

Editorial: Insulin-Like Growth Factor (IGF)-Binding Proteins in Serum—Do They Have Additional Roles besides Modulating the Endocrine IGF Actions?

Although the existence of insulin-like growth factor (IGF)-binding proteins (IGFBPs) in serum was suspected over 20 yr ago, it is only recently that we have begun to gain insight into the biological significance of the various IGFBPs. Six IGFBPs, designated IGFBP-1 through IGFBP-6 (24–50 kDa in size) have been found in serum, tissue extracts, and culture medium that was conditioned by a variety of cell types. The structural features that distinguish one IGFBP from another include an RGD cell matrix adhesion sequence, glycosylation sites, phosphorylation sites, and extracellular matrix binding (1, 2). In this editorial, we discuss the possible reasons for the redundancy in IGFBPs (why six IGFBPs instead of one?) and evaluate the potential roles that IGFBPs may play in regulating the endocrine and local actions of the IGFs.

Endocrine and local actions of IGFs

In adult humans, the total concentration of IGF-I and IGF-II in blood is about 800 $\mu\text{g/L}$ or 0.1 $\mu\text{mol/L}$, which is approximately 1000 times greater than the insulin concentration (3). Despite the fact that the insulin-like activity of IGFs is only 5% that of insulin, the IGFs could in theory contribute 50 times more insulin-like activity than insulin alone because of their abundance. However, this does not occur because the activity of IGFs is largely neutralized as a consequence of binding to the IGFBPs. Thus, without IGFBPs, the effect of IGFs would swamp any effect of insulin alone, and because IGF expression is not tightly controlled by blood glucose levels, glucose homeostasis would not be possible.

One obvious question that arises with regard to the circulating IGF concentration is why are the IGFs present in such abundance compared to other polypeptide growth factors in blood? Because the circulating level of the IGFs is largely determined by GH and nutrition and because IGFs have been shown to stimulate the growth and differentiation of a number of cell types, it is interesting to speculate that circulating IGFs may function to promote general growth. Thus, the reason why IGFs may circulate at high levels in blood, is so that a large readily available reserve is available for IGFs to act systematically in an endocrine manner. In addition to general systemic regulation, there is also a need for local regulation of growth in specific organs depending on local needs. For example, mechanical loading at a local site in bone stimulates local bone growth and not generalized

skeletal growth. The current evidence strongly suggests that IGFs may play a significant role in such a local regulation inasmuch as IGFs produced by one cell type act on the same cell in an autocrine manner or on a neighboring cell type in a paracrine manner (3). Thus, IGFs may function locally in a variety of cell types, including brain, muscle, kidney, lung, pancreas, and bone, in an autocrine/paracrine manner in addition to their systemic endocrine actions.

Tissue-specific regulation of IGF actions

If IGFs play a central role in the local regulation of a number of tissues, then how are the actions of IGFs regulated differently in various tissues? This is obviously an important question because IGFs are produced by most cell types, but their role in the regulation of cell proliferation and other functions appears to vary from one tissue to another. Thus, some means must exist for IGF signal molecules, which are common to so many tissues, to act on a given tissue in a specific manner. One means by which tissue specificity can be accomplished is by producing multiple forms of a given growth factor. For example, the fibroblast growth factors, the interleukins, and the transforming growth factor- β (TGF β) groups of growth factors each contain several members, some of which have been shown to function in a tissue-specific manner. In the case of IGFs, only two forms of IGFs are produced by many tissues. However, tissue-specific regulation may be accomplished partly by the presence of multiple forms of IGFBPs and their corresponding proteases. For IGFBPs to serve as efficient tissue-specific regulators of IGF activity, it can be predicted that multiple components will be required to serve multiple effector inputs from different tissues. In this regard, the following findings support the concept that the six IGFBPs and their corresponding proteases (unknown number) may provide a mechanism for tissue-specific regulation of IGF actions: 1) IGFBPs differ in their biological actions depending on the cell type studied; 2) IGFBPs are differentially expressed in different tissues in both fetal and adult life; and 3) extracellular fluids of certain tissues are enriched with specific IGFBPs (e.g. IGFBP-1 in human amniotic fluid, IGFBP-6 in human cerebrospinal fluid, and IGFBP-3 in human follicular fluid) (1, 2, 4).

IGFBPs as potential modulators of endocrine IGF actions

In the circulation an, about 75% of the IGFs are complexed with IGFBP-3 and an acid-labile subunit in a 150- to 200-kDa ternary complex, whereas the remainder of the circulating IGFs are bound to lower molecular mass IGFBPs (5–7). With the recent development and validation of RIAs for accurate measurements of various IGFBPs in biological fluids, it is

Received May 20, 1996. Revision received July 9, 1996. Accepted July 15, 1996.

Address all correspondence and requests for reprints to: Dr. Subburaman Mohan, Research Service (151), Jerry L. Pettis Veterans Administration Medical Center, 11201 Benton Street, Loma Linda, California 92357.

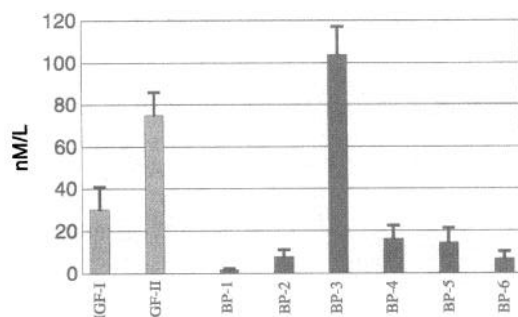


FIG. 1. Concentrations of IGFs and IGFBPs in adult human serum. IGF-I, IGF-II, IGFBP-3, IGFBP-4, and IGFBP-5 values were measured by RIA in the author's laboratory. Data for IGFBP-1, IGFBP-2, and IGFBP-6 were compiled from published literature. Values are the mean \pm SD.

now possible to evaluate the relative contributions of all six IGFBPs to the IGF-binding capacity of serum (Fig. 1). The total concentration of the small molecular mass IGFBPs (IGFBP-1, -2, -4, -5, and -6) add up to about 50% of the IGFBP-3 concentration (8–18). Based on the observation that there is a 50% molar excess of IGFBPs over IGFs, it is unlikely that there is much free IGFs in normal adult human serum.

If the majority of the IGFs circulate in serum as IGF-IGFBP complexes, it should be possible to modulate the endocrine actions of IGF by several mechanisms, including those that regulate the levels of IGFBPs (Table 1). In this regard, there is evidence that the relative concentrations of various IGFBPs in the serum can change depending on physiological and pathological situations, such as age, nutrition, serum GH levels, diabetes, puberty, pregnancy, *etc.* (1, 8–18). Studies of serum regulation of various IGFBPs to date reveal that the candidate IGFBP chosen as a primary regulator of IGF bioavailability may vary depending on the metabolic conditions. For example, IGFBP-1 appears to be the primary regulator of IGF bioavailability in response to acute changes in the circulating insulin level, whereas IGFBP-3 appears to be the primary regulator of IGF bioavailability in response to changes in the circulating GH level (5). In addition, serum levels of IGFBP-5, but not IGFBP-4, showed significant positive correlations with IGF-I and IGF-II, suggesting that different mechanisms may regulate the amount of inhibitory IGFBP-4 and stimulatory IGFBP-5 in human serum (8, 9). Although the findings that serum levels of one or more IGFBPs change acutely upon treatment with a number of physiological regulators and that they change in the right direction to explain the corresponding metabolic change are consistent with the general hypothesis that the endocrine IGF actions may in part be regulated by changes in serum IGFBP levels, further studies are needed to establish the cause and effect relationship.

As the IGF system in serum is made up of multiple components, it would be important for the stimulatory and inhibitory components of the IGF system to be regulated in a reciprocal manner to produce an optimal effect in response to changes in physiological or pathological conditions. Otherwise, parallel changes in inhibitory and stimulatory components of the IGF system would cancel each other out. An example of a situation in which a coordinated regulation of

TABLE 1. Potential mechanisms to increase IGF bioavailability in the target tissues

- 1) Increase the level of free IGFs
- 2) Increase the ratio of IGFs/inhibitory IGFBPs
- 3) Increase the ratio of stimulatory IGFBPs/inhibitory IGFBPs
- 4) Increase the rate of proteolysis of inhibitory IGFBPs in the target tissue
- 5) Decrease the rate of proteolysis of stimulatory IGFBPs in the target tissue
- 6) Increase IGF receptor abundance in the target tissue

TABLE 2. Age-related changes in IGF system components

Component	Function	Change (%)
IGF-I	Stimulatory	60 ↓
IGF-II	Stimulatory	15 ↓
IGFBP-1	Inhibitory	20 ↑
IGFBP-2	Inhibitory	35 ↑
IGFBP-3	Stimulatory/inhibitory	25 ↓
IGFBP-4	Inhibitory	35 ↓
IGFBP-5	Stimulatory	35 ↓
IGFBP-6	Inhibitory	?

Estimated percent change between 20–29 and more than 60 yr of age (see text for references).

multiple IGF system components occurs can be seen during aging (8–18). With advancing age, not only does the concentration of IGFs decrease, but the concentration of stimulatory IGFBP-5 decreases, whereas the concentrations of inhibitory IGFBP-1, IGFBP-2, and IGFBP-4 increase with age (Table 2). Thus, the multiple deficits in the IGF system components that occur as a function of age lead to a marked decrease in the ratio of stimulatory IGFs to inhibitory IGFBPs, which would tend to decrease the endocrine actions of IGFs.

Another potential control mechanism for regulation of endocrine IGF actions is by IGFBP-specific proteases. Since the discovery of IGFBP proteases in pregnancy serum several years ago (19, 20), scores of papers have been published identifying the IGFBP protease activities in the conditioned media of various cell types and in sera or body fluids of patients with a variety of pathophysiological conditions (21–25). Although it is not known at this time whether the various IGFBP proteases identified are absolutely specific to IGFBPs or whether there are multiple proteases for each IGFBP, the findings that the rate of IGFBP proteolysis can be regulated by a variety of systemic and local effectors raise interesting possibilities for the involvement of IGFBP proteases in regulating the endocrine actions of IGF. For example, IGFBP-3 protease has been proposed to play an important role in modulating the bioavailability of IGFs. As the ternary complex of ALS-IGFBP-3-IGF-I does not appear to cross the vascular endothelium, it has been proposed that an increase in IGFBP-3 proteolysis leads to the breakdown of the ternary complex and a corresponding increase in the formation of a lower molecular mass IGF-IGFBP complex, which is capable of crossing the vascular endothelial barrier. Consistent with this idea, the percentage of IGF-I was significantly reduced in the 150-kDa ternary complex, with a concomitant increase in the percentage of free IGF-I in pregnancy plasma (25). In contrast to these findings, Suikkari and Baxter (26) have shown that the proteolyzed IGFBP-3 from pregnancy serum

can bind to IGFs and form a ternary complex with ALS of normal affinity. Thus, the functional significance of pregnancy-associated serum proteolysis of IGFBP-3 has not been fully established.

IGFBPs as potential modulators of local IGF actions

In addition to modulating the endocrine actions of IGF as described above, IGFBPs may have other roles.

To increase IGF bioavailability in specific tissues. Recent evidence suggests that IGFBP proteases may also regulate the bioavailability of IGFs in a tissue-specific manner. For example, it is known that the IGFBP-4-IGF complex is inactive, as this complex does not bind to IGF receptors, and that proteolysis of IGFBP-4 can increase the amount of local IGF available for IGF receptor interaction. Based on these findings and the observation that a number of local growth factors (e.g. TGF β) can regulate IGFBP-4 proteolysis (23), it can be speculated that some effectors may increase the local production of IGFBP proteases, which, in turn, may degrade the inhibitory IGFBPs in extracellular fluid and thus increase the free level of IGFs for receptor interaction. Thus, tissue-specific regulation of IGFBP proteolysis may provide a mechanism to increase site-specific bioavailability of serum IGFs depending on local needs. In addition, the findings that IGFs regulate IGFBP levels by regulating proteolysis in a variety of cell types raise the interesting possibility that IGFs function to regulate IGFBPs, as well as *vice versa*. Future understanding of the identity, regulation, and roles of IGFBP proteases produced by various cell types will resolve these issues.

To facilitate storage of IGFs in extracellular matrixes. It is well known that growth factors, such as IGFs, are stored in relatively high abundance in certain extracellular matrixes, such as bone (27–29). Studies related to the mechanisms by which IGFs are stored in bone revealed that IGFBP-5, but not IGF-I or -II, can bind to hydroxyapatite and extracellular matrix proteins (27–29). These data suggest that IGFs are fixed in bone by means of IGFBP-5. The concept of a binding protein being involved in the storage of growth factors is not unique to IGFs, as TGF β appears to be stored in bone via its binding protein. Although, the extent to which the IGFBPs in serum contribute to tissue fixation of IGFs is not clear, it is interesting to speculate that serum IGFBP-5 or locally produced IGFBP-5 may help to localize IGFs in a given tissue by binding to extracellular matrix proteins, thereby creating a growth factor depot in certain tissues for future actions. In this regard, we and others have proposed that growth factors fixed in bone may be released during bone resorption to stimulate nearby osteoblasts so as to ensure that there is a site-specific replacement of the resorbed bone. Similarly, IGFs stored in the extracellular matrixes of certain other tissues may promote wound healing and tissue regeneration.

To exert IGF-independent effects on target cells. The explosion of IGFBP research during the last few years has led to the novel concept that IGFBPs are not simply transport proteins, but, in addition, either inhibit or potentiate IGF actions (2). Recent studies on the mechanisms by which IGFBPs may mediate their effects on target cells have revealed an interesting pos-

sibility that some IGFBPs may have IGF-independent effects besides modulating the actions of IGF. For example, IGFBP-1 has been shown to stimulate smooth muscle cell migration in an IGF-independent manner involving binding to integrin receptor (30). IGFBP-3 and IGFBP-5 have also been shown to mediate their effects on breast cancer cells and bone cells, respectively, by IGF-independent mechanisms, possibly involving IGFBP-specific cell surface binding sites (31–33). Valentinis *et al.* (34) have shown that overexpression of IGFBP-3 in fibroblasts derived from IGF-I receptor knockout mouse embryo inhibited cell growth, suggesting that the inhibitory effect of IGFBP-3 does not involve signal transduction via type I IGF receptor. Recent studies have also provided evidence for the presence of putative IGFBP-binding sites on the cell surface (31–33). These data support the unique possibility that serum IGFBPs may modulate their effects on target tissues depending on local needs by not only regulating the endocrine actions of IGF, but also by having IGF-independent direct effects on target cells. However, further studies involving the identification of signal-transducing IGFBP receptors and establishment of a cause and effect relationship between IGFBP binding to its potential receptor and IGFBP actions are needed to verify that IGFBPs may in part mediate their effects independent of IGFs. This is clearly an exciting area for future investigations.

In conclusion, all important regulatory systems in biology are complex, and the IGF regulatory system is no exception. The last few years have brought complexity, but also new vistas of insights into the IGF system with the discovery of new IGFBPs and new putative IGFBP proteases. One can envision very exciting times in the future as the scientific community seeks to advance the frontiers of both the molecular mechanisms of the IGF system and also the bigger picture of the function of the IGF system as a hormone and a local signaling system.

Subburaman Mohan, Ph.D David J. Baylink, M.D.
Jerry L. Pettis Veterans Administration Medical
Center
Loma Linda University
Loma Linda, California 92357

References

1. Rechler MM. 1993 Insulin-like growth factor binding proteins. *Vitam Horm.* 47:1–114.
2. Jones JJ, Clemmons DR. 1995 Insulin-like growth factors and their binding proteins: biological actions. *Endocr Rev.* 16:3–34.
3. Daughaday WH, Rotwein P. 1989 Insulin-like growth factors I and II. Peptide, messenger ribonucleic acid and gene structures, serum, and tissue concentrations. *Endocr Rev.* 10:68–91.
4. van Dessel HJHM, Chandrasekhar Y, Yap OWS, et al. 1996 Serum and follicular fluid levels of insulin-like growth factor-I (IGF-I), IGF-II and IGF binding protein-1 and -3 during the normal menstrual cycle. *J Clin Endocrinol Metab.* 81:1224–1231.
5. Baxter RC. 1994 Insulin-like growth factor binding proteins in the human circulation: a review. *Horm Res.* 42:140–144.
6. Guler HP, Zapf J, Schmid C, Froesch ER. 1989 Insulin-like growth factors (IGFs) I and II in healthy man. Estimation of half-lives and production rates. *Acta Endocrinol (Copenh).* 121:753–758.
7. Baxter RC, Martin JL. 1989 Structure of the 150,000 growth hormone-dependent insulin-like growth factor binding protein (IGFBP) complex: determination by reconstitution and affinity labeling. *Proc Natl Acad Sci USA.* 86:6898–6902.
8. Honda Y, Landale EC, Strong DD, Baylink DJ, Mohan S. 1996 Recombinant synthesis of insulin-like growth factor binding protein-4 (IGFBP-4): develop-

- ment, validation, and application of a radioimmunoassay for IGFBP-4 in human serum and other biological fluids. *J Clin Endocrinol Metab.* 81:1389–1396.
9. Mohan S, Libanati C, Dony C, Lang K, Srinivasan N, Baylink DJ. 1995 Development, validation, and application of a radioimmunoassay for insulin-like growth factor binding protein 5 in human serum and other biological fluids. *J Clin Endocrinol Metab.* 80:2638–2645.
 10. Hall K, Lundin G, Pova G. 1988 Serum levels of the low molecular weight form of insulin-like growth factor binding protein in healthy subjects and patients with growth hormone deficiency, acromegaly and anorexia nervosa. *Acta Endocrinol (Copenh).* 118:321–326.
 11. Busby WH, Snyder DK, Clemmons DR. 1988 Radioimmunoassay of a 26,000-dalton plasma insulin-like growth factor-binding protein: control by nutritional variables. *J Clin Endocrinol Metab.* 67:1225–1230.
 12. Clemmons DR, Snyder DK, Busby WH. 1991 Variables controlling the secretion of insulin-like growth factor binding protein-2 in normal human subjects. *J Clin Endocrinol Metab.* 73:727–733.
 13. Schwander J, Mary JL. 1992 The RIA for IGFBP-2 in man: a meagre catch. *Growth Regul.* 3:104–108.
 14. Blum WF, Horn N, Kratzsch J, et al. 1993 Clinical studies of IGFBP-2 by radioimmunoassay. *Growth Regul.* 3:100–104.
 15. Baxter RC, Martin JL. 1986 Radioimmunoassay of growth hormone-dependent insulin-like growth factor binding protein in human plasma. *J Clin Endocrinol Metab.* 78:1504–1512.
 16. Blum WF, Ranke MB, Kietzmann K, Gauggel F, Zeisel HJ, Bierich JR. 1990 A specific radioimmunoassay for the growth hormone(GH)-dependent somatomedin-binding protein: its use for diagnosis of GH deficiency. *J Clin Endocrinol Metab.* 70:1292–1298.
 17. Baxter RC, Sanders H. 1992 Radioimmunoassay of insulin-like growth factor binding protein-6 in human serum and other biological fluids. *J Endocrinol.* 134:133–139.
 18. Mohan S, Farley JR, Baylink DJ. 1995 Age-related changes in IGFBP-4 and IGFBP-5 levels in human serum and bone: implications for bone loss with aging. *Prog Growth Factor Res.* 6:465–473.
 19. Giudice LC, Farrel EM, Pham H, Lamson G, Rosenfeld RG. 1990 Insulin-like growth factor binding proteins in maternal serum throughout gestation in puerperium: effects of pregnancy associated serum protease activity. *J Clin Endocrinol Metab.* 71:806–816.
 20. Hossenlopp P, Sergovia B, Lassarrie C, Roghani M, Bredon M, Binoux M. 1990 Evidence of enzymatic degradation of insulin-like growth factor binding proteins in the 150 K complex during pregnancy. *J Clin Endocrinol Metab.* 71:797–805.
 21. Fowlkes J, Freemark M. 1992 Evidence for a novel insulin-like growth factor (IGF)-dependent protease regulating IGF-binding protein-4 in dermal fibroblasts. *Endocrinology.* 131:2071–2076.
 22. Kanzaki S, Hilliker S, Baylink DJ, Mohan S. 1994 Evidence that human bone cells in culture produce insulin-like growth factor binding protein-4 and -5 proteases. *Endocrinology.* 134:383–392.
 23. Durham SK, Riggs L, Conover C. 1994 The insulin-like growth factor-binding protein-4 (IGFBP-4)-IGFBP-4 protease system in normal human osteoblast-like cells: regulation by transforming growth factor- β . *J Clin Endocrinol Metab.* 79:1752–1758.
 24. Giudice LC. 1995 Editorial: IGF binding protein-3 protease regulation: how sweet it is! *J Clin Endocrinol Metab.* 80:2279–2281.
 25. Blat C, Villaudy J, Binoux M. 1994 *In vivo* proteolysis of serum insulin-like growth factor binding protein-3 results in increased availability of IGF to target cells. *J Clin Invest.* 93:2286–2290.
 26. Suikkari A-M, Baxter RC. 1992 Insulin-like growth factor binding protein-3 is functionally normal in pregnancy serum. *J Clin Endocrinol Metab.* 74:177–183.
 27. Mohan S, Jennings JC, Linkhart TA, Baylink DJ. 1988 Primary structure of human skeletal growth factor: homology with human insulin-like growth factor-II. *Biochim Biophys Acta.* 966:44–55.
 28. Nicolas V, Mohan S, Honda Y, et al. 1995 An age-related decrease in the concentration of insulin-like growth factor binding protein-5 in human cortical bone. *Calcif Tissue Int.* 57:206–212.
 29. Jones JJ, Gockerman A, Busby WH, Camacho-Hubner C, Clemmons DR. 1993 Extracellular matrix contains insulin-like growth factor binding protein-5: potentiation of the effects of IGF-I. *J Cell Biol.* 121:679–687.
 30. Jones JJ, Gockerman A, Busby WH, Wright G, Clemmons DR. 1993 Insulin-like growth factor binding protein-1 stimulates cell migration and binds to the $\alpha 1 \beta 5$ integrin by means of its Arg-Gly-Asp sequence. *Proc Natl Acad Sci USA.* 90:10553–10557.
 31. Oh Y, Muller HL, Lamson G, Rosenfeld RG. 1993 Insulin-like growth factor (IGF)-independent action of IGF binding protein-3 in Hs578T human breast cancer cells: cell surface IGF binding and growth inhibition. *J Biol Chem.* 268:14964–14971.
 32. Andress DL, Birnbaum RS. 1992 Human osteoblast-derived insulin-like growth factor binding protein-5 stimulates osteoblast mitogenesis and potentiates IGF action. *J Biol Chem.* 267:22467–22472.
 33. Mohan S, Nakao Y, Honda Y, et al. 1995 Studies on the molecular mechanisms by which insulin-like growth factor (IGF) binding protein-4 (IGFBP-4) and IGFBP-5 modulate IGF actions in bone cells. *J Biol Chem.* 270:20424–20431.
 34. Valentinis B, Bhala A, DeAngelis T, Baserga R, Cohen P. 1995 The human insulin-like growth factor (IGF) binding protein-3 inhibits the growth of fibroblasts with a targeted disruption of the IGF-I receptor gene. *Mol Endocrinol.* 9:361–367.