

Original papers

QJM

Clinical studies of multiple endocrine neoplasia type 1 (MEN1)

D. TRUMP¹, B. FARREN¹, C. WOODING¹, J.T. PANG¹, G.M. BESSER², K.D. BUCHANAN³, C.R. EDWARDS⁴, D.A. HEATH⁵, C.E. JACKSON⁶, S. JANSEN⁷, K. LIPS⁸, J.P. MONSON², D. O'HALLORAN⁹, J. SAMPSON¹⁰, S.M. SHALET⁹, M.H. WHEELER¹¹, A. ZINK¹², and R.V. THAKKER¹

From the ¹MRC Molecular Endocrinology Group, MRC Clinical Sciences Centre, Royal Postgraduate Medical School, Hammersmith Hospital, London, UK, ²Department of Endocrinology, St Bartholomew's Hospital, London, UK, ³Department of Medicine, The Queen's University of Belfast, Belfast, UK, ⁴Imperial College School of Medicine, London, UK, ⁵Division of Medicine, Selly Oak Hospital, Birmingham, UK, ⁶Division of Clinical & Molecular Genetics, Henry Ford Hospital, Detroit, USA, ⁷Faculty of Medicine, University of Orange Free State, Bloemfontein, South Africa, ⁸Ziekenhuis Utrecht, Utrecht, The Netherlands, ⁹Department of Diabetes & Endocrinology, Withington Hospital, Manchester, UK, ¹⁰Institute of Medical Genetics, University Hospital of Wales, Cardiff, UK, ¹¹Department of Surgery, Cardiff Royal Infirmary, Cardiff, UK, ¹²Abteilung Innere Medizin I, Klinikum der Universität Heidelberg, Heidelberg, Germany

Received 12 June 1996

Summary

Multiple endocrine neoplasia type 1 (MEN1) is an autosomal dominant disorder characterized by the combined occurrence of parathyroid, pancreatic islet and anterior pituitary tumours. To facilitate a screening programme for MEN1, we investigated 709 people (364 males and 345 females, age range 1–84 years) from 62 MEN1 families, and 36 non-familial MEN1 patients. Of those investigated, 220 (95 males and 125 females, age range 8–79 years) suffered from MEN1. Parathyroid, pancreatic and pituitary tumours occurred in 95%, 41% and 30% of the patients, respectively. Parathyroid tumours were the first manifestation of MEN1 in 87% of

patients, and amongst the pituitary and pancreatic tumours, somatotrophinomas and gastrinomas were more common in patients above the age of 40 years, whilst insulinomas occurred more frequently in patients below the age of 40 years. Biochemical screening indicated that the penetrance of MEN1 by the ages of 20, 35 and 50 years was 43%, 85% and 94%, respectively, and that the development of MEN1 was confined to first-degree relatives in 91% of patients and to second-degree relatives in 9% of patients. These findings have helped to define a proposed screening programme for MEN1.

Address correspondence to Professor R.V. Thakker, MRC Molecular Endocrinology Group, Collier Building, MRC Clinical Sciences Centre, Royal Postgraduate Medical School, Hammersmith Hospital, Du Cane Road, London W12 0NN

© Oxford University Press 1996

Introduction

Multiple endocrine neoplasia type 1 (MEN1)¹ is an autosomal dominant disorder characterized by the combined occurrence of tumours of the parathyroid glands, the pancreatic islet cells and the anterior pituitary.^{1–5} Parathyroid tumours are the most common manifestation of the disorder^{6–10} but patients may also suffer from pancreatic tumours which secrete gastrin, insulin, pancreatic polypeptide (PP), or glucagon, and anterior pituitary tumours which usually secrete prolactin, growth hormone (GH) or adrenocorticotrophin (ACTH).^{4–10} Adrenal cortical tumours,¹¹ carcinoid tumours,¹² lipomatous tumours⁷ and the McCune-Albright syndrome¹³ have also been observed in association with MEN1. The disease may arise in families, and an autosomal dominant inheritance has been established.^{2,3} Children of an affected individual are thus at a 50% risk of inheriting the mutant gene and of potentially developing these endocrine tumours. Earlier detection of these tumours by screening may help to reduce the morbidity and mortality in this high-risk population. However, screening for MEN1 is difficult as the combination of affected glands may differ in members of the same family.¹ In addition, the age-related penetrance (the proportion of gene carriers who have manifested symptoms or signs of the disease by a given age) has not been established. We used clinical and biochemical methods to investigate the members of 98 families with MEN1, aiming to characterize further its manifestations, to determine its age-related penetrance, and to define a suitable screening strategy.

Methods

Patients

Ninety-eight unrelated MEN1 patients were studied and their detailed family medical histories obtained. Patients and their family members were assessed for present and past manifestations of MEN1: polyuria, polydipsia, constipation, malaise, bone pains or nephrolithiasis suggesting hypercalcaemia, indigestion, recurrent peptic ulceration, hypoglycaemia, neuroglycopenia or diarrhoea suggesting the presence of a pancreatic islet-cell tumour, amenorrhoea, galactorrhoea, impotence or weight changes suggesting the presence of a pituitary tumour and a detailed questionnaire (Appendix I) completed. The medical records of deceased family members and of the known affected individuals were also examined.

Biochemical measurements

Venous blood samples were obtained from patients and family members, and serum calcium, albumin,

creatinine and alkaline phosphatase were determined using a multi-channel autoanalyser.¹⁴ Serum calcium concentrations were corrected to an albumin of 41 g/l¹⁴ (normal range 2.25–2.55 mmol/l). Serum prolactin was also determined in all individuals using the Serono immunoradiometric assay¹⁵ (normal range <500 mIU/l). Estimations of other anterior pituitary hormones, gastrointestinal hormones, serum insulin and glucose were made if indicated from the clinical history and examination.

Phenotype allocation

Individuals were taken to be affected if they had evidence of two or more MEN1-associated tumours, or if they were the relative of an affected individual and had evidence of one MEN1 tumour as determined by the following characteristics: (i) persistent hypercalcaemia (corrected calcium >2.55 mmol/l); (ii) persistent hyperprolactinaemia (defined by a serum prolactin of >800 mIU/l, so as to exclude stress-related hyperprolactinaemia) occurring in the absence of a known cause, such as pregnancy, hypothyroidism, or drugs, e.g. phenothiazines, together with other radiological or surgical evidence of a pituitary tumour; (iii) biochemical and/or radiological abnormalities demonstrating acromegaly or Cushing's disease (iv) biochemical and/or radiological abnormalities demonstrating a gastrinoma, insulinoma, PPoma, VIPoma or glucagonoma.

Kinship coefficient

Individuals were designated a kinship coefficient (KC) as an index of their proximity to affected members.⁷ KCs were assigned prior to biochemical screening, and known affected individuals were designated a KC of 1.0; first-degree unaffected relatives, *i.e.* siblings and children of an affected individual, were designated a KC of 0.5; and second-degree unaffected relatives, *i.e.* grandchildren, nephews and nieces, were assigned a KC of 0.25. Spouses, who were also screened for MEN1 to ensure that they were unaffected and therefore not transmitting MEN1 to their children, were assigned a KC of 0.

Data analysis

The clinical and biochemical information for each individual was entered into a database using the DATAEASE computer program on an IBM PC, and statistical analysis was performed with the MINITAB package. The incidence of insulinomas by age and its correlation with the number of mutations required for its development was assessed by plotting the proportion of patients not yet diagnosed (S) at a

given age against the age (t), as previously described for the development of retinoblastoma and Knudson's two-hit hypothesis.¹⁶ Thus, the incidence by age of tumours that develop after a single mutation, e.g. familial forms, would conform to a first-order equation $\log_{10} S = a - kt$, whereas that of tumours developing as a result of two mutations, e.g. the sporadic forms, would conform to a second-order equation $\log_{10} S = a - kt^2$. The ages at which insulinomas developed in familial MEN1 patients and sporadic non-MEN1 patients were determined from our study and a previously reported study,¹⁷ respectively, and used for these calculations.

Results

Details of patients and families

Clinical and biochemical results were obtained from 745 individuals (380 males and 365 females) aged 1–84 years, of whom 220 (95 males and 125 females) were affected with MEN1, and 525 (283 males and 242 females) were unaffected (Table 1). In the unaffected group, 364 (204 males and 160 females) were siblings and 161 (79 males and 82 females) were spouses. Spouses were included in the analysis to ensure that none was affected with MEN1. Of the 220 affected individuals, 184 were family members from 62 families (Figure 1 and Appendix II) and 36 individuals were sporadic cases (Table 1) in whom no family history of MEN1 could be established; 118 of the 184 familial MEN1 patients presented with symptoms and the remaining 66 individuals, who were asymptomatic, were detected by biochemical screening. The mean ages of the affected ($\mu \pm \sigma = 33.3 \pm 15.5$ years), the total unaffected (33.9 ± 17.9 years) and the unaffected sibling (28.9 ± 16.5 years) groups did not differ significantly (Table 1). However, the mean age of the screened asymptomatic group (24.8 ± 11.6 years) was significantly ($p < 0.001$) lower than that of the symptomatic (36.4 ± 15.7 years) and sporadic (37.7 ± 15.6)

groups, thereby demonstrating the value of biochemical screening in the earlier detection of MEN1. The male to female ratio of the affected group (M:F = 95:125) differed significantly ($p < 0.05$) from the whole, which may reflect a higher penetrance of MEN1 in females, and it is of interest that the two individuals in whom non-penetrance was observed above the age of 55 years (see below) were both males.

Analysis of tumour types

The 220 affected individuals had a total of 384 tumours (Appendix II); 120 of these patients had two or more tumours and the remaining 100 patients had one tumour. The distribution of the tumours is shown in Figure 2. Hypercalcaemia was found in 208 (94.5%) patients; the diagnosis of primary hyperparathyroidism had been confirmed in 87% of these patients by either parathyroidectomy (71%) or persistently raised serum PTH concentrations (16%). In the remaining 5.5% ($n = 12$) of MEN1 patients (2.2/91 I.1, 4/89 V.6, 8.7/87 II.1, 11/89 II.7, 11.2/90 III.4, 13.3/90 III.24, 13.3/92 III.1, 16/90 IV.20, 19/89 III.4, 19.2/92 II.1, S15, S19, Appendix II) who had no evidence of parathyroid tumours, five patients (4 males, 1 female) had a pancreatic tumour (2 insulinomas, 1 combined gastrinoma-glucagonoma, 2 non-secreting), six patients (2 males, 4 females) had a pituitary tumour (5 prolactinomas, 1 somatotrophinoma) and one patient (13.3/92 III.1, Appendix II) had an insulinoma and a non-functioning pituitary tumour. Of these 12 patients, eight were under 30 years of age and thus may develop parathyroid tumours which represent the most common manifestation of MEN1 (Figures 1 and 3) and which occurred as the sole endocrinopathy in 42.3% of MEN1 patients. Pancreatic tumours occurred in 40% of patients, with gastrinomas being the most common (63%) and with insulinomas being the next most common (27%) tumours; three of these tumours secreted both gastrin and insulin, and two of the

Table 1 Ages and sex ratios of MEN1 patients and families ($n = 745$)

| Age ($\mu \pm \sigma$) (years) M:F | Affected ($n = 220$) | | Unaffected ($n = 525$) | | |
|--|------------------------|-----------------|--------------------------|-----------------|-----------------|
| | 33.3 \pm 15.5 | | 33.9 \pm 17.9 | | |
| | 95:125 | | 283:242 | | |
| | Familial | | Sporadic | Siblings | Spouses |
| | Asymptomatic | Symptomatic | | | |
| n | 66 | 118 | 36 | 364 | 161 |
| Age ($\mu \pm \sigma$) (years) | 24.8 \pm 11.6 | 36.4 \pm 15.7 | 37.7 \pm 15.6 | 28.9 \pm 16.5 | 46.6 \pm 14.5 |
| M:F | 26:40 | 53:65 | 16:20 | 204:160 | 79:82 |

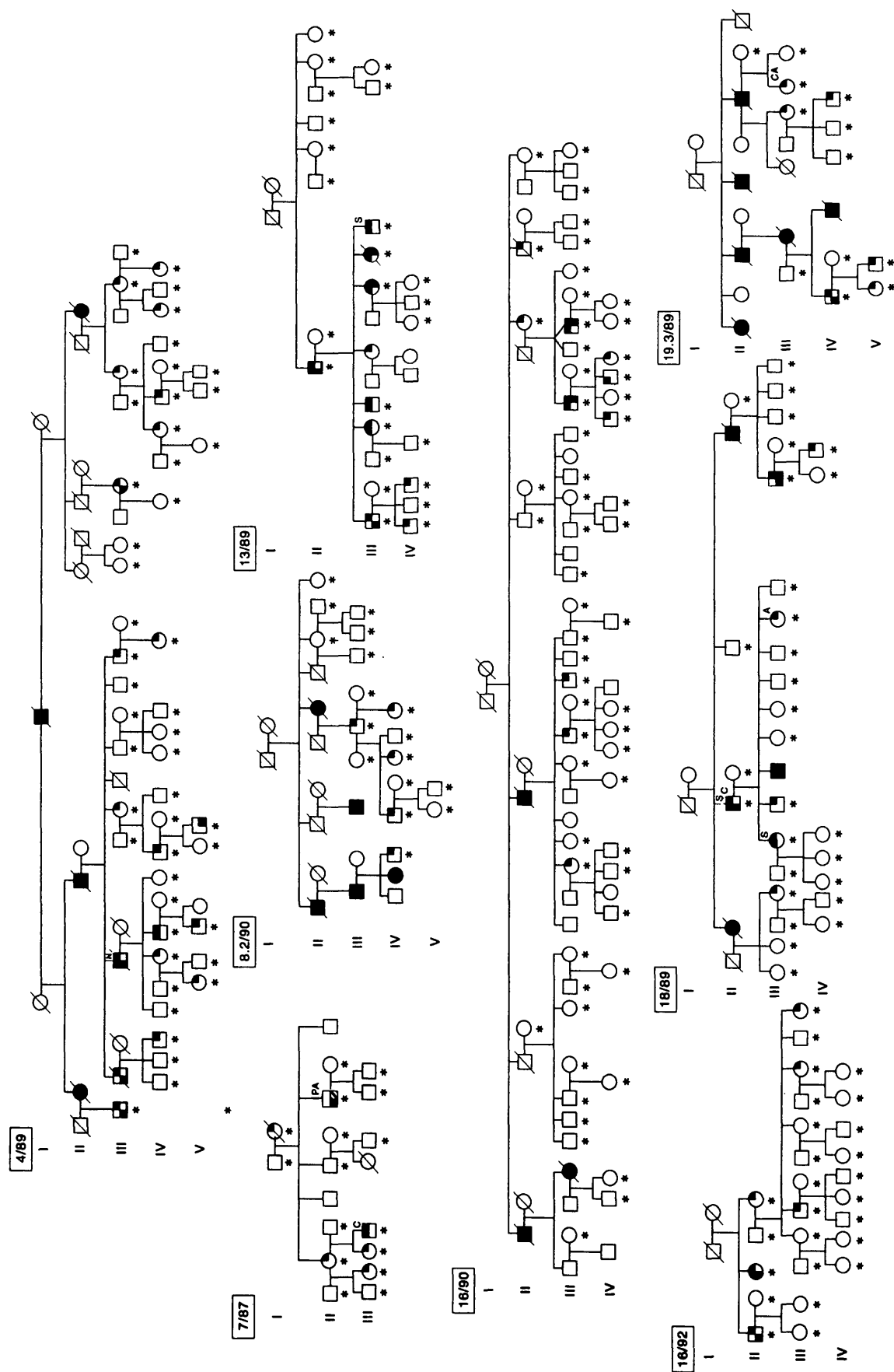


Figure 1. Eight MEN1 pedigrees in whom there were five or more affected members are shown. Males are indicated by a square, females by a circle and deceased family members are indicated by a diagonal line across their symbol. The identification code of each family is as indicated in Appendix II. The individuals from whom blood samples were obtained are denoted by (*). The clinical details of the six affected members of family 7/87, the nine affected members of family 18/89, 5/13 affected members of family 16/90, eight of the 24 affected members of family 4/89 have been previously reported.^{17,25} The presence of tumours is indicated as follows: Anterior pituitary tumour (for details of: prolactinoma; somatotrophinoma; corticotrophinoma; non-functioning Insulinoma, Pancreatic tumour: type unknown, Carcinoid tumour, Adrenocortical tumour. Unaffected members are represented by , and those affected members who were unavailable for this study but were reported to suffer from MEN1 tumours by their relatives are represented as .

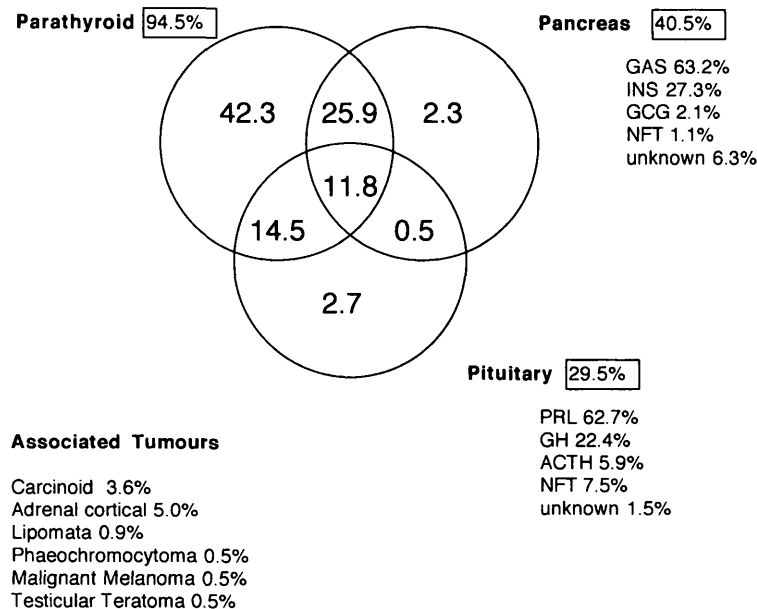


Figure 2. Schematic representation of the distribution of 384 MEN1 tumours in the 220 MEN1 patients. The proportions of patients in whom parathyroid, pancreatic or pituitary tumours occurred are shown in the respective boxes, for example, 94.5% of the patients had a parathyroid tumour. The Venn diagram indicates the proportions of patients with each combination of tumours, for example, 33.7% (25.9% + 11.8%) of the patients had both a parathyroid and pancreatic tumour, whereas 2.3% of the patients had a pancreatic tumour only. The hormones secreted by each of these tumours are indicated: GAS, gastrin; INS, insulin; GCG, glucagon; NFT, non-functioning tumour; PRL, prolactin; GH, growth hormone; ACTH, adrenocorticotrophic hormone. Thus, parathyroid tumours represent the most common form of MEN1 tumours.

gastrinomas also secreted glucagon. The gastrinomas were confirmed surgically or radiologically in 80% of patients and the insulinomas were confirmed surgically in 96% of patients. Pituitary tumours occurred in 30% of patients. Over three-fifths were prolactinomas and the majority of the remainder were somatotrophinomas. Ninety-six percent of the prolactinomas and 93% of the somatotrophinomas were confirmed by surgery or radiology. Additional endocrine tumours were found in 21 individuals: carcinoid tumours in eight patients (4%), adrenal

cortical tumours in 12 patients (5%), lipomata in two patients (1%), a pheochromocytoma in one patient, and a testicular teratoma and a malignant melanoma in one patient. Of these 21 patients, one patient had both a non-functioning adrenal tumour and a pheochromocytoma, one patient had both an adrenal tumour and a carcinoid tumour, and the patient with a testicular teratoma and a malignant melanoma also had a carcinoid tumour.

The combinations of these MEN1 tumours differed in members of the same family, and this variable expression of MEN1 is illustrated by family 13/89 (Figure 1). In this family the father (generation II) suffered from parathyroid tumours, an insulinoma and a prolactinoma. All of his six affected children

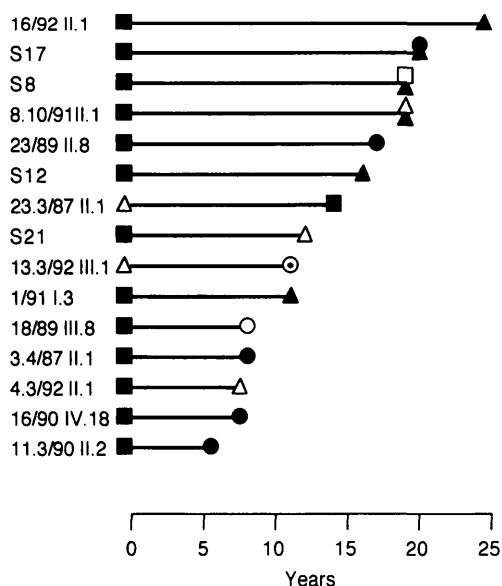


Figure 3. Order of tumour development in 15 MEN1 patients. The time interval between the occurrence of the first tumour (time=0 years) and subsequent tumours in each patient, who is identified by the family and individual number (Appendix II), are shown. The tumours are: ■, PTH-secreting; ▲, gastrinoma; △, insulinoma; ●, prolactinoma; ○, corticotrophinoma; □, somatotrophinoma; ⊙, non-functioning pituitary tumour. Parathyroid tumours were the first manifestation of MEN1 in 13/15 patients, and in the remaining two patients, insulinomas represented the first manifestation of MEN1. The time interval for the occurrence of the subsequent tumours ranged from 6 to 24 years and there was no correlation between the time interval and the tumour types.

(generation III) suffered from parathyroid tumours but three daughters and one son had a prolactinoma whilst another son had a somatotrophinoma. In addition, one of his daughters suffered from an insulinoma whereas a son suffered from a gastrinoma. The interval between the appearance of each tumour in a patient also varied and ranged from 6 to 24 years in the 15 MEN1 patients in whom the order of tumour development could be unequivocally established (Figure 3).

Age-related penetrance and a mutational model

The ages at which the first manifestation of an MEN1 tumour occurred could be established in 219/220

MEN1 patients and ranged from 8 to 79 years. Sixty-six of these 219 patients were asymptomatic and had been detected by biochemical screening; the remaining 153 patients had presented with symptoms. The cumulative percentages of patients who had developed MEN1 in the symptomatic group at the ages of 20, 35 and 50 years were 18%, 52% and 78%, respectively, whereas in the biochemically-screened asymptomatic group, these respective cumulative percentages were increased to 43%, 85% and 94% (Figure 4a). Thus biochemical screening detected an earlier onset ($p < 0.001$) of MEN1 in all age groups.

The age-related penetrance (the proportion of gene carriers who have manifested symptoms or signs of the disease by a given age) of MEN1 was determined

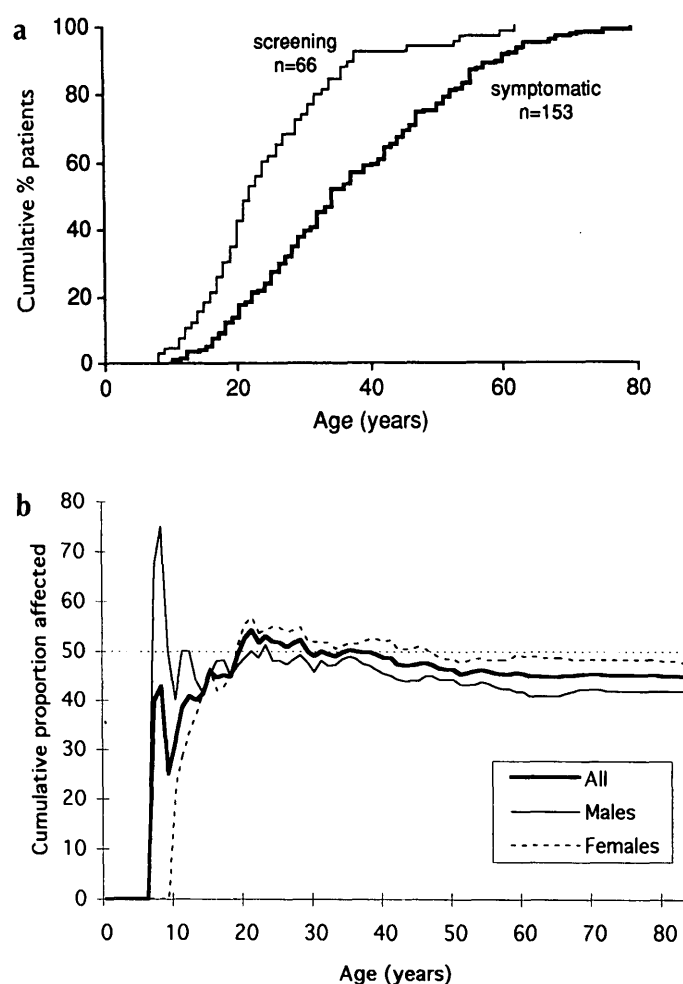


Figure 4. Age-related onset and penetrance of MEN1. The ages and cumulative percentages of patients (a) who had developed the first manifestation of MEN1 are shown for the symptomatic presenting group (bold line) and for the asymptomatic, biochemically detected group (faint line). The ratio of the affected individuals (■) to the total number (■ affected + □ unaffected) of individuals at different ages (b) represents an estimation of the penetrance of MEN1. The initial sharp rise in this ratio below the age of 10 years that was particularly observed in males is due to five individuals (4 males and 1 female) who are 4/89 V.3. 7/87 III.4, 13.3/92 III.1, 18/89 IV.7, and S7 (Appendix II). All four males had primary hyperparathyroidism, one also had an insulinoma and another also suffered from a corticotrophinoma; the one female, who suffered from an insulinoma and a non-functioning pituitary tumour, did not have hypercalcaemia. However, the expected 50% ratio for an autosomal dominant disorder was attained by 21 years and maintained until 40 years of age, after which it decreased to 42%. This decrease was attributed to the higher mortality associated with the gastrinomas and somatotrophinomas (Figure 5). These results indicate that MEN1 has a high penetrance by the age of 21 years.

and the results from 288 offspring (129 affected, 159 unaffected) of 101 affected parents are shown in Figure 4b. The 1:1 ratio for affected to unaffected individuals that would be expected for an autosomal dominant disorder was first achieved at 21 years, thereby indicating a near complete penetrance for MEN1 by this age. Interestingly, the ages of conversion from an unaffected to affected phenotype that were established for two individuals, 19/92 III.4 and 16/90 IV.20 (Appendix II), who had undergone annual biochemical screening from their early teens were found to be 20 and 21 years respectively; individual 19/92 III.4 developed primary hyperparathyroidism and underwent parathyroidectomy and individual 16/90 IV.20 developed a microprolactinoma and was treated with bromocriptine. However, the penetrance of MEN1 above the age of 21 years was not complete, as two male obligate carriers (the father of the affected male 7.2/89 III.1 and the father of the affected female 3/92 III.2, Appendix II) had no clinical or biochemical manifestations of MEN1 by the ages of 52 years and 53 years, respectively. Thus, 2/162 carriers remain unaffected, thereby indicating a 98.8% penetrance of the MEN1 gene by the age of 53 years.

The decline observed above the age of 40 years in the ratio of affected individuals to the total number of individuals could result either from an increased number of unaffected individuals or a decreased number of affected individuals. The former possibility is clearly implausible, and the latter possibility, which may be due to a greater death rate in MEN1 patients

above the age of 40 years was indirectly assessed by investigating the age distribution (Figure 5) of the 384 tumours (Figure 1) in the affected individuals. The results revealed that above the age of 40 years, there was a higher occurrence of gastrinomas and somatotrophinomas, which would be associated with a higher mortality. Such differences were not observed for the occurrence of parathyroid tumours and prolactinomas in the two age groups (Figure 5) but insulinomas were found to occur more frequently in the younger age group, and this may partly contribute to the decline in the ratio of affected individuals to the total between the ages of 13 and 17 years (Figure 4b).

This significantly ($p < 0.01$) higher occurrence of insulinomas in the younger age group (Figure 5) may also be due to their neuroglycopenic presentation, which would result in their earlier detection when compared with other MEN1 tumours. A further analysis of the ages at which the 26 MEN1 insulinomas occurred and their comparison to the previously reported ages at which 57 non-MEN1 insulinomas¹⁷ occurred helped to estimate the number of mutations which were likely to be required for their development. The mean age (\pm SD) at which insulinomas were detected in the MEN1 patients (28.86 ± 19.09) was significantly ($p < 0.04$) lower than that in the non-MEN1 patients (45.98 ± 16.96). An analysis of the age of incidence for MEN1 insulinomas and for non-MEN1 insulinomas gave similar results to those reported for bilateral and unilateral retinoblastomas,¹⁶ respect-

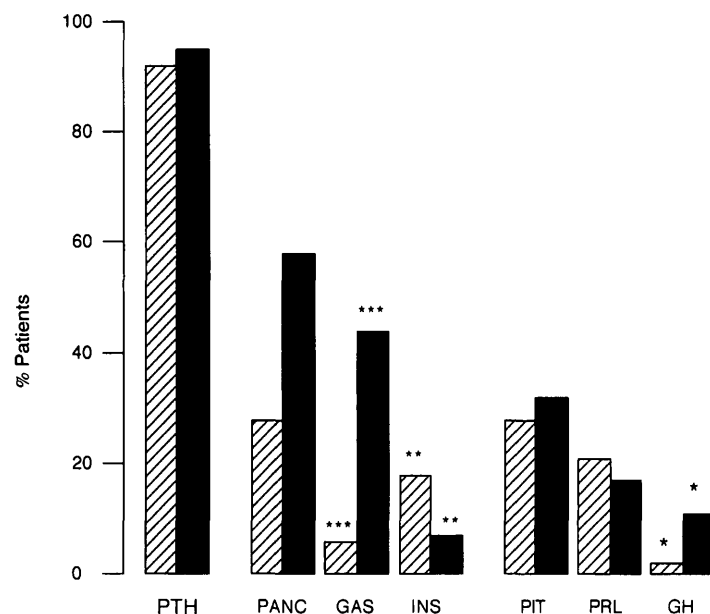


Figure 5. Development of 384 MEN1 tumours below (▨) and above (■) 40 years of age in 220 affected individuals (Figure 1). The proportion of patients developing parathyroid tumours (PTH), all pancreatic tumours (PANC), gastrinomas (GAS), insulinomas (INS), all pituitary tumours (PIT), prolactinomas (PRL) and somatotrophinomas (GH) is shown for each group. Significant differences (* $p < 0.05$, ** $p < 0.01$ and *** $p < 0.0001$) are indicated.

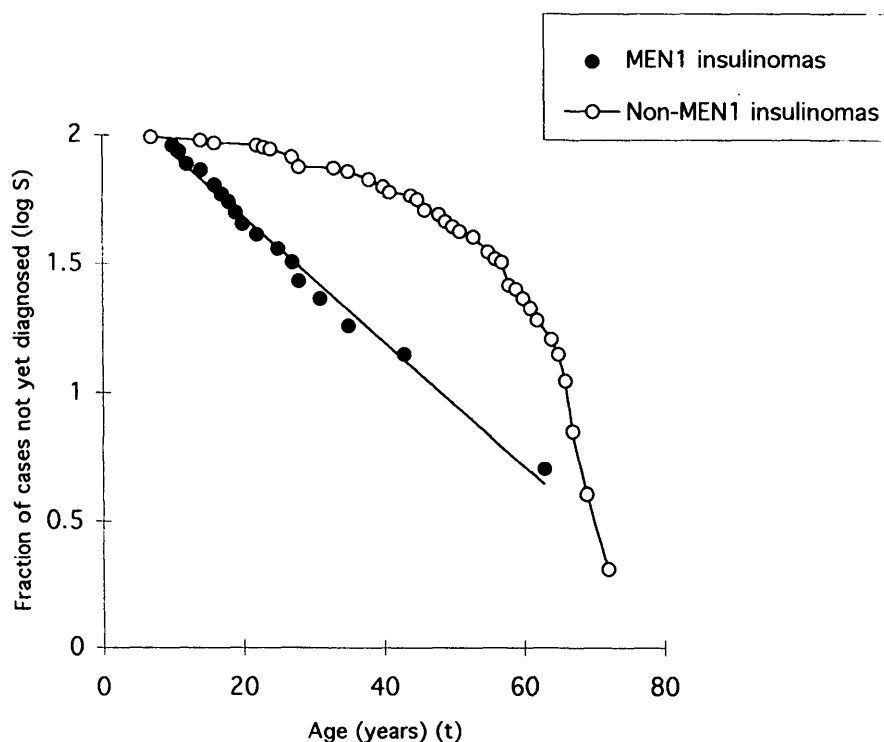


Figure 6. Age incidence of insulinoma in 26 familial MEN1 patients (●) and in 57 sporadic non-MEN1 patients¹⁷ (○), based on the Knudson retinoblastoma model.¹⁶ The age is plotted on the x axis and the corresponding $\log_{10}S$, which is the proportion of cases not yet diagnosed, is plotted on the y axis. The incidence by age of the familial MEN1 insulinomas ($n=26$) is best described by the first-order equation $y=0.024x+2.169$ ($r=0.994$), whereas that of the sporadic non-MEN1 insulinomas is best described by the second-order equation $y=-0.0004653x^2$ rather than a first-order equation ($p<0.001$). These results indicate that a single mutation is likely to be associated with the development of MEN1 insulinomas whereas two mutations are likely to be involved in the development of sporadic non-MEN1 insulinomas.

ively (Figure 6). Thus, the relationship between $\log_{10}S$ (the proportion of cases not yet diagnosed) and t (the age) was best described by a first-order equation for the MEN1 insulinomas and by a second-order equation for the non-MEN1 insulinomas. This situation is analogous to that for the retinoblastoma model in which two recessive mutations of a tumour suppressor gene are involved in oncogenesis; thus, MEN1 insulinomas require a single mutation, whereas non-MEN1 insulinomas require additional mutations.

Hypercalcaemia and coefficient of kinship

Parathyroid tumours are the most common and usually the first manifestation of MEN1 (Figures 2 and 3) and the detection of hypercalcaemia in members of MEN1 families represents a useful biochemical screening investigation. In order to assess which relatives of an MEN1 patient should be screened, we analysed the pre-treatment corrected serum calcium concentrations from 602 individuals according to their coefficient of kinship, KC (Figure 7). The corrected serum calcium concentrations for those individuals without previous parathyroid surgery (mean \pm SD) in the KC 1.0 group

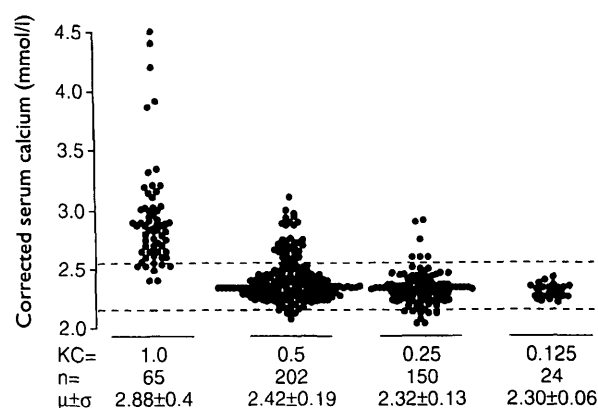


Figure 7. Corrected serum calcium concentrations by coefficient of kinship (KC). The normal range (2.20–2.55 mmol/l) for the corrected serum calcium is indicated by the broken lines. The mean corrected serum calcium concentrations and standard deviation (mean \pm SD) for each of the KC groups 1.0, 0.5, 0.25, and 0.125 are shown. Data from KC group 0, i.e. the spouses ($n=161$, mean \pm SD = 2.31 ± 0.09) are not shown. Hypercalcaemia was detected in only the first- (KC=0.5) and second- (KC=0.25) degree relatives of affected individuals (KC=1.0).

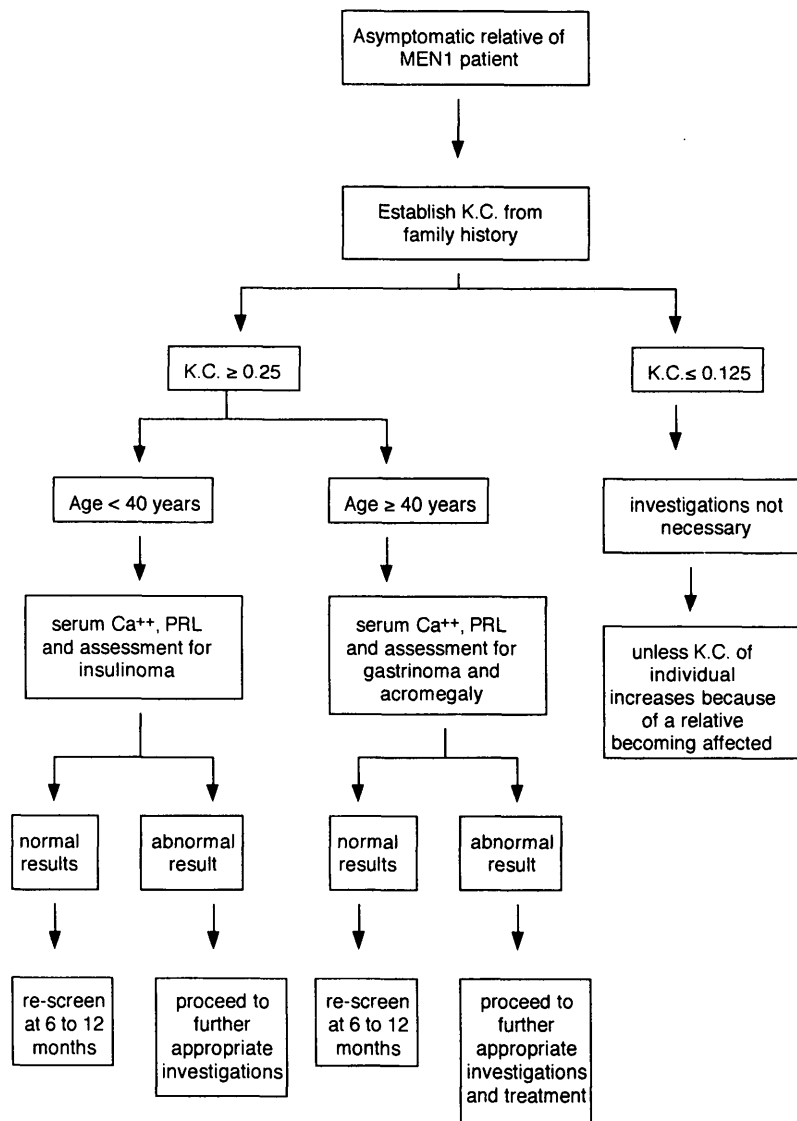


Figure 8. Screening protocol for an asymptomatic relative of a patient with MEN1. The relative should first have undergone a detailed clinical evaluation for MEN1-associated tumours to establish that the individual is asymptomatic. Relatives who are symptomatic should proceed to appropriate investigations and management. The KC refers to the coefficient of kinship (Figure 7), and helps to identify those members at a higher risk of developing MEN1. The MEN1 gene is located on chromosome 11q13^{25,26} and the molecular genetic markers²⁴ which are not yet widely available, will help to identify those individuals with an affected haplotype²⁷ who have the greater likelihood of developing MEN1 tumours. Individuals identified either by KC or molecular genetic analysis as at higher risk should be entered into the screening programme as outlined.

(2.88 ± 0.40 , $n=65$), was significantly ($p<0.001$) higher than in the KC 0.5 (2.42 ± 0.19 , $n=202$), KC 0.25 (2.32 ± 0.13 , $n=150$) and KC 0.125 (2.30 ± 0.06 , $n=24$) groups. In addition, hypercalcaemia was observed in 91%, 19%, and 4% of the individuals in KC 1.0, 0.5 and 0.25 groups, respectively, but in 0% of the KC 0.125 group. These results indicate that hypercalcaemia is a useful investigation for the biochemical screening of MEN1 which has a high penetrance, and that such screening could be restricted to the first (KC=0.5) and second (KC=0.25) degree relatives of affected individuals.

Discussion

Our extensive study of 220 MEN1 patients has helped to define the manifestations and age-related penetrance of this autosomal dominant disorder. Parathyroid tumours were the first and most common of the lesions to develop, occurring in 87% and 94% of patients, respectively (Figures 2 and 3). In addition, pancreatic islet-cell tumours and anterior pituitary adenomas occurred in 40% and 30% of patients, respectively, with gastrinomas and prolactinomas representing the majority of

tumours in each of these respective categories. The occurrence of carcinoid tumours, adrenal cortical tumours and lipomata was 4%, 5%, and 1% in the patients, respectively. Our results, which represent the largest study of MEN1 patients and are comparable to those of previous reports,^{1-4, 6-9, 18-20} also demonstrated the highly variable expression of MEN1. Thus, although the majority of patients had parathyroid tumours, the types of subsequent tumours originating from the pancreatic islets and the anterior pituitary showed considerable inter- and intra-familial variation (Figure 1). In addition, the time interval between the development of the first and subsequent tumours in a patient also varied considerably from 6 to 24 years (Figure 3). Our results are the first to demonstrate that gastrinomas and somatotrophinomas occur more frequently in MEN1 patients who are above the age of 40 years, that insulinomas occur more frequently in MEN1 patients who are below the age of 40 years (Figure 5), and that the genetic aetiology of MEN1 insulinomas (Figure 6) is consistent with Knudson's two-hit hypothesis.^{16,21-23} In addition, our defining of the age-related onset for MEN1 tumours, which were confined to the first ($KC=0.5$) and second ($KC=0.25$) degree relatives (Figure 7), helped in the estimation of residual risks (i.e. 100% — [age-related onset %], Figure 4a), for these individuals. Thus, the residual risks for developing MEN1 tumours in such unaffected relatives who are biochemically normal are estimated to be 57%, 15% and 6% at the ages of 20, 35, and 50 years, respectively.

These results have facilitated the development of a proposed screening programme to help in the management of members from MEN1 families (Figure 8). Screening should be initiated before 8 years of age, as affected children (III.4 from family 7/87 and IV.7 from family 18/89, Appendix II) have been observed by this age. Biochemical screening should be undertaken in those with a $KC \geq 0.25$. In addition, the use of molecular genetic markers that are close to the MEN1 gene²⁴ which is located on chromosome 11q13^{25,26} will help further to identify those individuals who have inherited the affected haplotype²⁷ and are therefore at an increased risk of developing the disease. Serum calcium and prolactin concentrations should be determined in all such asymptomatic individuals, as parathyroid tumours or prolactinomas occur alone or in combination in 97.7% of MEN1 patients. In addition, further clinical and biochemical assessments should be particularly performed for insulinomas in individuals below the age of 40 years and for gastrinomas and somatotrophinomas in individuals above the age of 40 years. Family members who remain asymptomatic and biochemically normal should be rescreened at 6- to

12-monthly intervals until the age of 50 years, by which age the disease will have developed in 94% of individuals at risk. An application of this proposed screening protocol which has been derived from our comprehensive analysis of MEN1 patients will help in the earlier detection of MEN1 tumours and thereby appropriate management of patients with this inherited endocrine disorder.

Acknowledgements

We are grateful to: The Medical Research Council (MRC), UK for support; to D.A. Anderson, P. Bouloux, D.P. Brenton, R.A. Norum and J.R.W. Yates for access to families, and to S. Goodburn, D. Ridout and J. Swinton for useful discussion. DT was an MRC Training Fellow and JTP was an MRC PhD student.

References

1. Thakker RV, Ponder BAJ. Multiple endocrine neoplasia. In: Sheppard MC, ed. *Clinical Endocrinology and Metabolism*, vol. 2. London, Bailliere Tindall, 1988: 1031-67.
2. Wermer P. Genetic aspects of adenomatosis of endocrine glands. *Am J Med* 1954; **16**:362-3.
3. Ballard HS, Frame B, Hartstock RJ. Familial multiple endocrine adenoma-peptic ulcer complex. *Medicine* 1964; **43**:481-515.
4. Lips CM, Vasen HFA, Lamers CBHW. Multiple endocrine neoplasia syndromes. *CRC Crit Rev Oncol Haematol* 1984; **2**:117-84.
5. Calender A, Giraud S, Cougard P, Chanson P, Lenoir G, Murat A, Hamon P, Proye C. Multiple endocrine neoplasia type 1 in France: clinical and genetic studies. *J Intern Med* 1995; **238**:263-8.
6. Benson L, Ljunghall S, Åkerström G, Oberg K. Hyperparathyroidism presenting as the first lesion in multiple endocrine neoplasia type 1. *Am J Med* 1987; **82**:731-7.
7. Marx SJ, Vinik AI, Santen RJ, Floyd JC, Mills JL, Green J. Multiple endocrine neoplasia type 1: Assessment of laboratory tests to screen for the gene in a large kindred. *Medicine* 1986; **65**(4):226-41.
8. Majewski JT, Wilson SD. The MEA-1 syndrome: an all or none phenomenon. *Surgery* 1979; **86**:474-84.
9. Eberle F, Grun R. Multiple endocrine neoplasia type 1 (MEN1). *Erbeg Inn Med Kinderheilkd* 1981; **46**:76-149.
10. Marx SJ, Spiegel AM, Levine MA, Rizzoli RE, Lasker RD, Santora AC, Downs RW Jr, Aurbach GD. Familial hypocalciuric hypercalcaemia; the relation to primary parathyroid hyperplasia. *N Engl J Med* 1982; **307**:416-26.
11. Skogseid B, Larsson C, Lindgren P-G, Kvanta E, Rastad J, Theodorsson E, Wide L, Wilander E, Oberg K. Clinical and genetic features of adrenocortical lesions in multiple endocrine neoplasia type 1. *J Clin Endocrinol Metab* 1992; **75**(1):76-81.
12. Duh Q-Y, Hybarger CP, Geist R, Gamsu G, Goodman PC, Gooding GAW, Clark OH. Carcinoids associated with

- multiple endocrine neoplasia syndromes. *Am J Surg* 1987; **154**:142–8.
13. O'Halloran DJO, Shalet SM. A family pedigree exhibiting features of both multiple endocrine neoplasia type 1 and McCune-Albright Syndromes. *J Clin Endocrinol Metab* 1994; **78**(3):523–5.
 14. Thakker RV, Fraher LJ, Adami S, Karmali R, O'Riordan JL. Circulating concentrations of 1,25-dihydroxyvitamin D3 in patients with primary hyperparathyroidism. *J Bone Min Res* 1986; **1**:137–44.
 15. Webster J, Pisciteelli G, Polli A, Ferrari CI, Ismail I, Scanlon MF. A comparison of cabergoline and bromocriptine in the treatment of hyperprolactinaemic amenorrhoea. *N Engl J Med* 1994; **331**:904–9.
 16. Knudson AG. Mutation and Cancer: Statistical study of retinoblastoma. *Proc Natl Acad Sci USA* 1971; **68**(4):820–3.
 17. Service FJ, Dale AJD, Elveback LR, Jiang N-S. Insulinoma: Clinical and diagnostic features of 60 cases. *Mayo Clinic Proc* 1976; **51**:417–29.
 18. Vasen HFA, Lamers CBHW, Lips CJM. Screening for the multiple endocrine neoplasia syndrome type 1. *Arch Intern Med* 1989; **149**:2717–22.
 19. Croisier JC, Azerod E, Lubetzki J. L'adenomatose polyendocrinienne (syndrome de Wermer). A propos d'une observation personnelle et revue de la literature. *Semin Hop Paris* 1971; **47**:494–525.
 20. Skogseid B, Eriksson B, Lundqvist G, Lorelius E, Rastad J, Wide L, Åkerstrom G, Oberg K. Multiple endocrine neoplasia type 1: A 10-year prospective screening study in four kindreds. *J Clin Endocrinol Metab* 1991; **73**(2):281–7.
 21. Knudson AG, Strong LC. Mutation and cancer: a model for Wilm's tumor of the kidney. *J Natl Cancer Inst* 1972; **48**:313–24.
 22. Knudson AG, Strong LC. Mutation and cancer: neuroblastoma and phaeochromocytoma. *Am J Hum Genet* 1972; **24**:514–32.
 23. Maher E, Yates JRW, Ferguson-Smith MA. Statistical analysis of the two-stage model in von Hippel Lindau disease, and in sporadic cerebellar and renal cell carcinoma. *J Med Genet* 1990; **27**:311–14.
 24. Pang JT, Lloyd SE, Wooding C, Farren B, Pottinger B, Harding B, Leigh SEA, Pook MA, Benham FJ, Gillett GT, Taggart RT, Thakker RV. Genetic mapping studies of 40 loci and 23 cosmids in chromosome 11p13-11q13, and exclusion of m-calpain as the multiple endocrine neoplasia type 1 gene. *Hum Genet* 1996; **97**: 732–41.
 25. Larsson C, Skogseid B, Oberg K, Nakamura Y, Nordenskjold M. Multiple endocrine neoplasia type 1 gene maps to chromosome 11 and is lost in insulinoma. *Nature* 1988; **332**:85–7.
 26. Thakker RV, Bouloux P, Wooding C, Chotal K, Broad PM, Spurr NK, Besser GM, O'Riordan JLH. Association of parathyroid tumours in multiple endocrine neoplasia type 1 with loss of alleles on chromosome 11. *N Engl J Med* 1989; **321**:218–24.
 27. Thakker RV. The role of molecular genetics in screening for multiple endocrine neoplasia type 1. *Endocrinol Metab Clin North Am* 1994; **23**(1):117–35.

Appendix I: MEN1 patient questionnaire

SERUM BIOCHEMISTRY RESULTS: (indicate “ND” if not done)

| | | | |
|------------------------|-------|-------|-------|
| Calcium | date: | date: | date: |
| Albumin | | | |
| Corrected calcium | | | |
| Ionised calcium | | | |
| Creatinine | | | |
| Phosphate | | | |
| PTH | | | |
| Prolactin | | | |
| GH | | | |
| ACTH | | | |
| Gastrin | | | |
| Insulin/glucose | | | |
| Pancreatic polypeptide | | | |
| Others | | | |

Please provide a **FAMILY TREE** below; include all affected and unaffected members, give dates of birth whether alive/dead (age at death) and maiden name or origin, if possible

Please return completed form to **Professor RV Thakker, MRC Molecular Endocrinology Group, Collier Building, Royal Postgraduate Medical School, Hammersmith Hospital, Du Cane Road, London W12 0NN, UK, tel: (44) (0)181 740 3014; fax (44) (0)181 749 8341**

Please supply details of patients with MEN1 (i.e. 2 or more tumours), or patients who have one tumour and a family history of MEN1, or any individual who has been screened in a family with MEN1.

NAME AND ADDRESS and/or identification code (e.g.hospital number)

DATE OF BIRTH: **SEX:**

SYMPTOMATIC/ASYMPTOMATIC **List order (1,2,3,...) Date/age of onset**

| | |
|--------------------|--|
| renal stones | yes/no |
| bone pain | yes/no |
| indigestion | yes/no |
| ulcers | yes/no |
| diarrhoea | yes/no |
| fits | yes/no |
| amenorrhoea | yes/no |
| galactorrhoea | yes/no |
| TREATMENT: | Drug Surgery pathology report |
| | (date) (date) (precis and reference numbers) |
| parathyroid tumour | |
| pancreatic tumour | |
| pituitary tumour | |
| renal stones | |
| ulcers | |

KNOWN FAMILY HISTORY yes/no

CONSULTANT/GP in charge:(name address and telephone number)

Blood obtained for: cell line (10-20 ml heparin) yes/no

DNA (10-20 ml EDTA) yes/no

Growth factor (10-20 ml each of clotted EDTA and heparin) yes/no

Appendix II: Clinical details of 220 MEN1 patients

| Family | Patient no. | Sex | Age of detection | Tumour | | | |
|--------|-------------|-----|------------------|-------------|-------------------|-------------------|------------------------------------|
| | | | | Parathyroid | Pancreatic | Pituitary | Other |
| 1/90 | III.4 | F | 18 | PTX | | | |
| 1/91 | I.3 | M | 28 | PTX | GAS _r | | |
| 1/92 | III.3 | M | 27 | PTX | GAS _s | | |
| 2/92 | II.3 | F | 21 | PTX | | | |
| 2.2/91 | I.1 | M | 45 | | TUM _{pm} | | ADR _{pm} |
| | II.1 | M | 34 | PTX | GAS _r | | |
| | III.1 | M | 11 | PTX | INS _s | | |
| 2.4/90 | II.8 | F | 62 | PTX | GAS _s | | |
| | II.10 | F | 60 | PTX | | | |
| 3/92 | II.5 | M | 33 | PTX | | GH _s | |
| | III.2 | F | 22 | PTX | | | |
| 3.3/87 | II.1 | M | 16 | Ca | GAS _r | | |
| 3.4/87 | II.1 | M | 23 | PTX | | PRL _r | |
| 3.5/87 | I.1 | M | 20 | Ca | INS _s | | |
| | II.1 | M | 34 | PTX | | | |
| 3.6/87 | II.1 | M | 29 | PTX | GAS _a | | |
| 4/89 | III.1 | M | 67 | PTH | GAS _r | | |
| | III.2 | M | 46 | PTX | GAS _s | | |
| | III.4 | F | 45 | PTX | GAS _{pm} | NFT _{pm} | |
| | III.7 | F | 42 | PTX | | | |
| | III.12 | M | 39 | Ca | | | |
| | III.17 | F | 54 | Ca | GAS _r | | |
| | III.19 | F | 20 | PTX | | | |
| | III.21 | F | 55 | Ca | | | |
| | IV.3 | M | 26 | Ca | | | |
| | IV.6 | F | 31 | Ca | | | |
| | IV.7 | M | 31 | PTH | | PRL _r | |
| | IV.10 | M | 24 | PTX | | | |
| | IV.16 | F | 21 | Ca | | | |
| | IV.19 | F | 36 | Ca | | | |
| | IV.20 | M | 27 | PTH | | | |
| | IV.23 | F | 38 | Ca | | | |
| | IV.25 | F | 23 | Ca | | | |
| 4.2/90 | V.1 | F | 15 | PTX | | | |
| | V.3 | M | 9 | PTX | | | |
| | V.6 | M | 14 | | INS _s | | |
| | I.2 | F | 36 | PTX | | | |
| 4.3/92 | II.2 | F | 18 | PTX | | | LIP _s |
| | II.3 | M | 20 | PTX | | | |
| | II.4 | M | 35 | PTX | INS _s | | |
| 5/87 | III.2 | F | 16 | PTX | INS _s | | |
| | II.1 | M | 37 | PTX | | | |
| 6/91 | II.1 | M | 25 | PTX | GAS _{gx} | | |
| | III.1 | F | 17 | Ca | | PRL _r | |
| 6.2/92 | I.1 | F | 55 | PTX | GAS _a | PRL _r | |
| | I.2 | M | 63 | PTX | TUM _{pm} | | |
| | II.1 | M | 32 | PTX | GAS _u | | ADR _s PHAE _s |
| | III.2 | M | 16 | PTH | | PRL _r | |
| | III.1 | M | 16 | PTH | | | |
| 6.4/91 | II.1 | F | 26 | PTX | GAS _r | PRL _r | |
| | II.2 | F | 62 | PTX | | | |
| | III.1 | F | 26 | PTX | | | |
| 7/87 | I.2 | F | 50 | PTX | | | |

| Family | Patient no. | Sex | Age of detection | Tumour | | | |
|---------|-------------|-----|------------------|-------------|-----------------------------------|-------------------|------------------|
| | | | | Parathyroid | Pancreatic | Pituitary | Other |
| 7.2/89 | II.2 | F | 38 | PTX | | | |
| | II.7 | M | 28 | PTX | TUM _{pm} | PRL _r | |
| | III.2 | F | 19 | PTX | | | |
| | III.3 | F | 14 | PTH | | | |
| | III.4 | M | 8 | PTH | | ACTH _r | |
| | II.4 | M | 29 | PTH | GAS _a | | CRN _s |
| | II.6 | F | 29 | Ca | | PRL _s | |
| 7.3/90 | III.1 | M | 22 | Ca | | | |
| | III.3 | F | 25 | PTX | | PRL _s | |
| | I.2 | F | 67 | PTX | | | |
| | II.2 | M | 29 | PTX | | | |
| 7.4/91 | III.1 | M | 18 | Ca | | | |
| | III.2 | M | 17 | Ca | | | |
| | II.1 | M | 42 | PTX | GAS _u | | |
| 8/89 | II.2 | F | 52 | PTX | GAS _u | | |
| | II.1 | M | 55 | PTX | GAS _r | | |
| | II.4 | F | 35 | PTX | | | |
| 8.2/90 | III.3 | F | 29 | PTH | | | |
| | III.4 | F | 26 | PTX | | PRL _r | |
| | III.5 | M | 39 | PTH | | | |
| | IV.3 | M | 24 | Ca | | | |
| | IV.4 | M | 35 | Ca | | | |
| 8.5/90 | IV.6 | F | 32 | PTX | | | |
| | IV.8 | F | 17 | PTH | | | |
| | II.1 | F | 51 | PTX | | | |
| 8.7/87 | II.1 | M | 75 | | TUM _r | | |
| | II.2 | F | 20 | Ca | | | |
| 8.10/91 | II.1 | F | 30 | PTX | INS _s GAS _s | | |
| 10/90 | I.1 | M | 45 | PTX | | PRL _s | |
| 11/89 | II.7 | F | 25 | | | PRL _r | |
| | II.5 | M | 42 | PTX | GAS _r | | |
| | III.5 | M | 18 | PTX | | PRL _s | |
| 11.2/90 | II.3 | M | 55 | PTH | | | |
| | II.14 | F | 21 | PTX | GAS _r | PRL _s | ADR _r |
| | III.2 | M | 32 | PTX | GAS _s | | |
| 11.3/90 | III.4 | F | 25 | | | PRL _r | |
| | II.2 | F | 22 | Ca | | | |
| | II.4 | F | 20 | PTX | | | |
| 12.3/87 | II.2 | F | 18 | PTX | | | |
| | III.1 | F | 52 | PTX | GAS _r | | |
| | II.1 | M | 33 | PTX | GAS _u | PRL _r | |
| 13/89 | III.1 | M | 27 | PTX | GAS _s | | |
| | III.4 | F | 37 | PTX | | PRL _g | |
| | III.5 | M | 32 | PTH | | PRL _r | |
| | III.7 | F | 33 | PTH | | | |
| | III.8 | F | 25 | Ca | INS _s | PRL _r | |
| 13/92 | III.10 | F | 15 | PTH | INS _s | PRL _r | |
| | III.11 | M | 20 | PTH | | GH _s | |
| | IV.1 | M | 17 | PTH | | | |
| | IV.3 | M | 12 | PTH | | | |
| | II.2 | F | 60 | PTX | | | |
| 13.2/90 | III.4 | F | 53 | Ca | | | |
| | III.2 | F | 53 | PTX | GAS _r | GH _s | |
| | II.1 | M | 47 | PTX | | | |
| 13.3/90 | II.3 | F | 54 | PTX | GAS _s | | |
| | III.1 | F | 15 | PTX | | | |
| | II.5 | F | 60 | PTX | | | |

| Family | Patient no. | Sex | Age of detection | Tumour | | | |
|----------|-------------|-----|------------------|-------------|-----------------------------------|-----------------------------------|--|
| | | | | Parathyroid | Pancreatic | Pituitary | Other |
| 13.3/92 | II.8 | M | 24 | PTX | | | |
| | III.24 | F | 11 | | | PRL _r | |
| | II.2 | M | 47 | PTX | GAS _r | | |
| | III.1 | F | 10 | | INS _s | NFT _s | |
| 13.10/87 | II.1 | F | 25 | PTX | GAS _{gx} | | |
| | III.1 | F | 20 | PTH | | | |
| 13.5/90 | III.1 | F | 18 | PTX | INS _s | PRL _r | |
| 13.7/87 | II.1 | F | 29 | PTX | GAS _s | | |
| 13.8/92 | I.2 | F | 20 | PTX | GAS _u | PRL _r | |
| | II.1 | F | 21 | PTX | | | |
| 16/90 | II.10 | F | 70 | PTX | | | |
| | II.11 | M | 68 | PTH | | | |
| | III.19 | M | 37 | PTH | | | |
| | III.21 | M | 39 | PTH | | | |
| 16/92 | III.14 | F | 46 | PTH | | | |
| | III.32 | M | 28 | PTX | GAS _r | PRL _r | |
| | III.35 | M | 32 | PTX | GAS _r | PRL _r | |
| | IV.18 | M | 14 | PTX | | | |
| | IV.20 | M | 21 | | | PRL _r | |
| | IV.21 | F | 13 | PTH | | | |
| | III.5 | M | 24 | PTX | | | |
| | III.10 | F | 23 | Ca | | | |
| | II.5 | F | 36 | PTX | | | |
| | III.12 | F | 32 | Ca | | | |
| | II.1 | M | 36 | PTX | GAS _r | | |
| | II.3 | F | 35 | PTX | GAS _r | PRL _r | |
| 16.3/87 | II.1 | M | 22 | PTX | TUM _s | | |
| 18/89 | II.3 | M | 23 | PTX | GAS _s | GH _r ACTH _r | NF |
| | III.4 | F | 30 | PTH | | | |
| | III.6 | F | 29 | PTX | | ACTH _r | |
| | III.7 | M | 24 | PTH | | | |
| 18.2/91 | III.15 | M | 32 | PTX | GAS _s INS _s | | |
| | III.13 | F | 19 | PTX | | | ADR _r |
| | IV.7 | M | 8 | PTH | | | |
| | II.4 | F | 26 | Ca | | PRL _r | |
| 19/89 | II.7 | F | 21 | PTX | | | |
| | II.2 | M | 41 | PTX | | TUM _s | |
| 19/92 | III.4 | F | 16 | | INS _s | | |
| | II.2 | M | 47 | PTX | | | CRN _{pm} [*] , ADR _{pm} |
| | II.3 | F | 39 | PTX | GAS _s INS _s | | ADR _{pm} |
| | III.4 | F | 20 | PTX | | | |
| 19.1/92 | III.6 | M | 17 | Ca | | | |
| | II.6 | F | 22 | PTH | | PRL _r | |
| | I.2 | F | 61 | PTX | | | |
| 19.2/89 | I.2 | F | 56 | PTX | GAS _r | NFT _r | |
| | II.2 | F | 28 | PTX | | PRL _r | |
| 19.2/92 | II.1 | M | 71 | | | PRL _h | |
| | III.2 | F | 35 | PTX | GAS _s | | |
| | IV.3 | F | 11 | PTX | | | |
| | III.5 | F | 41 | PTX | | | |
| 19.3/89 | III.6 | F | 40 | PTX | | | CRN _s |
| | IV.1 | M | 30 | PTX | GAS _s | | |
| | IV.6 | M | 35 | PTX | | | |
| | V.1 | F | 12 | PTX | | | |
| | V.2 | M | 12 | PTX | | | |
| | II.3 | F | 31 | PTX | INS _s | | |
| 19.4/91 | II.3 | F | 31 | PTX | | | |
| 19.6/87 | II.6 | F | 62 | PTX | | | |

| Family | Patient no. | Sex | Age of detection | Tumour | | | |
|---------|-------------|-----|------------------|-------------|-----------------------------------|----------------------------------|---|
| | | | | Parathyroid | Pancreatic | Pituitary | Other |
| 20/89 | II.7 | F | 56 | PTX | | | |
| | II.8 | F | 51 | PTX | | GH _s | |
| | I.2 | F | 40 | PTX | | | |
| | II.2 | F | 50 | PTX | GAS _s | | ADR _r |
| 23/89 | III.2 | M | 26 | PTH | | | |
| | II.6 | F | 60 | PTX | GAS _a | | |
| | II.8 | F | 51 | PTX | | PRL _r | |
| | III.12 | F | 20 | PTX | TUM _{pm} | NFT _{pm} | ADR _{pm} |
| | III.13 | F | 27 | PTX | NFT _s | | |
| 23.2/87 | I.1 | F | 44 | PTX | GAS _r | | |
| 23.3/87 | II.1 | F | 12 | PTX | INS _s | | |
| 23.5/87 | II.3 | F | 57 | PTX | GAS _u | | CRN _s [†] |
| 23.6/90 | II.2 | M | 35 | PTX | | GH _r | |
| 25/90 | III.6 | F | 19 | Ca | | PRL _r | |
| | V.4 | M | 20 | PTH | | | |
| 25.1/92 | II.1 | F | 34 | PTX | INS _s | PRL _r | |
| S1 | | F | 47 | PTH | GAS _s | | |
| S2 | | F | 59 | PTX | | GH _r | |
| S3 | | M | 37 | PTX | GAS _s | | ADR _s |
| S4 | | F | 27 | PTX | INS _s | | |
| S5 | | F | 79 | PTX | INS _s | | |
| S6 | | F | 52 | PTX | | GH _s | |
| S7 | | M | 10 | PTX | INS _s | | |
| S8 | | F | 48 | PTX | GAS _r | GH _g | |
| S9 | | F | 50 | PTX | | GH _s | |
| S10 | | M | 17 | Ca | INS _s | | |
| S11 | | F | 57 | PTH | | GH _r | |
| S12 | | M | 18 | PTX | GAS _r | | |
| S13 | | M | 47 | PTX | | GH _s | |
| S14 | | M | 42 | PTX | GAS _s GCG _s | PRL _s GH _s | CRN _s T _s MM _s |
| S15 | | M | 42 | | GAS _s GCG _s | | ADR _r |
| S16 | | M | 44 | PTX | GAS _{pm} | PRL _r | |
| S17 | | F | 30 | PTX | GAS _s | PRL _s | |
| S18 | | M | 46 | PTX | GAS _s | | |
| S19 | | F | 20 | | | GH _s | CRN _s |
| S20 | | F | 37 | PTX | GAS _s | | |
| S21 | | F | 31 | PTX | INS _s | | |
| S22 | | M | 47 | PTX | | | CRN _s |
| S23 | | F | 19 | PTX | INS _s | PRL _r | |
| S24 | | F | 63 | PTH | INS _s | | |
| S25 | | F | 32 | PTX | GAS _s | | |
| S26 | | F | 28 | PTX | INS _s | PRL _r | |
| S27 | | F | 43 | PTX | GAS _r | PRL _r | |
| S28 | | M | 12 | PTX | INS _s | | |
| S29 | | M | 44 | PTX | | NFT _s | |
| S30 | | F | 54 | PTX | | PRL _s | ADR _s [*] |
| S31 | | M | 43 | PTX | | | CRN _s [†] |
| S32 | | F | 24 | PTX | | ACTH _s | |
| S33 | | M | 32 | PTX | | GH _s | |
| S34 | | F | 18 | PTX | GAS _s INS _s | PRL _r | LIP _s |
| S35 | | M | 36 | PTX | GAS _s | | |
| S36 | | M | 22 | PTX | GAS _s | | |

Ca, persistent hypercalcaemia; PTH, high serum PTH level and hypercalcaemia; PTX, parathyroidectomy.

GAS, pancreatic tumour secreting gastrin confirmed on the basis of: _a acid output studies, _g gastrectomy, _u repeated gastric ulceration, _r radiology, _s surgery, _{pm} post-mortem.

INS, pancreatic tumour secreting insulin, confirmed by: _b biochemistry, _s surgery.

GCG, pancreatic tumour secreting glucagon: _s surgery.

TUM, tumour type unknown: _r radiology, _{pm} at post mortem.

NFT, non-functioning tumour: _s surgery.

PRL, prolactinoma on basis of: _g galactorrhoea, _h persistent hyperprolactinaemia and/or confirmed by: _r radiology, _s surgery, _{pm} post-mortem.

GH, somatotrophinoma on basis of biochemical data and: _s surgery, _r radiology.

ACTH, corticotrophinoma on basis of biochemical data and: _s surgery, _r radiology, _s glucose tolerance test.

NFT, non-functioning tumour found at: _s surgery, _r radiology, _{pm} post-mortem.

TUM, tumour of unknown type found at: _s surgery.

ADR, adrenocortical tumour found at: _r radiology, _s surgery, _{pm} post-mortem, *carcinoma.

PHAE, pheochromocytoma.

CRN, carcinoid tumour demonstrated biochemically and: _s surgery, _r radiology, _{pm} post-mortem, [†]gastric, ^{*}thymic.

LIP, lipomata: _s surgery.

T, testicular teratoma: _s surgery.

MM, malignant melanoma.

