human reproduction **OPINION**

Adjuncts in the IVF laboratory: where is the evidence for 'add-on' interventions?

Joyce Harper^{1,*}, Emily Jackson², Karen Sermon³, Robert John Aitken⁴, Stephen Harbottle⁵, Edgar Mocanu⁶, Thorir Hardarson⁷, Raj Mathur⁸, Stephane Viville⁹, Andy Vail¹⁰, and Kersti Lundin¹¹

¹ Embryology, IVF and Reproductive Genetics, Institute for Women's Health, University College London, London, UK ²Law Department, London School of Economics and Political Science, London, UK ³Research Group Reproduction and Genetics, Vrije Universiteit Brussel, Laarbeeklaan 101, 1090 Brussels, Belgium ⁴Priority Research Centre for Reproductive Science, University of Newcastle, NSW, Australia ⁵Cambridge IVF, Cambridge University Hospitals NHS Trust, Cambridge, UK ⁶Rotunda Hospital and RCSI, Parnell Square, Dublin I, Ireland ⁷Fertilitetscentrum, Carlanders Hospital, 402 29 Gothenburg, Sweden ⁸Department of Reproductive Medicine, Central Manchester University Hospitals NHS Foundation Trust, Manchester Academic Health Science Centre, Manchester M13 9WL, UK ⁹Institut de Parasitologie et Pathologie Tropicale, EA 7292, three rue Koeberlé, 67000 Strasbourg, France and Laboratoire de diagnostic génétique, UF3472-génétique de l'infertilité, Hôpitaux Universitaires de Strasbourg, 67000 Strasbourg, France ¹⁰Centre for Biostatistics, University of Manchester, Manchester Academic Health Science Centre, Manchester M13 9PL, UK ¹¹Reproductive Medicine, Sahlgrenska University Hospital, Goteborg, Sweden

*Correspondence address. Embryology, IVF and Reproductive Genetics, Institute for Women's Health, University College London, London, UK. E-mail: joyce.harper@ucl.ac.uk

Submitted on December 11, 2016; resubmitted on December 11, 2016; accepted on January 5, 2017

ABSTRACT: Globally, IVF patients are routinely offered and charged for a selection of adjunct treatments and tests or 'add-ons' that they are told may improve their chance of a live birth, despite there being no clinical evidence supporting the efficacy of the add-on. Any new IVF technology claiming to improve live birth rates (LBR) should, in most cases, first be tested in an appropriate animal model, then in clinical trials, to ensure safety, and finally in a randomized controlled trial (RCT) to provide high-quality evidence that the procedure is safe and effective. Only then should the technique be considered as 'routine' and only when applied to the similar patient population as those studied in the RCT. Even then, further pediatric and long-term follow-up studies will need to be undertaken to examine the long-term safety of the procedure. Alarmingly, there are currently numerous examples where adjunct treatments are used in the absence of evidence-based medicine and often at an additional fee. In some cases, when RCTs have shown the technique to be ineffective, it is eventually withdrawn from the clinic. In this paper, we discuss some of the adjunct treatments currently being offered globally in IVF laboratories, including embryo glue and adherence compounds, sperm DNA fragmentation, time-lapse imaging, preimplantation genetic screening, mitochondria DNA load measurement and assisted hatching. We examine the evidence for their safety and efficacy in increasing LBRs. We conclude that robust studies are needed to confirm the safety and efficacy of any adjunct treatment or test before they are offered routinely to IVF patients.

Key words: IVF adjuncts / sperm DNA fragmentation / embryo glue / adherence compounds / assisted hatching / PGS / mitochondria load / time-lapse imaging / RCT / live-birth rate

Introduction

IVF is a globally adopted technique supporting an extremely lucrative medical industry which has revolutionized human reproduction by offering hope of a family where none existed before. Patients routinely

pay large sums of money for treatment and many are willing to try anything that might help them improve their chances of having a baby.

The vast majority of IVF clinics want to help their patients achieve this objective as much as possible, which may involve undertaking unproven procedures and tests supported by anecdotal, low quality or

486 Harper et *al.*

unpublished evidence. In the last decade, a plethora of adjuncts or 'add-ons' have been introduced, many without any robust evidence that they increase the chances of a live birth or have any tangible benefit in terms of the health and well-being of the offspring (Nardo et al., 2015; Harper and Brison, 2013; Datta et al., 2015; Spencer et al., 2016).

The requirement that patients give informed consent to adjunct treatment in IVF, while necessary, may be insufficient to eliminate the over-selling or mis-selling of adjunct therapies for which the evidence of efficacy is poor or non-existent. In the UK, the Human Fertilization and Embryology (HFEA) Act 1990, as amended, requires patients to 'be provided with such relevant information as is proper' before embarking on treatment (Human Fertilization and Embryology Act 1990, sch. 3, para. 3(1)(b)). Patients should also be provided with 'a personalised cost treatment plan' (HFEA Code of Practice, 2012, para 4.3). Before an adjunct treatment is offered, the legislation requires clinics to provide open and honest information about the existence of robust evidence to support the particular intervention, along with information about costs. However, the 'therapeutic illusion' (Casarett, 2016), which commonly involves 'unjustified enthusiasm for treatment on the part of both patients and doctors' (Thomas, 1978), may mean that patients are not necessarily put off by low success rates or underpowered trial data, especially when simplistic explanations for reproductive failure circulate online and in the popular press. For example, a clinician might explain that the studies of immune therapy in assisted reproduction treatments to date have been poorly designed and that larger randomized controlled trials (RCTs) are necessary (Nardo et al., 2015), while patients might read newspaper articles with headlines such as 'The killer cells that robbed me of four babies' (Barber, 2015).

All IVF clinics need to consider the safety and efficacy of new technologies before introducing them and beginning to charge patients. In most cases, this should include preliminary work on animal models, followed by studies on human embryos donated for research and finally well-designed RCTs with a follow-up of all children born from the procedure (Harper et al., 2011). If such preliminary studies are not published, it is possible that technology bringing no clinical benefit or even leading to adverse health outcomes may be introduced.

There are several key factors affecting the validity and usefulness of any RCT performed for IVF. Validity can be assessed through risk of bias (Higgins et al., 2011) whereas the usefulness depends on the definition of the patient cohorts, the interventions compared, the primary outcome and the number of participants. Typically, demonstrating a clinical benefit will require many more participants than that required to demonstrate physiological effects.

Wilkinson et al. (2016) analyzed 142 IVF RCTs published in 2013 and 2014. They found that no consistent outcome measure was used. They suggest that initiatives to standardize outcome such as live birth rates (LBR) or cumulative LBR should be encouraged. Trials using implantation rate or clinical pregnancy rate (CPR) as outcome measures are only appropriate for preliminary studies. After any technique is brought into routine clinical practice, follow-up longitudinal studies should be undertaken to ensure the safety and efficacy of the intervention.

In 2009, the Policy and Practice Committee of the British Fertility Society reported on medical adjuncts in IVF and concluded that 'there is a need for good clinical trials in many of the areas surrounding medical adjuncts in IVF to resolve the empirical/evidence divide' (Nardo et al., 2015). Datta et al. (2015) reported on clinical and laboratory adjuncts and tests in IVF and stated that properly powered RCTs are

more valuable than a meta-analysis of a number of small heterogeneous RCTs. Spencer et al. (2016) carried out an audit of UK-based IVF clinic web sites and found that many were offering patients a large number of unproven adjuncts at additional cost.

In this paper, we describe some of the adjunct IVF laboratory treatments and tests that are currently being offered globally (Table I), often at a substantial cost for the patient. We describe the techniques and discuss the evidence for their safety and whether they increase LBR. The majority of the adjunct treatments listed here are included in the HFEA's recent addition to their website where they list evidence for 'add-on' treatments (due to go live in February 2017). Laboratory adjuncts that could have been included but were not due to space limitations are: ICSI for patients with non-male infertility (Grimstad et al., 2016), intracytoplasmic morphologically selected sperm injection, in vitro maturation, artificial oocyte activation, augmentation of mitochondria, intrauterine culture and elective freeze all embryos strategies.

Embryo glue and adherence compounds

The use of fibrin sealants to reduce ectopic pregnancy rate and increase LBRs was first proposed by Feichtinger et al. (1990) and the same author published further supportive data in Feichtinger et al. (1992). Despite this early promise, treatment using fibrin sealants never demonstrated reliable significant improvement in clinical outcomes and more recently, the focus has shifted to the use of a specific embryo transfer (ET) medium enriched with the glycoprotein hyaluronan (HA). It is well reported that HA is naturally present in the female reproductive tract and endometrium and forms a viscous solution which could enhance the ET process and prohibit embryo expulsion (Bontekoe et al., 2014).

The published data surrounding the use of adherence compounds are highly varied in quality and robustness of study design and as a result, the use of HA supplemented media for ET is still regarded as controversial (Bontekoe et al., 2014).

The latest Cochrane review of 3898 participants from 17 RCTs demonstrated moderate quality evidence for an improvement in CPR and LBR, with an associated increase in multiple pregnancy rate, when transfer medium was supplemented with HA (Bontekoe et al., 2014). The authors concluded that further high-quality studies were required, in particular where an elective single embryo transfer (eSET) procedure was performed, in part to alleviate concerns over the reported increase in the multiple pregnancy rate. A more recent RCT by Fancsovits et al. (2015) looked at 581 cycles and did not show a benefit in implantation rate, CPR or LBR, but found a higher birthweight in the HA group.

The reported increase in multiple pregnancy rate is suggestive of a need for clinics considering the use of a HA supplemented ET medium not only to re-evaluate their eSET policy and closely monitor their multiple pregnancy rate but also to ensure that patients are aware, not only of the possible increased chance of pregnancy, but also of the increased chance of multiple pregnancy when they are considering the number of embryos they wish to transfer.

The published evidence may be suggestive of a beneficial effect of the use of HA supplemented ET media. Before robust conclusions can be drawn, however, further RCTs are needed to evaluate the efficacy Adjuncts in IVF 487

Table I The current status of evidence relating to adjuncts used in the IVF laboratory. In all cases, further randomized controlled trials (RCTs) and long-term offspring and patient health follow-up studies are required.

Adjunct	Evidence for significant increase in live birth rate
Embryo glue and adherence compounds	Published evidence may be suggestive of a beneficial effect but further RCTs are needed regarding eSET and management of the multiple pregnancy rate
Sperm DNA fragmentation	Limited evidence
Time-lapse imaging	Limited evidence
Preimplantation genetic screening	Limited evidence
Mitochondria DNA load measurement	No evidence
Assisted hatching	No evidence

of HA as an adherence compound during ET with respect to eSET and the possibility of reducing the multiple pregnancy rate.

Sperm DNA fragmentation

Many clinics offer all their patients a sperm DNA fragmentation test. The assays include TUNEL, Comet, SCD assay, SCSA and 8-OHdG test (Shamsi et al., 2011). There are clear differences between assays in terms of the type of DNA damage being measured and their relative sensitivity (Smith et al., 2013). However, no particular assay has yet emerged as being of greater diagnostic value than any other. Ultimately, the purpose of such an assay is to indicate which treatments may be contraindicated for, or beneficial to, patients. This requires both diagnostic accuracy for the assay and evidence of effectiveness for the treatment(s). If, for example, the purpose of the assay is to determine whether antioxidant therapy is appropriate for the male partner then the measurement of 8-OHdG is of paramount importance and robust assays to assess this base adduct need to be developed and optimized (Muratori et al., 2015).

Three recent meta-analyses looked at measuring sperm DNA fragmentation in patients undergoing IVF and ICSI. Osman et al. (2015) performed a meta-analysis of six studies and found that, overall, men with low sperm DNA fragmentation had a higher LBR than those with high DNA fragmentation, but that the evidence was not sufficient to support this when ICSI was used. They concluded that further RCTs are needed to examine the role of ICSI versus IVF for men with high DNA fragmentation. Simon et al. (2016) looked at 8068 treatment cycles where DNA damage was measured using all four assays and found a modest but statistically significant association of DNA damage with CPR following IVF and/or ICSI. They found that the data varied depending on the assay used. Cissen et al. (2016) performed a systematic review and meta-analysis looking at the prognostic value of sperm DNA damage measurement, including 30 out of 658 studies. They concluded

that current tests have limited capacity to predict either the chance of conception after ART or which treatment method to choose, and that for now there is insufficient evidence to recommend sperm DNA testing.

The Practice Committee of the ASRM has concluded that 'current methods for assessing sperm DNA integrity do not reliably predict treatment outcomes and cannot be recommended routinely for clinical use' (Pfeifer et al., 2014).

However, a recent Cochrane report observed that low-quality evidence suggests that antioxidant therapy in the male might increase CPR and LBR in patients, where the spermatozoa are suffering from oxidative stress (Showell et al., 2014). In this context, accurate assessment of 8-OHdG levels could be of value in selecting a valid patient population. An RCT investigating the hypothesis that antioxidants can reverse oxidative DNA damage in spermatozoa is therefore urgently needed to address this possibility.

Time-lapse imaging

Taking pictures over time and reviewing them as a film, also known as time-lapse imaging (TL), is a technique that has been used for a century. Indeed, the first time TL imaging was reported as a tool to visualize early embryonic development was in 1929 (Lewis and Gregorgy, 1929). In that report, a remarkably detailed description of hamster embryonic development was described and the authors went on to speculate whether the observed timings in cleavage rate could predict 'embryonic potential'. More than 50 years later, human embryos were filmed using TL technology during their first 3 days of development (Eriksson et al. 1981). The next significant breakthrough was the work by Payne et al. (1997) who used TL imaging to describe the first events during fertilization, thus providing insight into how diverse and dynamic early embryonic development can be.

The first attempt to meaningfully use the unique information from different embryo cleavage timings and/or cleavage patterns was performed by Meseguer et al. (2011) based on data from 247 embryos known to have implanted. The latest prediction model was published by Petersen et al. (2016) but still requires extensive prospective testing and validation.

The usefulness of TL imaging in human IVF has been well debated. Among the proposed benefits that have been put forward are 'not missing important events during culture', quality control, teaching applications, more information to the patient and, of course, an increase in LBR.

Rubio et al. (2014) conducted the largest RCT to date that included 843 patients randomized mainly on Day 3 but also on Day 5. They reported a 9.7% increase in CPR compared to traditional culture and morphology assessments alone. This effect was diluted in the Cochrane review that also included two smaller trials under the intention to treat principle. The authors concluded that 'there is insufficient evidence of differences in live birth, miscarriage, stillbirth or clinical pregnancy to choose between [TL imaging] and conventional incubation' (Armstrong et al., 2015). However, more refined models are being continually developed as more data are being collected world-wide.

TL imaging serves so many other functions in the laboratory that its introduction will not be held back. It may be unthinkable in 5-10 years to still only be observing embryos by manually taking them out and looking at them. TL imaging is a tool which confers a number of practical benefits to the IVF laboratory. The future challenge for TL imaging

488 Harper et al.

is to find the best role in the IVF laboratory and to reduce implementation and consumable costs.

More RCTs are needed to distinguish whether there are clinical benefits of embryo selection algorithms based on TL information leading to an increase in LBR and whether there are benefits from uninterrupted embryo culture (Armstrong et al., 2014).

Preimplantation genetic screening

When in the 1990s, several studies demonstrated that cleavage stage embryos showed a high level of aneuploidy (Coonen et al., 1994; Munné et al., 1995), it was postulated that selection against these aneuploid embryos would improve LBRs. It was surprising that not only were meiotic abnormalities originating in the oocyte found, but also abnormalities occurring postzygotically. As a consequence, many embryos were mosaics, containing both normal and aneuploid cells, or several different lines of aneuploid cells. Thousands of IVF cycles were performed with preimplantation genetic screening (PGS), by biopsying one cell at Day 3 and performing fluorescent in situ hybridization (FISH) for five chromosomes. Eleven RCTs later, PGS was shown not to increase CPR or LBR and, in some cases, to decrease LBR (Harper et al., 2010; Geraedts and Sermon, 2016). It was realized that PGS at Day 3 was not effective because of the limited accuracy of FISH, the limited number of cells available for biopsy, and because at Day 3, cleavage stage embryos are at a peak of chromosomal abnormality/ mosaicism.

With the advent of new technology allowing comprehensive chromosome screening of Day 5 biopsied trophectoderm cells, PGS is now actively marketed as increasing implantation rates, and consequently decreasing time to pregnancy, recurrent miscarriages and repeated implantation failure (Sermon et al., 2016).

Despite these claims, only three RCTs have been published, all of which have been criticized because of poor study design. The pilot RCT by Yang et al. (2012) included a small sample size of 45 young, good prognosis patients. Scott et al. (2013) performed an RCT on 72 good prognosis patients between the ages of 21 and 42 years who were randomized quite late, i.e. if they had at least two blastocysts available for analysis. Although the authors claimed that PGS increased implantation and delivery rates, there was a fundamental methodological flaw in the study's failure to account for the difference between the unit of randomization (patients) and unit of analysis (individual embryos). The third RCT studied 89 patients aiming to compare PGS and SET with the transfer of two embryos (Forman et al., 2013). The same methodological problem encountered by the Scott trial was introduced and even so, the wide confidence interval for pregnancy did not demonstrate a beneficial effect.

Currently, two larger RCTs are underway and the results are expected soon. The ESTEEM study recruits patients of advanced maternal age and includes analyses of polar bodies using array-CGH, while the STAR study recruits all IVF patients and uses next generation sequencing on blastocyst biopsies. Other noteworthy differences are that the ESTEEM study has an intention-to-treat analysis, while STAR includes patients with two analyzable blastocysts as in the Scott and Forman studies. Furthermore, the ESTEEM outcome is cumulative LBR, while for STAR it is ongoing pregnancy rate after one transfer, an

outcome measure that has received much criticism and should be abandoned in favor of LBR (Griesinger, 2016).

Although these studies may serve to provide stronger evidence supporting PGS, the current RCTs do not provide sufficiently robust evidence to consider PGS as a proven and beneficial treatment.

Mitochondrial DNA load measurement

It has been estimated that metaphase II oocytes contain ~10⁵ mitochondrial DNA (mtDNA) copies, but since no replication of the mtDNA occurs until the blastocyst stage of embryonic development, the mtDNA molecules are divided over the cleaving cells (Fragouli and Wells, 2015). In 2015, two papers were published reporting an association between higher mtDNA level and lower implantation potential in blastocysts (Diez-Juan et al., 2015; Fragouli et al., 2015), pointing to disturbed energy provision and metabolic stress in embryos with a higher mtDNA content. While the paper of Diez-Juan et al. focused on euploid, transferred blastocysts, the other report also showed a relationship between aneuploidy of the blastocyst and a higher mtDNA load. According to both reports, euploid embryos that implanted after transfer had a mtDNA load below a data-derived threshold. Conversely, embryos that failed to implant, or that were aneuploid, showed a wide range of mtDNA load. This range overlapped with the implanting embryos at the low end, but the level of mtDNA at the high end was much higher in the non-implanting embryos. A threshold embryonic mtDNA load above which all embryos failed to implant could therefore be identified. Diez-Juan et al. reported that 52% (34/65) of the embryos below the identified threshold implanted compared to an implantation rate across the whole study population of 47% (34/72). For Fragouli et al., these figures were 59% (16/27) versus 38% (16/42), respectively.

Both groups have initiated an RCT. MitoScore is marketed by the group of Diez and is currently tested in RCT NCT02662686 (clinicaltrials.gov). MitogradeTM is marketed by Reprogenetics and is being tested in RCT NCT02673125.

Currently, there is no evidence that selection through mtDNA load measurement increases LBR. Application of the technique should therefore strictly be limited to participation in either one of RCTs, and this should clearly communicated to the patient.

Assisted hatching

In Cohen et al. (1990), proposed that making a breach in the zona pellucida may help implantation in some patients. Assisted hatching (AH) is usually performed on Day 3, 5 or 6 of embryo development using a non-contact laser, but mechanical or acidic solutions have also been used (Balaban et al., 2002). Clinics use AH for patients of advanced maternal age, smokers or patients with a raised FSH, or when transferring embryos that have been cryopreserved.

Three meta-analyses on AH have found a significant increase in CPR but no evidence for a difference in LBR. Martins et al. (2011) found a significant difference in CPR using frozen thawed embryos in unselected women and for patients with repeated IVF failure, but no evidence of benefit for subgroups of either older women or those with a good prognosis. They concluded that there were too few studies looking at

Adjuncts in IVF 489

LBR to draw conclusions. The Cochrane review by Carney et al. (2012) looked at 31 trials including 1992 clinical pregnancies in 5728 women. Nine of the 31 RCTs included data on LBR. There was no evidence of difference between the LBR in the AH and control groups. Li et al. (2016) looked at 36 RCTs with 6459 participants and found that AH gave a significant increase in CPR and multiple pregnancy rate but in the 15 RCTs that looked at LBR, there was no evidence of difference between the AH and control groups.

The National Institute for Clinical Excellence (NICE) guidelines (2013) state that 'assisted hatching is not recommended because it has not been shown to improve pregnancy rates'.

Duty of care toward the offspring

While we have considered evidence for increasing the chances of pregnancy and live birth, very few interventions in this field have considered the long-term health of the child. Individual clinics and national and international data collection bodies have a duty to evaluate data surrounding the use of adjuncts in IVF and collect long-term data pertaining to the health of any children born as a result of their use.

At the individual practitioner level, doctors and scientists recommending an unproven procedure to their patients must ensure that they provide comprehensive information surrounding the lack of evidence on the safety of the intervention for the resultant child. As best clinical practice dictates that professional guidelines are followed when managing patients, adjuncts that have not been proven to be beneficial should be used with caution, if used at all. Furthermore, regulatory bodies could insist that any empirical therapy prescribed must be accounted for, ideally with the establishment of clinical trials, to ensure long-term maternal and neonatal follow-up.

Among the techniques described in this paper, it is possible that some could have an impact upon the health of the embryo and the newborn. While some retrospective studies have been published, there are no RCTs on the impact of these technologies upon newborn health and child development.

The only Cochrane review on AH identified 2 studies out of 31 reporting on congenital anomalies and concluded that many unanswered questions remain about the perceived risks of the procedure, from embryo damage to chromosomal and congenital abnormalities (Carney et al., 2012).

Conclusion

IVF clinicians and scientists must recognize that appropriately powered, well-designed, peer-reviewed RCTs, with a LBR outcome measure which goes on to report on child health, are the gold standard of evidenced-based medicine.

Those advocating and recommending unproven procedures to their patients must ensure that they fully inform the patient of the evidence for its safety and effectiveness orally and in writing to ensure that people considering treatment using adjunct therapies are in a position to make an informed decision. It is also important that all procedures performed, including the adjunct treatments, are well-documented and followed up.

Regulators and professional bodies also have a role to play in ensuring that only suitable practices are used in the clinic.

Authors' roles

All authors contributed to planning, writing and revising article. J.H. conceived the idea for the paper and wrote the section on assisted hatching. E.J. wrote the legal section. K.S. wrote the sections on preimplantation genetic screening and mitochondria DNA load measurement. R.J.A. wrote the section on sperm DNA fragmentation. S.H. wrote the section on embryo glue and adherence compounds. E.M. wrote the section on duty of care toward the offspring. T.H. wrote the section on time-lapse imaging.

Funding

No funding was used to produce this paper.

Conflict of interest

R.J.A. has an honorary position on the Scientific Advisory Board of CellOxcess, a NJ-based biotechnology company dedicated to disease prevention through the diagnosis and treatment of Chronic Cell Oxidative Stress. R.M. has received hospitality at academic meetings from pharmaceutical companies Merck Serono, Ferring and Finox Biotech. A.V. is a statistical editor of the Cochrane Gynecology and Fertility Review Group. J.H. is Director of the Embryology and PGD Academy and Director of Global Women Connected. The remaining authors have no conflicts of interest to declare.

References

Armstrong S, Vail A, Mastenbroek S, Jordan V, Farquhar C. Time-lapse in the IVF-lab: how should we assess potential benefit? *Hum Reprod* 2014; doi: 10.1093/humrep/deu250.

Armstrong S, Arroll N, Cree LM, Jordan V, Farquhar C. Time-lapse systems for embryo incubation and assessment in assisted reproduction (Review). *Cochrane Database Syst Rev* 2015;**27**:CD011320. doi: 10.1002/14651858. CD011320.pub2.

Alaban B, Urman B, Alatas C, Mercan R, Mumcu A, Isiklar A. A comparison of four different techniques of assisted hatching. Hum Reprod 2002; 17:1239–1243

Barber, R. The killer cells that robbed me of four babies: TV star Sally Meen on her struggle to have two healthy daughters Daily Mail 2 January, 2015.

Bontekoe S, Johnson N, Blake D. Adherence compounds in embryo transfer media for assisted reproductive technologies. *Cochrane Database Syst Rev* 2014:CD007421. DOI:10.1002/14651858.CD007421.pub3.

Carney SK, Das S, Blake D, Farquhar C, Seif MM, Nelson L. Assisted hatching on assisted conception (in vitro fertilisation (IVF) and intracytoplasmic sperm injection (ICSI)). *Cochrane Database Syst Rev* 2012: CD001894. DOI:10.1002/14651858.CD001894.pub5.

Casarett D. The science of choosing wisely—overcoming the therapeutic illusion. N Engl J Med 2016;**374**:1203–1205.

Cissen M, Wely MV, Scholten I, Mansell S, Bruin JP, Mol BW, Braat D, Repping S, Hamer G. Measuring sperm DNA fragmentation and clinical outcomes of medically assisted reproduction: a systematic review and meta-analysis. *PLoS One* 2016;11:e0165125.

Cohen J, Elsner C, Kort H, Malter H, Massey J, Mayer MP. Impairment of the hatching process following IVF in the human and improvement of implantation by assisting hatching using micromanipulation. *Hum Reprod* 1990;**5**:7–13.

490 Harper et al.

- Coonen E, Harper JC, Ramaekers FCS, Delhanty JDA, Hopman AHN, Geraedts JPM, Handyside AH. Presence of chromosomal mosaicism in abnormal preimplantation embryos detected by fluorescent in situ hybridisation. *Hum Genet* 1994;**94**:609–615.
- Datta AK, Campbell S, Deval B, Nargund G. Add-ons in IVF programme Hype or Hope? Facts Views Vis Obgyn 2015;**7**:241–250.
- Diez-Juan A, Rubio C, Marin C, Martinez S, Al-Asmar N, Riboldi M, Diaz-Gimeno P, Valbuena D, Simon C. Mitochondrial DNA content as a viability score in human euploid embryos: less is better. *Fertil Steril* 2015; **104**:534–541
- Eriksson, Bo G, Hamberger L, Hillensjö T, Janson PO, Löfman C, Nilsson L, Nilsson, Lennart, Sjögren A, Wikland M. Microcinematorgraphic analysis of early cleavage stages in fertilized human ova. 3rd World Congress of Human Reproduction, Berlin, 1981.
- Fancsovits P, Lehner A, Murber A, Kaszas Z, Rigo J, Urbancsek J. Effect of hyaluronan-enriched embryo transfer medium on IVF outcome: a prospective randomized clinical trial. *Arch Gynecol Obstet* 2015;**291**:1173–1179.
- Feichtinger W, Barad D, Feinman M, Barg P. The use of two-component fibrin sealant for embryo transfer. *Fertil Steril* 1990;**54**:733–734.
- Feichtinger W, Strohmer H, Radner KM, Goldin M. The use of fibrin sealant for embryo transfer: development and clinical studies. *Hum Reprod* 1992;**7**:890–893.
- Forman EJ, Hong KH, Ferry KM, Tao X, Taylor D, Levy B, Treff NR, Scott RT Jr. In vitro fertilisation with single euploid blastocyst transfer: a randomised controlled trial. *Fertil* 2013;**100**:100–107.
- Fragouli E, Spath K, Alfarawati S, Kaper F, Craig A, Michel C-E, Kokocinski F, Cohen J, Munne S, Wells D. Altered levels of mitochondrial DNA are associated with female age, aneuploidy, and provide an independent measure of embryonic implantation potential. *PLoS Genet* 2015;11: e1005241.
- Fragouli E, Wells D. Mitochondrial DNA assessment to determine oocyte and embryo viability. Semin Reprod Med [Internet] 2015;33:401–409.
- Geraedts J, Sermon K. Preimplantation genetic screening 2.0: the theory. Mol Hum Reprod 2016;22:839–844.
- Griesinger G. Beware of the 'implantation rate'! Why the outcome parameter 'implantation rate' should be abandoned from infertility research. Hum Reprod 2016;31:249–251.
- Grimstad FW, Nangla AK, Luke B, Stern JE, Mak W. Use of ICSI in IVF cycles in women with tubal ligation does not improve pregnancy or live birth rates. *Hum Reprod* 2016;**31**:2750–2755. doi:10.1093/humrep/dew247.
- Harper J, Coonen E, De Rycke M, Fiorentino F, Geraedts J, Goossens V, Harton G, Pehlivan Budak T, Renwick P, Sengupta S et al. What next for preimplantation genetic screening (PGS)? A position statement from the ESHRE PGD Consortium steering committee. Hum Reprod 2010;25: 821–823.
- Harper JC, Brison D. Current evidence for ART practice: The Cochrane of Cochranes on optimising outcomes. *Evid Based Med* 2013;**19**. DOI:10. 1136/eb-2013-101571.
- Harper JC, Magli C, Lundin K, Barrat C, Brison D. When should new technology be introduced into the IVF lab? Hum Reprod 2011;27:303–313.
 Epub 2011 Dec 12.
- HFEA code of practice. 2012. http://www.hfea.gov.uk/docs/8th_Code_ of_Practice_Upto102013.pdf.
- Higgins JP, Altman DG, Gøtzsche PC, Jüni P, Moher D, Oxman AD, Savovic J, Schulz KF, Weeks L, Sterne JA. The Cochrane Collaboration's tool for assessing risk of bias in randomised trials. Cochrane Bias Methods Group; Cochrane Statistical Methods Group. *BMJ* 2011;343: d5928. doi:10.1136/bmj.d5928.
- Lewis WH, Gregorgy PW. Cinematographs of living developing rabbiteggs. Science 1929;69:226–229.

Li D, Yang D-L, An J, Jiao J, Zhou Y-M, Wu Q-J, Wang X-X. Effect of assisted hatching on pregnancy outcomes: a systematic review and meta-analysis of randomized controlled trials. *Sci Rep* 2016;**6**:31228.

- Martins WP, Rocha IA, Ferriani RA, Nastri CO. Assisted hatching of human embryos: a systematic review and meta-analysis of randomized controlled trials. *Hum Reprod* 2011;17:438–453.
- Meseguer M, Herrero J, Tejera A, Hilligsoe KM, Ramsing NB, Remohi J. The use of morphokinetics as a predictor of embryo implantation. *Hum Reprod* 2011;**26**:2658–2671.
- Munné S, Sultan KM, Weier HU, Grifo JA, Cohen J, Rosenwaks Z. Assessment of numeric abnormalities of X, Y, 18, and 16 chromosomes in preimplantation human embryos before transfer. *Am J Obstet Gynecol* 1995:172:1191–1199.
- Muratori M, Tamburrino L, Marchiani S, Cambi M, Olivito B, Azzari C, Forti G, Baldi E. Investigation on the origin of sperm DNA fragmentation: role of apoptosis, immaturity and oxidative stress. *Mol Med* 2015; **21**:109–122.
- Nardo LG, El-Toukhy T, Stewart J, Balen AH, Potdar N. British fertility society policy and practice committee: adjuvants in IVF: evidence for good clinical practice. *Hum Fertil* 2015;**18**:2–15. DOI:10.3109/14647273.2015.985454.
- NICE guidelines. (2013) https://www.nice.org.uk/donotdo/assisted-hatching-is-not-recommended-because-it-has-not-been-shown-toimprove-pregnancy-rates.
- Osman A, Alsomait H, Seshadri S, El-Toukhy T, Khalaf Y. The effect of sperm DNA fragmentation on live birth rate after IVF or ICSI: a systematic review and meta-analysis. *Reprod Biomed Online* 2015;**130**:120–127.
- Payne D, Flaherty SP, Barry MF, Matthews CD. Preliminary observations on polar body extrusion and pronuclear formation in human oocytes using time-lapse video cinematography. *Hum Reprod* 1997; 12:532–541.
- Petersen BM, Boel M, Montag M, Gardner DK. Development of a generally applicable morphokinetic algorithm capable of predicting the implantation potential of embryos transferred on Day 3. *Hum Reprod* 2016;31: 2231–2244.
- Pfeifer S, Goldberg J, Thomas M, Pisarska M, Widra E, Licht M, Sandlow J, Collins J, Cedars M, Rosen M et al. The clinical utility of sperm DNA integrity testing: a guideline. Practice Committee of the American Society for Reproductive Medicine. Erratum Fertil Steril 2014;101:884.
- Rubio I, Galan A, Larreategui Z, Ayerdi F, Bellver J, Herrero J, Meseguer M. Clinical validation of embryo culture and selection by morphokinetic analysis: a randomized, controlled trial of the EmbryoScope. Fertil Steril 2014;102:1287–1294.
- Scott RT, Upham KM, Forman EJ, Hong KH, Scott KL, Taylor D, Tao X, Treff NR. Blastocyst biopsy in CCS and fresh ET significantly increases IVF implantation and delivery rates; an RCT. *Fertil Steril* 2013;**100**: 687–703.
- Sermon K, Capalbo A, Cohen J, Coonen E, De Rycke M, De Vos A, Delhanty J, Fiorentino F, Gleicher N, Griesinger G et al. The why, the how and the when of PGS 2.0: current practices and expert opinions of fertility specialists, molecular biologists, and embryologists. *Mol Hum Reprod [Internet]* 2016;**22**:845–857.
- Shamsi MB, Imam SN, Dada R. Sperm DNA integrity assays: diagnostic and prognostic challenges and implications in management of infertility. [Assist Reprod Genet 2011; 28:1073–1085.]
- Showell MG, Mackenzie-Proctor R, Brown J, Yazdani A, Stankiewicz MT, Hart RJ. Antioxidants for male subfertility. *Cochrane Database Syst Rev* 2014:CD007411 doi: 10.1002/14651858.CD007411.pub3.
- Simon L, Zini A, Dyachenko A, Ciampi A, Carrell DT. A systematic review and meta-analysis to determine the effect of sperm DNA damage on in vitro fertilization and intracytoplasmic sperm injection outcome. *Asian J Androl* 2016;**18**:1–11; Fertil Steril 2013;99:673–677. doi:10.1016/j. fertnstert.2012.12.049.

Adjuncts in IVF 491

Smith TB, Dun MD, Smith ND, Curry BJ, Connaughton HS, Aitken RJ. The presence of a truncated base excision repair pathway in human spermatozoa that is mediated by OGG1. *J Cell Sci* 2013;**126**:1488–1497.

- Spencer EA, Mahtani KR, Goldacre B, Heneghan C. Claims for fertility interventions: a systematic assessment of statements on UK fertility centre websites. *BMJ Open* 2016;**6**:e013940. doi:10.1136/bmjopen-2016-013940.
- Thomas KB. The consultation and the therapeutic illusion. *Br Med J* 1978; 1:1327–1328.
- Wilkinson J, Roberts SA, Showell M, Brison DR, Vail A. No common denominator: a review of outcome measures in IVF RCTs. *Hum Reprod* 2016;**31**:2714–2722.

Yang Z, Liu J, Collins GS, Salem SA, Liu X, Lyle SS, Peck AC, Sills ES, Salem RD. Selection of single blastocysts for fresh transfer via standard morphology assessment alone and with array CGH for good prognosis IVF patients: results from a randomized pilot study. *Mol Cytogenet [Internet]* 2012;**5**:24.