den Camp 2002).

Little is known about the utilisation of sulfur compounds as energy sources in methanotrophs. Genomic comparison of verrucomicrobial methanotrophs revealed that all known strains carry a gene encoding methanethiol oxidase (annotated as selenium-binding protein 56) (Eyice et al. 2018) (Fig. 3). This enzyme oxidises methanethiol to form formaldehyde, hydrogen sulfide and hydrogen peroxide (H₂O₂) (Suylén et al. 1987). The oxidation of methanethiol could render the verrucomicrobial methanotrophs with useful products. Formaldehyde could be converted to formate via F_oLD and F_tFL and to CO₂ by the formate dehydrogenase (Fig. 4). H₂S can be used as a sulfur source and could even be used as an energy source. In addition, a

gene encoding sulfide:quinone oxidoreductase (SQR) is found in the genomes of all verrucomicrobial methanotrophs, and highly expressed in *Methylacidiphilum fumariolicum* SolV (Mohammadi et al. 2017a). More specifically, it contains a type III SQR of which little is known, that could be involved in aerobic H₂S respiration (Marcia, Ermler and Peng 2010). The third product of methanethiol oxidation, hydrogen peroxide, is toxic and could be converted to water and oxygen by the enzyme catalase (Zamocky, Furtmüller and Obinger 2008). A gene encoding catalase is only found in *Methylacidiphilum infernorum* V4 and all *Methylacidimicrobium* strains. However, several peroxidases have been found in the genomes that might substitute the catalase activity.

The effect of intracellular H₂S and methanethiol in methanotrophs is currently not well understood. In some microorganisms, methane oxidation seems to be enhanced or unaffected by the presence of methanethiol or hydrogen sulfide (Lee, Kim and Cho 2012), whereas in other strains methane oxidation is inhibited by these compounds (Börjesson 2001; Lee et al. 2015). Since all verrucomicrobial methanotrophs possess a gene encoding an SQR and because H₂S is generally emitted at high concentrations from geothermal systems (Chiodini et al. 2001), it is conceivable that also H₂S could be an important energy source for these microorganisms. Alternatively, sulfide-oxidising enzymes could be present as a means of detoxification. Future experiments have to resolve whether energy conservation from the oxidation of H₂S is indeed possible.

The electron transport chain and energy conservation

The main location for energy conservation in prokaryotic cells is the cytoplasmic membrane. Quinones inside the cytoplasmic membrane are essential parts of the electron transport chain and therefore essential to energy conservation (Kurosu and Begari 2010). Verrucomicrobial methanotrophs seem to synthesise menaquinones (MK) via an alternative pathway that involves the intermediate futasoline (Hiratsuka et al. 2008). However, the gene *mqnB* involved in menaquinone biosynthesis was not detected in any of the genomes of verrucomicrobial methanotrophs (Kruse et al. 2019), but pathway variations are known to exist (Arakawa et al. 2010).

All verrucomicrobial methanotrophs described in this review possess five different complexes involved in energy conservation (Fig. 6). The NADH:quinone oxidoreductase (Complex I) is a membrane-bound protein complex that oxidises NADH, transfers the freed electrons to the quinone pool, and couples this reaction to the translocation of protons across the membrane, contributing to a proton motive force (Friedrich and Scheide 2000). The bacterial enzyme typically consists of 14 subunits, which are all encoded by the verrucomicrobial methanotrophs (Berrisford, Baradaran and Sazanov 2016). Complex II, or succinate dehydrogenase, is involved in both the electron transport chain and the tricarboxylic acid (TCA) cycle (Cecchini et al. 2002). In this cycle, Complex II couples the oxidation of succinate to the reduction of quinone (Fig. 6). Whereas Complex I is also involved in the translocation of protons across the membrane, Complex II is only involved in electron transfer. The quinols produced from the reduction of quinones by both complexes are typically oxidised by the *bc*₁ complex (Complex III) (Trumpower 1990). In that case, electrons yielded from the oxidation of quinols are subsequently transferred via an additional cytochrome *c* protein to a cytochrome *c* oxidase (complex IV or terminal oxidase). However, the verrucomicrobial methanotrophs do not carry genes encoding Complex III. Instead, these

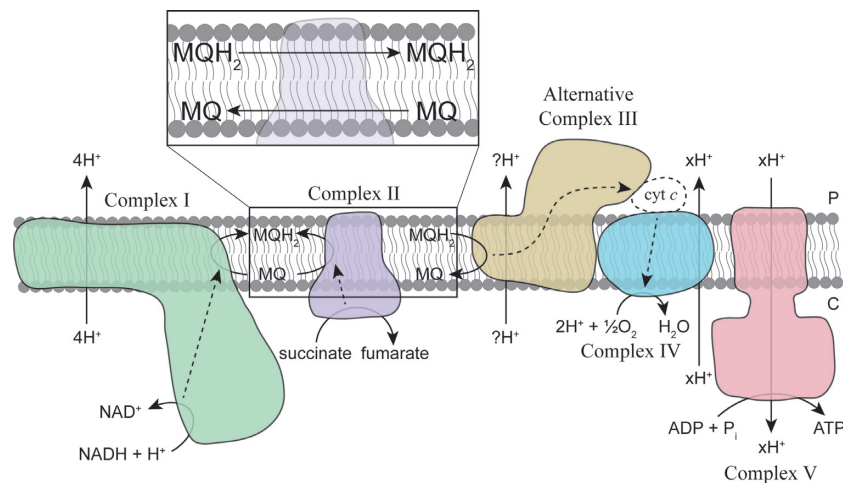


Figure 6. Schematic overview of the typical electron transport chain of verrucomicrobial methanotrophs. NADH generated through catabolic processes is oxidised by the NADH:quinone oxidoreductase (Complex I), transferring electrons to menaquinone (MQ) to form menaquinol (MQH₂) while translocating protons across the membrane to generate a proton motive force. The succinate dehydrogenase (Complex II) oxidises succinate to fumarate as a part of the tricarboxylic acid cycle and transfers electrons to MQ to form MQH₂. Alternative complex III oxidises MQH₂ back to MQ, transferring electrons to a cytochrome c oxidase (Complex IV), either via an external periplasmic cytochrome c protein (cyt c) or directly. Whether protons are translocated during this electron transfer is unknown. Ultimately, electrons are used to reduce O₂ to water, while protons are translocated over the membrane, contributing to the proton motive force that is used by an ATP synthase (Complex V) to synthesise ATP. Dashed lines indicate electron flow. Inset: transfer of oxidized and reduced quinones inside the bilayer (the quinone pool). C: cytoplasmic side of the membrane; P: periplasmic side of the membrane.

microorganisms possess genes encoding a structurally different protein complex with similar function: the Alternative Complex III (ACIII; Fig. 6) (Pereira et al. 2007; Refojo et al. 2010). The classical Complex III contributes to the proton motive force by translocating protons over the membrane via the quinone cycle. Whether ACIII also translocates protons across the membrane for energy conservation is under debate (Refojo, Teixeira and Pereira 2012; Sun et al. 2018). Ultimately, Complex IV reduces the terminal electron acceptor O₂ to water, with electrons retrieved from a c-type haem (Sun et al. 2018). All isolated verrucomicrobial methanotrophs, except for *Ca. 'Methylacidithermus pantelleriae'* PQ17, possess genes encoding three distinct Complexes IV: a *cbb*₃-type, an *aa*₃-type and a *ba*₃-type. These cytochrome c oxidases are classified based on the type of haems they contain and are known to have different affinities for O₂ and can be differentially used for various substrates (García-Horsman et al. 1994). In addition, *Methylacidiphilum fumarolicum* SolV and *Methylacidiphilum* sp. Phi possess a second *ba*₃-type cytochrome c oxidase. Interestingly, ACIII of *Flavobacterium johnsoniae* can form a supercomplex with Complex IV, in which an additional cytochrome c protein for electron transfer is not needed (Sun et al. 2018). With this in mind, it is interesting to note that in verrucomicrobial methanotrophs the genes encoding the different subunits composing ACIII are found directly adjacent to the genes encoding the *cbb*₃-type cytochrome c oxidase. Likewise, in other microorganisms the genes encoding ACIII are often found directly adjacent to genes encoding a cytochrome c oxidase (Refojo et al. 2010). However, only in verrucomicrobial methanotrophs and Opiritaceae a C-type cytochrome c oxidase is found together with ACIII, whereas typically these are A-type or B-type cytochrome c oxidases (Refojo et al. 2010). The different cytochrome c oxidases encoded by *Methylacidiphilum fumarolicum* SolV are differentially expressed, underscoring the metabolic versatility of verrucomicrobial methanotrophs (Khadem et al. 2012; Mohammadi et al. 2017a). Interestingly, the *ba*₃-type cytochrome c oxidase of *Aquifex aeolicus* is able to use ubiquinol as electron donor, reduced by electrons yielded from sulfide oxidation by the sulfide:quinone oxidoreductase (SQR)

(Gao et al. 2012). Since the verrucomicrobial methanotrophs encode for both a *ba*₃-type cytochrome c oxidase and SQR, these bacteria might utilise a specific system to conserve energy from H₂S oxidation, but this needs experimental validation. The proton motive force created by the transfer of electrons through the different membrane complexes is used for the synthesis of ATP via the ATP synthase (Complex V). The verrucomicrobial methanotrophs, except for *Ca. 'Methylacidithermus pantelleriae'* PQ17, carry genes encoding two ATP synthases (ATPases). One ATPase is related to ATPases of other Verrucomicrobia, while the other is related to the gammaproteobacterial ATPase (Hou et al. 2008; Kruse et al. 2019).

VERRUCOMICROBIAL METHANOTROPHS IN THE ENVIRONMENT

Verrucomicrobial methanotrophs are generally found in acidic volcanic ecosystems. Interestingly, global biogeographic distributions of closely related strains of verrucomicrobial methanotrophs suggest allopatric evolution over time (Erikstad et al. 2019). Additionally, evolutionary genomic analyses of verrucomicrobial methanotrophs revealed multiple horizontal gene transfer events (Sharp et al. 2013). Many genes have a relatively high similarity to those of the phylum Proteobacteria, but also to genes of Aquificae, Thermus/Deinococcus and archaea (Sharp et al. 2013; Schmitz et al. 2020b). Over the past 13 years, physiological and biochemical experiments have rendered valuable information that allows us to predict in which habitats we could encounter verrucomicrobial methanotrophs. Ecophysiological experiments could shed light on how verrucomicrobial methanotrophs interact with other microorganisms in the natural environment.

Acidic volcanic ecosystems are characterised by various morphological features. In general, these features are formed in places where mud, gas and water are expelled due to high fluid pressure from the subsurface, creating hot springs, mud pools and fumaroles (Cioni, Corazza and Marini 1984; Feyzullayev and Movsumova 2010; Benson et al. 2011). Where reduced sulfur

compounds are present, often sulfuric acid (H_2SO_4) is produced by (thermo)acidophilic microorganisms. The microbial production of sulfuric acid is environmentally important because it creates a low pH, which is characteristic for the Solfatara, several parts of Yellowstone National Park and various other geothermal ecosystems (Schoen and Rye 1970; Quatrini and Johnson 2018). A low pH seems to be a key determinant of verrucomicrobial methanotrophs to be present in geothermal habitats (Sharp et al. 2014). 16S rRNA and *pmoA* gene sequences of verrucomicrobial methanotrophs have so far not been detected in habitats with a pH > 5.0 and all isolates have a pH optimum around 3. Physicochemical parameters within geothermal habitats such as temperature and the oxygen concentration can differ locally within a few meters. Many of the thermophilic *Methylacidiphilum* strains were isolated from hot geothermal areas in close proximity to the moderate temperature geothermal areas where the *Methylacidimicrobium* strains were isolated from (Sharp et al. 2014). Although bogs and fens are also characterised by a high acidity (pH 3.3 to 4.9), no 16S rRNA or *pmoA* gene sequences of verrucomicrobial methanotrophs have been detected in these habitats, which may be explained by the relatively low concentration of lanthanides in peat bogs (Vodyanitskii et al. 2012; Sharp et al. 2014). Based on environmental sequencing and activity studies, the methanotrophic activity can be attributed to Proteobacteria, at a pH as low as 3.5 (Dunfield and Dedysh 2010). Proteobacteria such as strains of the genus *Acidithiobacillus* have been found at lower pH, but this does not include proteobacterial methanotrophs (Crognale et al. 2018). With current knowledge, methane-rich habitats below pH 3.5, including the aforementioned crown of rusted sewer pipes, seem to be better suited for verrucomicrobial methanotrophs than proteobacterial methanotrophs.

Methane and hydrogen gas are often emitted from geothermal habitats and these two compounds seem to be the major energy sources for verrucomicrobial methanotrophs. All verrucomicrobial methanotrophs were isolated on methane and several strains were shown to grow as autotrophs on H_2 (Carere et al. 2017; Mohammadi et al. 2017a; Mohammadi et al. 2019). Multiple verrucomicrobial methanotrophs possess more than one particulate methane monooxygenase. The three *pmo* operons within the *Methylacidiphilum* strains differ significantly in amino acid sequences, suggesting selection pressure for different functions and adaptation to changing environmental conditions (Op den Camp et al. 2009). In addition, short chain alkenes could be oxidised by one or more of the pMMOs found in *Methylacidiphilum* strains (Picone et al. 2020b). For methane oxidation to occur, a sufficient concentration of lanthanides must be present in the environment, because the verrucomicrobial methanotrophs only carry genes encoding the lanthanide-dependent XoxF-type methanol dehydrogenase. This does not necessarily mean that verrucomicrobial methanotrophs are only present in lanthanide-rich environments. If H_2 and oxygen are present at suitable concentrations, they could grow as Knallgas bacteria without the need for lanthanides (Carere et al. 2017; Mohammadi et al. 2017a; Mohammadi et al. 2019). The concentration of H_2 and O_2 in the environment could partially determine which strains of verrucomicrobial methanotrophs are present in the environment, because the different kinds of [NiFe] hydrogenases found in their genomes may have very different tolerances towards oxygen and possess different affinities for H_2 .

As example, *Methylacidimicrobium tartarophylax* 4AC only respire H_2 at microoxic conditions with the group 1b [NiFe] hydrogenase (Mohammadi et al. 2019), whereas *Methylacidiphilum fumarolicum* SolV can respire H_2 at wide range of oxygen concentrations catalysed by the group 1d and group 1h [NiFe] hydrogenases (Mohammadi et al. 2017a). In addition, all isolates carry a gene encoding a sulfide:quinone oxidoreductase (SQR), indicating that verrucomicrobial methanotrophs might even grow in acidic ecosystems where H_2S is present and CH_4 and H_2 are absent.

The main difference between *Methylacidiphilum* and *Methylacidimicrobium* strains seems to be their growth temperature, with the former being thermophiles and the being latter mesophiles. However, the isolation of *Methylacidimicrobium thermophilum* AP8 with an optimum growth temperature of 50°C muddles this clear division (Picone 2020). Still, *Methylacidiphilum* and *Methylacidimicrobium* strains are often found within a few meters of each other, but at different temperatures (Sharp et al. 2014). In the environment, the concentration of ammonium could be a key factor in determining which verrucomicrobial methanotrophs are present. In general, ammonia is a competitive inhibitor of pMMO, potentially reducing methane oxidation rates. Additionally, high ammonium concentrations can reduce the methane oxidation rate of microorganisms that are unable to detoxify the oxidised products of ammonia oxidation (D'Alessandro et al. 2009; Nyerges, Han and Stein 2010). Since only *Methylacidiphilum* strains possess a gene encoding a hydroxylamine oxidoreductase, they are well-equipped to detoxify hydroxylamine, intracellularly produced through ammonia oxidation by pMMO. Consequently, *Methylacidimicrobium* strains are expected to be primarily found at moderate temperatures and habitats with relatively low ammonium concentrations. However, different pMMOs could have different affinities for ammonia, or could not be able to oxidise ammonia at all (Nyerges and Stein 2009).

Little is known about the interaction of verrucomicrobial methanotrophs with other microorganisms in their natural geothermal habitat. Trough $\delta^{13}\text{C}$ analysis of CH_4 , the origin of this gas in geothermal systems can be deduced, since CH_4 produced by archaea is relatively light compared to thermogenic or abiotic methane (Op den Camp et al. 2009). Typically, methane emitted from geothermal ecosystems is from a non-microbial source, produced through a chemical reaction of H_2 and CO or by the non-microbial decomposition of buried organic matter (Giggenbach 1995; Etiope and Klusman 2002; Fiebig et al. 2007). However, methanogenic archaea have been found in anoxic parts of several geothermal ecosystems, but mostly at a pH of 5 or higher (Zeikus, Ben-Bassat and Hegge 1980; Berghuis et al. 2019). Remarkably, a metagenome-assembled genome (MAG) closely related to the methanogen *Methanocella conradii* was abundantly present in the metagenome of the hot, acidic, and methane-rich geothermal soil on the island Pantelleria, Italy (Picone et al. 2020a). This finding suggests that a larger part of methane emitted from hot and acidic geothermal ecosystems could be of microbial origin than initially thought (D'Alessandro et al. 2009). In contrast, H_2 utilised by verrucomicrobial methanotrophs does not seem to be produced by other microorganisms in their habitat, but rather through abiotic or thermogenic processes in the Earth's crust (Aragno 1992; Lindsay et al. 2019).

CONCLUDING REMARKS AND FUTURE PERSPECTIVES

Since the isolation of verrucomicrobial methanotrophs from hot and acidic geothermal ecosystems 13 years ago, significant progress had been made in our understanding of these microorganisms living in extreme environments. Verrucomicrobial methanotrophs are much more than their name suggests: these extremophiles are actually metabolically versatile microorganisms. In fact, the same could be true for their proteobacterial counterparts. Whereas several verrucomicrobial methanotrophs were shown to grow as Knallgas bacteria, hydrogen consumption by proteobacterial methanotrophs has only been demonstrated as part of a mixotrophic lifestyle. However, proteobacterial methanotrophs that possess the CBB cycle and an uptake hydrogenase could be Knallgas bacteria as well. In recent years, scientists have mainly focused on the metabolism of inorganic compounds by verrucomicrobial methanotrophs, not of organic compounds, as was done for several proteobacterial methanotrophs. Originally thought to be obligate methanotrophs, a large variety of multi-carbon compounds such as acetate, succinate and ethanol were shown to be consumed by alphaproteobacterial methanotrophs (Dunfield and Dedysch 2014). The first attempts to grow verrucomicrobial methanotrophs on several organic compounds such as glucose, citrate and malate were unsuccessful, but later *Methylacidiphilum fumarolicum* SolV was shown to grown on ethane and butane (Op den Camp et al. 2009; Picone et al. 2020b). The major questions for future research regarding metabolism are whether proteobacterial methanotrophs can grow as Knallgas bacteria and whether verrucomicrobial methanotrophs can incorporate organic compounds into their diet.

Since multiple verrucomicrobial methanotrophs were shown to be excellent hydrogen-oxidisers, it would be worthwhile to see if new enrichment strategies using hydrogen gas, carbon dioxide and low oxygen would result in novel verrucomicrobial isolates. The exciting discovery of 16S rRNA genes related to verrucomicrobial methanotrophs outside of terrestrial volcanic ecosystems suggests a distribution outside of geothermal habitats. Several factors such as pH and temperature, but also the oxygen and ammonium concentration and the presence of methane, hydrogen gas, hydrogen sulfide and lanthanides could be used as parameters to predict the presence of verrucomicrobial methanotrophs in nature. Methane and hydrogen gas seem to be the main energy sources for verrucomicrobial methanotrophs. In addition, hydrogen sulfide could be a potent energy source, but additional physiological and biochemical experiments are needed to clarify this. From soil it is known that up to 90% of methane produced in these systems is consumed before it reaches the atmosphere, thereby contributing significantly to the mitigation of greenhouse gas emissions from the biosphere (Oremland and Culbertson 1992; Singh et al. 2010). To what extent verrucomicrobial methanotrophs alleviate methane emissions from geothermal environments is unknown. With the annual seepage of 40 to 60 Tg methane from mud volcanoes, fumaroles and hydrothermal vents (Etiope 2009), it would be interesting to investigate verrucomicrobial methanotrophy in the environment, especially if the use alternative energy sources such as hydrogen gas and hydrogen sulfide could enhance this process.

SUPPLEMENTARY DATA

Supplementary data are available at [FEMSRE](https://femsre.onlinelibrary.wiley.com/doi/10.1111/femsre.125968) online.

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REFERENCES

- Acton SD, Baggs EM. Interactions between N application rate, CH₄ oxidation and N₂O production in soil. *Biogeochemistry* 2011;**103**:15–26.
- Allan W, Struthers H, Lowe DC. Methane carbon isotope effects caused by atomic chlorine in the marine boundary layer: Global model results compared with Southern Hemisphere measurements. *J Geophys Res* 2007;**112**:D04306.
- Angel R, Claus P, Conrad R. Methanogenic archaea are globally ubiquitous in aerated soils and become active under wet anoxic conditions. *ISME J* 2012;**6**:847–62.
- Anthony C. The c-type cytochromes of methylotrophic bacteria. *Biochim Biophys Acta* 1992;**1099**:1–15.
- Anthony C. The quinoprotein dehydrogenases for methanol and glucose. *Arch Biochem Biophys* 2004;**428**:2–9.
- Anvar SY, Frank J, Pol A et al. The genomic landscape of the verrucomicrobial methanotroph *Methylacidiphilum fumarolicum* SolV. *BMC Genomics* 2014;**15**:914.
- Aragno M. Thermophilic, aerobic, hydrogen-oxidizing (Knallgas) bacteria. In: Balows A, Trüper HG, Dworkin M, Harder W, Schleifer KH (eds). *The Prokaryotes*. New York: Springer, 1992, 3917–33.
- Arakawa C, Kuratsu M, Furihata K et al. Diversity of the early step of the futasoline pathway. *Antimicrob Agents Chemother* 2010;**55**:913–6.
- Balch WE, Fox GE, Magrum LJ et al. Methanogens: reevaluation of a unique biological group. *Microbiol Rev* 1979;**43**:260–96.
- Barnes RO, Goldberg ED. Methane production and consumption in anoxic marine sediments. *Geology* 1976;**4**:297–300.
- Bastviken D, Cole JJ, Pace ML et al. Fates of methane from different lake habitats: Connecting whole-lake budgets and CH₄ emissions. *J Geophys Res Biogeo* 2008;**113**:G02024.
- Baxter NJ, Hirt RP, Bodrossy L et al. The ribulose-1,5-bisphosphate carboxylase/oxygenase gene cluster of *Methylococcus capsulatus* (Bath). *Arch Microbiol* 2002;**177**:279–89.
- Beal EJ, House CH, Orphan VJ. Manganese- and iron-dependent marine methane oxidation. *Science* 2009;**325**:184–7.
- Benson CA, Bizzoco RW, Lipson DA et al. Microbial diversity in nonsulfur, sulfur and iron geothermal steam vents. *FEMS Microbiol Ecol* 2011;**76**:74–88.
- Berghuis BA, Yu FB, Schulz F et al. Hydrogenotrophic methanogenesis in archaeal phylum Verstraetearchaeota reveals the shared ancestry of all methanogens. *Proc Natl Acad Sci USA* 2019;**116**:5037–44.
- Birmingham A, Derrick JP. The folic acid biosynthesis pathway in bacteria: evaluation of potential for antibacterial drug discovery. *Bioessays* 2002;**24**:637–48.
- Berney M, Greening C, Hards K et al. Three different [NiFe] hydrogenases confer metabolic flexibility in the obligate aerobe *Mycobacterium smegmatis*. *Environ Microbiol* 2014;**16**:318–30.

- Berrisford JM, Baradaran R, Sazanov LA. Structure of bacterial respiratory complex I. *Biochim Biophys Acta* 2016;1857:892–901.
- Bižić M, Klintzsch T, Ionescu D et al. Aquatic and terrestrial cyanobacteria produce methane. *Sci Adv* 2020;6: eaax5343.
- Bodelier PLE, Gillisen MJ, Hordijk K et al. A reanalysis of phospholipid fatty acids as ecological biomarkers for methanotrophic bacteria. *ISME J* 2009;3:606–17.
- Bodelier PLE, Pérez G, Veraart AJ et al. Methanotroph ecology, environmental distribution and functioning. In: Lee E (ed). *Methanotrophs: Microbiology Fundamentals and Biotechnological Applications*. Cham: Springer, 2019, 1–38.
- Bodelier PLE, Steenbergh AK. Interactions between methane and the nitrogen cycle in light of climate change. *Curr Opin Environ Sustain* 2014;10:26–36.
- Boden R, Cunliffe M, Scanlan J et al. Complete genome sequence of the aerobic marine methanotroph *Methylomonas methanica* MC09. *J Bacteriol* 2011;193:7001–2.
- Bodrossy L, Kovács KL, McDonald IR et al. A novel thermophilic methane-oxidising γ -Proteobacterium. *FEMS Microbiol Lett* 1999;170:335–41.
- Boetius A, Ravensschlag K, Schubert CJ et al. A marine microbial consortium apparently mediating anaerobic oxidation of methane. *Nature* 2000;407:623–6.
- Bowman JP, Sly LI, Nichols PD et al. Revised taxonomy of the methanotrophs: description of *Methylobacter* gen. nov., emendation of *Methylococcus*, validation of *Methylosinus* and *Methylocystis* species, and a proposal that the family *Methylococcaceae* includes only the group I methanotrophs. *Int J Syst Bacteriol* 1993;43:735–3.
- Bridgham SD, Cadillo-Quiroz H, Keller JK et al. Methane emissions from wetlands: biogeochemical, microbial, and modeling perspectives from local to global scales. *Glob Chang Biol* 2013;19:1325–46.
- Brune A, Frenzel P, Cypionka H. Life at the oxic-anoxic interface: microbial activities and adaptations. *FEMS Microbiol Rev* 2000;24:691–710.
- Brune A. Methanogens in the digestive tract of termites. In: Hackstein J (ed). *(Endo)symbiotic Methanogenic Archaea*. Heidelberg: Springer, 2010, 81–100.
- Buffett BA. Clathrate Hydrates. *Annu Rev Earth Planet Sci* 2000;28:477–507.
- Bédard C, Knowles R. Physiology, biochemistry, and specific inhibitors of CH₄, NH₄⁺, and CO oxidation by methanotrophs and nitrifiers. *Microbiol Rev* 1989;53:68–84.
- Börjesson G. Inhibition of methane oxidation by volatile sulfur compounds (CH₃SH and CS₂) in landfill cover soils. *Waste Manag Res* 2001;19:314–9.
- Cai C, Leu AO, Xie G-J et al. A methanotrophic archaeon couples anaerobic oxidation of methane to Fe(III) reduction. *ISME J* 2018;12:1929–39.
- Cai Y, Zheng Y, Bodelier PLE et al. Conventional methanotrophs are responsible for atmospheric methane oxidation in paddy soils. *Nat Comm* 2016;7:11728.
- Campbell MA, Nyerges G, Kozlowski JA et al. Model of the molecular basis for hydroxylamine oxidation and nitrous oxide production in methanotrophic bacteria. *FEMS Microbiol Lett* 2011;322:82–9.
- Caranto JD, Lancaster KM. Nitric oxide is an obligate bacterial nitrification intermediate produced by hydroxylamine oxidoreductase. *Proc Natl Acad Sci USA* 2017;114:8217–22.
- Carere CR, Hards K, Houghton KM et al. Mixotrophy drives niche expansion of verrucomicrobial methanotrophs. *ISME J* 2017;11:2599–610.
- Carlson KM, Gerber JS, Mueller ND et al. Greenhouse gas emissions intensity of global croplands. *Nat Clim Change* 2017;7:63–8.
- Castaldi S, Tedesco D. Methane production and consumption in an active volcanic environment of Southern Italy. *Chemosphere* 2005;58:131–9.
- Cecchini G, Schröder I, Gunsalus RP et al. Succinate dehydrogenase and fumarate reductase from *Escherichia coli*. *Biochim Biophys Acta* 2002;1553:140–57.
- Chang J, Peng S, Ciais P et al. Revisiting enteric methane emissions from domestic ruminants and their $\delta^{13}\text{C}_{\text{CH}_4}$ source signature. *Nat Comm* 2019;10:4320.
- Chen Y, Crombie A, Rahman MT et al. Complete genome sequence of the aerobic facultative methanotroph *Methylocella silvestris* BL2. *J Bacteriol* 2010;192:3840–41.
- Chiodini G, Frondini F, Cardellini C et al. CO₂ degassing and energy release at Solfatara volcano, Campi Flegrei, Italy. *J Geophys Res Solid Earth* 2001;106:16213–21.
- Chistoserdova L, Kalyuzhnaya MG, Lidstrom ME. The expanding world of methylotrophic metabolism. *Annu Rev Microbiol* 2009;63:477–99.
- Chistoserdova L, Kalyuzhnaya MG. Current trends in methylotrophy. *Trends Microbiol* 2018;26:703–14.
- Chistoserdova L. Modularity of methylotrophy, revisited. *Environ Microbiol* 2011;13:2603–22.
- Cioni R, Corazza E, Marini L. The gas/steam ratio as indicator of heat transfer at the Solfatara fumaroles, Phlegraean Fields (Italy). *Bull Vulcanol* 1984;47:295–302.
- Cobo-Simón M, Tamames J. Relating genomic characteristics to environmental preferences and ubiquity in different microbial taxa. *BMC Genomics* 2017;18:499.
- Conrad R, Seiler W. Field measurements of hydrogen evolution by nitrogen-fixing legumes. *Soil Biol Biochem* 1979;11:689–90.
- Conrad R. Methane production in soil environments – anaerobic biogeochemistry and microbial life between flooding and desiccation. *Microorganisms* 2020;8:881.
- Conrad R. The global methane cycle: recent advances in understanding the microbial processes involved. *Environ Microbiol Rep* 2009;1:285–92.
- Constant P, Chowdhury SP, Hesse L et al. Genome data mining and soil survey for the novel group 5 [NiFe]-hydrogenase to explore the diversity and ecological importance of presumptive high-affinity H₂-oxidizing bacteria. *Appl Environ Microbiol* 2011;77:6027–35.
- Constant P, Chowdhury SP, Pratscher J et al. Streptomycetes contributing to atmospheric molecular hydrogen soil uptake are widespread and encode a putative high-affinity [NiFe]-hydrogenase. *Environ Microbiol* 2010;12:821–9.
- Constant P, Poissant L, Villemur R. Isolation of *Streptomyces* sp. PCB7, the first microorganism demonstrating high-affinity uptake of tropospheric H₂. *ISME J* 2008;2:1066–76.
- Cremers G, Pol A, Jetten MSM et al. Draft genome sequences of two acidophilic, mesophilic verrucomicrobial methanotrophs contain only one *pmoCAB* operon. *Microbiol Res Announc* 2020;9:e00315–20.
- Crognale S, Venturi S, Tassi F et al. Microbiome profiling in extremely acidic soils affected by hydrothermal fluids: the case of the Solfatara Crater (Campi Flegrei, southern Italy). *FEMS Microbiol Ecol* 2018;94: fyy90.
- Crowther GJ, Kosály G, Lidstrom ME. Formate as the main branch point for methylotrophic metabolism in *Methylobacterium extorquens* AM1. *J Bacteriol* 2008;190:5057–62.

- Csáki R, Hanczár T, Bodrossy L et al. Molecular characterization of structural genes coding for a membrane bound hydrogenase in *Methylococcus capsulatus* (Bath). *FEMS Microbiol Lett* 2001;**205**:203–7.
- D'Alessandro W, Bellomo S, Brusca L et al. Hydrothermal methane fluxes from the soil at Pantelleria island (Italy). *J Volcanol Geotherm Res* 2009;**187**:147–57.
- Daumann LJ. Essential and ubiquitous: The emergence of lanthanide metallobiochemistry. *Angew Chem Int Ed* 2019;**58**:12795–802.
- Davies SL, Whittenbury R. Fine structure of methane and other hydrocarbon-utilizing bacteria. *J Gen Microbiol* 1970;**61**:227–32.
- Dean JF, Middelburg JJ, Röckmann T et al. Methane feedbacks to the global climate system in a warmer world. *Rev Geophys* 2018;**56**:207–50.
- De Bont JAM. Nitrogen fixation by methane-utilizing bacteria. *Antonie Van Leeuwenhoek* 1976;**42**:245–53.
- Dedysh SN, Panikov NS, Liesack W et al. Isolation of acidophilic methane-oxidizing bacteria from northern peat wetlands. *Science* 1998;**282**:281–4.
- Deng YW, Ro SY, Rosenzweig AC. Structure and function of the lanthanide-dependent methanol dehydrogenase XoxF from the methanotroph *Methylomicrobium buryatense* 5GB1C. *J Biol Inorg Chem* 2018;**23**:1037–47.
- Dixon R, Kahn D. Genetic regulation of biological nitrogen fixation. *Nat Rev Microbiol* 2004;**2**:621–31.
- Duine JA, Frank J, Jr. Studies on methanol dehydrogenase from *Hyphomicrobium* X. Isolation of an oxidized form of the enzyme. *Biochem J* 1980;**187**:213–9.
- Dunfield PF, Dedysh SN. Acidic environments. In: Timmis KN (ed). *Handbook of Hydrocarbon and Lipid Microbiology*, Berlin, Heidelberg: Springer, 2010, 2181–92.
- Dunfield PF, Dedysh SN. *Methylocella*: a gourmand among methanotrophs. *Trends Microbiol* 2014;**22**:368–9.
- Dunfield PF, Yuryev A, Senin P et al. Methane oxidation by an extremely acidophilic bacterium of the phylum Verrucomicrobia. *Nature* 2007;**450**:879–82.
- Eadsforth TC, Maluf FV, Hunter WN. *Acinetobacter baumannii* FOLD ligand complexes - potent inhibitors of folate metabolism and a re-evaluation of the structure of LY374571. *FEBS J* 2012;**279**:4350–60.
- Edgar RC. Search and clustering orders of magnitude faster than BLAST. *Bioinformatics* 2010;**26**:2460–1.
- Erikstad H-A, Ceballos RM, Smestad NB et al. Global biogeographic distribution patterns of thermoacidophilic Verrucomicrobia methanotrophs suggest allopatric evolution. *Front Microbiol* 2019;**10**:1129.
- Etheridge DM, Steele LP, Francey RJ et al. Atmospheric methane between 1000 A.D. and present: Evidence of anthropogenic emissions and climatic variability. *J Geophys Res* 1998;**103**:15979–93.
- Etioppe G, Klusman RW. Geologic emissions of methane to the atmosphere. *Chemosphere* 2002;**49**:777–89.
- Etioppe G. Natural emissions of methane from geological seepage in Europe. *Atmos Environ* 2009;**43**:1430–43.
- Etmann M, Myhre G, Highwood EJ et al. Radiative forcing of carbon dioxide, methane, and nitrous oxide: A significant revision of the methane radiative forcing. *Geophys Res Lett* 2016;**43**:12614–23.
- Ettwig KF, Butler MK, Le Paslier D et al. Nitrite-driven anaerobic methane oxidation by oxygenic bacteria. *Nature* 2010;**464**:543–8.
- Ettwig KF, Shima S, van de Pas-Schoonen KT et al. Denitrifying bacteria anaerobically oxidize methane in the absence of Archaea. *Environ Microbiol* 2008;**10**:3164–73.
- Ettwig KF, Speth DR, Reimann J et al. Bacterial oxygen production in the dark. *Front Microbiol* 2012;**3**:273.
- Ettwig KF, Zhu B, Speth D et al. Archaea catalyze iron-dependent anaerobic oxidation of methane. *Proc Natl Acad Sci USA* 2016;**113**:12792–96.
- Evans PN, Boyd JA, Leu AO et al. An evolving view of methane metabolism in the Archaea. *Nat Rev Microbiol* 2019;**17**:219–32.
- Eyice Ö, Myronova N, Pol A et al. Bacterial SBP56 identified as a Cu-dependent methanethiol oxidase widely distributed in the biosphere. *ISME J* 2018;**12**:145–60.
- Feyzullayev AA, Movsumova UA. The nature of the isotopically heavy carbon of carbon dioxide and bicarbonates in the waters of mud volcanoes in Azerbaijan. *Geochem Int* 2010;**48**:517–22.
- Fiebig J, Woodland AB, Spangenberg J et al. Natural evidence for rapid abiogenic hydrothermal generation of CH₄. *Geochim Cosmochim Acta* 2007;**71**:3028–39.
- Friedrich T, Scheide D. The respiratory complex I of bacteria, archaea and eukarya and its module common with membrane-bound multisubunit hydrogenases. *FEBS Lett* 2000;**479**:1–5.
- Gagliano AL, D'Alessandro W, Tagliavia M et al. Methanotrophic activity and diversity of methanotrophs in volcanic geothermal soils at Pantelleria (Italy). *Biogeosciences* 2014;**11**:5865–75.
- Gao Y, Meyer B, Sokolova L et al. Heme-copper terminal oxidase using both cytochrome c and ubiquinol as electron donors. *Proc Natl Acad Sci USA* 2012;**109**:3275–80.
- García-Horsman JA, Barquera B, Rumbley J et al. The superfamily of heme-copper respiratory oxidases. *J Bacteriol* 1994;**176**:5587–600.
- Giggenbach WF. Variations in the chemical and isotopic composition of fluids discharged from the Taupo Volcanic Zone, New Zealand. *J Volcanol Geotherm Res* 1995;**68**:89–116.
- Good NM, Fellner M, Demirel K et al. Lanthanide-dependent alcohol dehydrogenases require an essential aspartate residue for metal coordination and enzymatic function. *J Biol Chem* 2020;**295**:8272–84.
- Good NM, Moore RS, Suriano CJ et al. Contrasting *in vitro* and *in vivo* methanol oxidation activities of lanthanide-dependent alcohol dehydrogenases XoxF1 and ExaF from *Methylobacterium extorquens* AM1. *Sci Rep* 2019;**9**:4248.
- Greening C, Biswas A, Carere CR et al. Genomic and metagenomic surveys of hydrogenase distribution indicate H₂ is a widely utilised energy source for microbial growth and survival. *ISME J* 2015;**10**:761–77.
- Hakobyan A, Zhu J, Glatter T. Hydrogen utilization by *Methylocystis* sp. strain SC2 expands the known metabolic versatility of type IIa methanotrophs. *Metab Eng* 2020;**61**:181–96.
- Hanaki K, Portugal-Pereira J. The effect of biofuel production on greenhouse gas emission reductions. In: Takeuchi K, Shiroshima H, Saito O, Matsuura M. (eds). *Biofuels and Sustainability*. Tokyo: Springer, 2018, 53–71.
- Hanczár T, Csáki R, Bodrossy L et al. Detection and localization of two hydrogenases in *Methylococcus capsulatus* (Bath) and their potential role in methane metabolism. *Arch Microbiol* 2002;**177**:167–72.
- Hanson RS, Hanson TE. Methanotrophic bacteria. *Microbiol Rev* 1996;**60**:439–71.
- Hao WM, Ward DE. Methane production from global biomass burning. *J Geophys Res* 1993;**98**:20657–61.

- Haroon MF, Hu S, Shi Y et al. Anaerobic oxidation of methane coupled to nitrate reduction in a novel archaeal lineage. *Nature* 2013;**500**:567–70.
- Heede R. Tracing anthropogenic carbon dioxide and methane emissions to fossil fuel and cement producers, 1854–2010. *Clim Change* 2014;**122**:229–41.
- He H, Noor E, Ramos-Parra PA et al. *In vivo* rate of formaldehyde condensation with tetrahydrofolate. *Metabolites* 2020;**10**:65.
- He Z, Cai C, Wang J et al. A novel denitrifying methanotroph of the NC10 phylum and its microcolony. *Sci Rep* 2016;**6**:32241.
- Hiratsuka T, Furihata K, Ishikawa J et al. An alternative menaquinone biosynthetic pathway operating in microorganisms. *Science* 2008;**321**:1670–3.
- Hirayama H, Suzuki Y, Abe M et al. *Methylothermus subterraneus* sp. nov., a moderately thermophilic methanotroph isolated from a terrestrial subsurface hot aquifer. *Int J Syst Evol Microbiol* 2011;**61**:2646–53.
- Hoefman S, van der Ha D, Boon N. Niche differentiation in nitrogen metabolism among methanotrophs within an operational taxonomic unit. *BMC Microbiol* 2014;**14**:83.
- Hogendoorn C, Pol A, Nuijten GHL et al. Methanol production by “*Methylacidiphilum fumariolicum*” SolV under different growth conditions. *Appl Environ Microbiol* 2020;**86**:e01188–20.
- Hogendoorn C. *Autotrophic microorganisms in acidic volcanic ecosystems*. [dissertation]. Nijmegen, The Netherlands: Radboud University Nijmegen; 2020.
- Holloway JM, Nordstrom DK, Böhlke JK et al. Ammonium in thermal waters of Yellowstone National Park: Processes affecting speciation and isotope fractionation. *Geochim Cosmochim Acta* 2011;**75**:4611–36.
- Holmes AJ, Roslev P, McDonald IR et al. Characterization of methanotrophic bacterial populations in soils showing atmospheric methane uptake. *Appl Environ Microbiol* 1999;**65**:3312–8.
- Houghton KM, Carere CR, Stott MB et al. Thermophilic methanotrophs: In hot pursuit. *FEMS Microbiol Ecol* 2019;**95**: fuz125.
- Hou S, Makarova KS, Saw JHW et al. Complete genome sequence of the extremely acidophilic methanotroph isolate V4, *Methylacidiphilum infernorum*, a representative of the bacterial phylum Verrucomicrobia. *Biol Direct* 2008;**3**:26.
- Hughes MN. Chemistry of nitric oxide and related species. *Methods Enzymol* 2008;**436**:3–19.
- Islam T, Jensen S, Reigstad LJ et al. Methane oxidation at 55°C and pH 2 by a thermoacidophilic bacterium belonging to the Verrucomicrobia phylum. *Proc Natl Acad Sci USA* 2008;**105**:300–4.
- Iversen N, Jorgensen BB. Anaerobic methane oxidation rates at the sulfate-methane transition in marine sediments from Kattegat and Skagerrak (Denmark). *Limnol Oceanogr* 1985;**30**:944–55.
- Jahn B, Pol A, Lumpe H et al. Similar but not the same: First kinetic and structural analyses of a methanol dehydrogenase containing a europium ion in the active site. *ChemBioChem* 2018;**19**:1147–53.
- Jensen BB. Methanogenesis in monogastric animals. *Environ Monit Assess* 1996;**42**:99–112.
- Kallen RG, Jencks WP. The mechanism of the condensation of formaldehyde with tetrahydrofolic acid. *J Biol Chem* 1966;**241**:5851–63.
- Kalyuzhnaya MG, Puri AW, Lidstrom ME. Metabolic engineering in methanotrophic bacteria. *Metab Eng* 2015;**29**:142–52.
- Kaplan NO, Ciotti MM. Competitive inhibition of hydroxylamine on alcohol dehydrogenase. *J Biol Chem* 1953;**201**:785–94.
- Kaserer H. Über die Oxidation des Wasserstoffes und des Methans durch Mikroorganismen. *Z landw Versuchsw in Österreich* 1905;**8**:789–92.
- Kato S, Takashino M, Igarashi K et al. Isolation and genomic characterization of a proteobacterial methanotroph requiring lanthanides. *Microbes Environ* 2020;**35**:ME19128.
- Kelly DP, Anthony C, Murrell JC. Insights into the obligate methanotroph *Methylococcus capsulatus*. *Trends Microbiol* 2005;**13**:195–8.
- Keltjens JT, Pol A, Reimann J et al. PQQ-dependent methanol dehydrogenases: rare-earth elements make a difference. *Appl Microbiol Biotechnol* 2014;**98**:6163–83.
- Khadem AF, Pol A, Jetten MSM et al. Nitrogen fixation by the verrucomicrobial methanotroph “*Methylacidiphilum fumariolicum*” SolV. *Microbiology* 2010;**156**:1052–9.
- Khadem AF, Pol A, Wieczorek A et al. Autotrophic methanotrophy in Verrucomicrobia: *Methylacidiphilum fumariolicum* SolV uses the Calvin-Benson-Bassham cycle for carbon dioxide fixation. *J Bacteriol* 2011;**193**:4438–46.
- Khadem AF, Pol A, Wieczorek AS et al. Metabolic regulation of “*Ca. Methylacidiphilum fumariolicum*” SolV cells grown under different nitrogen and oxygen limitations. *Front Microbiol* 2012;**3**:266.
- Khmelenina VN, Murrell CJ, Smith TJ et al. Physiology and Biochemistry of the Aerobic Methanotrophs. In: Rojo F (ed). *Aerobic Utilization of Hydrocarbons, Oils and Lipids. Handbook of Hydrocarbon and Lipid Microbiology*. Cham: Springer, 2018, 1–25.
- Kip N, Ouyang W, van Winden J et al. Detection, isolation, and characterization of acidophilic methanotrophs from *Sphagnum* mosses. *Appl Environ Microbiol* 2011;**77**:5643–54.
- Kirschke S, Bousquet P, Ciais P et al. Three decades of global methane sources and sinks. *Nature Geosci* 2013;**6**:813–23.
- Kits KD, Klotz MG, Stein LY. Methane oxidation coupled to nitrate reduction under hypoxia by the Gammaproteobacterium *Methylomonas denitrificans*, sp. nov. type strain FJG1. *Environ Microbiol* 2015;**17**:3219–32.
- Klotz MG, Stein LY. Nitrifier genomics and evolution of the nitrogen cycle. *FEMS Microbiol Lett* 2008;**278**:146–56.
- Kostka JE, Weston DJ, Glass JB. The *Sphagnum* microbiome: new insights from an ancient plant lineage. *New Phytol* 2016;**211**:57–64.
- Kox MAR, Aalto SL, Penttilä T et al. The influence of oxygen and methane on nitrogen fixation in subarctic *Sphagnum* mosses. *AMB Express* 2018;**8**:76.
- Kruse T, Ratnadevi CM, Erikstad H-A et al. Complete genome sequence analysis of the thermoacidophilic verrucomicrobial methanotroph “*Candidatus Methylacidiphilum kamchatkense*” strain Kam1 and comparison with its closest relatives. *BMC Genomics* 2019;**20**:642.
- Kurosu M, Begari E. Vitamine K₂ in electron transport system: Are enzymes involved in vitamin K₂ biosynthesis promising drug targets? *Molecules* 2010;**15**:1531–53.
- Kvenvolden KA, Rogers BW. Gaia’s breath—global methane exhalations. *Mar Pet Geol* 2005;**22**:579–90.
- La H, Hettiaratchi JPA, Achari G et al. Biofiltration of methane. *Bioresour Technol* 2018;**268**:759–72.
- Lee E-H, Moon K-E, Kim TG et al. Inhibitory effects of sulfur compounds on methane oxidation by a methane-oxidizing consortium. *J Biosci Bioeng* 2015;**120**:670–6.
- Lee J-H, Kim TG, Cho K-S. Isolation and characterization of a facultative methanotroph degrading malodor-causing volatile sulfur compounds. *J Hazard Mater* 2012;**235–236**:224–9.

- Lees V, Owens NP, Murrell JC. Nitrogen metabolism in marine methanotrophs. *Arch Microbiol* 1991;157:60–5.
- Lehtovirta-Morley LE. Ammonia oxidation: Ecology, physiology, biochemistry and why they must all come together. *FEMS Microbiol Lett* 2018;365:fny058.
- Letunic I, Bork P. Interactive Tree Of Life (iTOL): an online tool for phylogenetic tree display and annotation. *Bioinformatics* 2007;23:127–8.
- Leu AO, Cai C, Mcllroy SJ et al. Anaerobic methane oxidation coupled to manganese reduction by members of the *Methanoperedenaceae*. *ISME J* 2020;14:1030–41.
- Levicán G, Ugalde JA, Ehrenfeld N et al. Comparative genomic analysis of carbon and nitrogen assimilation mechanisms in three indigenous bioleaching bacteria: predictions and validations. *BMC Genomics* 2008;9:581.
- Lieberman RL, Rosenzweig AC. Crystal structure of a membrane-bound metalloenzyme that catalyses the biological oxidation of methane. *Nature* 2005;434:177–82.
- Lim S, Franklin SJ. Lanthanide-binding peptides and the enzymes that Might Have Been. *Cell Mol Life Sci* 2004;61:2184–8.
- Lindsay MR, Colman DR, Amenabar MJ et al. Probing the geological source and biological fate of hydrogen in Yellowstone hot springs. *Environ Microbiol* 2019;21:3816–30.
- Lomans BP, Pol A, Op den Camp HJM. Microbial cycling of volatile organic sulfur compounds. *Cell Mol Life Sci* 2002;59:575–88.
- Lubitz W, Ogata H, Rüdiger O et al. Hydrogenases. *Chem Rev* 2014;114:4081–148.
- Marcia M, Ermler U, Peng G. A new structure-based classification of sulfide:quinone oxidoreductases. *Proteins* 2010;78:1073–83.
- Marx CJ, Laukel M, Vorholt JA. Purification of the formate-tetrahydrofolate ligase from *Methylobacterium extorquens* AM1 and demonstration of its requirement for methylotrophic growth. *J Bacteriol* 2003;185:7169–75.
- Marx CJ, van Dien SJ, Lidstrom ME. Flux analysis uncovers key role of functional redundancy in formaldehyde metabolism. *PLoS Biol* 2005;3:e16.
- Metcalfe WW, Griffin BM, Cicchillo RM et al. Synthesis of methylphosphonic acid by marine microbes: a source for methane in the aerobic ocean. *Science* 2012;337:1104–7.
- Mohammadi SS, Pol A, van Alen TA et al. Ammonia oxidation and nitrite reduction in the verrucomicrobial methanotroph *Methylacidiphilum fumarolicum* SolV. *Front Microbiol* 2017b;8:1901.
- Mohammadi SS, Pol A, van Alen TA et al. *Methylacidiphilum fumarolicum* SolV, a thermoacidophilic ‘Knallgas’ methanotroph with both an oxygen-sensitive and -insensitive hydrogenase. *ISME J* 2017a;11:945–58.
- Mohammadi SS, Schmitz RA, Pol A et al. The acidophilic methanotroph *Methylacidimicrobium tartarophylax* 4AC grows as autotroph on H₂ under microoxic conditions. *Front Microbiol* 2019;10:2352.
- Moreno-Vivián C, Cabello P, Martínez-Luque M et al. Prokaryotic nitrate reduction: molecular properties and functional distinction among bacterial nitrate reductases. *J Bacteriol* 1999;181:6573–84.
- Murrell JC, Dalton H. Ammonia assimilation in *Methylococcus capsulatus* (Bath) and other obligate methanotrophs. *Microbiology* 1983b;129:1197–206.
- Murrell JC, Dalton H. Nitrogen fixation in obligate methanotrophs. *Microbiology* 1983a;129:3481–86.
- Nakagawa T, Mitsui R, Tani A et al. A catalytic role of XoxF1 as La³⁺-dependent methanol dehydrogenase in *Methylobacterium extorquens* strain AM1. *PLoS One* 2012;7:e50480.
- Novelli PC, Lang PM, Masarie KA et al. Molecular hydrogen in the troposphere: Global distribution and budget. *J Geophys Res Atmos* 1999;104:30427–44.
- Nyerges G, Han SK, Stein LY. Effects of ammonium and nitrite on growth and competitive fitness of cultivated methanotrophic bacteria. *Appl Environ Microbiol* 2010;76:5648–51.
- Nyerges G, Stein LY. Ammonia cometabolism and product inhibition vary considerably among species of methanotrophic bacteria. *FEMS Microbiol Lett* 2009;297:131–6.
- Op den Camp HJM, Islam T, Stott MB et al. Environmental, genomic and taxonomic perspectives on methanotrophic Verrucomicrobia. *Environ Microbiol Rep* 2009;1:293–306.
- Op den Camp HJM, Mohammadi SS, Pol A et al. Verrucomicrobial methanotrophs. In: Kalyuzhnaya MG, Xing X-H (eds). *Methane biocatalysis: Paving the way to sustainability*. Cham: Springer, 2018, 43–55.
- Oremland RS, Culbertson CW. Importance of methane-oxidizing bacteria in the methane budget as revealed by the use of a specific inhibitor. *Nature* 1992;356:421–3.
- Pagaling E, Yang K, Yan T. Pyrosequencing reveals correlations between extremely acidophilic bacterial communities with hydrogen sulphide concentrations, pH and inert polymer coatings at concrete sewer crown surfaces. *J Appl Microbiol* 2014;117:50–64.
- Pawelek PD, MacKenzie RE. Methenyltetrahydrofolate cyclohydrolase is rate limiting for the enzymatic conversion of 10-formyltetrahydrofolate to 5,10-methylenetetrahydrofolate in bifunctional dehydrogenase-cyclohydrolase enzymes. *Biochemistry* 1998;37:1109–15.
- Pereira MM, Refojo PN, Hreggvidsson GO et al. The alternative complex III from *Rhodothermus marinus* – A prototype of a new family of quinol:electron acceptor oxidoreductases. *FEBS Lett* 2007;581:4831–5.
- Pester M, Brune A. Hydrogen is the central free intermediate during lignocellulose degradation by termite gut symbionts. *ISME J* 2007;1:551–65.
- Picone N, Hogendoorn C, Cremers G et al. Geothermal gases shape the microbial community of the volcanic soil of Pantelleria, Italy. *mSystems* 2020a;5:e00517–20.
- Picone N, Mohammadi SS, Waajen AC et al. More than a methanotroph: a broader substrate spectrum for *Methylacidiphilum fumarolicum* SolV. *Front Microbiol* 2020b;11:604485.
- Picone N, Op den Camp HJM. Role of rare earth elements in methanol oxidation. *Curr Opin Chem Biol* 2019;49:39–44.
- Picone N. *Novel methane- and ammonia-oxidizing microorganisms from acidic environments*. [dissertation]. Nijmegen, The Netherlands: Radboud University Nijmegen; 2020.
- Pol A, Barends TRM, Dietl A et al. Rare earth metals are essential for methanotrophic life in volcanic mudpots. *Environ Microbiol* 2014;16:255–64.
- Pol A, Heijmans K, Harhangi HR et al. Methanotrophy below pH 1 by a new Verrucomicrobia species. *Nature* 2007;450:874–8.
- Pol A, Op den Camp HJM, Mees SG et al. Isolation of a dimethylsulfide-utilizing *Hyphomicrobium* species and its application in biofiltration of polluted air. *Biodegradation* 1994;5:105–12.
- Price MN, Dehal PS, Arkin AP. FastTree 2—approximately maximum-likelihood trees for large alignments. *PLoS One* 2010;5:e9490.
- Pruesse E, Peplies J, Glöckner FO. SINA: accurate high-throughput multiple sequence alignment of ribosomal RNA genes. *Bioinformatics* 2012;28:1823–9.
- Prusiner S, Stadtman ER. *The enzymes of glutamine metabolism*. New York: Academic Press, Inc, 1973.

- Quatrini R, Johnson DB. Microbiomes in extremely acidic environments: functionalities and interactions that allow survival and growth of prokaryotes at low pH. *Curr Opin Microbiol* 2018;**43**:139–47.
- Raghoebarsing AA, Pol A, van de Pas-Schoonen KT et al. A microbial consortium couples anaerobic methane oxidation to denitrification. *Nature* 2006;**440**:918–21.
- Raghoebarsing AA, Smolders AJ, Schmid MC et al. Methanotrophic symbionts provide carbon for photosynthesis in peat bogs. *Nature* 2005;**436**:1153–6.
- Rasigraf O, Kool DM, Jetten MS et al. Autotrophic carbon dioxide fixation via the Calvin-Benson-Bassham cycle by the denitrifying methanotroph “*Candidatus Methylomirabilis oxyfera*”. *Appl Environ Microbiol* 2014;**80**:2451–60.
- Reeburgh WS. Methane consumption in Cariaco Trench waters and sediments. *Earth Planet Sci Lett* 1976;**28**:337–44.
- Reeburgh WS. Oceanic methane biogeochemistry. *Chem Rev* 2007;**107**:486–513.
- Reeves EP, McDermott JM, Seewald JS et al. The origin of methanethiol in midocean ridge hydrothermal fluids. *Proc Natl Acad Sci USA* 2014;**111**:5474–9.
- Refojo PN, Sousa FL, Teixeira M et al. The alternative complex III: A different architecture using known building modules. *Biochim Biophys Acta* 2010;**1797**:1869–76.
- Refojo PN, Teixeira M, Pereira MM. The Alternative complex III: properties and possible mechanisms for electron transfer and energy conservation. *Biochim Biophys Acta* 2012;**1817**:1852–9.
- Rigby M, Montzka SA, Prinn RG et al. Role of atmospheric oxidation in recent methane growth. *Proc Natl Acad Sci USA* 2017;**114**:5373–7.
- Rogers KL, Schulte MD. Organic sulfur metabolisms in hydrothermal environments. *Geobiology* 2012;**10**:320–32.
- Ross MO, MacMillan F, Wang J et al. Particulate methane monooxygenase contains only mononuclear copper centers. *Science* 2019;**364**:566–70.
- Ross MO, Rosenzweig AC. A tale of two methane monooxygenases. *J Biol Inorg Chem* 2017;**22**:307–19.
- Saunio M, Jackson RB, Bousquet P et al. The growing role of methane in anthropogenic climate change. *Environ Res Lett* 2016;**11**:120207.
- Saunio M, Stavert AR, Poulter B et al. The global methane budget 2000–2017. *Earth Syst Sci Data* 2020;**12**:1561–623.
- Schmitz RA, Dietl A, Müller M et al. Structure of the 4-hydroxy-tetrahydrodipicolinate synthase from the thermoacidophilic methanotroph *Methylacidiphilum fumariolicum* SolV and the phylogeny of the aminotransferase pathway. *Acta Cryst F* 2020b;**76**:199–208.
- Schmitz RA, Pol A, Mohammadi SS et al. The thermoacidophilic methanotroph *Methylacidiphilum fumariolicum* SolV oxidizes subatmospheric H₂ with a high-affinity, membrane-associated [NiFe] hydrogenase. *ISME J* 2020a;**14**:1223–32.
- Schoen R, Rye RO. Sulfur isotope distribution in Solfataras, Yellowstone National Park. *Science* 1970;**170**:1082–4.
- Schäfer C, Bommer M, Hennig SE et al. Structure of an actinobacterial-type [NiFe]-hydrogenase reveals insight into O₂-tolerant H₂ oxidation. *Structure* 2016;**24**:285–92.
- Schäfer C, Friedrich B, Lenz O. Novel, oxygen-insensitive group 5 [NiFe]-hydrogenase in *Ralstonia eutropha*. *Appl Environ Microbiol* 2013;**79**:5137–45.
- Semrau JD, DiSpirito AA, Yoon S. Methanotrophs and copper. *FEMS Microbiol Rev* 2010;**34**:496–531.
- Sharp CE, Op den Camp HJM, Tamas I et al. Unusual members of the PVC superphylum: the methanotrophic Verrucomicrobia genus “*Methylacidiphilum*”. In: Fuerst J (ed). *Planctomycetes: cell structure, origins and biology*. Berlin: Springer-Verlag, 2013, 211–27.
- Sharp CE, Smirnova AV, Graham JM et al. Distribution and diversity of Verrucomicrobia methanotrophs in geothermal and acidic environments. *Environ Microbiol* 2014;**16**:1867–78.
- Sherwood Lollar B, Lacrampe-Couloume G, Slater GF. Unraveling abiogenic and biogenic sources of methane in the Earth’s deep subsurface. *Chem Geol* 2006;**226**:328–39.
- Siliakus MF, van der Oost J, Kengen SWM. Adaptations of archaeal and bacterial membranes to variations in temperature, pH and pressure. *Extremophiles* 2017;**21**:651–70.
- Singh BK, Bardgett RD, Smith P et al. Microorganisms and climate change: terrestrial feedbacks and mitigation options. *Nat Rev Microbiol* 2010;**8**:779–90.
- Spiro PA, Jacob DJ, Logan JA. Global inventory of sulfur emissions with 1° × 1° resolution. *J Geophys Res* 1992;**97**:6023–36.
- Stein LY, Klotz MG. Nitrifying and denitrifying pathways of methanotrophic bacteria. *Biochem Soc Trans* 2011;**39**:1826–31.
- Stein LY, Roy R, Dunfield PF. Aerobic methanotrophy and nitrification: processes and connections. *eLS* 2012; DOI: 10.1002/9780470015902.a0022213.
- Stein LY, Yoon S, Semrau JD et al. Genome sequence of the obligate methanotroph *Methylosinus trichosporium* strain OB3b. *J Bacteriol* 2010;**192**:6497–8.
- Stein LY. Proteobacterial methanotrophs, methylotrophs, and nitrogen. In: Kalyuzhnaya MG, Xing X-H (eds). *Methane biocatalysis: Paving the way to sustainability*. Cham: Springer, 2018, 57–66.
- Stein LY. The long-term relationship between microbial metabolism and greenhouse gases. *Trends in Microbiol* 2020;**28**:500–11.
- Stolper DA, Martini AM, Clog M et al. Distinguishing and understanding thermogenic and biogenic sources of methane using multiply substituted isotopologues. *Geochim Cosmochim Acta* 2015;**161**:219–47.
- Strong PJ, Xie S, Clarke WP. Methane as a resource: Can the methanotrophs add value? *Environ Sci Technol* 2015;**49**:4001–18.
- Sun C, Benlekbir S, Venkatakrisnan P et al. Structure of the alternative complex III in a supercomplex with cytochrome oxidase. *Nature* 2018;**557**:123–6.
- Suylen GMH, Large PJ, van Dijken JP et al. Methyl mercaptan oxidase, a key enzyme in the metabolism of methylated sulphur compounds by *Hyphomicrobium* EG. *Microbiology* 1987;**133**:2989–97.
- Svenning MM, Hestnes AG, Wartiaainen I et al. Genome sequence of the Arctic methanotroph *Methylobacter tundripaludum* SV96. *J Bacteriol* 2011;**193**:6418–9.
- Söhngen NL. Ueber Bakterien, welche Methan als Kohlenstoffnahrung und Energiequelle gebrauchen. *Zentralbl Bakteriol B* 1906;**15**:513–7.
- Søndergaard D, Pedersen C, Greening C. HydDB: A web tool for hydrogenase classification and analysis. *Sci Rep* 2016;**6**:34212.
- Tassi F, Fiebig J, Vaselli O et al. Origins of methane discharging from volcanic-hydrothermal, geothermal and cold emissions in Italy. *Chem Geol* 2012;**310–311**:36–48.
- Tavormina PL, Orphan VJ, Kalyuzhnaya MG et al. A novel family of functional operons encoding methane/ammonia monooxygenase-related proteins in gammaproteobacterial methanotrophs. *Environ Microbiol Rep* 2011;**3**:91–100.

- Taylor SC, Dalton H, Dow CS. Ribulose-1,5-bisphosphate carboxylase/oxygenase and carbon assimilation in *Methylococcus capsulatus* (Bath). *Microbiology*, 1981;122:89–94.
- Themelis NJ, Ulloa PA. Methane generation in landfills. *Renew Energy* 2007;32:1243–57.
- Trumpower BL. Cytochrome *bc*₁ complexes of microorganisms. *Microbiol Rev* 1990;54:101–29.
- Turner AJ, Frankenberg C, Kort EA. Interpreting contemporary trends in atmospheric methane. *Proc Natl Acad Sci USA* 2019;116:2805–13.
- Tveit AT, Hestnes AG, Robinson SL et al. Widespread soil bacterium that oxidizes atmospheric methane. *Proc Natl Acad Sci USA* 2019;116:8515–24.
- Udert KM, Larsen TA, Gujer W. Chemical nitrite oxidation in acid solutions as a consequence of microbial ammonium oxidation. *Environ Sci Technol* 2005;39:4066–75.
- Van Heeswijk WC, Westerhoff HV, Booger FC. Nitrogen assimilation in *Escherichia coli*: Putting molecular data into a systems perspective. *Microbiol Mol Biol Rev* 2013;77:628–95.
- Van Teeseling MCF, Pol A, Harhangi HR et al. Expanding the verucomicrobial methanotrophic world: Description of three novel species of *Methylacidimicrobium* gen. nov. *Appl Environ Microbiol* 2014;80:6782–91.
- Vasilakos C, Maggos T, Bartzis JG et al. Determination of atmospheric sulfur compounds near a volcanic area in Greece. *J Atmos Chem* 2005;52:101–16.
- Vasileva A, Moiseenko K. Methane emissions from 2000 to 2011 wildfires in Northeast Eurasia estimated with MODIS burned area data. *Atmos Environ* 2013;71:115–21.
- Venturi S, Tassi F, Magi F et al. Carbon isotopic signature of interstitial soil gases reveals the potential role of ecosystems in mitigating geogenic greenhouse gas emissions: Case studies from hydrothermal systems in Italy. *Sci Total Environ* 2019;655:887–98.
- Versantvoort W, Guerrero-Cruz S, Speth DR et al. Comparative genomics of *Candidatus Methylomirabilis* species and description of *Ca. Methylomirabilis lanthanidiphila*. *Front Microbiol* 2018;9:1672.
- Versantvoort W, Pol A, Daumann LJ et al. Characterization of a novel cytochrome *c* as the electron acceptor of XoxF-MDH in the thermoacidophilic methanotroph *Methylacidiphilum fumariolicum* SolV. *Biochim Biophys Acta Protein Proteom* 2019;1867:595–603.
- Versantvoort W, Pol A, Jetten MSM et al. Multiheme hydroxylamine oxidoreductases produce NO during ammonia oxidation in methanotrophs. *Proc Natl Acad Sci USA* 2020;117:24459–63.
- Vodyanitskii YN, Savichev AT, Trofimov SY et al. Accumulation of heavy metals in oil-contaminated peat soils. *Eurasian Soil Sci* 2012;45:977–82.
- Vorholt JA, Marx CJ, Lidstrom ME et al. Novel formaldehyde-activating enzyme in *Methylobacterium extorquens* AM1 required for growth on methanol. *J Bacteriol* 2000;182:6645–50.
- Vorholt JA. Cofactor-dependent pathways of formaldehyde oxidation in methylotrophic bacteria. *Arch Microbiol* 2002;178:239–49.
- Vu HN, Subuyuj GA, Vijayakumar S et al. Lanthanide-dependent regulation of methanol oxidation systems in *Methylobacterium extorquens* AM1 and their contribution to methanol growth. *J Bacteriol* 2016;198:1250–9.
- Vuilleumier S, Khmelenina VN, Bringel F et al. Genome sequence of the haloalkaliphilic methanotrophic bacterium *Methylomicrobium alcaliphilum* 20Z. *J Bacteriol* 2012;194:551–2.
- Wang J, Yan D, Dixon R et al. Deciphering the principles of bacterial nitrogen dietary preferences: a strategy for nutrient containment. *mBio* 2016;7:e00792–16.
- Ward N, Larsen Ø, Sakwa J et al. Genomic insights into methanotrophy: the complete genome sequence of *Methylococcus capsulatus* (Bath). *PLoS Biol* 2004;2:e303.
- Whittenbury R, Phillips KC, Wilkinson JF. Enrichment, isolation and some properties of methane-utilizing bacteria. *Microbiology* 1970;61:205–18.
- Wu ML, Wessels H, Pol A et al. XoxF-type methanol dehydrogenase from the anaerobic methanotroph “*Candidatus Methylomirabilis oxyfera*”. *Appl Environ Microbiol* 2015;81:1442–51.
- Yanpirat P, Nakatsuji Y, Hiraga S et al. Lanthanide-dependent methanol and formaldehyde oxidation in *Methylobacterium aquaticum* strain 22A. *Microorganisms* 2020;8:822.
- Yoon K-S, Ueda Y, Ishii M et al. NADH:ferredoxin reductase and NAD-reducing hydrogenase activities in *Hydrogenobacter thermophilus* strain TK-6. *FEMS Microbiol Lett* 1996;139:139–42.
- Zalkin H, Smith JL. Enzymes utilizing glutamine as an amide donor. *Adv Enzymol Relat Areas Mol Biol* 1998;72:87–144.
- Zamocky M, Furtmüller PG, Obinger C. Evolution of catalases from bacteria to humans. *Antioxid Redox Signal* 2008;10:1527–48.
- Zeikus JG, Ben-Bassat A, Hegge PW. Microbiology of methanogenesis in thermal, volcanic environments. *J Bacteriol* 1980;143:432–40.
- Zheng Y, Huang J, Zhao F et al. Physiological effect of XoxG(4) on lanthanide-dependent methanotrophy. *mBio* 2018;9:e02430–17.