

132. Lass P, Buscombe JR, Harber M *et al.* Cognitive impairment in patients with renal failure is associated with multiple-infarct dementia. *Clin Nuclear Medicine* 1999; 24: 561–565
133. Gowdak LHW, Arantes RL, de Paula FJ *et al.* Underuse of American College of Cardiology/American Heart Association Guidelines in hemodialysis patients. *Ren Fail* 2007; 29: 559–565
134. Gowdak LH, Arantes RL, de Paula FJ *et al.* beta-Blocker use in long-term dialysis patients: association with hospitalized heart failure and mortality. *Arch Intern Med* 2004; 164: 2465–471
135. Cice G, Ferrara L, D'Andrea A *et al.* Carvedilol increases two year survival in dialysis patients with dilated cardiomyopathy, a prospective placebo controlled trial. *J Am Coll Cardiol* 2003; 41: 1438–1444
136. Sigrist MK, Taal MW, Bungay P *et al.* Progressive vascular calcification over 2 years is associated with arterial stiffening and increased mortality in patients with stages 4 and 5 chronic kidney disease. *Clin J Am Soc Nephrol* 2007; 2: 1241–1248
137. Takei T, Otsubo S, Uchida K *et al.* Effects of sevelamer on the progression of vascular calcification in patients on chronic hemodialysis. *Nephron Clin Pract* 2008; 108: c278–c288
138. Spalding EM, Chandna SM, Davenport A *et al.* Kt/V underestimates the haemodialysis dose in women and small men. *Kidney Int* 2008; 74: 348–355
139. Banerjee A, Davenport A. Changing patterns of pericardial disease in patients with end-stage renal disease. *Hemodial Int* 2006; 10: 249–255
140. Tordoir J, Canaud B, Haage P *et al.* European Best Practice Guidelines on vascular access. *Nephrol Dial Transplant* 2007; 22: 88–117
141. Basile C, Lomonte C, Vernaglione L *et al.* The relationship between the flow of arteriovenous fistula and cardiac output in haemodialysis patients. *Nephrol Dial Transplant* 2008; 23: 282–287
142. Cridlig J, Selton-Suty C, Alla F *et al.* Cardiac impact of the arteriovenous fistula after kidney transplantation: a case-controlled, matched study. *Transpl Int* 2008; 21: 948–954
143. Levin NW, Kotanko P, Eckardt KU *et al.* Blood pressure in chronic kidney disease stage 5D-report from a Kidney Disease: Improving Global Outcomes Controversies Conference. *Kidney Int* 2010; 77: 273–284

Received for publication: 18.11.09; Accepted in revised form: 15.4.10

Nephrol Dial Transplant (2010) 25: 2089–2098  
doi: 10.1093/ndt/gfq231

## Post-transplant lymphoproliferative disorder in view of the new WHO classification: a more rational approach to a protean disease?

Krzysztof Mucha<sup>1,2</sup>, Bartosz Foroniewicz<sup>1,2</sup>, Bogna Ziarkiewicz-Wróblewska<sup>3</sup>, Marek Krawczyk<sup>4</sup>, Jan Lerut<sup>5</sup> and Leszek Pączek<sup>1</sup>

<sup>1</sup>Transplantation Institute, Department of Immunology, Transplantology and Internal Medicine, Warsaw Medical University, Warsaw, Poland, <sup>2</sup>Institute of Biochemistry and Biophysics Polish Academy of Sciences, Warsaw, Poland, <sup>3</sup>Department of Pathology, Warsaw Medical University, Warsaw, Poland, <sup>4</sup>Department of General, Transplant and Liver Surgery, Medical University of Warsaw, Warsaw, Poland and <sup>5</sup>Unit of Abdominal Transplantation, Department of Abdominal and Transplantation Surgery, Université catholique de Louvain-UCL, Cliniques Universitaires Saint-Luc, Brussels, Belgium

Correspondence and offprint requests to: Krzysztof Mucha; E-mail: kjmucha@gmail.com

### Abstract

Post-transplant lymphoproliferative disorders (PTLDs) are serious, life-threatening complications of solid-organ transplantation (SOT) and bone marrow transplantation leading to a high mortality (30–60%). PTLD represents a heterogeneous group of lymphoproliferative diseases. They become clinically relevant because of the expansion of transplantation medicine together with the development of potent immunosuppressive drugs. Although the diagnostic morphological criteria of different forms of PTLD are commonly known, rapid and correct diagnosis is not always easy. Because of the limited number of clinical trials, a consensus is lacking on the optimal treatment of PTLD. This review focuses on incidence, risk factors, clinical

picture of the disease and diagnostic tools including histopathology relating to the new classification introduced in 2008 by the World Health Organisation (WHO) and treatment of PTLD.

**Keywords:** EBV; histopathology; PTLD; SOT; transplantation

### Introduction

The term ‘post-transplant lymphoproliferative disorder’ or disease (PTLD) was first introduced in 1984 by Starzl [1]. Today, it represents a heterogeneous group of lymphoproliferative

**Table 1.** The incidence and location of PTLD in solid-organ transplant recipients

Type of transplanted organ	Location and frequency [%]							References
	Kidney	Lung	Liver	CNS	Lymph nodes	GI tract	Disseminated	
Kidney	10.3–32	4.4	4.9	11.7	9.5	15.3	14	[4–7,16,23,32]
Liver		4.2	21.8–33	4.2	9.7	12.1	13.3	
Heart	0.6	16.0	8.9	4.0	4.4	14.3	14.5	
Lung and heart–lung	1.4	50–80	4.8	3.4	2.1	4.8	10.3	

ferative diseases, ranging from Epstein–Barr virus (EBV)-associated polyclonal proliferation to highly aggressive monomorphic proliferations, such as diffuse large B-cell lymphoma (DLBCL) [2,3]. The reported incidence of PTLD is very variable as is its related mortality (30–60%). The clinical picture, intensity of immunosuppression (IS), primary and co-existing diseases and PTLD location are also quite variable. For all these reasons, the diagnosis of PTLD is not easy; moreover, the pathologist frequently faces unusual cases of lymphoid proliferations that do not fit any recognized PTLD type. Therefore, WHO introduced in 2008 a new PTLD classification, aimed at improving diagnosis and consequently treatment modalities. So far, immunosuppression reduction (IR) is the only accepted first-line therapy; however, poor response in many types of PTLD requires other approaches such as antiviral therapies, chemotherapy, monoclonal antibodies, proliferation signal inhibitors (PSIs) and finally surgery.

### Incidence and risk factors

The incidence of PTLD, ranging from 1% to 20%, clearly relates to the type of transplanted organ, IS, underlying disease, age, viral infections including EBV, cytomegalovirus and hepatitis C virus (HCV), and length of post-transplant follow-up. Although the occurrence decreases after the first post-transplant year, the cumulative incidence increases with the time elapsed since transplantation. The highest incidence from 5% to 20% is reported following lung and intestinal transplantation; in contrast, renal transplant recipients (RTRs) have an incidence of 1–3% [4,5]. In liver transplant recipients, the occurrence ranges from 2% to 10%, from 2–3% in adults to 10% in children [4,6,7] (Table 1).

Induction IS using anti-T-cell antibodies such as OKT3® (Janssen-Cilag, New Brunswick, USA) or Thymoglobulin® can lead to an increased risk of PTLD. In contrast, ATG® (Fresenius Biotech GmbH, Germany) or interleukin 2-receptor (IL-2R) antibodies induction (Simulect®, Novartis Pharma AG, Basel, Switzerland) do not increase PTLD risk [4,8–10]. On the other hand, the treatment of rejection episodes during the first post-transplant year with OKT3 or ATG enhances the PTLD risk in patients who did not receive antibody induction. Moreover, in patients who also received antibody induction, rejection therapy with OKT3 or ATG adds to the already increased lymphoma risk [4]. The PTLD risks associated with the use of the calcineurin inhibitors (CNI) tacrolimus (Prograf®, Astellas Pharma Europe B.V., Leiderdorp, The

Netherlands) and cyclosporin A (Neoral®, Novartis Pharma AG, Basel, Switzerland), and PSI: sirolimus (Rapamune®, Pfizer, New York, USA) and everolimus (Certican®, Novartis Pharma AG, Basel, Switzerland) remains uncertain [11–13]. It is, however, clear that the intensity of IS, rather than the use of any particular agent, is critical for development of PTLD.

EBV infection is a risk factor and a cause of PTLD present in more than 80% of B-lymphocyte phenotypic disorders and, less commonly, in T-lymphocytic proliferations [9,14]. After infection, B cells incorporate EBV DNA into the cellular genome, decreasing the rate of apoptotic cell death through bcl-2 induction and stimulating extensive proliferation of B cells, possibly leading to lymphoblastoid transformation [10,15]. EBV-seronegative recipients receiving transplants from EBV-seropositive donors are at particular risk for PTLD development. Paediatric recipients who are very frequently EBV seronegative before transplantation are especially prone to develop PTLD.

Cytomegalovirus and HCV are also, albeit less, involved in the pathogenesis of PTLD [16–18]; however, their role as risk factors for PTLD is controversial, as is that of herpes simplex or simian virus infections [19].

Human leukocyte antigen (HLA) matching is another risk factor in the pathogenesis of PTLD in RTRs; HLA-B or HLA-DR mismatches especially seem to be critical, independently of the type of IS. The number of HLA mismatches parallels with an increased risk of PTLD. HLA-B mismatches increase the risk of lymphoma in the kidney, whereas HLA-DR locus mismatches are exposed to a higher risk of non-Hodgkin lymphoma, namely located in the kidney and the central nervous system [20,21].

### Clinical picture

The clinical picture of PTLD differs from that of lymphomas observed in the general population. Aggressiveness and outcome of the PTLD depends on the histological type and/or the supplementary fact that transplant recipients are more susceptible to develop complications after lymphoma treatment.

Symptoms may be mild, such as fever, mononucleosis-like syndrome, lymphadenopathy, recurrent infections of unknown origin resistant to antibiotics or severe organ dysfunction. The variable manifestation of PTLD depends on numerous factors, including the type of transplanted organ or IS used, histopathology and time elapsed since transplantation. The incidence during the first year varies depending on the reported series, and one must be cautious,

as the mean time to diagnosis changes with the duration of follow-up. It has been shown that the PTLTD incidence at 1 year was only one-fifth of the cumulative 10-year incidence and that the median time of occurrence reaches about 5 years [4]. A case of PTLTD observed as early as 15 days after kidney–pancreas grafting has been reported [22]. The location of the lesions also relates to the type of transplant and the time span elapsed since transplantation. In lung recipients, more than 50% of all PTLTDs develop during the first post-transplant year in the allograft itself in contrast to other organs [4,23]. The large European registry of over 200 000 SOT recipients estimated that the respective allograft was affected in 10.3% of kidney, 16% of heart, 21.8% of liver and 42.7% of heart–lung recipients (Table 1) [4]. The exact mechanism leading to a preferential allograft localization remains unclear today; chronic antigen stimulation, passenger lymphocytes in the graft or development of lymphoma from donor lymphocytes have all been mentioned in relation to this.

Other frequent sites of PTLTD include the gastrointestinal tract (jejunum more often than colon), lymph nodes and central nervous system. The involvement of these locations also varies between types of transplanted organ [4] and depends on the age of the recipient too. Skin and tonsils are considered rare PTLTD locations in adults, whereas in children, the lymph nodes of the Waldeyer's ring and tonsils are very common target organs. Regardless of the graft type, patients with lymph node localization have a relatively good outcome; disseminated disease in contrast has a poor prognosis.

## Diagnosis

PTLTD often presents in a nonspecific way. Medical history, thorough physical examination and different endoscopic and imaging techniques are crucial in making a prompt diagnosis. Fluorodeoxyglucose-positron emission tomography has proved superior to conventional methods of PTLTD visualization. The final diagnosis is always based on histopathology.

Assessment of EBV DNA load is important for early identification and appropriate monitoring of high-risk recipients [24]. Detection of an increased EBV load alone is not always predictive of PTLTD, a fact that may be explained by the concomitant increase in EBV-specific cellular responses. Because there are no established threshold values of the number of EBV DNA copies, the dynamics of increasing EBV DNA levels is a helpful guide to decrease IS or start cytotoxic T-cell-based or anti-CD20 therapies [25]. Some authors have suggested that the detection of increased EBV DNA load in combination with reduced EBV-specific T-cell counts could allow a more precise prediction of the risk of PTLTD development, particularly in EBV-seronegative recipients [26]. Additionally, the EBV particles can be detected in examined tissues by immunohistochemistry (IHC) and molecular methods including in situ hybridization [27]. Detection of antibodies against the EBV latent membrane protein 1 (LMP1) or EBV nuclear antigen 2 may be applied in frozen or paraffin-embedded tissues.

Polymorphisms of interferon- $\gamma$ , tumour necrosis factor- $\alpha$  and IL-2 related to a low cellular immune response, as well as increased levels of IL-10 acting as an autocrine growth factor for EBV-transformed B cells are associated with an increased risk of PTLTD. They all represent other measures that could potentially supplement diagnostics of EBV-related PTLTD [28–30].

## Histopathological classification

As the morphological picture of PTLTD is variable, several classifications of PTLTD have been put forward. According to the newest WHO classification, introduced in 2008, four basic histological types of PTLTD have been identified [31].

### *Typical morphological picture of different forms of PTLTD*

Plasmacytic hyperplasia (PH), mononucleosis-like PTLTD, and polymorphic PTLTD (P-PTLTD) are specific for transplant recipients, whereas the other types can also be diagnosed in immunocompetent individuals.

**1. Early lesions.** The architecture of the involved tissue in PH is generally retained, and nodal sinuses are preserved. Reactive follicles in the periphery of the lymph node are often seen. The large number of polyclonal plasma cells (VS38c+,  $\kappa$ +,  $\lambda$ +) may be found as single cells, small groups or large sheets (Figure 1A–C) together with lymphocytes and occasional immunoblasts.

The microscopic picture of the infectious mononucleosis-like PTLTD typically appears with the expansion of the T zone with numerous immunoblasts and plasma cells (Figure 1D).

Cytologic atypia in both types of early lesions is minimal. Early PTLTD is usually diagnosed in children, young adults and patients with primary EBV infection [31,32].

**2. P-PTLTD.** The architecture of involved tissues is affected by an infiltrate consisting of various cells: small and medium-sized lymphoid cells, centrocyte-like cells, plasma cells and immunoblasts. Atypical lymphoid cells and cells resembling Reed-Sternberg cells (RS) may be observed. It is the most common type in children, usually related to primary EBV infection [33] (Figure 1E).

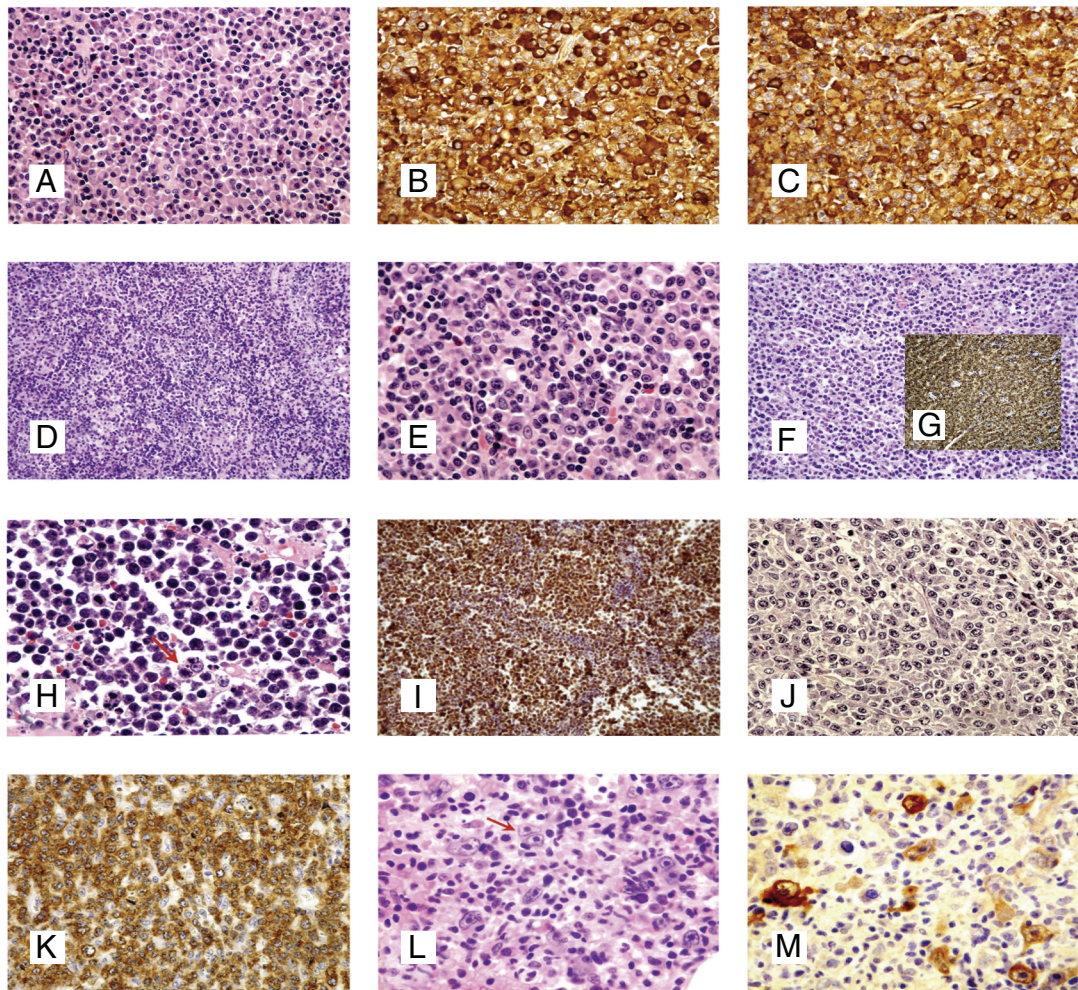
**3. Monomorphic PTLTD (M-PTLTD)** includes all T/natural killer neoplasms and most B-cell lymphomas.

**4. Classical Hodgkin lymphoma (HL)** is diagnosed according to the same criteria as in immunocompetent patients. The most frequent form is of mixed cellularity [34]. Classic RS cells are seen in a reactive inflammatory background (small lymphocytes, histiocytes, plasma cells, eosinophils). Diagnostic cells are typically CD30 and CD15 positive and CD20 marker variable.

### *Typical and common forms of PTLTD*

The first three types of PTLTD are relatively common and usually develop with the typical morphological features





**Fig. 1.** Histopathology of PTLD. (A) PH. Haematoxylin and eosin (H&E) stain. Objective magnification  $\times 40$ . Large number of plasma cells admixed with small lymphocytes and singular eosinophils. (B) PH. Immunohistochemical stain with kappa immunoglobulin light-chain antibody. Objective magnification  $\times 60$ . (C) PH. Immunohistochemical stain with lambda immunoglobulin light-chain antibody. Objective magnification  $\times 60$ . (D) Infectious mononucleosis-like PTLD. H&E stain. Proliferation of small lymphoid cells, plasma cells and immunoblasts. Objective magnification  $\times 10$ . (E) P-PTLD. H&E stain. Many plasma cells, immunoblasts (large cells with vesicular nuclei and clearly visible centrally located nucleoli, singular eosinophils). Objective magnification  $\times 60$ . (F) M-PTLD, DLBCL. H&E stain. Atypical large cells with large irregular nuclei, abundant cytoplasm, and two to three peripherally located nucleoli. Objective magnification  $\times 20$ . (G) M-PTLD, DLBCL. Immunohistochemical stain. Positive reaction of lymphoma cells with CD20. Objective magnification  $\times 20$ . (H) M-PTLD, Burkitt's lymphoma. H&E stain. Medium-sized B cells with multiple inconspicuous nuclei and scattered tingible-body macrophages containing cellular debris. Objective magnification  $\times 60$ . (I) M-PTLD, Mantle cell lymphoma. Immunohistochemical stain with cyclin D1 (SP4). Typical nuclear staining. Objective magnification  $\times 20$ . (J) M-PTLD, PTCL, unspecified. H&E stain. Polymorphic medium-sized and large cells, many with irregular vesicular nuclei and distinct nucleoli. Objective magnification  $\times 40$ . (K) M-PTLD, PTCL, unspecified. Positive immunohistochemical stain with CD3 antibody. Objective magnification  $\times 40$ . (L) M-PTLD, DLBCL, Hodgkin-like. H&E stain. Objective magnification  $\times 60$ . Numerous Hodgkin-like and RS-like cells, with abundant cytoplasm, multilobular nucleus and one to two prominent nucleoli. (M) M-PTLD, DLBCL, Hodgkin-like. Immunohistochemical stain. Objective magnification  $\times 60$ . Positive reaction with EBV (LMP1).

described above. More than 85% of PTLDs derive from B cells, 14% from T cells and about 1% from natural killer cells [31,35–37].

Among M-PTLDs, the most common is DLBCL. The term 'monomorphic' does not reflect similarity of the cells, which are often different in shape and size. RS-like, multinucleated and plasmacytoid forms may be encountered. According to the 2008 WHO classification, DLBCL is classified as a centroblastic or immunoblastic variant. Centroblastic DLBCL presents with diffuse proliferation of atypical large cells with large irregular nuclei and two to three nucleoli located peripherally (Figures 1F and G).

The immunoblastic variant has large neoplastic cells with large nuclei and centrally located prominent nucleoli. IHC distinguishes two main groups of DLBCL: those originating from germinal centre cells and those of non-germinal-centre phenotype. The proliferation index of DLBCL is high; Ki-67 exceeds 40% of lymphoma cells [38].

Other types of B-cell lymphomas such as Burkitt's lymphomas are diagnosed less frequently [39,40]. Burkitt's lymphoma presents with the same pathomorphology as in immunocompetent patients: monomorphic medium-sized B cells infiltrate with extremely high mitotic activity, often with the presence of many tingible-body macro-

phages, containing cellular debris ('starry sky' pattern) (Figure 1H).

Another group of M-PTLDs are the plasma cell neoplasms: multiple myeloma or extramedullary plasmacytoma, which are morphologically and immunophenotypically identical to the forms in immunocompetent patients [16,41,42].

#### *Rare forms of PTLT*

Although common in immunocompetent patients, some types of lymphoma such as mantle cell lymphoma develop rarely in transplant recipients. Centrocyte-like cells with irregular nuclei infiltrate the lymph node diffusely, and IHC shows a positive reaction with cyclin D1 (SP4) (Figure 1I) [37]. The T/natural killer-cell lymphomas are rare forms of PTLT. They develop late after transplantation, usually in extranodal sites, and are more aggressive than B-cell neoplasms [31,43]. Peripheral T-cell lymphoma (PTCL), not specified, and hepatosplenic T-cell lymphoma belong to the most frequent types (Figure 1J and K) [44].

#### *'Mixed' PTLT*

Different morphological types of PTLT can be observed in the same patient. Overlapping forms of PTLT or cases in which histological changes are differentially advanced in various organs may be diagnosed. Reactive PH in one tonsil and a more advanced form of P-PTLT in the second or partial involvement of the tonsil by P-PTLT and partial by classic DLBCL have been described [37,45]. It should be emphasized that final diagnosis corresponds always to the more aggressive lesion.

#### *Atypical PTLT*

Atypical forms of PTLT are morphologically or immunophenotypically different from classical forms and are therefore difficult to classify. An example is PH with depletion of lymphocytes diagnosed in the lymph node of a 54-year-old after liver transplant. The node architecture was partially effaced with sinus dilatation and fibrosis. It was accompanied by a decreased number of lymphocytes, especially of T phenotype. Numerous polyclonal plasma cells were present. The course of the disease was rapidly progressive, and the patient died due to generalized infection and systemic lymphadenopathy [46].

Certain types of PTLT initially do not fit a classification and demand complementary IHC or molecular techniques as showed in one of our RTR [47]. His bone marrow biopsy revealed diffuse dense infiltration formed by two populations of lymphoid cells. The first consisted of small- to medium-sized cells with slightly irregular nuclei and the second of large polymorphic RS-like cells, with a multilobular nucleus and prominent one to two nucleoli (CD30+, EBV/LMP1+) but negative for CD15 and B-cell markers (Figure 1L and M). There was also no fibrosis and no reactive inflammatory background characteristic for HL; therefore, the generally accepted diagnostic criteria of HL were not fulfilled. Because basic

IHC stains for B lymphocytes were negative, B-cell Hodgkin-like lymphoma was also excluded, and anaplastic large-cell lymphoma was firstly diagnosed. However, the IHC stains for BOB1 and OCT2 confirmed the diagnosis of B-cell Hodgkin-like lymphoma (WHO 2001), currently DLBCL (WHO 2008).

Analysis of the literature shows that the differences between WHO 2001 and 2008 classifications are few and lead to the maintenance of the four main categories of PTLT with small changes in their terminology. Two significant changes concern Burkitt-like variant of Burkitt lymphoma and HL-like PTLT. Many of Burkitt-like lymphoma falls into the new category: unclassifiable B-cell lymphoma, with features intermediate between DLBCL and Burkitt lymphoma. HL-like PTLT, belonging to category 4 of WHO 2001 classification, is currently considered DLBCL and belongs to M-PTLT group. It may be microscopically identical to classical HL, but large RS-like cells, active forms of immunoblasts, are LCA, CD30 and B-cell marker positive and CD15 negative [31,48–50]. This change greatly impacts on the choice of treatment, which is different for each type of lymphoma. The possibility of applying anti-CD20 antibodies gives the chance of successful treatment in case of B-cell origin lymphomas.

According to the newest classification, indolent small B-cell lymphomas such as follicular lymphomas and mucosa-associated lymphoid tissue lymphomas (MALT) are not anymore considered PTLTs [31]. It remains unclear whether other non-aggressive B-cell lymphomas, such as chronic lymphocytic leukaemia/small lymphocytic lymphoma, are still recognized as PTLT.

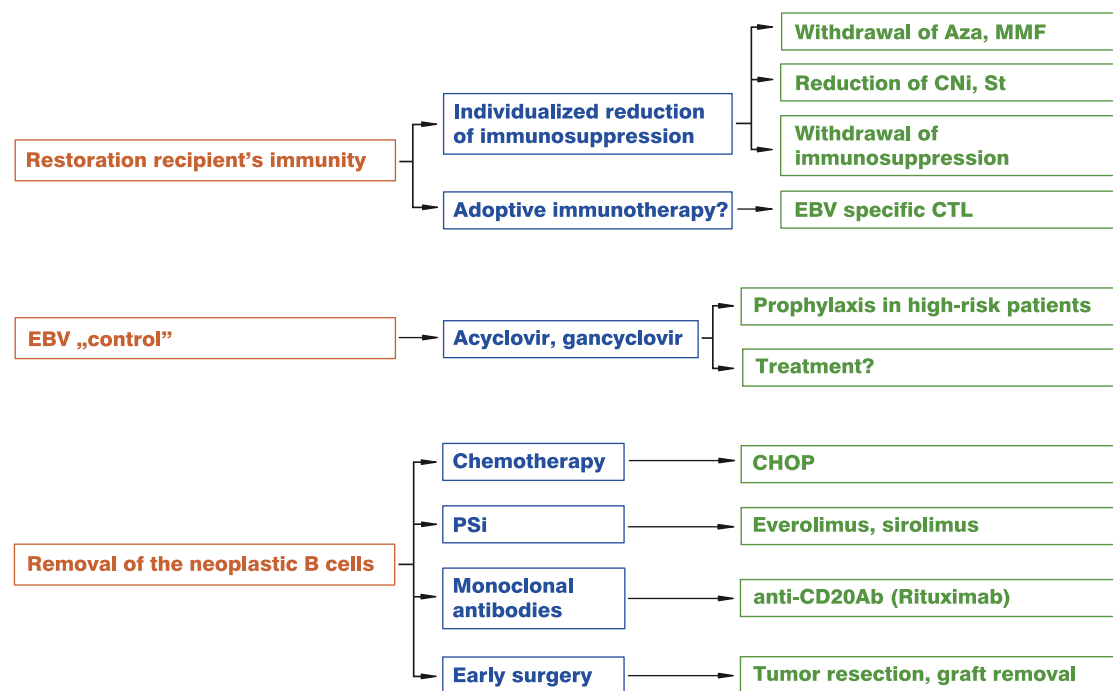
### **Treatment modalities and prognosis**

There is no consensus about the optimal treatment of PTLT. Several therapeutic approaches are currently used; the limited number of treated patients, however, precludes a standardized therapeutic algorithm. It is generally agreed that three major strategies should be applied: restoration of the recipient's immunity (to limit the EBV infection), elimination EBV and removal of neoplastic B cells [12,51,52] (Figure 2).

#### *Restoring the recipient's immunity*

**Reduction of immunosuppression.** The evidence that immunosuppression of cytotoxic T lymphocytes enables proliferation of (EBV-transformed B) cells favours reduction of IS in patients with (EBV-related PTLT). IR or even withdrawal remains the first-line treatment [53]. The withdrawal or reduction of azathioprine, mycophenolate mofetil and CNI has been reported to be effective [53]; however, the clinical outcome is highly variable and depends on the type and lineage of PTLT (Table 2). Early lesions, especially in children, usually regress with IR [19]. The majority of M-PTLTs, however, do not respond to reduction of IS, and only about 50% of polymorphic PTLTs do. One of the factors predicting poor response to IR is the presence of *BCL6* gene mutations [54]. Association with EBV predicts a better outcome when compared with EBV-negative





#### Legend:

Aza – azathioprine; CNi – calcineurin inhibitor; CHOP – cyclophosphamide, doxorubicin, vincristine and prednisone; EBV – Epstein-Barr virus; St – steroids; MMF – mycophenolate mofetil; PSi – proliferation signal inhibitor

**Fig. 2.** Proposed treatment approaches in patients with PTLD.

**Table 2.** Efficacy of various PTLD treatment options after SOT

Response rate to treatment (%)					
Immunosuppression reduction alone	Radiotherapy and/or surgery	CTL infusion	Chemotherapy	Rituximab	References
23–100 <sup>a</sup>	Variable, depending on the location and the aggressiveness	48–100	24–65 <sup>b</sup>	44–68	[19,45,55,57–62,66–78]

CTL, cytotoxic T lymphocytes.

<sup>a</sup>Depending on the type of PTLD and the association with EBV infection.

<sup>b</sup>Depending on the type of transplanted organ.

PTLD; however, subsets of EBV-negative PTLD patients have also been reported to regress with IR [55].

**Adoptive immunotherapy.** Because of the high mortality rate (ranging from 50% to 90%) when conventional IS reduction fails [56], better therapeutic approaches have been sought. One of the most promising strategies is adoptive immunotherapy with EBV-specific cytotoxic T lymphocytes (CTLs) [57]. The pathogenesis of PTLD provides a rationale for such an approach. In healthy people, virus-induced proliferation is kept under control by cell-mediated immunity elicited at the moment of primary infection. As immuno-compromised transplant recipients lack appropriate EBV-specific cell-mediated immunity, restoration can be obtained by administration of selected, ex vivo expanded, virus-specific T cells [57,58]. The requirement for genera-

tion of autologous lymphocytes results from the fact that more than 90% of PTLDs arising after SOT derive from recipient B cells. PTLDs usually arise from the donor B cells in bone marrow or haematopoietic stem cell recipients; therefore, donor lymphocyte infusions were applied in this latter population [59]. Unfortunately, in rapidly progressive forms of PTLD, the 2 or 3 months' time span required for generation of autologous CTL implies that allogeneic CTL in this setting is unrealistic.

Both auto- and allogeneic CTL administration have been shown to be safe, well-tolerated and effective methods of PTLD prophylaxis and treatment [60–62] (Table 2). Prophylactic use of CTL may be considered in a high-risk population, especially in EBV-negative heart or liver recipients who receive organs from EBV-positive donors. Although the results of the adoptive therapy are encourag-

**Table 3.** Efficacy of Rituximab in patients with PTLD after SOT

	Number of patients	Complete remission (%)	Partial remission	Overall survival (%/years)	Mean time of follow-up (months)
Rituximab					
Oertel [73]	17	52.9	5.9	56/3	24.2
Choquet [74]	43	44.2	16.2	67/1	12
Blaes [75]	11	54.5	9.1	54.5/1	10
Jain [76]	17	60	20	64/1	60
				47/3	
				35/5	
Milpied [77]	26	57.6	7.6	73/1	8
Rituximab alone or with chemotherapy					
Events [78]	59	73/3	40		

ing, only a limited number of centres applied this therapy. Several questions remain to be answered in relation to this approach, such as: the destination and survival of infused CTLs; migration to the target lesions or rather unspecific circulation; and influence of IS on these cells. For all these reasons, adoptive immunotherapy cannot yet be proposed as the first-line option in the majority of transplant recipients.

#### *EBV elimination*

**Antiviral agents.** Bearing in mind the pathophysiology of PTLD, it is unlikely that antivirals such as acyclovir or ganciclovir, even given in high doses, will be effective in the treatment of PTLD, particularly when used as a single agent. Because the EBV genome is incorporated into the infected B cell, these cells express a limited number of viral proteins that could be eliminated by these agents. Prophylaxis rather than treatment is currently indicated for high-risk patients (EBV+ donor and EBV- recipient pair), but the limited number and non-randomized character of related trials preclude definitive conclusions [63]. To date, it is believed that these agents can be useful in case of prevention of PTLD, particularly in EBV-seronegative patients and/or overimmunosuppressed recipients as well as in EBV-replicative forms of PTLD such as lymphoid hyperplasia [64].

#### *Removal of the neoplastic B cells*

**Radiotherapy and surgery.** Depending on the location and the aggressiveness of the lymphoid proliferation, surgery and radiotherapy may both be of value. These treatment modalities are recommended especially when PTLD is limited to a single lesion. Radiotherapy may be applied in patients with central nervous system involvement and in the rare cases of the extranodal natural killer/T-cell lymphomas, which represents the only form of T-cell-derived lymphoma in which radiotherapy as a primary treatment appears to yield favourable outcomes [65].

Prompt surgery of localized lesions, such as tonsillectomy or lung or liver resection (this means eventually re-transplantation), combined with reduction of IS is critical for successful treatment of PTLD [45,66] (Table 2).

**Chemotherapy.** If there is no therapeutic response to reduction of IS, chemotherapy may be an option. Overall survival improves with multidrug regimens and is more ef-

fective than single-agent chemotherapy. The most frequently used combination includes cyclophosphamide, doxorubicin, vincristine and prednisone (CHOP). In a recent retrospective analysis with a median follow-up of 8.8 years, response rate to standard CHOP reached 65%; median overall and progression-free survival was 13.9 and 42 months, respectively [67]. Some forms of the disease, especially those derived from T cells, respond badly to chemotherapy [65].

When comparing various chemotherapeutic options, one should keep in mind that most reports concern small patient cohorts presenting with a heterogeneous spectrum of PTLD subtypes and of chemotherapy regimens [68,69]. Because chemotherapy is known, particularly in immuno-compromised patients, to be associated with significant toxicity and mortality, it should be reserved for disseminated forms of PTLD that are unresponsive to other types of treatment, especially in the era of availability of anti-CD20 antibodies.

**Rituximab.** Because the majority of PTLDs originate from B lymphocytes, monoclonal antibodies directed against B-cell surface antigens have become an interesting weapon in the treatment of PTLD. Rituximab® (Roche, Basel, Switzerland) is a mouse/human chimeric anti-CD20 IgG monoclonal antibody, consisting of human constant Fc regions linked to murine variable domains [70]. Murine Fab domains of rituximab bind the CD20 antigen, a transmembrane protein located on the surface of both malignant and normal, mature B lymphocytes. Its mechanisms of action include apoptosis and complement-mediated and antibody-dependent cell-mediated cytotoxicity of the targeted lymphocytes. The resulting activation of the effector cell ends up in cellular killing of lymphoma cells [71]. Rituximab has proven to be effective and safe in numerous retrospective and prospective studies on PTLD treatment, especially when combined with chemotherapy. However, one should take into consideration that the largest trials contain only from 11 to 59 patients (Table 3) [72–78].

**Proliferation signal inhibitors.** Despite the fact that sirolimus and everolimus were found to have antiproliferative potential in PTLD-derived cell lines in vitro as well as in solid tumours in a mouse in vivo model of PTLD, one must be careful when transposing these conclusions into the treatment of human PTLD [12,79]. Indeed, despite the po-

tential of PSI in the management of PTLT, the UNOS study unexpectedly reported a 2-fold increase in PTLT in RTRs treated with sirolimus after transplantation [13,80]. It is therefore difficult to draw definitive conclusions in relation to the use of PSI in the treatment of PTLT.

## Summary

PTLT represents a serious problem after SOT. As the majority of risk factors cannot yet be influenced, prompt diagnosis and treatment are key objectives. Although the diagnostic and morphological criteria of different forms of PTLT are known, these objectives are not always reached. The 2008 WHO classification of PTLT was expected to refine the diagnosis and to increase its contribution to the treatment. Even though several changes were introduced in comparison to the former 2001 WHO classification, the judgment of the experienced pathologist still frequently allows to make a correct diagnosis. Does new classification mean better treatment? Sometimes yes, even small changes in classification may have an impact on therapy, especially in the era of modern drugs. Unfortunately, for the majority of PTLT patients, no significant impact on treatment options has arisen so far from these changes. Continuously, IS reduction remains the first-line therapy in the treatment of PTLT. As monomorphic or more aggressive forms of lymphoma do not respond well to IR alone, rituximab and/or multidrug chemotherapy should be added to the therapeutic algorithm of PTLT. In selected patients, IR along with the introduction of a PSI might be considered. High-risk populations may profit from preventive antiviral therapy. Further studies are necessary to establish the role of CTL infusion in cases of EBV-related PTLT. As the origin of PTLT is not exactly known, it is difficult to generate new treatments, and therefore large, randomized trials should be set up in order to further refine the therapeutic algorithm of PTLT in organ transplant recipients.

**Acknowledgements.** We thank Krzysztof Król for professional assistance in editing tables and figures.

**Conflict of interest statement.** None declared.

## References

- Starzl TE, Nalesnik MA, Porter KA *et al.* Reversibility of lymphomas and lymphoproliferative lesions developing under cyclosporin-steroid therapy. *Lancet* 1984; 1: 583–587
- Nalesnik MA. The diverse pathology of post-transplant lymphoproliferative disorders: the importance of a standardized approach. *Transpl Infect Dis* 2001; 3: 88–96
- Tanner JE, Alfieri C. The Epstein–Barr virus and posttransplant lymphoproliferative disease: interplay of immunosuppression, EBV, and the immune system in disease pathogenesis. *Transpl Infect Dis* 2001; 3: 60–69
- Opelz G, Dohler B. Lymphomas after solid organ transplantation: a collaborative transplant study report. *Am J Transplant* 2003; 4: 222–230
- Opelz G, Henderson R. Incidence of non-Hodgkin lymphoma in kidney and heart transplant recipients. *Lancet* 1993; 342: 1514–1516
- Leblond V, Choquet S. Lymphoproliferative disorders after liver transplantation. *J Hepatol* 2004; 40: 728–735
- Jain A, Nalesnik M, Reyes J *et al.* Posttransplant lymphoproliferative disorders in liver transplantation: a 20-year experience. *Ann Surg* 2002; 236: 429–436
- Opelz G, Naujokat C, Daniel V *et al.* Disassociation between risk of graft loss and risk of non-hodgkin lymphoma with induction agents in renal transplant recipients. *Transplantation* 2006; 81: 1227–1233
- Caillard S, Lelong C, Pessione F *et al.* Post-transplant lymphoproliferative disorders occurring after renal transplantation in adults: report of 230 cases from the French Registry. *Am J Transplant* 2006; 6: 2735–2742
- Guppy AE, Rawlings E, Madrigal JA *et al.* A quantitative assay for Epstein–Barr Virus-specific immunity shows interferon-gamma producing CD8+ T cells increase during immunosuppression reduction to treat posttransplant lymphoproliferative disease. *Transplantation* 2007; 84: 1534–1539
- Wiesner RH. A long-term comparison of tacrolimus (FK 506) versus cyclosporine in liver transplantation: a report of the United States FK506 Study Group. *Transplantation* 1998; 66: 493–499
- Krams SM, Martinez OM. Epstein–Barr virus, rapamycin, and host immune responses. *Curr Opin Organ Transplant* 2008; 13: 563–568
- Kirk AD, Cherikh WS, Ring M *et al.* Dissociation of depletion induction and posttransplant lymphoproliferative disease in kidney recipients treated with alemtuzumab. *Am J Transplant* 2007; 7: 2619–2625
- Hoshida Y, Li T, Dong Z *et al.* Lymphoproliferative disorders in renal transplant patients in Japan. *Int J Cancer* 2001; 91: 869–875
- Marshall WL, Yim C, Gustafson E *et al.* Epstein–Barr virus encodes a novel homolog of the bcl-2 oncogene that inhibits apoptosis and associates with Bax and Bak. *J Virol* 1999; 73: 5181–5185
- Caillard S, Agodoa LY, Bohen EM *et al.* Myeloma, Hodgkin disease, and lymphoid leukemia after renal transplantation: characteristics, risk factors and prognosis. *Transplantation* 2006; 81: 888–895
- Mañez R, Breinig MC, Linden P *et al.* Posttransplant lymphoproliferative disease in primary Epstein–Barr virus infection after liver transplantation: the role of cytomegalovirus disease. *J Infect Dis* 1997; 176: 1462–1467
- McLaughlin K, Wajstau S, Marotta P *et al.* Increased risk for post-transplant lymphoproliferative disease in recipients of liver transplants with hepatitis C. *Liver Transpl* 2000; 6: 570–574
- Tsao L, His ED. The clinicopathologic spectrum of posttransplantation lymphoproliferative disorders. *Arch Pathol Lab Med* 2007; 131: 1209–1218
- Bakker NA, van Imhoff GW, Verchuuren EA *et al.* HLA antigens and post renal transplant lymphoproliferative disease: HLA-B matching is critical. *Transplantation* 2005; 80: 595–599
- Opelz G, Döhler B. Impact of HLA mismatching on incidence of posttransplant non-hodgkin lymphoma after kidney transplantation. *Transplantation* 2010; 89: 567–572
- Kroes AC, van der Pijl JW, van Tol MJ *et al.* Rapid occurrence of lymphoproliferative disease after pancreas-kidney transplantation performed during acute primary Epstein–Barr virus infection. *Clin Infect Dis* 1997; 24: 339–343
- Bakker NA, van Imhoff GW, Verchuuren EA *et al.* Early onset post-transplant lymphoproliferative disease is associated with allograft localization. *Clin Transplant* 2005; 19: 327–334
- Stevens SJ, Verschuuren EA, Verkuujlen SA *et al.* Role of Epstein–Barr virus DNA load monitoring in prevention and early detection of post-transplant lymphoproliferative disease. *Leuk Lymphoma* 2007; 43: 831–840
- Gustafsson A, Levitsky V, Zou JZ *et al.* Epstein–Barr virus (EBV) load in bone marrow transplant recipients at risk to develop post-transplant lymphoproliferative disease: prophylactic infusion of EBV-specific cytotoxic T cells. *Blood* 2000; 95: 807–814
- Smets F, Latine D, Bazin H *et al.* Ratio between Epstein–Barr viral load and anti-Epstein–Barr virus specific T-cell response as a predictive marker of posttransplant lymphoproliferative disease. *Transplantation* 2002; 73: 1603–1610
- Tsai DE, Douglas L, Andreadis C *et al.* EBV PCR in the diagnosis and monitoring of posttransplant lymphoproliferative disorder:



- results of a two-arm prospective trial. *Am J Transplant* 2008; 8: 1016–1024
28. Lee TC, Savoldo B, Barshes NR *et al.* Use of cytokine polymorphisms and Epstein–Barr virus viral load to predict development of post-transplant lymphoproliferative disorder in paediatric liver transplant recipients. *Clin Transplant* 2006; 20: 389–393
  29. McAulay KA, Haque T, Crawford DH. Tumour necrosis factor gene polymorphism: a predictive factor for the development of post-transplant lymphoproliferative disease. *Br J Cancer* 2009; 101: 1019–1027
  30. Beatty PR, Krams SM, Martinez OM. Involvement of IL-10 in the autonomous growth of EBV-transformed B cell lines. *J Immunol* 1997; 158: 4045–4051
  31. Swerdlow SH, Webber SA, Chadburn A *et al.* Post-transplant lymphoproliferative disorders. *WHO Classification of Tumours of Haematopoietic and Lymphoid Tissues*. Lyon: IARC Press, 2008
  32. Katz BZ, Pahl E, Crawford SE *et al.* Case-control study of risk factors for the development of post-transplant lymphoproliferative disease in a pediatric heart transplant cohort. *Pediatr Transplant* 2007; 11: 58–65
  33. Webber SA, Naftel DC, Fricker FJ *et al.* Lymphoproliferative disorders after paediatric heart transplantation: a multi-institutional study. *Lancet* 2006; 367: 233–239
  34. Hsi ED. *Hematopathology*. Philadelphia: Churchill Livingstone, 2007
  35. Leblond V, Sutton L, Dorent R *et al.* Lymphoproliferative disorders after organ transplantation: a report of 24 cases observed in a single center. *J Clin Oncol* 1995; 13: 961–968
  36. Bustillo M, Perez MC, Otero GlzA *et al.* High grade lymphoma in a post-renal transplant patient. Description of a case and literature review. *Nephron* 2000; 84: 189–191
  37. Ziarkiewicz-Wróblewska B, Górnicka B, Suleiman W *et al.* Posttransplant lymphoproliferative disorder: morphological picture and diagnostic difficulties. *Transplant Proc* 2006; 38: 168–172
  38. Johnson LR, Nalesnik MA, Swerdlow SH. Impact of Epstein–Barr virus in monomorphic B-cell posttransplant lymphoproliferative disorders: a histogenetic study. *Am J Surg Pathol* 2006; 30: 1604–1612
  39. Baccarani U, Adani GL, Montanaro D *et al.* De novo malignancies after kidney and liver transplantations: experience on 582 consecutive cases. *Transplant Proc* 2006; 38: 1135–1137
  40. Lohrisch CA, Nevill TJ, Barnett MJ *et al.* Development of a biologically distinct EBV-related lymphoproliferative disorder following autologous bone marrow transplantation for an EBV-negative post-renal allograft Burkitt's lymphoma. *Leuk Lymphoma* 2000; 39: 195–201
  41. Knowles DM, Cesarman E, Chadburn A *et al.* Correlative morphologic and molecular genetic analysis demonstrates three distinct categories of posttransplantation lymphoproliferative disorders. *Blood* 1995; 85: 552–565
  42. Tchong WY, Said J, Hall T *et al.* Post-transplant multiple myeloma in a pediatric renal transplant patient. *Pediatr Blood Cancer* 2006; 47: 218–223
  43. Hanson MN, Morrison VA, Peterson BA *et al.* Posttransplant T-cell lymphoproliferative disorders—an aggressive, late complication of solid-organ transplantation. *Blood* 1996; 88: 3626–3633
  44. Swerdlow SH. T-cell and NK-cell posttransplantation lymphoproliferative disorders. *Am J Clin Pathol* 2007; 127: 887–895
  45. Mucha K, Foronczewicz B, Niemczyk K *et al.* Tonsil enlargement after liver transplantation in adults—reason enough for tonsillectomy? Two cases of tonsillar posttransplantation lymphoproliferative disease. *Liver Transpl* 2007; 13: 918–923
  46. Ziarkiewicz-Wróblewska B, Górnicka B, Oldakowska U *et al.* Plasmacytic hyperplasia—the early form of posttransplant lymphoproliferative disorder—with atypical morphology and clinical course in patient after liver transplantation: a case report. *Transplant Proc* 2003; 35: 2320–2322
  47. Ziarkiewicz-Wróblewska B, Górnicka B, Gieriej B *et al.* Hodgkin-like lymphoma, simulating anaplastic large cell lymphoma in the patient after renal transplantation—unusual case report and literature review. *J Pol Pathol* 2008; 59: 63–69
  48. Pitman SD, Huang Q, Zuppan CW *et al.* Hodgkin lymphoma-like posttransplant lymphoproliferative disorder (HL-like PTLD) simulates monomorphic B-cell PTLD both clinically and pathologically. *Am J Surg Pathol* 2006; 30: 470–476
  49. Schlieper G, Kurschat C, Donner A *et al.* Hodgkin disease-like post-transplantation lymphoproliferative disorder of donor origin in a renal allograft recipient. *Am J Kidney Dis* 2006; 47: e37–e41
  50. Ranganathan S, Webber S, Ahuja S *et al.* Hodgkin-like posttransplant lymphoproliferative disorder in children: does it differ from posttransplant Hodgkin lymphoma? *Pediatr Dev Pathol* 2004; 7: 348–360
  51. Gottschalk S, Heslop HE, Rooney CM. Adoptive immunotherapy for EBV-associated malignancies. *Leuk Lymphoma* 2005; 46: 1–10
  52. Everly MJ, Bloom RD, Tsai DE *et al.* Posttransplant lymphoproliferative disorder. *Ann Pharmacother* 2007; 41: 1850–1858
  53. Frey NV, Tsai DE. The management of posttransplant lymphoproliferative disorder. *Med Oncol* 2007; 24: 125–136
  54. Cesarman E, Chadburn A, Liu YF *et al.* BCL-6 gene mutations in posttransplantation lymphoproliferative disorders predict response to therapy and clinical outcome. *Blood* 1998; 92: 2294–2302
  55. Nelson BP, Nalesnik MA, Bahler DW *et al.* Epstein–Barr virus-negative posttransplant lymphoproliferative disorders: a distinct entity? *Am J Surg Pathol* 2000; 24: 375–385
  56. Taylor AL, Marcus R, Bradley JA. Post-transplant lymphoproliferative disorders (PTLD) after solid organ transplantation. *Crit Rev Oncol Hematol* 2005; 56: 155–167
  57. Merlo A, Turrini R, Dolcetti R *et al.* Adoptive cell therapy against EBV-related malignancies: a survey of clinical results. *Expert Opin Biol Ther* 2008; 8: 1265–1294
  58. Bollard CM, Gottschalk S, Leen AM *et al.* Complete responses of relapsed lymphoma following genetic modification of tumor-antigen presenting cells and T-lymphocyte transfer. *Blood* 2007; 110: 2838–2845
  59. Papadopoulos EB, Ladanyi M, Emanuel D *et al.* Infusions of donor leukocytes to treat Epstein–Barr virus-associated lymphoproliferative disorders after allogeneic bone marrow transplantation. *N Engl J Med* 1994; 330: 1185–1191
  60. Comoli P, Maccario R, Locatelli F *et al.* Treatment of EBV-related post-renal transplant lymphoproliferative disease with a tailored regimen including EBV-specific T cells. *Am J Transplant* 2005; 5: 1415–1422
  61. Sherritt MA, Bharadwaj M, Burrows JM *et al.* Reconstitution of the latent T-lymphocyte response to Epstein–Barr virus is coincident with long-term recovery from posttransplant lymphoma after adoptive immunotherapy. *Transplantation* 2003; 75: 1556–1560
  62. Haque T, Wilkie GM, Jones MM *et al.* Allogeneic cytotoxic T-cell therapy for EBV-positive posttransplantation lymphoproliferative disease: results of a phase 2 multicenter clinical trial. *Blood* 2007; 110: 1123–1131
  63. Darenkov IA, Marcarelli MA, Basadonna GP *et al.* Reduced incidence of Epstein–Barr virus-associated posttransplant lymphoproliferative disorder using preemptive antiviral therapy. *Transplantation* 1997; 64: 848–852
  64. Funch DP, Walker AM, Schneider G *et al.* Ganciclovir and acyclovir reduce the risk of post-transplant lymphoproliferative disorder in renal transplant recipients. *Am J Transplant* 2005; 5: 2894–2900
  65. Rodriguez-Abreu D, Filho VB, Zucca E. Peripheral T cell lymphomas, unspecified (or not otherwise specified): a review. *Hematol Oncol* 2008; 26: 8–20
  66. Foronczewicz B, Mucha K, Usiekiewicz J *et al.* Posttransplant lymphoproliferative disorder of the lung in a renal transplant recipient treated successfully with surgery. *Transplant Proc* 2006; 38: 173–176
  67. Choquet S, Trappe R, Leblond V *et al.* CHOP-21 for the treatment of post-transplant lymphoproliferative disorders following solid organ transplantation. *Haematologica* 2007; 92: 273–274
  68. Garrett TJ, Chadburn A, Barr ML *et al.* Posttransplantation lymphoproliferative disorders treated with cyclophosphamide-doxorubicin-vincristine-prednisone chemotherapy. *Cancer* 1993; 72: 2782–2785
  69. Taylor AL, Bowles KM, Callaghan CJ *et al.* Anthracycline-based chemotherapy as first-line treatment in adults with malignant post-transplant lymphoproliferative disorder after solid organ transplantation. *Transplantation* 2006; 82: 375–381

70. Lee JJ, Lam MS, Rosenberg A. Role of chemotherapy and rituximab for treatment of posttransplant lymphoproliferative disorder in solid organ transplantation. *Ann Pharmacother* 2007; 41: 1648–1659
71. Svoboda J, Kotloff R, Tsai DE. Management of patients with post-transplant lymphoproliferative disorder: the role of rituximab. *Transpl Int* 2006; 19: 259–269
72. Trappe R, Hinrichs C, Appel U *et al.* Treatment of PTLD with rituximab and CHOP reduces the risk of renal graft impairment after reduction of immunosuppression. *Am J Transplant* 2009; 9: 2331–2337
73. Oertel SH, Verschuuren E, Reinke P *et al.* Effect of anti- CD 20 antibody rituximab in patients with post-transplant lymphoproliferative disorder (PTLD). *Am J Transplant* 2005; 5: 2901–2906
74. Choquet S, Leblond V, Herbrecht R *et al.* Efficacy and safety of rituximab in B-cell post-transplantation lymphoproliferative disorders: results of a prospective multicenter phase 2 study. *Blood* 2006; 107: 3053–3057
75. Blaes AH, Peterson BA, Barlett N *et al.* Rituximab therapy is effective for posttransplant lymphoproliferative disorders after solid organ transplantation. *Cancer* 2005; 104: 1661–1667
76. Jain AB, Marcos A, Pokharna R *et al.* Rituximab (Chimeric anti-CD20 antibody) for posttransplant lymphoproliferative disorder after solid organ transplantation in adults: long-term experience from a single center. *Transplantation* 2005; 80: 1692–1698
77. Milpied N, Vasseur B, Parquet N *et al.* Humanized anti-CD20 monoclonal antibody (Rituximab) in post transplant B-lymphoproliferative disorder: a retrospective analysis on 32 patients. *Ann Oncol* 2000; 11: 113–116
78. Evens AM, David KA, Helenowski I *et al.* Multicenter analysis of 80 solid organ transplantation recipients with post-transplantation lymphoproliferative disease: outcomes and prognostic factors in the modern era. *J Clin Oncol* 2010; 28: 1038–1046
79. Cullis B, D'Souza R, McCullagh P *et al.* Sirolimus-induced remission of posttransplantation lymphoproliferative disorder. *Am J Kidney Dis* 2006; 47: e67–e72
80. Pascual J. Post-transplant lymphoproliferative disorder—the potential of proliferation signal inhibitors. *Nephrol Dial Transplant* 2007; 22: i27–i35

Received for publication: 30.9.09; Accepted in revised form: 6.4.10