

The *INO2* gene of *Saccharomyces cerevisiae* encodes a helix-loop-helix protein that is required for activation of phospholipid synthesis

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In the yeast *Saccharomyces cerevisiae*, biochemical and genetic evidence have established that a number of phospholipid biosynthetic enzymes are coordinately regulated in response to the soluble precursors inositol and choline and a common set of regulatory factors (1). The *ino2* and *ino4* mutants show pleiotropic defects in phospholipid metabolism. Recessive mutations at the *INO2* locus lead to reduced phosphatidylcholine synthesis and inositol auxotrophy due to an inability to derepress expression of the *INO1* structural gene (which encodes inositol-1-phosphate synthase) (2), (3). *Ino2* mutant extracts also lack a specific DNA-protein complex that is present in wildtype extracts (4). Thus, the *INO2* locus encodes a positive regulatory factor required for derepression of the coregulated phospholipid biosynthetic enzymes.

The wildtype *INO2* gene was isolated by functional complementation of the inositol auxotrophy in an *ino2* mutant. Upon transformation with a partial *Sau3A* genomic library, one plasmid harboring a 1.8 kilobase *SmaI*-*XbaI* insert restored inositol prototrophy. Integrative transformation established linkage to the *ino2-21* mutation. In a cross between an integrant and wildtype strain 48 tetrads showed 4:0 segregation for the *Ino*⁺ phenotype, confirming that the cloned DNA represents the authentic *INO2* locus. The nucleotide sequence of the *INO2* gene was determined on both strands by the Sanger dideoxy chain termination method (5). Computer-assisted sequence analysis revealed 912 base pair open reading frame, capable of encoding a 304 amino acid protein with a predicted molecular mass of 34,234 daltons. The *Ino2* protein (*Ino2p*) is largely hydrophilic and acidic (pI = 5.76). Proline residues comprise 8.5% of the protein. A potential structural similarity between the carboxy-

terminus of *Ino2p* and the helix-loop-helix (HLH) domain of the proto-oncogene *c-myc* mapped to residues 253–291 of *Ino2p* (see Figure 1) (6). Basic residues precede the HLH domain of *Ino2p*. Interestingly, the *Ino4* protein, which encodes a known transcriptional activator of phospholipid synthesis shares sequence similarity to *Ino2p* in a region that is restricted to the 68 amino acid HLH structural domain (9). Extracts prepared from *ino4* mutants lack the same DNA-protein complex that is missing in extracts prepared from *ino2* mutants (4). Thus, it is tempting to speculate that *Ino2p* and *Ino4p* may be dimerization partners that associate in a DNA-protein complex to regulate the expression of phospholipid structural genes.

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REFERENCES

- Nikoloff,D.M. and Henry,S.A. (1991) *Annu. Rev. Genet.* **25**, 559–583.
- Hirsch,J.P. and Henry,S.A. (1986) *Mol. Cell. Biol.* **6**, 3320–3328.
- Loewy,B.S. and Henry,S.A. (1984) *Mol. Cell. Biol.* **4**, 2479–2485.
- Lopes,J.M. and Henry,S.A. (1991) *Nucleic Acids Res.* **19**, 3987–3994.
- Sanger,F. *et al.*, (1977) *Proc. Natl. Acad. Sci. USA* **74**, 5463–5467.
- Murre,C., McCaw,P.S. and Baltimore,D. (1989) *Cell* **56**, 777–783.
- Cai,M. and Davis,R. (1990) *Cell* **61**, 437–446.
- Berben,G., Legrain,M., Gilliquet,V. and Hilger,F. (1990) *Yeast* **6**, 451–454.
- Hoshizaki,D.K., *et al.*, (1990) *J. Biol. Chem.* **265**, 4736–4745.

Myc consensus	NDKKRTNNVLEBQRBNELKSSFFALRDOVP	...EL...ENNEKAPRVVLEKATEVILLSLOAD
	BASIC / HELIX I /	LOOP / HELIX II
<i>Ino2</i>	KYRQWKNVOMEKIERINTKEAPERLIKSVR	TPPK...ENKRIKPHILLTCVMNDIKSERBA
<i>Ino4</i>	QGIKINHYSSSEKRRERLERAIIDELVAVVP	DLQ...QKRSRLTIYLKLSLVSLSWLYER
<i>Cbf1</i>	KQRDSEKKEVERREKRENTAIVNLSOLLP	...VSRPKAAILARAARVYQKLEKTEDE
<i>Pho4</i>	DERESKHAFOAQRNRLAVALHFLASLIP	FWKO...ONVSAATPKAAILARAARVYQKLEKTE
<i>Mu MyoD</i>	ADRRKAATNREERLVLKSYNEAFETLKRCTI	SNP...NORLPRVVEILLNAILRVYIEGL
<i>Hu E12</i>	KFRIVANMAEPEELIVVDINCAFELCPMCO	LHI V...SEKPTKELIILHQAVSVVILLNLEQVY
<i>Hu E47</i>	RFRSMANAREEIVVDINCAFELCKMCO	MHI I...SDRAQTKELIILQOAVGVILLGIFQVY
<i>Hu c-Myc</i>	NDKRTNNVLEBQRBNELKSSFFALRDOVP	ENNEKAPRVVLEKATEVILLSLOAD
H-L-H Motif	RR N ER R V F I V	EV IR VL AV IV

Figure 1. Alignment of several representative helix-loop-helix regulatory proteins. Amino acid similarity between the HLH domain of *Ino2p* and *Ino4p* (9), *Cbf1* (7), *Pho4* (8), *MyoD* (6), *E12* (6), *E47* (6) and *c-myc* (6). Identity is indicated in bold face; conservative substitution is denoted by an underline; Y represents hydrophobic amino acid residues.

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