

# Illusion of flight? Absence, evidence and the age of winged insects

SANDRA R. SCHACHAT<sup>1,\*</sup>, PAUL Z. GOLDSTEIN<sup>2</sup>, ROB DESALLE<sup>3</sup>, DEAN M. BOBO<sup>3,4</sup>, C. KEVIN BOYCE<sup>1</sup>, JONATHAN L. PAYNE<sup>1</sup> and CONRAD C. LABANDEIRA<sup>5,6,7</sup>

<sup>1</sup>*Department of Geological Sciences, Stanford University, Stanford, CA, USA*

<sup>2</sup>*Systematic Entomology Laboratory, USDA, National Museum of Natural History, Smithsonian Institution, Washington, DC, USA*

<sup>3</sup>*American Museum of Natural History, Sackler Institute for Comparative Genomics, New York, NY, USA*

<sup>4</sup>*Department of Ecology, Evolution, and Environmental Biology, Columbia University, New York, NY, USA*

<sup>5</sup>*Department of Paleobiology, National Museum of Natural History, Smithsonian Institution, Washington, DC, USA*

<sup>6</sup>*Department of Entomology and Behavior, Ecology, Evolution, and Systematics Program, University of Maryland, College Park, MD, USA*

<sup>7</sup>*Capital Normal University, School of Life Sciences, Beijing, China*

Received 31 March 2022; revised 2 October 2022; accepted for publication 3 October 2022

The earliest fossils of winged insects (Pterygota) are mid-Carboniferous (latest Mississippian, 328–324 Mya), but estimates of their age based on fossil-calibrated molecular phylogenetic studies place their origin at 440–370 Mya during the Silurian or Devonian. This discrepancy would require that winged insects evaded fossilization for at least the first ~50 Myr of their history. Here, we examine the plausibility of such a gap in the fossil record, and possible explanations for it, based on comparisons with the fossil records of other arthropod groups, the distribution of first occurrence dates of pterygote families, phylogenetically informed simulations of the fossilization of Palaeozoic insects, and re-analysis of data presented by Misof and colleagues using updated fossil calibrations under a variety of prior probability settings. We do not find support for the mechanisms previously suggested to account for such an extended gap in the pterygote fossil record, including sampling bias, preservation bias, and body size. We suggest that inference of an early origin of Pterygota long prior to their first appearance in the fossil record is probably an analytical artefact of taxon sampling and choice of fossil calibration points, possibly compounded by heterogeneity in rates of sequence evolution or speciation, including radiations or ‘bursts’ during their early history.

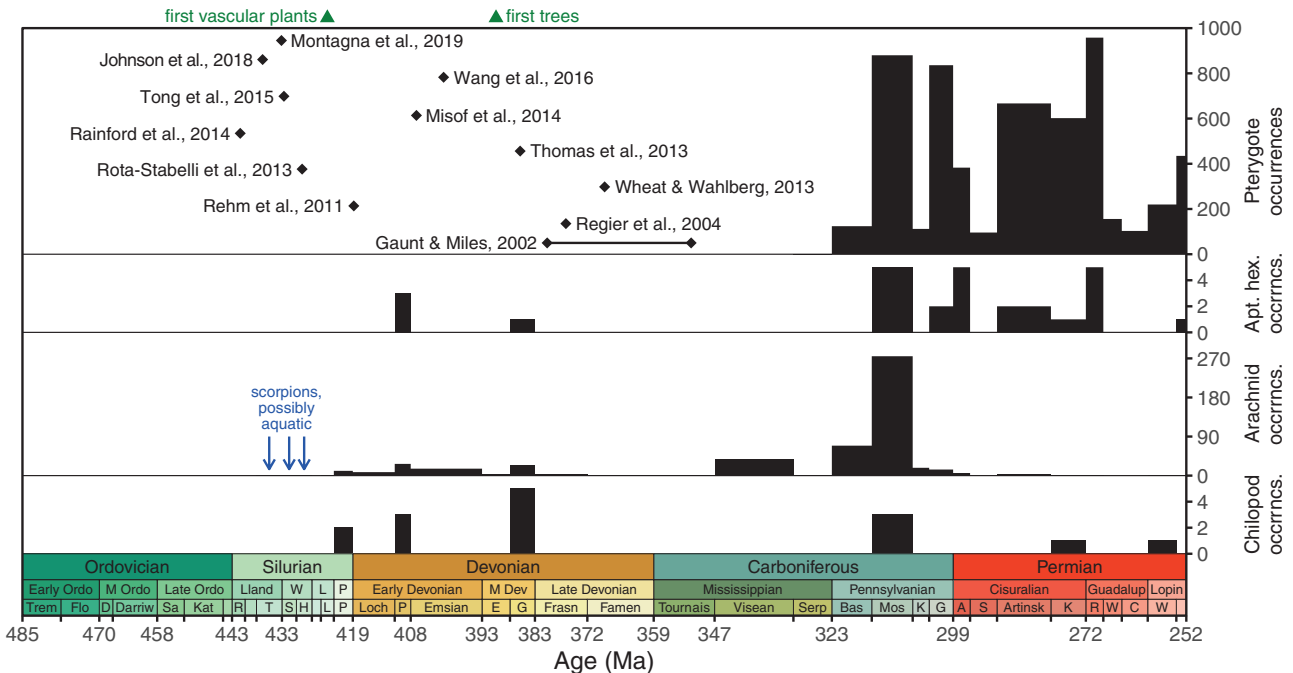
ADDITIONAL KEYWORDS: Carboniferous – Devonian – diversification – Palaeozoic – phylogeny.

## INTRODUCTION

Winged insects (Pterygota) contain more described species than the rest of all multicellular life (Grimaldi & Engel, 2005), and ascertaining the age of pterygotes is key to reconstructing the evolution of terrestrial ecosystems. Although the pterygote fossil record extends back as far as the mid-Carboniferous (328–324 Mya), age estimates for the origin of Pterygota derived from fossil-calibrated molecular phylogenetic studies have

ranged from the Late Devonian, ~370 Mya (Wheat & Wahlberg, 2013), to the Silurian, ~430 Mya (Johnson *et al.*, 2018). These estimates highlight broader questions about the completeness of the insect fossil record, as each requires a substantial gap between the most recent common ancestor of pterygotes and their oldest known fossil representative (Fig. 1). Understanding this discrepancy requires that we evaluate not only the robustness of the insect fossil record itself but also the extent to which choice of fossil calibration points and parameters impact the reliability of divergence time estimates. In this paper, we examine the plausibility of a major gap in the fossil record of winged insects,

\*Corresponding author. E-mail: [sschachat@schmidtsciencefellows.org](mailto:sschachat@schmidtsciencefellows.org)



**Figure 1.** Fossil occurrences of terrestrial arthropods during the Palaeozoic. Estimated origin dates for Pterygota are from various molecular phylogenetic studies (Gaunt & Miles, 2002; Regier *et al.*, 2004; Rehm *et al.*, 2011; Rota-Stabelli *et al.*, 2013; Thomas *et al.*, 2013; Wheat & Wahlberg, 2013; Misof *et al.*, 2014; Rainford *et al.*, 2014; Tong *et al.*, 2015; Wang *et al.*, 2016; Johnson *et al.*, 2018; Montagna *et al.*, 2019). Apt. hex., apterygote hexapoda. Non-chilopod myriapods are not included due to exoskeleton mineralization in most diplopods that renders them less comparable to insects.

and whether re-analyses using novel calibrations may affect its estimated duration.

Discordance between age estimates derived from time-calibrated phylogenetic studies and estimates based directly on the fossil record is not uncommon (Ksepka *et al.*, 2014; Beaulieu *et al.*, 2015; Brown & Smith, 2018). In addition to winged insects, significant discrepancies have been noted for Metazoa, angiosperms, and mammals (Phillips & Fruciano, 2018; Budd & Mann, 2020; Ramrez-Barahona *et al.*, 2020) as well as myriapods (Fernandez *et al.*, 2018) and arachnids (Lozano-Fernandez *et al.*, 2020). The fossil record of Pterygota is not nearly as complete as those of brachiopods, bivalves, gastropods, or cephalopods (Kidwell & Flessa, 1996; Foote & Sepkoski, 1999), but far more so than those of tardigrades, nematodes, and various other invertebrates (De Baets *et al.*, 2021; Mapalo *et al.*, 2021). Although the pterygote record is not sufficient to pinpoint the origins of most extant taxa with precision, here we examine the plausibility of such an extended gap in the fossil record for such a speciose group. The more widely distributed or ubiquitous a given group, the less likely that it will evade fossilization entirely through a given interval of geological time. Along with holometaboly, wings are considered a key innovation driving the tremendous

diversification of insects (Nicholson *et al.*, 2014), and pterygotes outnumber all other terrestrial arthropods combined among fossils from the Bashkirian onwards (323 Mya; Fig. 1). Although there is little consensus as to when Pterygota originated, the possibility that crown-group pterygotes became widespread within a few million years of their origin cannot be discounted.

The primary goal of this contribution is to evaluate the plausibility of a gap during the first 50+ Myr of pterygote evolution by examining possible explanations and the possible impacts of fossil calibrations, prior schemes, and parameterizations on re-analyses of phylogenomic data. After reviewing the insect fossil record in general terms and exploring possible explanations for the absence of pre-Carboniferous pterygote fossils, we perform four analyses. First, we use a comparative palaeontological approach to assess the likelihood of systematically overlooked pterygote fossils. Second, we use palaeontological data to directly estimate the age of Pterygota. Third, we use different fossil calibration compendia and calibration schemes to evaluate the molecular dating scheme proposed by Misof *et al.* (2014). Fourth and finally, we use simulations to estimate expected gap lengths in the fossil record of pterygotes.

Node-dating procedures are sensitive not only to analytical parameters, including choices of fossil

calibration points, and dataset-specific features such as taxon sampling and missing sequence data, but also to locus- and taxon-specific histories of rate heterogeneity. At issue, therefore, are the evolutionary, geological, and taphonomic explanations of the putative gap itself, and the sensitivity of its inference to choices of parameterization and fossil calibration.

### BACKGROUND: INFERENCE OF AND EXPLANATIONS FOR AN APPARENT FOSSIL GAP

The Palaeozoic fossil record of terrestrial arthropods can be divided into two phases. First, from the appearance of the first terrestrial arthropods during the Silurian, through the Middle Mississippian (Viséan; Edgecombe, 2011; Dunlop *et al.*, 2018), definitive fossil pterygotes are absent (Haug & Haug, 2017), and apterygote hexapod fossils appear even more rarely than myriapods and arachnids (Whalley & Jarzembowski, 1981; Shear *et al.*, 1984; Labandeira *et al.*, 1988; Fayers & Trewin, 2005) – the rarity of all groups driven largely by Lagerstätten effects (Edgecombe & Legg, 2013). The Late Mississippian (Serpukhovian; 330.9–323.2 Mya) marks an inflection point during which no described chilopods and only one possible arachnid are known, but multiple pterygotes appear by the end of the interval (Brauckmann & Schneider, 1996; Prokop *et al.*, 2005). In the second phase, from the Pennsylvanian onwards, apterygote hexapods remain relatively rare while pterygote fossils outnumber those of all other terrestrial arthropods. Here we summarize aspects of the pterygote fossil record that bear on the plausibility of an extended gap between the first known winged insects and inferred ages of origin, and review explanations put forth to account for their ‘sudden’ appearance in the fossil record. These include scenarios involving rapid changes in rates of evolution and empirical bias against pterygote fossils, especially small ones.

Both the calibration and the interpretation of dated phylogenies depend not only on the inferred ages of fossils used as calibration points but on the differentiation of crown- vs. stem-groups among those fossils. ‘Crown-group’ refers to the smallest monophyletic group that includes all extant members of a given clade, their most recent common ancestor, and any extinct descendants thereof. Crown-group Hemiptera, for example, includes all extant and extinct descendants of the most recent common ancestor of Sternorrhyncha, Coelorrhyncha, Auchenorrhyncha, and Heteroptera. ‘Stem-group’ refers to extinct taxa that are phylogenetically adjacent to but outside the crown-group. Stem-group Hemiptera includes all taxa that are more closely related to Hemiptera than to any

other extant order, but are not descended from the most recent common ancestor of extant Hemiptera. ‘Total-group’ encompasses the stem and crown of a given group.

The fossil record contains unambiguous stem representatives of most pterygote orders (Willmann, 1989; Ansoerge, 2002; Rasnitsyn & Quicke, 2002; Grimaldi & Engel, 2005; Engel *et al.*, 2009; Huang & Nel, 2009; Shang *et al.*, 2011; Nel *et al.*, 2013; Delsol *et al.*, 2016; Pohl & Beutel, 2016) with the three possible exceptions of Zoraptera, Megaloptera, and Lepidoptera. Plausible explanations exist for each of these: Zoraptera are the least speciose and least frequently encountered of all extant winged insect orders; Megaloptera is the second-least speciose of the holometabolous orders, and its proximity to Neuroptera complicates the task of distinguishing stem Megaloptera from stem Neuropterida; and Lepidoptera have thickly scaled wings and bodies that prevent them from sinking from water surfaces to the sediments where fossilization can occur (Grimaldi & Engel, 2005). Although the fossil record contains various potential stem representatives of each of these three orders, none has been conclusively excluded from their crown-groups (Engel, 2004; Mashimo *et al.*, 2014; Sohn *et al.*, 2015).

As various studies have reiterated (Smith *et al.*, 2006; Coiro *et al.*, 2019; Budd & Mann, 2020), the appearance of stem- and crown-group fossils in the sequence predicted by phylogenetic reconstruction evinces the fossil record’s general reliability. Primitively wingless insects (Shear *et al.*, 1984; Fayers & Trewin, 2005) appear in the fossil record tens of millions of years earlier than winged insects. Palaeoptera (Brauckmann & Schneider, 1996) and Polyneoptera (Prokop *et al.*, 2005) appear earlier than Paraneoptera (Laurentiaux, 1952). Paraneoptera are paraphyletic with respect to Holometabola (Whiting *et al.*, 1997; Ishiwata *et al.*, 2011; Misof *et al.*, 2014; Johnson *et al.*, 2018) and appear before them (Nel *et al.*, 2007, 2013). Holometabola, in turn, appear well before the crown-groups of its component orders (Haug *et al.*, 2015; Wolfe *et al.*, 2016).

Obviously the fossil record is not sufficiently complete to expect that it recapitulates phylogeny with fine-tuned precision or accuracy, beyond the very broad-brush sequence of major taxa that are well represented as fossils; younger taxonomic units are expected to be less well represented as fossils, much less captured sequentially in the fossil record. Two kinds of comparisons are especially informative when utilizing the potential correspondence between fossils and phyletic sequence to evaluate the completeness of the fossil record. First are comparisons among taxa with different preservation biases. Second are comparisons of stem groups to their corresponding crown groups.

The appearance of Apterygota and Pterygota in phyletic sequence is a particularly informative example due to the disparities in preservation potential among these two groups. The Pennsylvanian–Permian fossil record contains over 5500 described occurrences of Pterygota, but fewer than 20 described occurrences of apterygote insects (Durden, 1975; Rasnitsyn *et al.*, 2004; Rinehart *et al.*, 2005; Engel, 2009; Staniczek *et al.*, 2014; Mendes, 2018; Liu *et al.*, 2021). Because apterygote insects are so much less likely to become fossilized, their presence in the fossil record during an extended interval from which pterygotes are unknown can be reasonably interpreted as true evidence of pterygote absence. Findings of apterygote insects from Early (Fayers & Trewin, 2005) and Middle Devonian (Shear *et al.*, 1984) deposits constitute a canonical example of major groups appearing in phyletic sequence.

Some discussions of fossil gaps have centred on macroevolutionary explanations to account for whether a group for which a purported gap exists, pterygotes in this case, underwent a rapid radiation or evolutionary ‘burst’ early in their history, followed by a slowdown or stasis (Foote *et al.*, 1999; Beck & Lee, 2014; Beaulieu *et al.*, 2015). Although the presence of stem taxa for most major winged insect groups speaks to the strength of the fossil record, the earliest known fossil representatives of Holometabola – which appeared after several major pterygote lineages had arisen (Misof *et al.*, 2014) – are barely 10 Myr younger than the earliest known Palaeoptera and Neoptera (Nel *et al.*, 2013; Haug *et al.*, 2015). This timing would suggest a rapid radiation among crown Pterygota immediately following their appearance. Such an evolutionary burst might be expected in light of the manifold advantages that wings confer, such as locating food resources, avoiding predators, regulating body temperature, finding mates, and other advantages of colonizing a three-dimensional environment (Grimaldi & Engel, 2005). Although such scenarios would be reflected in highly asymmetrical rates of speciation and/or molecular evolution, these are difficult if not impossible to parse from neontological data alone and the ‘early burst’ hypothesis is thus equally difficult to test using molecular data, even with denser sampling of taxa and loci (Tihelka *et al.*, 2021; Craig *et al.*, 2022).

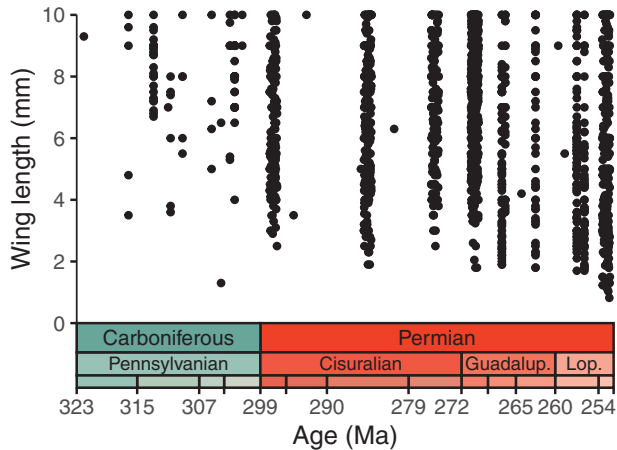
In addition to macroevolutionary explanations, statistical and intrinsic biological explanations have been variously invoked to account for gaps in the fossil records of groups other than insects, primarily sampling bias (Gardner *et al.*, 2019; Harper *et al.*, 2019), preservation bias (Doyle, 2012; Dos Reis *et al.*, 2015), and small size (Wray, 2015; Han *et al.*, 2017; Li *et al.*, 2019). None of these explains the lack of pterygote fossils before the Pennsylvanian.

The interpretation of the fossil gap as a sampling artefact would rest on the idea that insufficient collecting effort has been dedicated to Devonian and Mississippian insects. However, a number of productive arthropod compression localities are known, such as Alken an der Mosel (Poschmann & Dunlop, 2010), Campbellton (Shear *et al.*, 1996), Gilboa (Shear & Selden, 2001), and the Rhenish Slate Mountains (Poschmann & Dunlop, 2011), and an abundance of macerated material has been sifted (Shear *et al.*, 1984; Labandeira *et al.*, 1988). The case is simply that none of this material contains winged insects despite wings being the most abundantly preserved insect body part (Karr & Clapham, 2015). Similarly, the few putative fossil Pterygota that have been described from as far back as the Silurian (Agnus, 1904; Rohdendorf, 1961, 1970; Engel & Grimaldi, 2004; Garrouste *et al.*, 2012) have all subsequently been shown to belong to different arthropod groups, such as centipedes and crustaceans (Rohdendorf, 1972; Hornschemeyer *et al.*, 2013; Haug & Haug, 2017). Again, terrestrial arthropod fossils are present, just not winged insects.

With respect to preservation bias, no mechanism has been suggested to explain how pterygotes might have been excluded from lowland habitats in which arachnids and myriapods were fossilized during the Silurian–Mississippian. Arachnids and myriapods diversified considerably during this interval and reached depositional settings where they became preserved (Edgecombe, 2011; Dunlop *et al.*, 2018), occasionally alongside fragments of apterygote hexapods (Shear *et al.*, 1984; Labandeira *et al.*, 1988).

Finally, minute body size has been proposed as an explanation of how crown representatives of groups such as early Bilateria, which ostensibly diversified during the Ediacaran, could have evaded fossilization (Budd & Mann, 2020). In the case of insects, however, homologies among the primary wing veins suggest that the common ancestors of the major groups of pterygotes all had wings at least 5 mm long (Polilov, 2016). The primary wing veins have been recognized as homologous across all Pterygota (Schubnel *et al.*, 2020), but are nearly always lost in very small insects (Polilov, 2015), suggesting that extant pterygote orders diverged from common ancestors with body lengths significantly greater than those of the minute arachnids recovered from Silurian–Mississippian deposits (Dunlop, 2019). Moreover, the two oldest known pterygote fossils represent both Palaeoptera and Neoptera but each has wings 11 mm long (Brauckmann & Schneider, 1996; Prokop *et al.*, 2005); as noted previously (Rasnitsyn, 2003), minimum known wing lengths decrease from the Pennsylvanian through the Permian and into the Triassic (Fig. 2).





**Figure 2.** Recorded lengths of Pennsylvanian and Permian insect wings below 10 mm. Data compiled by Clapham & Karr (2012).

#### DIGGING DEEPER INTO THE FOSSIL RECORD OF HEMIPTERA

The origin of Hemiptera (true bugs, aphids, cicadas, scale insects), the most ancient and the only hemimetabolous of the ‘big five’ pterygote orders, presents an example of stem and crown groups appearing in phyletic sequence, at odds with one or more divergence time estimates that greatly pre-date first appearances in the fossil record. The earliest unequivocal hemipteran fossils – the only definitive stem-group hemipteran, followed by the first possible crown-group hemipteran (probably in the stem-group of Euhemiptera) – appear within 16 Myr in the late Carboniferous (Pennsylvanian) (Nel *et al.*, 2013; Szwedo, 2016). During the Permian, however, hemipteran fossils become much more common. Representatives of all three non-heteropteran suborders, and five of the seven non-heteropteran infraorders, are known from this period (Szwedo, 2016). This implies that (1) Hemiptera diversified quickly and (2) diversification of crown-group Hemiptera, if not total-group Hemiptera, occurred from the Pennsylvanian onwards.

The explosive diversification of Hemiptera during the Permian has long been noted in the palaeontological literature (Wootton, 1981). Of the 1023 known pterygote fossils older than 304 Mya (pre-Gzhelian), two are Hemiptera (Laurentiaux, 1952; Nel *et al.*, 2013). Of the 842 pterygote occurrences known from the Gzhelian (304–299 Mya), two are putative hemipterans but their affinities within the order remain unknown (Szwedo, 2016). In contrast, 531 of the 3636 known Permian occurrences of Pterygota (14.6%) have been assigned to Hemiptera, and 94% of these Hemiptera have been assigned to the total-group of an extant infraorder.

Several published time-calibrated phylogenies bear on the Hemiptera (Misof *et al.*, 2014; Johnson *et al.*, 2018; Montagna *et al.*, 2019; Ye *et al.*, 2022). Using BEAST (Drummond & Rambaut, 2007), Misof *et al.* (2014) inferred an Early Permian age for Hemiptera, and suggested that piercing-and-sucking feeding damage on Devonian liverworts (~380 Mya) might be attributable to hemipteroid insects, although Labandeira *et al.* (2013) had expressed doubts on palaeontological grounds that Hemipteroidea could have originated over 70 Myr before their first appearance in the Pennsylvanian. The divergence times inferred by Johnson *et al.* (2018) reconstructed with MCMCTree (Yang, 2007) are noteworthy in several respects. Their divergence estimates included an age for crown Hemiptera near the midpoint of the Devonian (419–359 Mya), with all four hemipteran suborders diverging during the Mississippian, and two of these > 20 Myr before the first appearance of winged insects in the fossil record. Their analyses further depict nine of 11 infraordinal divergences occurring by the end of the Permian (299–252 Mya), among them the two former ‘homopteran’ infraorders (Auchenorrhyncha, including Fulgoroidea, and Sternorrhyncha), which are inferred to have radiated primarily during the Pennsylvanian – a period from which only four total-group Hemipteran fossils are known. This scenario requires either: (1) that these infraorders diversified during the Pennsylvanian, but evaded fossilization, while other Pterygota such as Palaeodictyoptera and Orthopteroidea became fossilized in considerable numbers; or (2) that these hemipteran infraorders were excluded from depositional environments by a mechanism that did not impact other winged insects. Any such mechanisms (or bias) would then also have had to disappear during the Permian to account for the prevalence of Hemiptera in the entomofauna of that period. Given that there are over 500 hemipteran fossils known from the Palaeozoic, none of which are heteropterans, and that the first fossils appeared in a phylogenetically predictable sequence, the Palaeozoic hemipteran fossil record is not deeply suspect, and it is therefore reasonable to question whether the divergence of Heteroptera from ‘Homoptera’ could have been so strongly discordant with the fossil record, occurring before the appearance even of Pterygota in the fossil record.

The timing of infraordinal diversification within the Heteroptera is especially problematic. The only Palaeozoic heteropteran fossil is dubious, and the first unequivocal heteropteran appears in the Mesozoic (Fig. 3; Szwedo, 2016). The estimates of Johnson *et al.* include a divergence between Heteroptera and Auchenorrhyncha in the Late Devonian, well before any wings appear in the fossil record, with the basal

heteropteran infraorders diverging by the Permian/Triassic boundary. In other words, whereas the fossil record shows explosive radiations of ‘Homoptera’ confined to the Permian following basal splits in the Pennsylvanian, Johnson and colleagues infer a Palaeozoic diversification history of Hemiptera that began almost 100 Myr earlier, and encompassed not only ‘Homoptera’ but also Heteroptera – the latter of which appear in the fossil record from the Triassic onwards.

On an infraorder-by-infraorder basis, Hemiptera do not appear in the fossil record in perfect phyletic sequence (Fig. 3). For example, Dipsocoromorpha appear before Gerromorpha, and both Cimicomorpha and Pentatomomorpha appear before Leptopodomorpha. Aphidomorpha and Psyllodea appear before Aleyrodomorpha. However, the ubiquity of Scytinopteromorpha (putative stem-group Heteroptera; Shcherbakov, 2021) during the Permian, and the absence of crown-group Heteroptera during most if not all of this period, suggests that few if any divergences among crown-group Heteroptera occurred before the Triassic. Likewise, Inguromorpha (the proposed stem-group of Coleorrhyncha; Wang *et al.*, 2009; Ziade *et al.*, 2011) appears well before crown-group Coleorrhyncha. The indeterminate Sternorrhyncha illustrated in Figure 3 include many relatives of Aleyrodomorpha and Coccidomorpha (stem-group Sternorrhyncha, Aphidiformes, and Psylliformes; Drohojowska *et al.*, 2020) accounting for the lack of definitive Palaeozoic representatives of these infraorders. In summary, we see stem groups appearing in the fossil record before crown groups for Hemiptera, Heteroptera, Coleorrhyncha, Sternorrhyncha, and probably Euhemiptera.

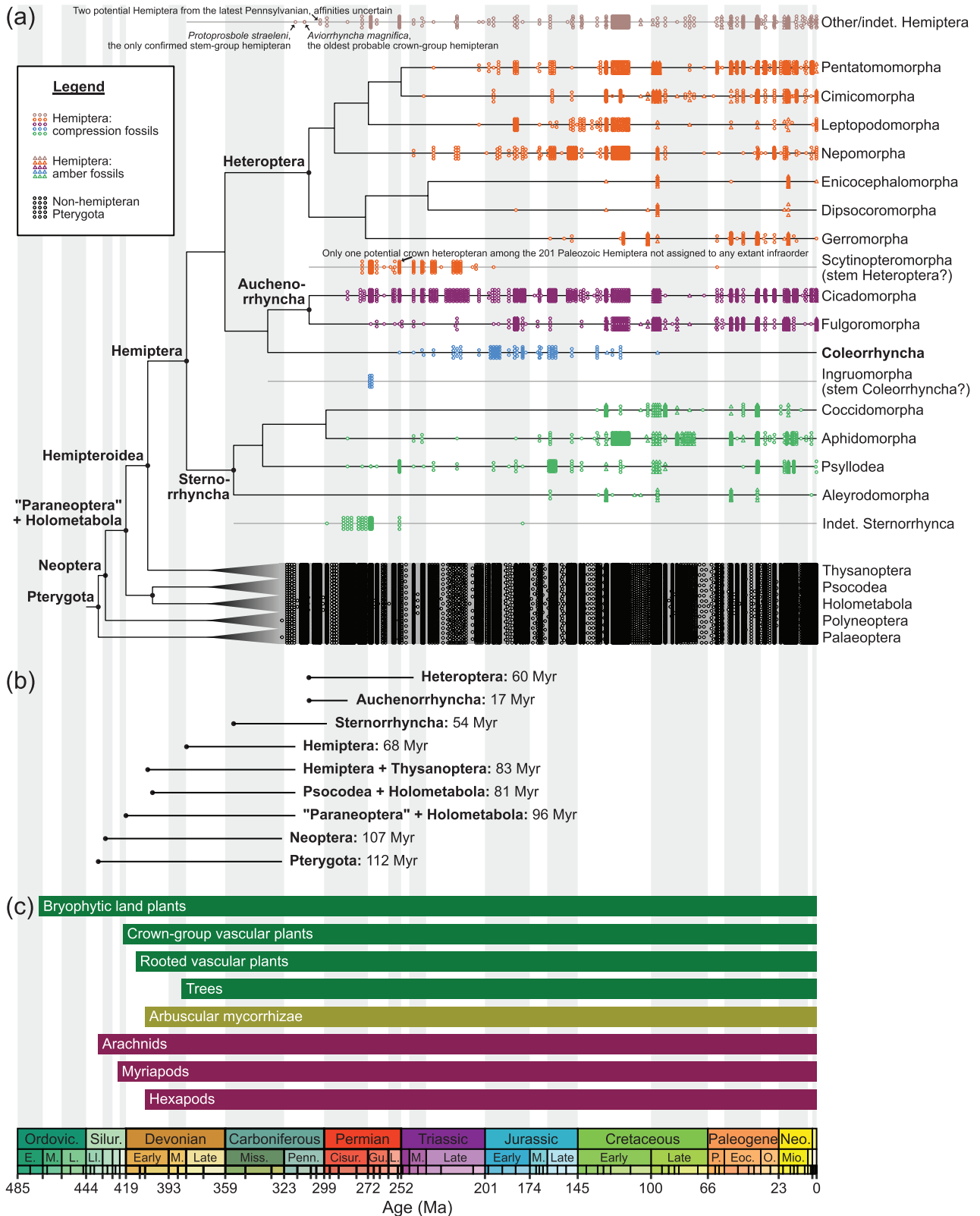
#### EMPIRICAL FACTORS AFFECTING DIVERGENCE TIME ESTIMATES

Estimating and assigning divergence times to nodes on a phylogenetic tree is deceptively straightforward (Brown & Smith, 2018), and general interactions between prior probability settings and underlying taxon sampling, intrinsic rate heterogeneity, and missing data are difficult to evaluate based on individual cases. (Of note, our discussion of dating analyses is limited to node-dating, wherein fossils are used to date divergences, rather than tip-dating, wherein fossil taxa are scored for characters and incorporated into a phylogenetic study as terminals.) We differentiate analytical input parameters such as fossil choice, prior probability settings, taxon sampling, and the extent of missing data from underlying factors intrinsic to the data under analysis, such as the extent of rate heterogeneity among loci. Beyond analytical input parameters, rate heterogeneity is perhaps the

most intractable phenomenon potentially giving rise to over-estimates of ages. Beaulieu *et al.* (2015) found that clade-specific rate heterogeneity can cause the age of a group to be overestimated by tens of millions of years. Building in part on these findings, Brown & Smith (2018) noted that the joint prior, generated by combining a branching process prior with the age distributions for fossil calibrations, may yield age estimates far older than those implied by the fossil record.

The choice of fossil calibrations clearly impacts divergence time estimates, in some cases perhaps more so than missing sequence data (Zheng & Wiens, 2015). Such impacts appear to be a function of phylogenetic position and the accuracy of a given fossil’s assignment to a stem vs. a crown group. Unfortunately, the choice of any given fossil calibration point in a node-dating analysis represents assumptions that are not themselves testable through that analysis. Even with partitioned analyses, an analysis based on fossil calibrations tests neither the wisdom of a given calibration point nor the accuracy of the fossil record writ large. As such, the fossil calibration points used to calibrate molecular phylogenetic studies warrant continuous scrutiny.

An additional factor complicates attempts to date the earliest divergences among winged insects: the paucity of fossils that can be used to calibrate deeper nodes in the hexapod tree. Few pre-Mississippian apterygote hexapods have been recognized (Whalley & Jarzembowski, 1981; Shear *et al.*, 1984; Labandeira *et al.*, 1988; Fayers & Trewin, 2005). Of these, only *Rhyniella praecursor* Hirst and Maulik 1926 can be confidently assigned at the level of order (Dunlop & Garwood, 2018). Stem-group Hexapoda, believed to have lived in aquatic habitats, have not been identified in the fossil record (Edgecombe *et al.*, 2020). The pancrustacean topology recovered by Misof *et al.* (2014) differs from topologies that have emerged more recently (Schwentner *et al.*, 2017; Giribet & Edgecombe, 2019; Lozano-Fernandez *et al.*, 2019), complicating the inclusion of various potential fossil calibrations. Misof *et al.* calibrated their tree with two non-hexapod pancrustacea: the Silurian ostracod *Pauline avibella* Siveter *et al.* 2013 and the Jurassic/Cretaceous cladoceran *Daphnia* sp. The fossil calibration compendium of Wolfe *et al.* (2016) and recent addenda include three non-hexapod pancrustaceans that can be used to calibrate Misof *et al.*’s tree as well as three non- and pre-pancrustacean fossils: the Devonian branchiopod *Lepidocaris rhyniensis* Scourfield & Calman 1926, the Silurian malacostracan *Cinerocaris magnifica* Briggs *et al.* 2004, the Ordovician/Silurian ostracod *Luprisca incuba* Siveter *et al.* 2014, the Cambrian crown-group pancrustacean *Yicaris dianensis* Zhang *et al.* 2007,



Downloaded from https://academic.oup.com/biolinnean/article/138/2/143/6936511 by guest on 25 April 2024

**Figure 3.** A timeline of hemipteran evolution. A, fossil occurrences plotted onto the time-calibrated phylogeny of [Johnson et al. \(2018\)](#). B, gaps between the origins of different groups, as inferred by [Johnson et al. \(2018\)](#), and their first appearances in the fossil record. C, ranges of relevant groups of the terrestrial biota.

the Silurian diplopod *Casiogrammus ichthyeros* Fernandez *et al.* 2018, and the Cambrian chelicerate *Wisangocaris barbarahardyae* Jago *et al.* 2016.

## METHODS

We conduct four analyses. The first compares occurrences of Palaeozoic Pterygota to those of arachnids, chilopods, and primitively wingless hexapods. The aim of this analysis is to evaluate the plausibility of a scenario in which Pterygota existed, but evaded fossilization, during the Silurian through Mississippian. The second analysis also uses fossil data, but only for Pterygota, to estimate the age for this group. The third analysis uses two fossil datasets to explore node-calibration procedures in the phylogenetic analysis of Misof *et al.* (2014): that of Wolfe *et al.* (2016) and that of Misof *et al.* itself. In these analyses, the divergence time estimates of Misof *et al.* (2014) were re-analysed using these two fossil datasets and a variety of parameter sets. The fourth analysis utilizes the phylogenetic topology recovered by Misof *et al.* (2014) to simulate the pterygote fossil record, with the aim of estimating the completeness of the fossil record by comparing various simulations to known occurrence data.

### PALAEONTOLOGICAL ANALYSES

The first two analyses, both palaeontological, were conducted with Palaeozoic data downloaded from the Paleobiology Database (PBDB; paleobiodb.org). These data were primarily compiled by Clapham *et al.* (2016). One insect occurrence was removed from the dataset: a roachoid whose possible age range listed in the PBDB spans 60 Myr (Martynov, 1936).

#### *Bootstrapping analysis*

To evaluate the plausibility of a gap in the pterygote record, and specifically to explore the probability that winged insects existed during various intervals before the Pennsylvanian, we bootstrapped arachnid and chilopod occurrence data from Palaeozoic intervals before and after the appearance of Pterygota in the fossil record. Occurrence data for Palaeozoic arachnids, chilopods, and Pterygota were downloaded from the PBDB; we verified that all Silurian–Mississippian chilopod occurrences listed in Edgecombe (2011) and arachnid genera listed in Dunlop *et al.* (2018) were included in the PBDB. We added missing occurrences of *Allobuthus pescei* Vachon and Heyler 1985, *Parageralinura neerlandica* Laurentiaux-Viera and Laurentiaux 1961, *Gondwanarachne argentinensis* Pinto and

Hunicken 1980, *Saccogulus seldeni* Dunlop *et al.* 2006, *Crussolum* sp. Shear *et al.* 1998, and *Crussolum crusserratum* Shear *et al.* 1998. The only taxon missing from the PBDB and from our dataset is the scorpion *Paraisobuthus* sp. Kjellesvig-Waering 1986 from the Pennsylvanian Mina la Mora locality of León, Spain (Gutierrez-Marco *et al.*, 2005), which we were unable to include due to insufficient locality and collection data. We downloaded final datasets from the PBDB on 10 February 2021. The midpoint between the minimum and maximum age for each specimen was assigned as its age.

The results presented here are primarily confined to the fossil records of insects and arachnids. Diplopods were not included in our bootstrapping analyses because they have mineralized exoskeletons (Plotnick, 1990), which confer a greater potential for preservation than those of most other terrestrial arthropods (Edgecombe & Legg, 2013). Arthropleureans were excluded from the dataset as well due to their proposed phylogenetic affinity with diplopods (Kraus & Brauckmann, 2003). [*Arthropleura* appears not to have had a mineralized exoskeleton (Davies *et al.*, 2022); the exclusion of all arthropleureans constitutes a conservative approach, as the resulting *P*-values would be lower if arthropleureans were included.] Only a handful of chilopod occurrences are known from the Palaeozoic, most of which are over 400 Myr old (Shear & Edgecombe, 2010; Haug & Haug, 2017) and the rest occurring at Mazon Creek (Mundel, 1979; Shear & Edgecombe, 2010) alongside many arachnids and insects.

Specimen and measurement datasets for Pterygota were downloaded for the purpose of determining which fossil occurrences were preserved with wings. For winged insects, the only specimens considered ‘winged’ are those for which a measurement of wing length is available (including wing fragments, nymphal wing pads, and structures derived from wings such as tegmina and elytra); none of these specimens belong to secondarily wingless lineages and all represent individuals that survived long enough for wings to develop sufficiently to be recognized as such. These include isolated wings that were preserved without other body parts. We chose a conservative approach, and did not include presumptive pterygote specimens with at least one body part, but no wings, measured. Such specimens might belong to secondarily wingless lineages, represent larval or nymphal pterygotes that died before their wings developed, or be fragments of a disarticulated winged insect such as an isolated mandible or femur. Ichnofossils such as insect galls on leaves were not included. For many Pterygota, measurements were not available, so it is unknown whether those individual specimens include fossilized wings.



The bootstrapping procedure was used only on contiguous intervals that (1) pre-date the Pennsylvanian, by which point winged insects had indisputably originated, and (2) encompass the Late Mississippian (Serpukhovian Stage), because winged insects are known only from the end of this interval. The bootstrapping procedure was performed for each of the three epochs/stages of the Mississippian, the entirety of the Devonian, and the entirety of the Silurian. Because 39 of the 41 Mississippian arachnid occurrences are from the Middle Mississippian, this epoch/stage was further subdivided. (The Mississippian is the only geological period discussed here for which each epoch contains only one stage, and thus epochs and stages are interchangeable.) The 'late-Middle' Mississippian includes the three arachnid localities whose midpoint ages are 331.2 Mya, only 0.2 Myr below the Middle/Late Mississippian boundary; all of these localities have Late Mississippian minimum ages. The 'mid-Middle' Mississippian includes the four arachnid localities whose midpoint ages are less than 340 Mya. Because only one arachnid specimen has been tentatively assigned to the Late Mississippian, this interval cannot be subdivided further for additional bootstrapping analyses.

The bootstrapping procedure was conducted as follows. First, the number of arachnid and chilopod occurrences during the interval was summed. Then, the same number of arachnid and chilopod occurrences was sampled with replacement from the Pennsylvanian and Permian. This procedure was repeated for 1 000 000 iterations. The resulting *P*-value represents the proportion of iterations in which none of the resampled Pennsylvanian–Permian arachnids and chilopods occur at localities that also include winged insect specimens. This *P*-value represents the probability of recovering the known fossil record of arachnids and chilopods for a given interval without sampling a single winged insect at any of the localities where the arachnids and chilopods were found. This probability is premised on the assumptions that: (1) winged insects existed during that interval; (2) winged insects were no less abundant than arachnids and chilopods; and (3) the probability of fossilization and recovery for pterygotes, relative to other terrestrial arthropods, did not change from the Silurian–Mississippian to Pennsylvanian–Permian.

### *PyRate*

Our second analysis used *PyRate* (Silvestro *et al.*, 2019) to estimate the age of Pterygota. *PyRate* uses a Bayesian framework (reversible jump Markov chain Monte Carlo) to estimate origination and extinction rates in deep time from fossil occurrence data, and the age of the clade under consideration. We used

family-level data for this analysis. We prepared ten input files, setting the seed from 1 to 10 with the `set.seed()` function in R. The use of ten replicates addresses the uncertainty associated with the sampling of random dates within the age range for each family. Each replicate input file was analysed separately, thus receiving a different random seed in python.

We used the `-qShift` option to independently estimate preservation rates for geological intervals as follows: 323.2 and 300.5 Mya mark inflection points in the rise and fall of Palaeozoic coal (Nelsen *et al.*, 2016); 130.1 Mya, after which insect inclusions in amber become prevalent in the fossil record with the appearance of Lebanese amber (Maksoud & Azar, 2020); and 2.589 Mya marks the point after which insects preserved in unlithified and poorly lithified sediments become prevalent (Böcher, 1995; Kiselev & Nazarov, 2009). We used the `-pP 1.5 0` option for the vector of preservation rates. The first value confirms the default shape prior of the gamma distribution (1.5) and the second value indicates estimation of the rate prior from the data. We analysed each replicate input file for 20 000 000 iterations, sampling every 1000 iterations.

We read the results into R with the *tracer* package (Bildrbeek & Etienne, 2018) and determined the number of iterations to discard as burn-in by iteratively calculating the effective sample size for different proportions of burn-in, using the `ess` function in the *mcmcse* package (Flegal *et al.*, 2021). Among the results that *PyRate* generates is an estimate of the root age for the clade under consideration, in this case Pterygota. As discussed below, *PyRate* does not account for variability in the completeness of the fossil record before the appearance of the fossils under consideration. *PyRate* also generates estimates of origination, extinction, and diversification rates through time, which bear on the plausibility of an early burst of evolution.

### NODE-DATING ANALYSES

The analysis of Misof *et al.* (2014) represents the most complete phylogenomic analysis of insect orders to date that employs fossil calibrations to estimate divergence times. Misof *et al.* used 37 fossil calibrations spanning chelicerates, myriapods, crustaceans, and hexapods, 18 of which represent non-apterygote arthropod lineages. We revisited the choice of fossil calibrations and prior probability assignments to examine their impacts on age estimates for Pterygota. Doubts have been raised about the use of *Rhyniognatha hirsti* as a calibration point (Haug & Haug, 2017). This species is known from a fossilized head capsule fragment from the Rhynie Chert, and pre-dates any definitive Pterygota by ~80 Myr. *R. hirsti* has been interpreted to possess

odonate affinities, placing it within Pterygota and crown-group Dicondylia (Engel & Grimaldi, 2004), and was therefore treated as a fossil calibration for Dicondylia by Misof *et al.* (2014) and as a fossil calibration for Insecta by Wolfe *et al.* (2016). However, a recent re-examination with advanced imaging techniques suggests this specimen is a myriapod (Haug & Haug, 2017). We re-analysed the divergence time estimates of Misof *et al.*, with and without *R. hirsti*, to evaluate its impact on estimated origination dates (see below) and additionally using the data of Wolfe *et al.* (2016). All analyses were run under the generally preferred lognormal prior probability settings and uniform (flat) priors to examine whether the estimates would be comparable, as well as under specific constraints based on defensible calibration points.

Relevant fossil calibrations from Wolfe *et al.* (2016) were updated as follows. *Casiogrammus ichthyeros* Wilson 2005 was used to calibrate the minimum age of Diplopoda following Fernández *et al.* (2018). The minimum age of *Oligotypus huangheensis* Ren *et al.* 2008 was updated according to the findings of Trumper *et al.* (2020). *Palaeotaeniopteryx elegans* Sharov 1961 was removed as a calibration for crown Plecoptera following Evangelista *et al.* (2019) and replaced with *Pronemoura shii* Yushuang *et al.* 2011. Of note, two insect calibrations in Wolfe *et al.* (2016) are housed in private collections, but both of the nodes calibrated with these fossils can be calibrated with other fossils from the same deposit that are housed in public museums (Mendes & Wunderlich, 2013; Chen *et al.*, 2019). A few nodes mentioned in Wolfe *et al.* (2016) can be calibrated with geologically older fossils described since 2016 (e.g. Fikacek *et al.*, 2020; Qvarnstrom *et al.*, 2021; Yang *et al.*, 2021). However, in order to compare our results to those of Misof *et al.* (2014) we chose to retain all fossils from the Wolfe *et al.* (2016) compendium believed to represent the relevant crown group.

We also examined the effect of using a maximum age for Pterygota of 331.1 Mya corresponding roughly to the maximum age of the Viséan/Serpukhovian boundary at  $330.9 \pm 0.2$  Mya (Richards, 2013), and a maximum age of 324 Mya corresponding to the age of the oldest verifiable pterygote fossils (Brauckmann & Schneider, 1996; Prokop *et al.*, 2005), which were deposited just below the Mississippian/Pennsylvanian boundary at  $323.2 \pm 0.4$  Mya (Richards, 2013). We rounded the age of this boundary up from 323.6 to 324 Mya because the pterygote fossils themselves slightly pre-date this boundary.

All sequence data were downloaded from <https://science.sciencemag.org/content/suppl/2014/11/05/346.6210.763.DC1>. We obtained 102 separate partitions from the download site.

Some of these were further sub-partitioned (e.g. p1 into p1A to p1E; p3 into p3A and p3B; p7 into p7A to p7D; p14A to p14I; p36 into p36A and p36B). In the original publication the sub-partitions for these five gene regions were combined and treated as a single partition. We have treated these data likewise, but we found that treating these sub-partitions as full partitions had no significant impact on the results of our analyses. In total there were 85 gene partitions we could obtain from the data archives for the study, ranging up to 3500 amino acids in length. Each partition was treated as described in the original publication with respect to model usage (the majority used the LG amino acid substitution matrix). However, we focused our analyses on the ages of four clades: Pterygota, Holometabola, Diptera, and Neuroptera.

Calibration dates from table S8 of Misof *et al.* (2014) are referred to in the figures as ‘Misof’. ‘Pterygote calibration time’ refers to the 331 Mya constraint and prior-setting scheme. Additional calibration dates obtained from Wolfe *et al.* (2016) (see Supporting Information, Appendix S1) are referred to as Wolfe followed by the pterygote calibration time and prior setting. For instance, MisofLogN405 indicates an analysis scheme using the Misof calibration of 405 Mya and LogN (lognormal) settings (the original setting for analysis in Misof *et al.*). Table S2 shows the analyses used for these comparisons.

XML files were produced using BEAUti from the BEAST package (Drummond & Rambaut, 2007). Partitions were analysed separately as in the original study to facilitate reasonable run times. For the Misof LogN runs we used the same prior setting as in the original study for the MisofLogN405 run. The number of generations run in each Bayesian analysis ranged from 10 million to 30 million.

All analyses were constrained with the original Misof *et al.* tree. The runs were executed using a linux-based (Centos 7) computer cluster at the AMNH which consists of 1024 hyper-threaded cores over 16 compute nodes. We used BEAST v.1.10.4, Beagle 3.1.2, and Oracle Java 8. Beagle was run with CPU resources only, multithreading was disabled with *-beagle* threading off, and scaling was set to ‘always’. The resulting .log files were combined using the LogCombiner software in the BEAST package. Individual .log files were analysed using Tracer v.1.7.1 also from the Beast package. Results were graphed using Tracer v.1.7.1 or Excel.

We used an uncorrelated relaxed clock with a relaxed lognormal distribution, replicating in part the analysis of Misof *et al.* (2014). Because of the large amount of sequence in the original dataset, partitions were analysed separately, resulting in estimates for divergence times for each partition. There are several ways that an overall estimate can be obtained,

and we chose the approach outlined by Misof *et al.* (2014), which is based on the median estimate from the distribution of individual partition estimates. We present the results of our analyses for Pterygota as well as those for Holometabola, Diptera, and Neuroptera as exemplar nodes in the tree.

To compare density distributions of the Misof and Wolfe datasets we used Tracer v.1.7.1. We loaded the combined logfiles for each of the two datasets and graphed the density distributions of user priors against lognormal posteriors.

#### SIMULATING THE FOSSIL RECORD

For our fourth analysis, a simulation procedure was used to quantify Budd & Mann's (2020) argument that the appearance of major groups in phylogenetic sequence implies that the fossil record is not so incomplete as to plausibly contain gaps of many tens of millions of years. This procedure is intended to quantify the gap lengths one can expect in the fossil record based on the sequence in which different groups appear. The tree from Misof *et al.* (2014) for all insects, truncated at the Permian/Triassic boundary, was imported into R.

For each of the three increases in the sampling rate mentioned below, the following procedure was repeated 5000 times. First, the branch lengths were randomly altered while constraining the tree as ultrametric, so that all terminal branches end at the Permian/Triassic boundary. A timescale was assigned to each tree by choosing a random multiplier that fulfilled two criteria: first, that crown-group pterygotes be at least 71 Myr older than the Permian/Triassic boundary, as observed in the fossil record; and second, that crown-group insects be no more than 200 Myr older than the Permian/Triassic boundary (which a direct reading of the fossil record, and all fossil-calibrated molecular phylogenetic studies that we are aware of, have shown). The random multiplier was applied to all branches of the phylogeny to retain ultrametricity.

The `sim.fossils.intervals()` function of the FossilSim package (Barido-Sottani *et al.*, 2019) was used to simulate a fossil record. To account for Pterygota being less abundant – and therefore less likely to be known from the fossil record – when they first originated, the sampling rate was simulated to increase linearly per geological stage by a total of 2×, 10×, and 50× from the origin of Pterygota until the Permian/Triassic boundary. Sampling rates were specified for each geological stage. The sampling rate for any stage in which insects were present but in which pterygotes had not yet originated was set to the minimum sampling rate mentioned above: 1/2, 1/10 or 1/50 of the sampling rate for the final stage of the Permian. The sampling rate for the Late Mississippian (Serpukhovian) was set to zero

to reflect the particularly depauperate record from this interval. Sampling rates did not vary by lineage. For each of the 5000 time-calibration simulations, and for each of the three protocols for increased sampling, a fossil record was simulated from the tree with the following maximum sampling rates: 0.02, 0.04, 0.06, 0.08, 0.1, 0.2, 0.3, 0.4, 0.5, 1, 1.5, 2, 2.5, 3, 3.5, 4, 4.5, 5, 5.5, 6, 6.5, 7, 7.5, 8, 8.5, 9, 9.5, and 10.

The above procedure was then repeated with an additional clustering procedure to mimic the Lagerstätte-driven nature of the insect fossil record. Using the `iNEXT` function in the `iNEXT` package in R (Hsieh *et al.*, 2016), we estimated the number of fossil assemblages that we expect per number of insect fossils. These estimates were generated with the above-mentioned Palaeozoic dataset from the PBDB. Then, while running the simulation procedure, we used the base-R function `kmeans` to cluster the fossil occurrences into the expected number of assemblages.

Each simulation was evaluated on the basis of two criteria: (1) whether Apterygota, Palaeoptera, Polyneoptera, Hemiptera, Holometabola, and Coleoptera appear and, (2) if so, whether they appear in the correct phyletic sequence (Apterygota no later than Palaeoptera and Polyneoptera; Palaeoptera and Polyneoptera no later than Hemiptera and Holometabola; and non-coleopteran Holometabola no later than Coleoptera). We also recorded the total number of insect fossils in each simulated fossil record, as well as the temporal lag between the origin of crown-group pterygotes and their appearance, the temporal lag between the first and tenth appearances of crown-group pterygotes, and the temporal lag between the first and hundredth appearances of crown-group pterygotes.

## RESULTS

### BOOTSTRAPPING ANALYSIS OF FOSSIL OCCURRENCE DATA

Our bootstrapping procedure found significant differences in the taxonomic composition of the Silurian through Early Mississippian fossil record of terrestrial arthropods, as compared to the Pennsylvanian and Permian record. Regardless of whether the threshold for significance is 0.05 or 0.005, the results of the bootstrapping procedure (Table 1) are consistent with diversification of crown-group pterygotes only after the start of the Late Mississippian, 331 Mya. Because only one arachnid specimen has been tentatively assigned to the Late Mississippian, this interval cannot be subdivided further for additional bootstrapping analyses. [The trigonotarbid *Anthracomartus* sp. (Copeland, 1957) has been tentatively assigned to the Mabou group

**Table 1.** Results of the bootstrapping procedure: the interval examined, the age range of the interval, the number of arachnids and chilopods described from the interval, and the fraction of bootstrapping iterations in which the same number of arachnids and chilopods were sampled from the Pennsylvanian and Permian without ever co-occurring with winged insects

Interval	Age (Mya)	<i>N</i>	<i>P</i>
Late Mississippian	331–323	1	0.7182
'late-Middle'–Late Mississippian	332–323	20	0.0036**
'mid-Middle'–Late Mississippian	340–323	28	0.0003***
Middle–Late Mississippian	347–323	40	< 0.0001***
Mississippian	359–323	40	< 0.0001***
Devonian–Mississippian	419–323	131	< 0.0001***
Silurian–Mississippian	444–323	151	< 0.0001***

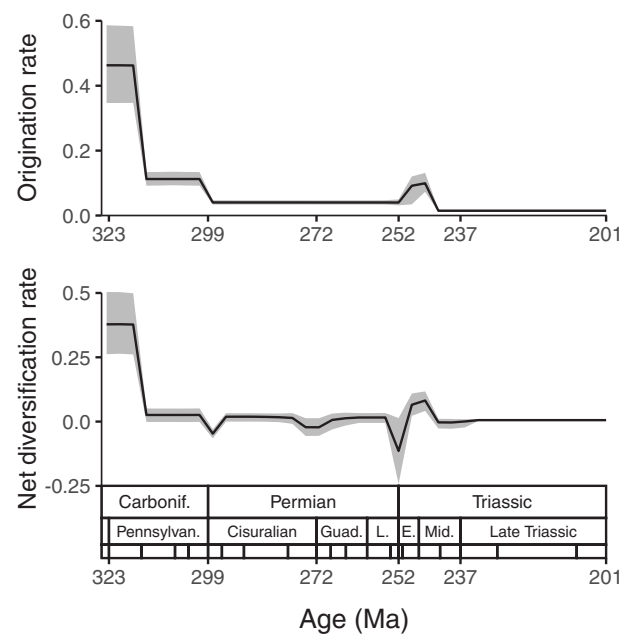
Asterisks denote \*\*\**P* < 0.001 and \*\**P* < 0.01.

(Miller & Forbes, 2001), whose Brigantian age spans the Viséan and Serpukhovian Stages with a midpoint in the Serpukhovian. Though the only option at present is to treat this occurrence of *Anthracomartus* sp. as belonging to the Mabou group, and to treat the Mabou group as dating to the Serpukhovian, neither of these designations can be made with any confidence.]. Whereas the *P*-value for the Late Mississippian is not significant (*P* = 0.72), the *P*-values for all of the longer intervals tested are significant. The results of the bootstrapping procedure imply that winged insects were absent during the Middle Mississippian (Viséan Stage), and therefore originated during the Late Mississippian (Serpukhovian Stage) – the latest interval in which they could have plausibly arisen given the presence of both Palaeoptera and Neoptera at the Mississippian/Pennsylvanian boundary.

#### PYRATE

Six of the ten replicate PyRate analyses reached convergence, or 'stationarity': i.e. estimates of the age of Pterygota remained within a narrow range of 10 Myr or less as the analysis progressed (Supporting Information, Table S1). Of these, the five with the highest effective sample sizes yielded a median age estimate of crown Pterygota between 326.09 and 326.60 Mya. The replicate with the lowest effective sample size yielded a slightly lower median age estimate for crown Pterygota, 324.96 Myr. For these six replicates, the lower bound of the 95% highest posterior density interval was above 324 Mya and the upper bound was below 330 Myr. The origination and net diversification rates, as estimated with PyRate, peaked early in the evolutionary history of Pterygota and then declined precipitously from ~317.5 to 313.4 Mya (Fig. 4).

Results of the PyRate analysis, like those of the bootstrapping analysis, suggest that Pterygota



**Figure 4.** Origination and net diversification rates for Pterygota at the family level as estimated with PyRate, from the Pennsylvanian to Triassic.

originated during the Late Mississippian – despite the two procedures using different kinds of data. The bootstrapping analysis depends entirely on the fossil records of terrestrial arthropods other than pterygotes, whereas PyRate does not account for non-ptyerygotes in any way. PyRate accounts for the family-level affinities of fossil pterygotes, and for their post-Palaeozoic fossil records, whereas the bootstrapping analysis accounts for neither. The bootstrapping analysis utilizes the co-occurrence of fossils belonging to different taxa and incorporates all fossil occurrences, whereas PyRate ignores co-occurrence/locality data, and accounts for only the first and last appearances of each family.



## NODE-DATING ANALYSES

In our re-analyses of Misof *et al.*'s node-dating, estimates of pterygote origins vary according to the choice of fossil calibration as well as the assignment of priors (lognormal vs. uniform distribution settings) and the enforcement of constraints. Although estimates based on the calibrations of Misof *et al.* were consistently older than those employing the Wolfe data, the prior settings of uniform/lognormal distributions appear to impact the results of using these two datasets in opposite ways, with lognormal settings resulting in older estimates with the Wolfe *et al.* data and the uniform settings resulting in older estimates with the Misof *et al.* data. The parsed summarized node age values for Pterygota, Neuroptera, Holometabola, and Diptera can be found in 'INSECT PUB EXCEL.xlsx'. While we focus primarily on the Pterygota node, we also summarize the Neuroptera, Holometabola, and Diptera nodes for comparison. In addition, the raw .log files contain node estimates for all nodes in the original (Misof *et al.*, 2014) tree and these are available from the authors on request.

Misof *et al.*'s estimate of 405 Mya for Pterygota is recovered using their settings (Supporting Information, Figs S2, S3). The oldest divergence time estimates (412 and 420 Mya) are obtained from applying uniformly distributed priors to the Misof data when the pterygote calibration is set at 324–450 Mya and fixed at 324 Mya, respectively. Under the Misof calibrations, uniform priors result in an older estimate for the age of Pterygota relative to lognormal priors; uniform priors increase the pterygote divergence time estimate by 15 Myr when the calibration is set at 324 Mya and by 7 Myr when set at 324–450 Myr. When no calibration is assigned (n) the estimated age of Pterygota (386 Mya) represents a reduction of 19 Myr, which is 9 Myr less than the 377 Mya obtained under lognormal prior settings. When the calibration is set at 324 Mya under lognormally distributed priors, the obtained age estimate for pterygotes (386 Mya) is 34 Myr less than when uniform priors are imposed. Using the Wolfe *et al.* calibrations, this effect is reversed and lognormal priors result in older estimates of pterygote age than do uniform priors: 368 vs. 363 Mya (a reduction of 37–42 Myr from 405 Mya) when the pterygote calibration is removed; and 358 vs. 355 Mya (a reduction of 47–50 Myr from 405 Mya) when the pterygote calibration is set at 331 Myr. The most recent estimate from the Misof *et al.* data (377 Mya) comes from removing the calibration for pterygotes entirely under lognormally distributed priors.

The Wolfe *et al.* (2016) calibrations consistently yield more recent origin times for Pterygota than the Misof compendium (Figs 5, 6) as well as shorter gaps between the inferred origin of Pterygota and the origins of Holometabola, Diptera, and Neuroptera

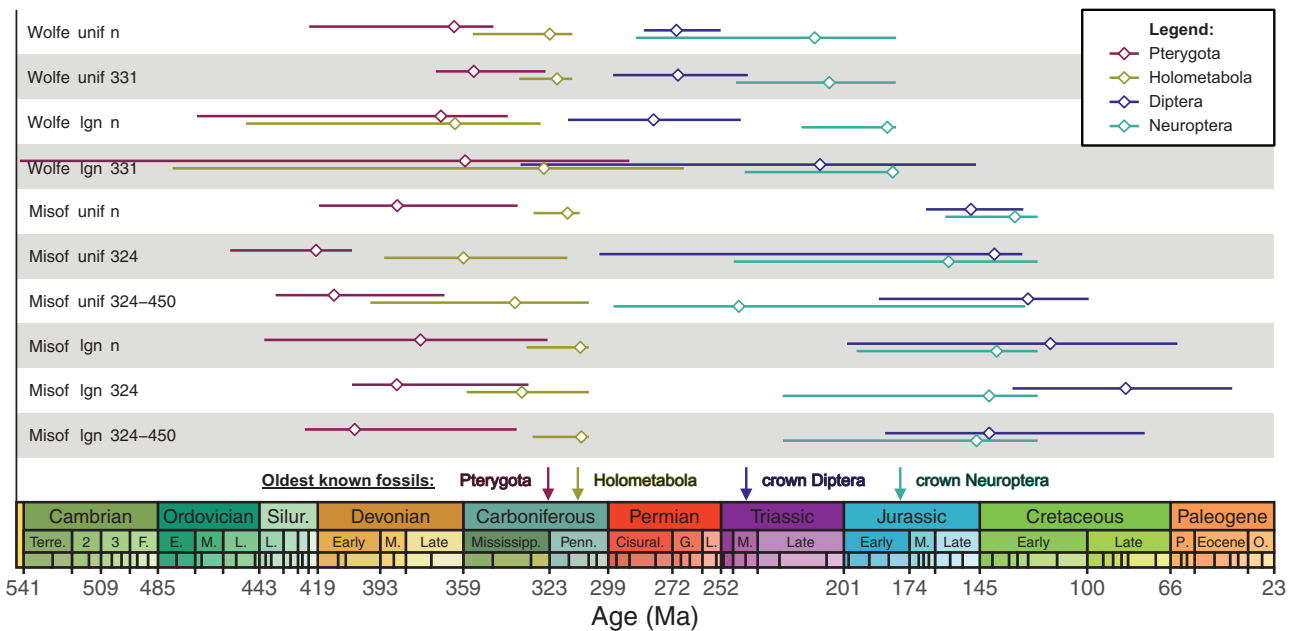
(Fig. 5). Under uniform prior settings, use of the Wolfe calibrations reduces the age estimate for Pterygota by 42–50 Myr regardless of whether the pterygote calibration is removed entirely or set at 331.1 Mya. Under lognormal prior settings, the estimate is reduced by 37–47 Myr whether the pterygote calibration is removed entirely or set at 331.1 Mya. Applying a strict clock (removing the need for priors) reduces the estimate by 23 Myr (382 Mya).

Following Brown & Smith (2018), we graphed the prior and posterior density distributions for the Pterygota node (Fig. 7). The posterior distributions were noticeably different from the prior distributions, suggesting that any discrepancies in the estimated age of pterygotes are not primarily attributable to insufficiency of molecular data to update the prior. This finding also holds for the other three nodes we examined: Holometabola, Diptera, and Neuroptera (Supporting Information, Figs S4, S5).

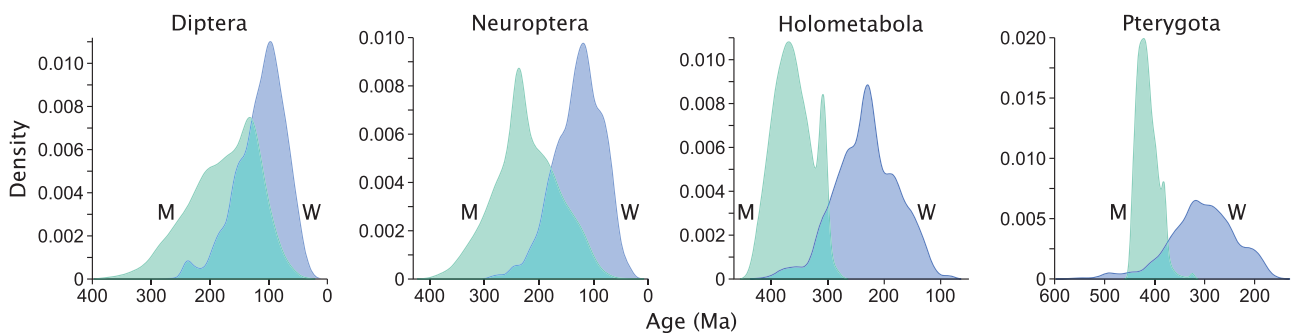
## RESULTS OF SIMULATIONS

Nearly all of our simulations of the fossil record that yielded at least 1000 Palaeozoic insect fossils share four characteristics. First, in 93.99% of such simulations, the major groups of insects (Apterygota, Palaeoptera, Polyneoptera, Hemiptera, Holometabola, and Coleoptera) appear in the fossil record in the expected phylogenetic sequence (Fig. 8A). We focus here on the simulations with 5000–6000 fossils, because there are currently around 5500 fossil pterygote occurrences described from the Palaeozoic fossil record. Among these simulations, an average of 1.07 Myr passed between the origin of pterygotes and their appearance in the fossil record; an average of 4.01 Myr passed between the appearance of the first and the tenth pterygote fossils; and an average of 20.05 Myr passed between the appearance of the first and the hundredth pterygote fossils. This is broadly similar to what is seen in the fossil record, with fewer than ten pterygotes known from the Late Mississippian (Brauckmann & Schneider, 1996; Prokop *et al.*, 2005) followed by various pterygote assemblages dating to the early Bashkirian such as Malanzán (Riek & Kukulova-Peck, 1984; Pinto, 1986), Ningxia (Gu *et al.*, 2011; Su *et al.*, 2012), and Hagen-Vorhalle (Ilger & Brauckmann, 2008; Brauckmann *et al.*, 2010).

The fossil record, as it is currently known, has a gap of ~5 Myr between the first and tenth pterygote occurrences, and a gap of ~7.5 Myr between the first and hundredth fossil occurrences. It contains ~5500 pterygote fossils from the Palaeozoic, and the major groups appear in the sequence predicted from phylogeny. The simulation results suggest that, when all these conditions hold, there is a gap of less than



**Figure 5.** Median age estimates and 95% highest posterior density intervals calculated from re-analysis of the [Misof \*et al.\* \(2014\)](#) data. Each calibration scheme is labelled with the fossil compendium used ('M', Misof; 'W', Wolfe), the prior distribution ('lgn', lognormal; 'unif', uniform), and the maximum age of Pterygota imposed as a constraint ('n' signifies no maximum age).



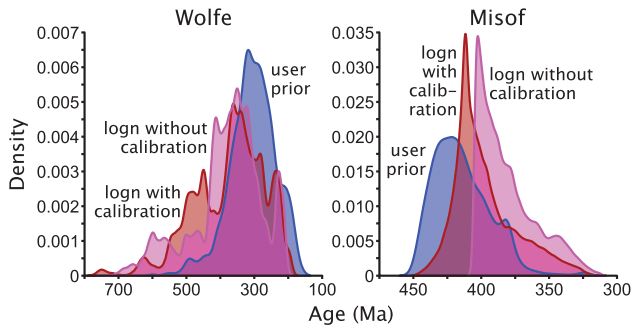
**Figure 6.** User-defined lognormal prior distributions for Wolfe (W) and Misof (M) fossil calibrations. Density is on the y-axis and divergence time is on the x-axis in all panels.

10 Myr between the origin of crown-group Pterygota and their appearance in the fossil record. (In contrast to the bootstrapping analysis discussed above, the final number of fossils is not fixed in our simulations.) These simulations suggest, however, that the fossil record is not so sparse that it fails to capture extended periods at the dawn of pterygote history (Fig. 8B–D) – especially if thousands of insect fossils are known.

## DISCUSSION

While the estimation of phylogenetic timing relies on the suitability of fossil calibration points, the

plausibility of estimates that appear to conflict with the fossil record depends on the plausibility of both their underlying assumptions and the evolutionary scenarios they require. The inference of a pterygote origin during either the Devonian ([Misof \*et al.\*, 2014](#)) or Silurian ([Montagna \*et al.\*, 2019](#)) requires that pterygotes evaded fossilization for at least 80 Myr, during which various arachnids, the occasional chilopod, and the occasional apterygote hexapod were preserved. It also requires that the mechanism(s) preventing pterygotes from becoming fossilized did not impede the detection of major pterygote groups in the sequence predicted by phylogenetic reconstruction.



**Figure 7.** Density distributions for user-defined lognormal priors, and lognormal posterior distributions for datasets with Pterygote calibration and without Pterygote calibration for the Pterygote node. Density is on the y-axis and divergence time is on the x-axis in all panels. Left panel shows the density distributions for the Wolfe calibrations and right panel shows the density distributions for the Misof calibrations.

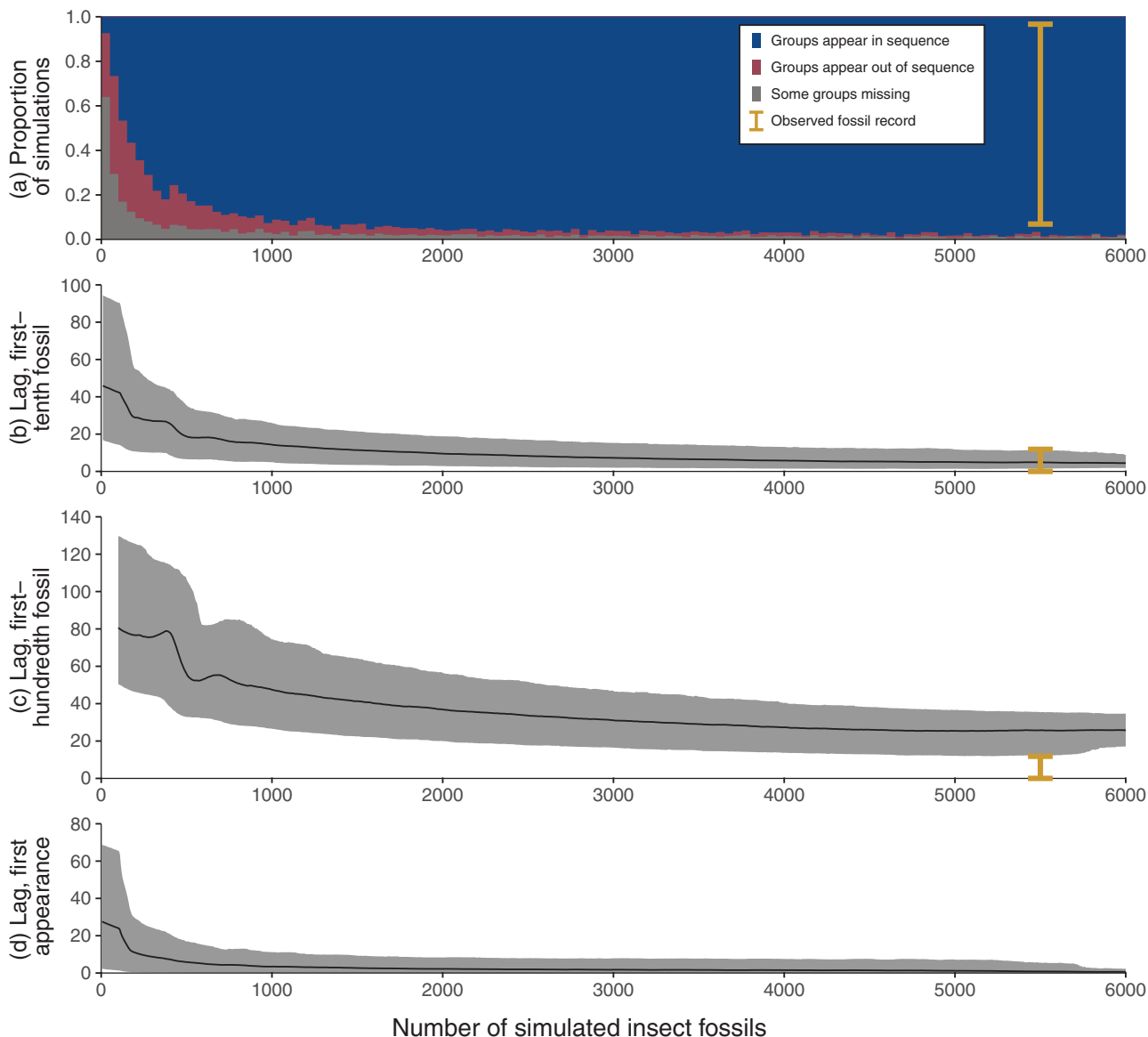
#### IMPLICATIONS FOR PTERYGOTA

The scenario in which divergences among crown-group Pterygota pre-date their known occurrence in the fossil record by tens of millions of years (Fig. 1) begs for explanation. This scenario requires that all pterygotes evaded fossilization for tens of millions of years despite having diversified considerably within this interval. None of the mechanisms by which pterygotes could have evaded fossilization for so long – small body size, low abundance, or confinement to upland habitats where the potential for fossilization was significantly reduced – could have applied to arachnids, for example, which have been found in various Silurian, Devonian, and Mississippian deposits. Furthermore, because the major groups of pterygotes appear in the fossil record in phyletic sequence, a Silurian or Devonian origin of Pterygota would require not only that major pterygote groups diverged long before their appearance in the fossil record, but also that they persisted for an extended period under the conditions that hindered their fossilization before ‘emerging’ into the fossil record in the same phyletic sequence as they evolved millions of years prior.

In the publications reporting a Silurian–Devonian origin of Pterygota, no explanation is proffered for this series of events. A proposed Silurian–Devonian origin of Pterygota raises the question of why pterygotes specifically might have been confined to upland habitats, numerically rare, or constrained to small body sizes while arachnids flourished in lowland habitats at large body sizes, for tens of millions of years. One might wonder why all suborders of Hemiptera, for example (Fig. 3), were unable to invade lowland habitats or increase in body size at the same time as Palaeoptera and Polyneoptera, despite already having existed for millions of years.

The gap between divergence dates estimated directly from the fossil record vs. from fossil-calibrated phylogenetic studies has been attributed specifically to the incompleteness of the fossil record (Barba-Montoya *et al.*, 2018; Morris *et al.*, 2018; Li *et al.*, 2019; Lozano-Fernandez *et al.*, 2020). Although the fossil record is not perfect (Holland, 2017), we are not aware of any previous efforts dedicated to quantifying precisely how long of a gap in the pterygote fossil record can be attributed to its incompleteness. Our bootstrapping analysis suggests that if Palaeozoic pterygotes were as likely to be fossilized as arachnids and chilopods, they simply were not present on the landscape during the Silurian, Devonian, and most of the Mississippian. The findings of Karr & Clapham (2015) suggest that Palaeozoic pterygotes would have been at least as readily fossilized as arachnids and chilopods: whereas pterygote heads, thoraxes, abdomens, and legs probably have preservation potentials comparable to those of equivalent structures in arachnids and chilopods, wings are by far the most common pterygote body part in the pre-amber fossil record.

The exceptional sparsity of the Late Mississippian fossil record of terrestrial arthropods in particular cautions against overinterpretation of the PyRate results. Although none of the 95% credible intervals extend as far back as the earliest 1 Myr of the Late Mississippian (329.9–330.9 Mya), the fossil record from that interval is not complete enough to preclude the possibility that crown-group pterygotes originated during that time. However, especially when considered together, the PyRate and bootstrapping analyses cast doubt upon the possibility that crown-group Pterygota would have originated during the Silurian or Devonian. Neither PyRate nor the simulation analysis make any assumptions about whether pterygotes fossilize as readily as arachnids and chilopods. The narrow 95% highest posterior density interval for the age of crown-group pterygotes obtained from the PyRate analysis falls almost entirely within the second half of the Late Mississippian (Serpukhovian). Because no fossil arachnids or chilopods are known from the first half of the Late Mississippian, there is little evidence to support the notion that crown-group pterygotes post-date the midpoint of this interval. However, because PyRate and the bootstrapping analysis rely on different data and assumptions to arrive at the same conclusion – a Late Mississippian origin of crown-group pterygotes – fossil evidence does not corroborate an origin for crown-group pterygotes older than the late-Middle to Late Mississippian. This conclusion is also supported by the simulation exercise which suggests that, because many thousands of Palaeozoic pterygotes are known from the fossil record and because the major groups appeared in the sequence suggested by their phylogenetic relationships, there



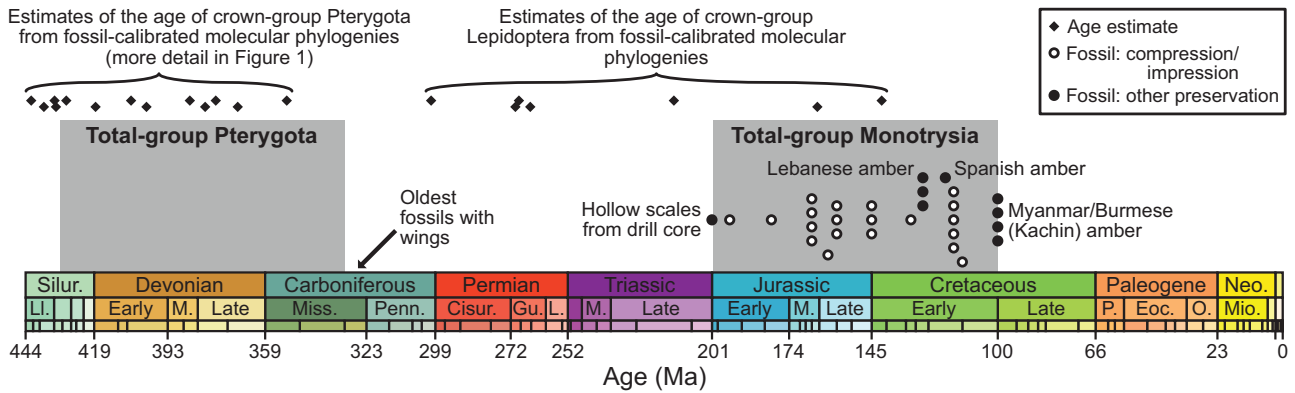
**Figure 8.** Results of the fossil simulation procedure. A, proportion of simulations in which the total-groups of Apterygota, Palaeoptera, Polyneoptera, Hemiptera, Holometabola, and Coleoptera did not appear in the simulated Palaeozoic insect fossil records, the proportion of simulations in which these groups did not appear in the expected phylogenetic sequence, and the proportion of simulations in which these groups did appear in the expected phylogenetic sequence. B, temporal lag (Myr) between the first and tenth appearances of crown-group Pterygota in the fossil record. C, temporal lag (Myr) between the first and hundredth appearances of crown-group Pterygota in the fossil record. D, temporal lag (Myr) between the origin of crown-group Pterygota and their appearance in the fossil record. Panels B–D show mean values and 95% confidence intervals for bins that contain 20 or more simulated records within a given category. For the sake of brevity, here we present only one of six versions of this analysis: that in which the initial sampling rate is 10% of the final sampling rate at the Permian/Triassic boundary, and in which occurrences were not clustered into Lagerstätten. This same illustration is available in the [Supporting Information](#) for the other five versions of the analysis (Figs S6–S10).

would have been a lag of only a few millions of years between the origin of crown-group pterygotes and their appearance in the fossil record.

A comparison with Lepidoptera (Fig. 9) further highlights the implausibility of a scenario in which

crown Pterygota diversified, but evaded fossilization, for an extended period of their early history. Lepidoptera appear sporadically in the fossil record (Sohn *et al.*, 2015). The Jurassic and Early Cretaceous span 101 Myr, post-dating widely accepted evidence





**Figure 9.** Comparison of the fossil records of total-group Pterygota and total-group Monotryisia (Lepidoptera). The grey rectangles span 100 Myr each. The grey rectangle spanning much of the Silurian to Middle Mississippian represents an interval in which Pterygota were already diversifying, according to some of the studies cited in Figure 1, but no wings are known from the fossil record. The second grey rectangle spans the first 100 Myr of the known fossil record of total-group Lepidoptera, which contains only Monotryisia. Compression/impression fossil occurrences are represented by white circles, and non-compression/impression fossils are represented by black circles and are labelled. Data for Lepidoptera are from Sohn *et al.* (2012); Necrotauliidae and *incertae sedis* taxa are not shown. Estimated ages for crown-group Lepidoptera are extracted, from oldest to youngest, from Kawahara *et al.* (2019), Montagna *et al.* (2019), Rainford *et al.* (2014), Tong *et al.* (2015), Wahlberg *et al.* (2013), Thomas *et al.* (2020), and Misof *et al.* (2014).

of crown Lepidoptera (Whalley, 1986; van Eldijk *et al.*, 2018). Although the angiosperms that have been largely credited with facilitating the diversification of this order (Grimaldi, 1999) were rare if not absent throughout this time (Coiro *et al.*, 2019), there is unambiguous evidence of monotryisian (basal) Lepidoptera throughout the entire interval (Kozlov, 1989; Ansonge, 2002) amounting to well over a dozen compression/impression fossils, plus amber and scales from a drill core (Zhang *et al.*, 2013; Sohn *et al.*, 2015).

The sparsity of the lepidopteran fossil record is to be expected for a few reasons. First, the scales on the wings of Lepidoptera are hydrophobic (Kim *et al.*, 2020), causing the wings to float and thus preventing them from sinking into a depositional environment where they can be preserved as fossils. Second, the most ancient families of Lepidoptera – the Monotryisia, which comprise the entire known lepidopteran fossil record until the deposition of Myanmar amber (Sohn *et al.*, 2015) – account for less than 2% of all lepidopteran diversity today (Nieuwerkerken *et al.*, 2011), with no evidence to suggest that they were previously more diverse or abundant than they are currently. In other words, even though there are plausible mechanisms that account for the poor lepidopteran fossil record, we still see an average of one compression/impression fossil lepidopteran per 5 Myr during the first ~100 Myr of their known history. In contrast, we see no fossil Pterygota during the > 100-Myr interval of the mid-Silurian to mid-Late Mississippian in which various publications (Rota-Stabelli *et al.*, 2013; Tong *et al.*, 2015; Johnson *et al.*, 2018; Montagna *et al.*, 2019) found Pterygota to have diversified.

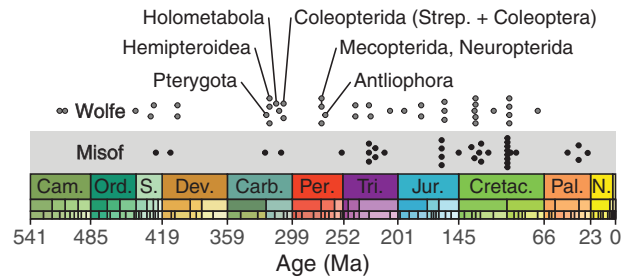
None of the mechanisms we find that could explain the near-absence of Lepidoptera from the fossil record during their early evolutionary history apply to Pterygota or to Hemiptera. The earliest crown Pterygota possessed one of the two features credited with this group's diversity and abundance, namely wings (Nicholson *et al.*, 2014). In contrast, Monotryisia lack most of the features credited with the diversification and abundance of Lepidoptera: female gonopore separate from the copulatory orifice, tympana, and, in the case of the non-glossotan moths that dominate the Jurassic–Early Cretaceous record, a siphoning proboscis with intrinsic musculature, and a wing coupling mechanism (Grimaldi & Engel, 2005). The complete absence of fossil Pterygota and Hemiptera from tens of millions of years of their purported evolutionary history, therefore appears even more implausible in light of the presence of Monotryisia in a number of Jurassic and Early Cretaceous deposits (Zhang *et al.*, 2013; Sohn *et al.*, 2015).

#### POTENTIAL CAUSES OF DISCORDANT AGE ESTIMATES

The lack of evidence for Devonian to mid-Mississippian pterygotes raises the question of why the age estimates generated in many recent studies (illustrated in Fig. 1) are so old. We examined the choice of specific fossil calibrations, and in the course of using multiple prior probability settings we identified dataset-specific behaviours under different priors, which we differentiate from intrinsic attributes of the sequence data, such as the distribution of missing

data, and rate variation among specific clades, which we did not explore. In our analyses, eliminating the controversial calibration point of *Rhyniognatha* results in younger age estimates, as did the systemic substitution of the Wolfe *et al.* (2016) calibration data. As one might expect, the Wolfe data responded to the maximum age constraint for Pterygota by yielding more recent age estimates (Fig. 5). The Misof data counterintuitively produced older dates when a maximum age constraint was imposed for Pterygota. Moreover, the Wolfe data consistently yielded an origin of crown-group Neuroptera prior to the oldest definitive fossil representative of this group, as well as for Diptera in three out of four node-dating analyses. The Misof data, in contrast, always yielded estimates for both groups that post-date their oldest known fossil representatives.

Although substituting the Wolfe data consistently retrieved more recent ages, we noted peripherally that the impact of choosing lognormal relative to uniform prior settings was reversed under alternative sets of fossil calibrations: lognormal prior settings yielded older estimates than uniform prior settings based on the Wolfe *et al.* (2016) calibration data, but older estimates based on the original Misof *et al.* (2014) calibration data obtained from uniform prior settings. We interpret this as a function of how the priors interact with distributions of the input fossil calibrations, and specifically how the uniform prior setting may exaggerate possible disparities in the sampling of those points. The Misof calibration points are less uniformly distributed than those of Wolfe, and specifically they are more heavily concentrated away from the time period when Pterygota could have originated such that there are effectively twice as many data points in the critical range of the Wolfe dataset than in the Misof dataset. Of 36 Wolfe calibration points above 100 Mya, 20 (55%) are in the range 200–400 Mya vs. five of 22 (23%) of those of Misof; and 13 of 22 (60%) of Misof's calibrations above 100 Mya are under 200 Mya vs. 16 of 36 (38%) of Wolfe's (Fig. 10). Counter-intuitively, the dataset more skewed towards younger calibration points may have yielded artificially old divergence time estimates that appear even more pronounced under uniform prior settings. When Montagna *et al.* (2019) re-calibrated the Misof *et al.* phylogeny by adding eight additional mid-Triassic (240 Mya) fossil calibrations, they recovered an even greater age for Pterygota, pushing the estimate back by 27 Myr, from 406.5 to 433.5 Mya. [Montagna *et al.* explained this result with the idea that insect wings originated during an interval of high  $pO_2$  (which facilitates flight; Dudley, 2000), and recent reconstructions of Palaeozoic  $pO_2$  tentatively identified a peak from ~440 to 410 Mya (Schachat *et al.*, 2018)].



**Figure 10.** Ages of relevant fossil calibrations from the Misof *et al.* (2014) and Wolfe *et al.* (2016) compendia. Palaeozoic fossils from the Wolfe *et al.* (2016) compendium used to calibrate supraordinal pterygote clades are labelled.

Whereas relaxed-clock methods are increasingly able to accommodate evolutionary rates that vary among lineages and among regions of the genome ('site' and 'lineage' effects), temporal variability in evolutionary rates ('epoch effects' *sensu* Lee & Ho, 2016) remains far more difficult if not impossible to detect, much less control for (Ho, 2020). Epoch effects are especially intractable because they do not necessarily cause patterns of genomic diversity that violate the null predictions based on assumptions of rate homogeneity (Lee & Ho, 2016).

Beaulieu *et al.* (2015) suggest that the inability to accommodate rate heterogeneity accurately has resulted in overestimates of divergence times among angiosperms, specifically by assuming rate homogeneity for groups that underwent high rates of diversification and molecular evolution early in their evolutionary history followed by a decrease in both rates – something plausible but inherently untestable. Budd & Mann (2020) suggested that hyperdiverse groups may be inherently vulnerable to age overestimates based on certain analyses. Drawing on insight from birth–death modelling, they suggested that, because many groups become extinct within tens to hundreds of millions of years of their origin, an early burst of diversification may account for the persistence of many hyperdiverse groups throughout the Phanerozoic (Phillimore & Price, 2008). (Whereas high rates of molecular evolution and high rates of net speciation are distinct phenomena, various authors have found them to be correlated; Duchene *et al.*, 2017; Hua & Bromham, 2017; Bromham, 2020.)

In the absence of molecular or palaeontological tests for epoch effects, the increased rates of molecular evolution that may accompany heightened rates of net diversification may predispose current methods for using fossil-calibrated phylogenetic data to overestimating ages of groups that experienced an early burst of evolution. Consequently, despite the increasing evidence of the ubiquity of initial bursts of diversification (McPeck, 2008; Phillimore & Price,

2008; Morlon *et al.*, 2010; Moen & Morlon, 2014), these cannot be inferred, much less pinpointed, unless the fossil record of a particular group is complete enough for a tree prior to be generated with an epoch-specific birth–death branching process (May *et al.*, 2021).

## CONCLUSIONS

Winged insects are hardly the only group for which age estimates drastically pre-date their appearance in the fossil record. However, the discrepancy between the fossil record and various recent fossil-calibrated phylogenetic studies is particularly noteworthy in the case of winged insects for at least two reasons. First is the length of the purported gap in their fossil record: 100 Myr is nearly a third of the duration of the known pterygote fossil record. Second, pterygotes overwhelm the fossil record of terrestrial arthropods as soon as they appear. One might expect the first winged insects to have been rare on the landscape briefly, but an extended fossil gap is less plausible given the transformative nature of insect flight, their high abundance following their appearance in the fossil record, and the available fossil data from related insect and arthropod groups. Because the potential explanations for winged insects' absence from the early- to mid-Palaeozoic fossil record (minute body size, exclusion from lowland habitats) are no less applicable to other terrestrial arthropods, we suggest that the absence of Pterygota from Silurian–Mississippian deposits that contain arachnids, centipedes, Entognatha, and Apterygota constitutes meaningful evidence of their actual absence prior to the Carboniferous. The phylogenetic recalibration exercises presented here suggest that the discordance among age estimates for Pterygota may become less severe as fossil calibration datasets improve.

## ACKNOWLEDGEMENTS

We thank Russell Garwood for comments that greatly improved the manuscript. This contribution was made possible by contributors to the Paleobiology Database. We thank Joanna Wolfe, Andrew Leslie, Mike D'Antonio, and Pulkit Singh for valuable discussion. S.R.S. was funded by the Chair's Fellowship, Harriet Benson Fellowship, and Coleman F. Fung Graduate Research Fellowship at Stanford University. Mention of trade names or commercial products in this publication is solely for the purpose of providing specific information and does not imply recommendation or endorsement by the USDA; USDA is an equal opportunity provider and employer. We have no conflicts of interest to declare.

## DATA AVAILABILITY

All data used as input for our analyses are publicly available from the Paleobiology Database (paleobiodb.org) and from the supplemental information of Misof *et al.* (2014). The raw log files from the BEAST and PyRate analyses are available upon request.

## REFERENCES

- Agnus A. 1904.** *Palaeoblattina douvillei*, considéré d'abord comme un insecte est une pointe générale de trilobite. *Comptes Rendus de l'Académie des Sciences, Serie Generale, la Vie des Sciences* **138**: 398.
- Ansorge J. 2002.** Revision of the 'Trichoptera' described by Geinitz and Handlirsch from the Lower Toarcian of Dobbertin (Germany) based on new material. *Proceedings of the 10th International Symposium on Trichoptera - Nova Supplementa Entomologica* **15**: 55–74.
- Barba-Montoya J, Reis M, Schneider H, Donoghue PCJ, Yang Z. 2018.** Constraining uncertainty in the timescale of angiosperm evolution and the veracity of a Cretaceous Terrestrial Revolution. *New Phytologist* **218**: 819–834.
- Barido-Sottani J, Pett W, O'Reilly JE, Warnock RCM. 2019.** FossilSim: an R package for simulating fossil occurrence data under mechanistic models of preservation and recovery. *Methods in Ecology and Evolution* **10**: 835–840.
- Beaulieu JM, O'Meara BC, Crane P, Donoghue MJ. 2015.** Heterogeneous rates of molecular evolution and diversification could explain the Triassic age estimate for angiosperms. *Systematic Biology* **64**: 869–878.
- Beck RMD, Lee MSY. 2014.** Ancient dates or accelerated rates? Morphological clocks and the antiquity of placental mammals. *Proceedings of the Royal Society B: Biological Sciences* **281**: 20141278.
- Bilderbeek RJC, Etienne RS. 2018.** babette: BEAUti 2, BEAST2 and tracer for R. *Methods in Ecology and Evolution* **9**: 2034–2040.
- Böcher J. 1995.** *Palaeoentomology of the Kap København Formation, a Plio-Pleistocene sequence in Peary Land, North Greenland*. Copenhagen: Museum Tusulanum Press.
- Brauckmann C, Gröning E, Ilger JM. 2010.** Von den ältesten Insekten [On the most ancient insects]. *Entomologie heute* **22**: 17–40.
- Brauckmann C, Schneider J. 1996.** Ein unter-karbonisches Insekt aus dem Raum Bitterfeld/Delitzsch (Pterygota, Arnsbergium, Deutschland). A Lower Carboniferous insect from the Bitterfeld/Delitzsch area (Pterygota, Arnsbergian, Germany). *Neues Jahrbuch für Geologie und Paläontologie-Monatshefte* **1996**: 17–30.
- Briggs DEG, Sutton MD, Siveter DJ, Siveter DJ. 2004.** A new phyllocarid (Crustacea: Malacostraca) from the Silurian Fossil-Lagerstätte of Herefordshire, UK. *Proceedings of the Royal Society of London. Series B: Biological Sciences* **271**: 131–138.



- Bromham L.** 2020. Causes of variation in the rate of molecular evolution. In: Ho SYW, ed. *The molecular evolutionary clock: theory and practice*. Cham: Springer International Publishing, 45–64.
- Brown JW, Smith SA.** 2018. The past sure is tense: on interpreting phylogenetic divergence time estimates. *Systematic Biology* **67**: 340–353.
- Budd GE, Mann RP.** 2020. Survival and selection biases in early animal evolution and a source of systematic overestimation in molecular clocks. *Interface Focus* **10**: 20190110.
- Chen M, Strömberg CAE, Wilson GP.** 2019. Assembly of modern mammal community structure driven by Late Cretaceous dental evolution, rise of flowering plants, and dinosaur demise. *Proceedings of the National Academy of Sciences of the United States of America* **116**: 9931–9940.
- Clapham ME, Karr JA.** 2012. Environmental and biotic controls on the evolutionary history of insect body size. *Proceedings of the National Academy of Sciences of the United States of America* **109**: 10927–10930.
- Clapham ME, Karr JA, Nicholson DB, Ross AJ, Mayhew PJ.** 2016. Ancient origin of high taxonomic richness among insects. *Proceedings of the Royal Society B: Biological Sciences* **283**: 20152476.
- Coiro M, Doyle JA, Hilton J.** 2019. How deep is the conflict between molecular and fossil evidence on the age of angiosperms? *New Phytologist* **223**: 83–99.
- Copeland MJ.** 1957. The arthropod fauna of the Upper Carboniferous rocks of the Maritime Provinces. *Geological Survey of Canada Memoir* **286**: 1–110.
- Craig JM, Kumar S, Hedges SB.** 2022. Limitations of phylogenomic data can drive inferred speciation rate shifts. *Molecular Biology and Evolution* **39**: msac038.
- Davies NS, Garwood RJ, McMahon WJ, Schneider JW, Shillito AP.** 2022. The largest arthropod in Earth history: insights from newly discovered *Arthropleura* remains (Serpukhovian Stainmore Formation, Northumberland, England). *Journal of the Geological Society* **179**: jgs2021–jgs2115.
- De Baets K, Dentzien-Dias P, Harrison GWM, Littlewood DTJ, Parry LA.** 2021. Fossil constraints on the timescale of parasitic Helminth evolution. In: De Baets K, Huntley JW, eds. *The evolution and fossil record of parasitism: identification and macroevolution of parasites*. Cham: Springer International Publishing, 231–271.
- Delclòs X, Peñalver E, Arillo A, Engel MS, Nel A, Azar D, Ross A.** 2016. New mantises (Insecta: Mantodea) in Cretaceous ambers from Lebanon, Spain, and Myanmar. *Cretaceous Research* **60**: 91–108.
- Dos Reis M, Thawornwattana Y, Angelis K, Telford MJ, Donoghue PC, Yang Z.** 2015. Uncertainty in the timing of origin of animals and the limits of precision in molecular timescales. *Current Biology* **25**: 2939–2950.
- Doyle JA.** 2012. Molecular and fossil evidence on the origin of angiosperms. *Annual Review of Earth and Planetary Sciences* **40**: 301–326.
- Drohojowska J, Szewo J, Żyła D, Huang DY, Müller P.** 2020. Fossils reshape the Sternorrhyncha evolutionary tree (Insecta, Hemiptera). *Scientific Reports* **10**: 11390.
- Drummond AJ, Rambaut A.** 2007. BEAST: Bayesian evolutionary analysis by sampling trees. *BMC Evolutionary Biology* **7**: 214.
- Duchêne DA, Hua X, Bromham L.** 2017. Phylogenetic estimates of diversification rate are affected by molecular rate variation. *Journal of Evolutionary Biology* **30**: 1884–1897.
- Dudley R.** 2000. *The biomechanics of insect flight*. Princeton, NJ: Princeton University Press.
- Dunlop JA.** 2019. Miniaturisation in Chelicerata. *Arthropod Structure & Development* **48**: 20–34.
- Dunlop JA, Fayers SR, Hass H, Kerp H.** 2006. A new arthropod from the early Devonian Rhynie chert, Aberdeenshire (Scotland), with a remarkable filtering device in the mouthparts. *Paläontologische Zeitschrift* **80**: 296–306.
- Dunlop JA, Garwood RJ.** 2018. Terrestrial invertebrates in the Rhynie chert ecosystem. *Philosophical Transactions of the Royal Society B: Biological Sciences* **373**: 20160493.
- Dunlop JA, Penney D, Jekel D.** 2018. *A summary list of fossil spiders and their relatives*. Bern: Natural History Museum Bern.
- Durden CJ.** 1975. *A dasyleptid from the Permian of Kansas, Lepidodasypus sharovi n. gen., n. sp. (Insecta: Thysanura: Monura)*. Austin: Texas Memorial Museum, The University of Texas at Austin.
- Edgecombe GD.** 2011. Chilopoda–fossil history. In: Minelli A, ed. *Treatise on zoology-anatomy, taxonomy, biology. The Myriapoda*. Leiden: Brill, 355–361.
- Edgecombe GD, Legg DA.** 2013. The arthropod fossil record. In: Minelli A, Boxshall G, Fusco G, eds. *Arthropod biology and evolution: molecules, development, morphology*. Berlin: Springer, 393–415.
- Edgecombe GD, Strullu-Derrien C, Góral T, Hetherington AJ, Thompson C, Koch M.** 2020. Aquatic stem group myriapods close a gap between molecular divergence dates and the terrestrial fossil record. *Proceedings of the National Academy of Sciences* **112**: 8966–8972.
- Engel MS.** 2004. The alderflies of Kansas (Megaloptera: Sialidae). *Transactions of the Kansas Academy of Science* **107**: 119–125.
- Engel MS.** 2009. A new Lower Permian bristletail from the Wellington Formation in Kansas (Archaeognatha: Dasyleptidae). *Transactions of the Kansas Academy of Science* **112**: 40–44.
- Engel MS, Grimaldi DA.** 2004. New light shed on the oldest insect. *Nature* **427**: 627–630.
- Engel MS, Grimaldi DA, Krishna K.** 2009. Termites (Isoptera): their phylogeny, classification, and rise to ecological dominance. *American Museum Novitates* **2009**: 1–27.
- Evangelista DA, Wipfler B, Béthoux O, Donath A, Fujita M, Kohli MK, Legendre F, Liu S, Machida R, Misof B, Peters RS, Podsiadlowski L, Rust J, Schuette K, Tollenaar W, Ware JL, Wappler T, Zhou X, Meusemann K, Simon S.** 2019. An integrative phylogenomic approach illuminates the evolutionary history of cockroaches and termites (Blattodea). *Proceedings of the Royal Society B: Biological Sciences* **286**: 20182076.



- Fayers SR, Trewin NH. 2005.** A hexapod from the early Devonian Windyfield chert, Rhynie, Scotland. *Paleontology* **48**: 1117–1130.
- Fernández R, Edgecombe GD, Giribet G. 2018.** Phylogenomics illuminates the backbone of the Myriapoda Tree of Life and reconciles morphological and molecular phylogenies. *Scientific Reports* **8**: 83.
- Fikáček M, Beutel RG, Cai C, Lawrence JF, Newton AF, Solodovnikov A, Šlipiński A, Thayer MK, Yamamoto S. 2020.** Reliable placement of beetle fossils via phylogenetic analyses – Triassic *Leehermania* as a case study (Staphylinidae or Myxophaga?). *Systematic Entomology* **45**: 175–187.
- Flegel JM, Hughes J, Vats D, Dai N, Gupta K, Maji U, Vats MD. 2021.** Package ‘mcmcse’. <https://search.r-project.org/CRAN/refmans/mcmcse/html/mcmcse-package.html>.
- Foote M, Hunter JP, Janis CM, Sepkoski JJJ. 1999.** Evolutionary and preservational constraints on origins of biologic groups: divergence times of eutherian mammals. *Science* **283**: 1310–1314.
- Foote M, Sepkoski JJ. 1999.** Absolute measures of the completeness of the fossil record. *Nature* **398**: 415–417.
- Gardner JD, Surya K, Organ CL. 2019.** Early tetrapodomorph biogeography: controlling for fossil record bias in macroevolutionary analyses. *Comptes Rendus Palevol* **18**: 699–709.
- Garroute R, Clément G, Nel P, Engel MS, Grandcolas P, D’Haese C, Lagebro L, Denayer J, Gueriau P, Lafaite P, Olive S, Prestianni C, Nel A. 2012.** A complete insect from the Late Devonian period. *Nature* **488**: 82–85.
- Gaunt MW, Miles MA. 2002.** An insect molecular clock dates the origin of the insects and accords with palaeontological and biogeographic landmarks. *Molecular Biology and Evolution* **19**: 748–761.
- Giribet G, Edgecombe GD. 2019.** The phylogeny and evolutionary history of arthropods. *Current Biology* **29**: R592–R602.
- Grimaldi D. 1999.** The co-radiations of pollinating insects and angiosperms in the Cretaceous. *Annals of the Missouri Botanical Garden* **18**: 373–406.
- Grimaldi D, Engel MS. 2005.** *Evolution of the insects*. New York: Cambridge University Press.
- Gu JJ, Béthoux O, Ren D. 2011.** *Longzhua loculata* n. gen. n. sp., one of the most completely documented Pennsylvanian Archaeorthoptera (Insecta; Ningxia, China). *Journal of Paleontology* **85**: 303–314.
- Gutiérrez-Marco JC, Tetlie OE, Arillo A, Rábano I. 2005.** El escorpión más antiguo de la Península Ibérica: *Parisobuthus* sp., del Estefaniense de la cuenca de Villablino (León). *Jornadas de Paleontología* **22**: 127–129.
- Han J, Morris SC, Ou Q, Shu D, Huang H. 2017.** Meiofaunal deuterostomes from the basal Cambrian of Shaanxi (China). *Nature* **542**: 228–231.
- Harper DAT, Topper TP, Cascales-Miñana B, Servais T, Zhang YD, Ahlberg P. 2019.** The Furongian (late Cambrian) Biodiversity Gap: real or apparent? *Palaeoworld* **28**: 4–12.
- Haug C, Haug JT. 2017.** The presumed oldest flying insect: more likely a myriapod? *PeerJ* **5**: e3402.
- Haug JT, Labandeira CC, Santiago-Blay JA, Haug C, Brown S. 2015.** Life habits, hox genes, and affinities of a 311 million-year-old holometabolon larva. *BMC Evolutionary Biology* **15**: 208.
- Ho SYW. 2020.** The molecular clock and evolutionary rates across the tree of life. In: Ho SYW, ed. *The molecular evolutionary clock: theory and practice*. Cham: Springer International Publishing, 3–23.
- Holland SM. 2017.** Structure, not bias. *Journal of Paleontology* **91**: 1315–1317.
- Hörschemeyer T, Haug JT, Béthoux O, Beutel RG, Charbonnier S, Hegna TA, Koch M, Rust J, Wedmann S, Bradler S, Willmann R. 2013.** Is *Strudiella* a Devonian insect? *Nature* **494**: E3–4; discussion E4–5.
- Hsieh TC, Ma KH, Chao A. 2016.** iNEXT: an R package for rarefaction and extrapolation of species diversity (Hill numbers). *Methods in Ecology and Evolution* **7**: 1451–1456.
- Hua X, Bromham L. 2017.** Darwinism for the genomic age: connecting mutation to diversification. *Frontiers in Genetics* **8**: 12.
- Huang DY, Nel A. 2009.** Oldest web-spinners from the Middle Jurassic of Inner Mongolia, China (Insecta: Embioidea). *Zoological Journal of the Linnean Society* **156**: 889–895.
- Ilger JM, Brauckmann C. 2008.** Evidence for prothoracic winglets in *Kochopteron hoffmannorum* from Hagen-Vorhalle (‘basal Neoptera’; early Late Carboniferous; Germany). *Entomologia Generalis* **31**: 279–286.
- Ishiwata K, Sasaki G, Ogawa J, Miyata T, Su Z. 2011.** Phylogenetic relationships among insect orders based on three nuclear protein-coding gene sequences. *Molecular Phylogenetics and Evolution* **58**: 169–180.
- Jago JB, García-Bellido DC, Gehling JG. 2016.** An early Cambrian chelicerate from the Emu Bay Shale, South Australia. *Palaeontology* **59**: 549–562.
- Johnson KP, Dietrich CH, Friedrich F, Beutel RG, Wipfler B, Peters RS, Allen JM, Petersen M, Donath A, Walden KKO, Kozlov AM, Podsiadlowski L, Mayer C, Meusemann K, Vasilikopoulos A, Waterhouse RM, Cameron SL, Weirauch C, Swanson DR, Percy DM, Hardy NB, Terry I, Liu S, Zhou X, Misof B, Robertson HM, Yoshizawa K. 2018.** Phylogenomics and the evolution of hemipteroid insects. *Proceedings of the National Academy of Sciences of the United States of America* **115**: 12775–12780.
- Karr JA, Clapham ME. 2015.** Taphonomic biases in the insect fossil record: shifts in articulation over geologic time. *Paleobiology* **41**: 16–32.
- Kawahara AY, Plotkin D, Espeland M, Meusemann K, Toussaint EFA, Donath A, Gimnich F, Frandsen PB, Zwick A, dos Reis M, Barber JR, Peters RS, Liu S, Zhou X, Mayer C, Podsiadlowski L, Storer C, Yack JE, Misof B, Breinholt JW. 2019.** Phylogenomics reveals the evolutionary timing and pattern of butterflies and moths. *Proceedings of the National Academy of Sciences* **116**: 201907847.
- Kidwell SM, Flessa KW. 1996.** The quality of the fossil record: populations, species, and communities. *Annual Review of Earth and Planetary Sciences* **24**: 433–464.

- Kim S, Wu Z, Esmaili E, Dombroskie JJ, Jung S. 2020.** How a raindrop gets shattered on biological surfaces. *Proceedings of the National Academy of Sciences of the United States of America* **117**: 13901–13907.
- Kiselev S, Nazarov V. 2009.** Late Cenozoic insects of northern Eurasia. *Paleontological Journal* **43**: 732–850.
- Kjellesvig-Waering EN. 1986.** A restudy of the fossil Scorpionida of the world. *Palaeontographica Americana* **54**: 1–28.
- Kozlov MV. 1989.** New upper Jurassic and Lower Cretaceous Lepidoptera (Papilionida). *Paleontological Journal* **23**: 34–39.
- Kraus O, Brauckmann C. 2003.** Fossil giants and surviving dwarfs. Arthropleurida and Pselaphognatha (Atelocerata, Diplopoda): characters, phylogenetic relationships and construction. *Verhandlungen des naturwissenschaftlichen Vereins in Hamburg (Neue Folge)* **40**: 5–50.
- Ksepka DT, Ware JL, Lamm KS. 2014.** Flying rocks and flying clocks: disparity in fossil and molecular dates for birds. *Proceedings of the Royal Society B: Biological Sciences* **281**: 20140677.
- Labandeira CC, Beall BS, Hueber FM. 1988.** Early insect diversification: evidence from a Lower Devonian bristletail from Quebec. *Science* **242**: 913–916.
- Labandeira CC, Tremblay SL, Bartowski KE, VanAller Hernick L. 2013.** Middle Devonian liverwort herbivory and antiherbivore defence. *New Phytologist* **200**: 247–258.
- Laurentiaux D. 1952.** Découverte d'un Homoptère Prosboloïde dans le Namurien Belge. *Association pour l'Étude de la Paléontologie et de la Stratigraphie Houillères Publication* **14**: 1–16.
- Laurentiaux-Viera F, Laurentiaux D. 1961.** *Prothelyphonus neerlandicus*, nov. sp., Uropyge du Westphalien du Limbourg Hollandais. *Mededelingen van de Geologische Stichting, NS* **13**: 29–34.
- Lee MSY, Ho SYW. 2016.** Molecular clocks. *Current Biology* **26**: R399–R402.
- Li HT, Yi TS, Gao LM, Ma PF, Zhang T, Yang JB, Gitzendanner MA, Fritsch PW, Cai J, Luo Y, Wang H, van der Bank M, Zhang SD, Wang QF, Wang J, Zhang ZR, Fu CN, Yang J, Hollingsworth PM, Chase MW, Soltis DE, Soltis PS, Li DZ. 2019.** Origin of angiosperms and the puzzle of the Jurassic gap. *Nature Plants* **5**: 461–470.
- Liu GC, Zhang H, Cai CY, Yuan AH, Zheng YQ, Huang DY. 2021.** The bristletail genus *Dasyleptus* (Insecta: Archaeognatha: Dasyleptidae) from the Upper Permian–Lower Triassic Kayitou formation of SouthWestern China. *Historical Biology* **33**: 1292–1296.
- Lozano-Fernandez J, Giacomelli M, Fleming JF, Chen A, Vinther J, Thomsen PF, Glenner H, Palero F, Legg DA, Iliffe TM, Pisani D, Olesen J. 2019.** Pancrustacean evolution illuminated by taxon-rich genomic-scale data sets with an expanded Remipede sampling. *Genome Biology and Evolution* **11**: 2055–2070.
- Lozano-Fernandez J, Tanner AR, Puttick MN, Vinther J, Edgecombe GD, Pisani D. 2020.** A Cambrian–Ordovician terrestrialization of arachnids. *Frontiers in Genetics* **11**.
- Maksoud S, Azar D. 2020.** Lebanese amber: latest updates. *Palaeontology* **3**: 125–155.
- Mapalo MA, Robin N, Boudinot BE, Ortega-Hernández J, Barden P. 2021.** A tardigrade in Dominican amber. *Proceedings of the Royal Society B: Biological Sciences* **288**: 20211760.
- Martynov A. 1936.** On some new materials of Arthropoda from Kuznetsk-Basin. *Izvestiya Akademii Nauk SSSR: Seriya Biologicheskaya* **6**: 1251–1264.
- Mashimo Y, Matsumura Y, Machida R, Dallai R, Gottardo M, Yoshizawa K, Friedrich F, Wipfler B, Beutel RG. 2014.** 100 years Zoraptera – a phantom in insect evolution and the history of its investigation. *Insect Systematics & Evolution* **45**: 371–393.
- May MR, Contreras DL, Sundue MA, Nagalingum NS, Looy CV, Rothfels CJ. 2021.** Inferring the total-evidence timescale of marattialean fern evolution in the face of model sensitivity. *Systematic Biology* **70**: 1232–1255.
- McPeck MA. 2008.** The ecological dynamics of clade diversification and community assembly. *The American Naturalist* **172**: E270–E284.
- Mendes LF. 2018.** Biodiversity of the thysanurans (Microcoryphia and Zygentoma). In: Footitt RG, Adler PH, eds. *Insect biodiversity*. Chichester: John Wiley & Sons, Ltd, 155–198.
- Mendes LF, Wunderlich J. 2013.** New data on thysanurans preserved in Burmese amber (Microcoryphia and Zygentoma Insecta). *Soil Organisms* **85**: 11–22.
- Miller RF, Forbes WH. 2001.** An Upper Carboniferous trigonotarbid, *Aphantomartus pustulatus* (Scudder, 1884), from the Maritimes Basin (Euramerican Coal Province), New Brunswick, Canada. *Atlantic Geology* **37**: 191–196.
- Misof B, Liu S, Meusemann K, Peters RS, Donath A, Mayer C, Frandsen PB, Ware J, Flouri T, Beutel RG, Niehuis O, Petersen M, Izquierdo-Carrasco F, Wappler T, Rust J, Aberer AJ, Aspöck U, Aspöck H, Bartel D, Blanke A, Berger S, Böhm A, Buckley TR, Calcott B, Chen J, Friedrich F, Fukui M, Fujita M, Greve C, Grobe P, Gu S, Huang Y, Jermiin LS, Kawahara AY, Krogmann L, Kubiak M, Lanfear R, Letsch H, Li Y, Li Z, Li J, Lu H, Machida R, Mashimo Y, Kapli P, McKenna DD, Meng G, Nakagaki Y, Navarrete-Heredia JL, Ott M, Ou Y, Pass G, Podsiadlowski L, Pohl H, von Reumont BM, Schütte K, Sekiya K, Shimizu S, Slipinski A, Stamatakis A, Song W, Su X, Szucsich NU, Tan M, Tan X, Tang M, Tang J, Timelthaler G, Tomizuka S, Trautwein M, Tong X, Uchifune T, Walz MG, Wiegmann BM, Wilbrandt J, Wipfler B, Wong TKF, Wu Q, Wu G, Xie Y, Yang S, Yang Q, Yeates DK, Yoshizawa K, Zhang Q, Zhang R, Zhang W, Zhang Y, Zhao J, Zhou C, Zhou L, Ziesmann T, Zou S, Xu X, Yang H, Wang J, Kjer KM, Zhou X. 2014.** Phylogenomics resolves the timing and pattern of insect evolution. *Science* **346**: 763–767.
- Moen D, Morlon H. 2014.** Why does diversification slow down? *Trends in Ecology & Evolution* **29**: 190–197.
- Montagna M, Tong KJ, Magoga G, Strada L, Tintori A, Ho SYW, Lo N. 2019.** Recalibration of the insect evolutionary time scale using Monte San Giorgio fossils suggests survival of key lineages through the End-Permian Extinction.

- Proceedings of the Royal Society B: Biological Sciences* **286**: 20191854.
- Morlon H, Potts MD, Plotkin JB. 2010.** Inferring the dynamics of diversification: a coalescent approach. *PLoS Biology* **8**: e1000493.
- Morris JL, Puttick MN, Clark JW, Edwards D, Kenrick P, Pressel S, Wellman CH, Yang Z, Schneider H, Donoghue PCJ. 2018.** The timescale of early land plant evolution. *Proceedings of the National Academy of Sciences* **115**: E2274–E2283.
- Mundel P. 1979.** The centipedes (Chilopoda) of the Mazon Creek. In: Nitecki MH, ed. *Mazon Creek fossils*. New York: Academic Press, 361–378.
- Nel A, Roques P, Nel P, Prokin AA, Bourgoïn T, Prokop J, Szwedo J, Azar D, Desutter-Grandcolas L, Wappler T, Garrouste R, Coty D, Huang D, Engel MS, Kirejtshuk AG. 2013.** The earliest known holometabolous insects. *Nature* **503**: 257–261.
- Nel A, Roques P, Nel P, Prokop J, Steyer JS. 2007.** The earliest holometabolous insect from the Carboniferous: a ‘crucial’ innovation with delayed success (Insecta Protomeropina Protomeropidae). *Annales de la Société entomologique de France* **43**: 349–355.
- Nelsen MP, DiMichele WA, Peters SE, Boyce CK. 2016.** Delayed fungal evolution did not cause the Paleozoic peak in coal production. *Proceedings of the National Academy of Sciences* **113**: 2442–2447.
- Nicholson DB, Ross AJ, Mayhew PJ. 2014.** Fossil evidence for key innovations in the evolution of insect diversity. *Proceedings of the Royal Society B: Biological Sciences* **281**.
- Nieukerken EJV, Kaila L, Kitching IJ, Kristensen NP, Lees DC, Minet J, Mitter C, Mutanen M, Regier JC, Simonsen TJ, Wahlberg N, Yen SH, Zahiri R, Adamski D, Baixeras J, Bartsch D, Bengtsson BÅ, Brown JW, Bucheli SR, Davis DR, de Prins J, De Prins W, Epstein ME, Gentili-Poole P, Gielis C, Hättenschwiler P, Hausmann A, Holloway JD, Kallies A, Karsholt O, Kawahara AY, Koster S, Kozlov MV, Lafontaine JD, Lamas G, Landry JF, Lee S, Nuss M, Park KT, Penz C, Rota J, Schintlmeister A, Schmidt BC, Sohn JC, Solis MA, Tarmann GM, Warren AD, Weller S, Yakovlev RV, Zolotuhin VV, Zwick A. 2011.** Order Lepidoptera Linnaeus, 1758. In: Zhang ZQ, ed. *Animal biodiversity: an outline of higher-level classification and survey of taxonomic richness*. *Zootaxa* **3703**: 212–221.
- Phillimore AB, Price TD. 2008.** Density-dependent cladogenesis in birds. *PLoS Biology* **6**: e71.
- Phillips MJ, Fruciano C. 2018.** The soft explosive model of placental mammal evolution. *BMC Evolutionary Biology* **18**: 104.
- Pinto ID. 1986.** Carboniferous insects from Argentina III- Familia Xenopteridae Pinto, nov. Ordo Megasecoptera. *Pesquisas em Geociências* **18**: 23–29.
- Pinto ID, Hünicken MA. 1980.** *Gondwanarachne* a new genus of the order Trigonotarvida (Arachnida) from Argentina. *Boletín de la Academia Nacional de Ciencias Córdoba* **53**: 307–315.
- Plotnick RE. 1990.** Paleobiology of the arthropod cuticle. *Short Courses in Paleontology* **3**: 177–196.
- Pohl H, Beutel RG. 2016.** †*Kinzelbachilla ellenbergeri* – a new ancestral species, genus and family of Strepsiptera (Insecta). *Systematic Entomology* **41**: 287–297.
- Polilov AA. 2015.** Small is beautiful: features of the smallest insects and limits to miniaturization. *Annual Review of Entomology* **60**: 103–121.
- Polilov AA. 2016.** *At the size limit - effects of miniaturization in insects*. Cham: Springer International Publishing.
- Poschmann M, Dunlop JA. 2010.** Trigonotarbid arachnids from the Lower Devonian (Lower Emsian) of Alken an der Mosel (Rhineland-Palatinate, SW Germany). *Paläontologische Zeitschrift* **84**: 467–484.
- Poschmann M, Dunlop JA. 2011.** Trigonotarbid arachnids from the Lower Devonian (Siegenian) of Bürdenbach (Lahrbach Valley, Westerwald area, Rhenish Slate Mountains, Germany). *Paläontologische Zeitschrift* **85**: 433–447.
- Prokop J, Nel A, Hoch I. 2005.** Discovery of the oldest known Pterygota in the Lower Carboniferous of the Upper Silesian Basin in the Czech Republic (Insecta: Archaeorthoptera). *Geobios* **38**: 383–387.
- Qvarnström M, Fikáček M, Vikberg Wernström J, Huld S, Beutel RG, Arriaga-Varela E, Ahlberg PE, Niedźwiedzki G. 2021.** Exceptionally preserved beetles in a Triassic coprolite of putative dinosauriform origin. *Current Biology* **31**: 3374–3381.e5.
- Rainford JL, Hofreiter M, Nicholson DB, Mayhew PJ. 2014.** Phylogenetic distribution of extant richness suggests metamorphosis is a key innovation driving diversification in insects. *PLoS One* **9**: e109085.
- Ramírez-Barahona S, Sauquet H, Magallón S. 2020.** The delayed and geographically heterogeneous diversification of flowering plant families. *Nature Ecology & Evolution* **4**: 1232–1238.
- Rasnitsyn AP. 2003.** On the skimming hypothesis of the origin of insect flight. *Acta Zoologica Cracoviensia* **46**: 85–88.
- Rasnitsyn AP, Aristov DS, Gorochov AV, Rowland JM, Sinitshenkova ND. 2004.** Important new insect fossils from Carrizo Arroyo and the Permo-Carboniferous faunal boundary. *New Mexico Museum of Natural History and Science Bulletin* **25**: 215–246.
- Rasnitsyn AP, Quicke DLJ. 2002.** *History of insects*. Dordrecht: Kluwer.
- Regier JC, Shultz JW, Kambic RE. 2004.** Phylogeny of basal hexapod lineages and estimates of divergence times. *Annals of the Entomological Society of America* **97**: 411–419.
- Rehm P, Borner J, Meusemann K, von Reumont BM, Simon S, Hadrys H, Misof B, Burmester T. 2011.** Dating the arthropod tree based on large-scale transcriptome data. *Molecular Phylogenetics and Evolution* **61**: 880–887.
- Ren D, Nel A, Prokop J. 2008.** New early griffenfly, *Sinomeganeura huangheensis* from the Late Carboniferous of northern China (Meganiptera: Meganeuridae). *Insect Systematics & Evolution* **39**: 223–229.
- Richards BC. 2013.** Current status of the international Carboniferous time scale. *The Carboniferous–Permian Transition, Bulletin* **60**: 348–353.
- Riek EF, Kukalová-Peck J. 1984.** A new interpretation of dragonfly wing venation based upon Early Upper



- Carboniferous fossils from Argentina (Insecta: Odonatoidea) and basic character states in pterygote wings. *Canadian Journal of Zoology* **62**: 1150–1166.
- Rinehart LF, Rasnitsyn AP, Lucas SG, Heckert AB. 2005.** Instar sizes and growth in the Middle Permian monuran *Dasyleptus brongniarti* (Insecta: Machilida: Dasyleptidae). *Bulletin – New Mexico Museum of Natural History and Science* **30**: 270–272.
- Rohdendorf BB. 1961.** The description of the first winged insect from the Devonian beds of the Timan. *Entomological Review* **40**: 484–489.
- Rohdendorf BB. 1970.** Vtoraya nakhodka ostatkov krylatykh devonskikh nasekomykh. *Entomologicheskoe Obozrenie* **49**: 835–837.
- Rohdendorf B. 1972.** Devonian eopterids were not insects but Eumalacostraca. *Entomological Review* **51**: 96–97.
- Rota-Stabelli O, Daley AC, Pisani D. 2013.** Molecular timetrees reveal a Cambrian colonization of land and a new scenario for ecdysozoan evolution. *Current Biology* **23**: 392–398.
- Schachat SR, Labandeira CC, Saltzman MR, Cramer BD, Payne JL, Boyce CK. 2018.** Phanerozoic  $pO_2$  and the early evolution of terrestrial animals. *Proceedings of the Royal Society B: Biological Sciences* **285**: 20172631.
- Schubnel T, Desutter-Grandcolas L, Legendre F, Prokop J, Mazurier A, Garrouste R, Grandcolas P, Nel A. 2020.** To be or not to be: postcubital vein in insects revealed by microtomography. *Systematic Entomology* **45**: 327–336.
- Schwentner M, Combosch DJ, Pakes Nelson J, Giribet G. 2017.** A phylogenomic solution to the origin of insects by resolving crustacean-hexapod relationships. *Current Biology* **27**: 1818–1824.e5.
- Scourfield DJ, Calman WT. 1926.** V. On a new type of crustacean from the old red sandstone (Rhynie Chert Bed, Aberdeenshire) – *Lepidocaris rhyniensis*, gen. et sp. nov. *Philosophical Transactions of the Royal Society of London. Series B, Containing Papers of a Biological Character* **214**: 153–187.
- Shang L, Bethoux O, Ren D. 2011.** New stem-Phasmatodea from the Middle Jurassic of China. *European Journal of Entomology* **108**: 677–685.
- Sharov AG. 1961.** Otryad Plecoptera (Order Plecoptera) in Paleozojskoe nasekomye Kuznetskovo bassejna (Paleozoic insects from Kuznetsk basin). *Trudy Paleontologicheskogo instituta, Akademiya Nauk SSSR* **85**: 225–234.
- Shcherbakov DE. 2021.** New Curvicutitidae and Paraknightiidae (Homoptera: Cicadomorpha) from the Triassic of Central Asia. *Russian Entomological Journal* **30**: 129–134.
- Shear WA, Bonamo PM, Grierson JD, Rolfe WD, Smith EL, Norton RA. 1984.** Early land animals in North America: evidence from Devonian age arthropods from Gilboa, New York. *Science* **224**: 492–494.
- Shear WA, Edgecombe GD. 2010.** The geological record and phylogeny of the Myriapoda. *Arthropod Structure and Development* **39**: 174–190.
- Shear WA, Gensel PG, Jeram AJ. 1996.** Fossils of large terrestrial arthropods from the Lower Devonian of Canada. *Nature* **384**: 555–557.
- Shear WA, Jeram AJ, Selden PA. 1998.** Centiped legs (Arthropoda, Chilopoda, Scutigleromorpha) from the Silurian and Devonian of Britain and the Devonian of North America. *American Museum Novitates* **3231**: 1–16.
- Shear W, Selden PA. 2001.** Rustling in the undergrowth: animals in early terrestrial ecosystems. In: Gensel PG, Edwards D, eds. *Plants invade the land*. New York: Columbia University Press, 29–51.
- Silvestro D, Salamin N, Antonelli A, Meyer X. 2019.** Improved estimation of macroevolutionary rates from fossil data using a Bayesian framework. *Paleobiology* **45**: 546–570.
- Siveter DJ, Briggs DEG, Siveter DJ, Sutton MD, Joomun SC. 2013.** A Silurian myodocope with preserved soft-parts: cautioning the interpretation of the shell-based ostracod record. *Proceedings of the Royal Society B: Biological Sciences* **280**: 20122664.
- Siveter DJ, Tanaka G, Farrell UC, Martin MJ, Siveter DJ, Briggs DEG. 2014.** Exceptionally preserved 450-million-year-old Ordovician ostracods with brood care. *Current Biology* **24**: 801–806.
- Smith AB, Pisani D, Mackenzie-Dodds JA, Stockley B, Webster BL, Littlewood DTJ. 2006.** Testing the molecular clock: molecular and paleontological estimates of divergence times in the Echinoidea (Echinodermata). *Molecular Biology and Evolution* **23**: 1832–1851.
- Sohn JC, Labandeira CC, Davis DR. 2015.** The fossil record and taphonomy of butterflies and moths (Insecta, Lepidoptera): implications for evolutionary diversity and divergence-time estimates. *BMC Evolutionary Biology* **15**: 12.
- Sohn JC, Labandeira CC, Davis DR, Mitter C. 2012.** An annotated catalog of fossil and subfossil Lepidoptera (Insecta: Holometabola) of the world. *Zootaxa* **3286**: 1–132.
- Staniczek AH, Sroka P, Bechly G. 2014.** Neither silverfish nor fowl: the enigmatic Carboniferous *Carbotriplura kukulovae* Kluge, 1996 (Insecta: Carbotriplurida) is the putative fossil sister group of winged insects (Insecta: Pterygota). *Systematic Entomology* **39**: 619–632.
- Su Y, Zhang Z, Hong Y. 2012.** Two new ancient griffenflies (Insecta: Odonatoptera) from the Namurian of Ningxia, China. *Insect Systematics & Evolution* **43**: 1–10.
- Szwedo J. 2016.** The unity, diversity and conformity of bugs (Hemiptera) through time. *Earth and Environmental Science Transactions of the Royal Society of Edinburgh* **107**: 109–128.
- Thomas JA, Frandsen PB, Prendini E, Zhou X, Holzenthal RW. 2020.** A multigene phylogeny and timeline for Trichoptera (Insecta). *Systematic Entomology* **45**: 670–686.
- Thomas JA, Trueman JWH, Rambaut A, Welch JJ. 2013.** Relaxed phylogenetics and the Palaeoptera problem: resolving deep ancestral splits in the insect phylogeny. *Systematic Biology* **62**: 285–297.
- Tihelka E, Cai C, Giacomelli M, Lozano-Fernandez J, Rota-Stabelli O, Huang D, Engel MS, Donoghue PCJ, Pisani D. 2021.** The evolution of insect biodiversity. *Current Biology* **31**: R1299–R1311.
- Tong KJ, Duchêne S, Ho SYW, Lo N. 2015.** Comment on ‘Phylogenomics resolves the timing and pattern of insect evolution’. *Science* **349**: 487–487.



- Trümper S, Schneider JW, Nemyrovska T, Korn D, Linnemann U, Ren D, Béthoux O. 2020.** Age and depositional environment of the Xiaheyan insect fauna, embedded in marine black shales (Early Pennsylvanian, China). *Palaeogeography, Palaeoclimatology, Palaeoecology* **538**: 109444.
- Vachon M, Heyler D. 1985.** Description d'une nouvelle espèce de Scorpion: *Buthiscorpius pescei* (Stéphanien de Montceau-les-Mines, France). Remarques sur la classification des Scorpions (Arachnida) du Carbonifère. *Bulletin de la Société d'Histoire Naturelle d'Autun* **113**: 29–47.
- van Eldijk TJB, Wappler T, Strother PK, van der Weijst CMH, Rajaei H, Visscher H, van de Schootbrugge B. 2018.** A Triassic–Jurassic window into the evolution of Lepidoptera. *Science Advances* **4**: e1701568.
- Wahlberg N, Wheat CW, Peña C. 2013.** Timing and patterns in the taxonomic diversification of Lepidoptera (butterflies and moths). *PLoS One* **8**: e80875.
- Wang Y-h, Engel MS, Rafael JA, Wu H, Rédei D, Xie Q, Wang G, Liu X, Bu W. 2016.** Fossil record of stem groups employed in evaluating the chronogram of insects (Arthropoda: Hexapoda). *Scientific Reports* **6**: 38939.
- Wang B, Szwedo J, Zhang H. 2009.** Jurassic Progonocimicidae (Hemiptera) from China and phylogenetic evolution of Coleorrhyncha. *Science in China Series D: Earth Sciences* **52**: 1953–1961.
- Whalley P. 1986.** The systematics and palaeogeography of the Lower Jurassic insects of Dorset, England. *Bulletin of the British Museum (Geology)* **39**: 107–189.
- Whalley P, Jarzembowski EA. 1981.** A new assessment of *Rhyniella*, the earliest known insect, from the Devonian of Rhynie, Scotland. *Nature* **291**: 317–317.
- Wheat CW, Wahlberg N. 2013.** Critiquing blind dating: the dangers of over-confident date estimates in comparative genomics. *Trends in Ecology & Evolution* **28**: 636–642.
- Whiting MF, Carpenter JC, Wheeler QD, Wheeler WC. 1997.** The Strepsiptera problem: phylogeny of the holometabolous insect orders inferred from 18S and 28S ribosomal DNA sequences and morphology. *Systematic Biology* **46**: 1–68.
- Willmann R. 1989.** Rediscovered: *Permotipula patricia*, the oldest known fly. *Die Naturwissenschaften* **76**: 375–377.
- Wilson HM. 2005.** Zosterogrammida, a new order of millipedes from the Middle Silurian of Scotland and the Upper Carboniferous of Euramerica. *Palaeontology* **48**: 1101–1110.
- Wolfe JM, Daley AC, Legg DA, Edgecombe GD. 2016.** Fossil calibrations for the arthropod Tree of Life. *Earth-Science Reviews* **160**: 43–110.
- Wootton RJ. 1981.** Palaeozoic insects. *Annual Review of Entomology* **26**: 319–344.
- Wray GA. 2015.** Molecular clocks and the early evolution of metazoan nervous systems. *Philosophical Transactions of the Royal Society B: Biological Sciences* **370**: 20150046.
- Yang Z. 2007.** PAML 4: phylogenetic analysis by maximum likelihood. *Molecular Biology and Evolution* **24**: 1586–1591.
- Yang H, Shi C, Engel MS, Zhao Z, Ren D, Gao T. 2021.** Early specializations for mimicry and defense in a Jurassic stick insect. *National Science Review* **8**: 20200402.
- Ye F, Kment P, Rédei D, Luo JY, Wang YH, Kuechler SM, Zhang WW, Chen PP, Wu HY, Wu YZ, Sun XY, Ding L, Wang YR, Xie Q. 2022.** Diversification of the phytophagous lineages of true bugs (Insecta: Hemiptera: Heteroptera) shortly after that of the flowering plants. *Cladistics* **38**: 403–428.
- Yushuang L, Sinitshenkova ND, Dong R, Chungkun S. 2011.** Pronemouridae fam. nov. (Insecta: Plecoptera), the stem group of Nemouridae and Notonemouridae, from the Middle Jurassic of Inner Mongolia, China. *Palaeontology* **54**: 923–933.
- Zhang W, Shih C, Labandeira CC, Sohn JC, Davis DR, Santiago-Blay JA, Flint O, Ren D. 2013.** New fossil Lepidoptera (Insecta: Amphiesmenoptera) from the Middle Jurassic Jiulongshan Formation of Northeastern China. (D Frayer, Ed.). *PLoS One* **8**: e79500.
- Zhang X, Siveter DJ, Waloszek D, Maas A. 2007.** An epipodite-bearing crown-group crustacean from the Lower Cambrian. *Nature* **449**: 595–598.
- Zheng Y, Wiens JJ. 2015.** Do missing data influence the accuracy of divergence-time estimation with BEAST? *Molecular Phylogenetics and Evolution* **85**: 41–49.
- Ziadé K, Szwedo J, Azar D. 2011.** The first Progonocimicidae (Insecta: Hemiptera: Coleorrhyncha) from Lower Cretaceous Lebanese amber. *Insect Systematics & Evolution* **42**: 161–177.

## SUPPORTING INFORMATION

Additional Supporting Information may be found in the online version of this article at the publisher's web-site:

**Table S1.** Results of the PyRate analyses, which were run on ten replicate input datasets. ‘Seed (R)’ is the value entered into the set.seed function in R while producing the input dataset. ‘Seed (p)’ is the random seed used when running PyRate in python. ‘Initial age’ is the first sampled root age estimated for Pterygota, sampled after 1000/20 000 000 iterations. We note whether each PyRate run reached convergence. For those that did, we note the proportion of iterations discarded as burn-in, the effective sample size (ESS) and the estimated age of Pterygota: the median value and upper and lower limits of the 95% highest posterior density interval.

**Table S2.** List of analyses undertaken in this study. The first column gives the name of the fossil dataset, the second column the prior setting for the analysis and the third column the calibration date for the pterygote node. A zero in the third column indicates that tree priors without a calibration date were used for the pterygote node.

**Table S3.** Age estimates for the origin of crown Pterygota derived from different calibration schemes using the Misof *et al.* (2014) dataset. The ‘Difference’ column denotes the difference between each age estimate and the age estimate published by Misof *et al.* (2014).

**Figure S1.** Estimates for the age of Pterygota (Mya) from the six PyRate analyses that reached convergence. The interval from 324 to 330 Mya is shaded in blue.

**Figure S2.** Age estimates for Pterygota based on 11 combinations of datasets and priors. Abbreviations: M = Misof *et al.* age estimate priors; W = Wolfe age estimate priors; lgn = lognormal prior settings (with uncorrelated relaxed clock); unif = uniform prior settings (with uncorrelated relaxed clock); 324, 331 and 324–450 refer to Mya settings for pterygote priors; strict = strict molecular clock setting.

**Figure S3.** Scatter plots of median values for the 85 gene regions in the study. Abbreviations: M = Misof *et al.* age estimate priors; W = Wolfe age estimate priors; lgn = lognormal prior settings (with uncorrelated relaxed clock); unif = uniform prior settings (with uncorrelated relaxed clock); 324, 331 and 324–450 refer to Mya settings for pterygote priors; strict = strict molecular clock setting.

**Figure S4.** Density distributions for user-defined lognormal priors, and lognormal posterior distributions for Misof datasets with pterygote calibration and without Pterygote calibration for the Diptera, Neuroptera, Holometabola and Pterygote nodes. Density is on the *y*-axis and divergence time is on the *x*-axis in all panels.

**Figure S5.** Density distributions for user-defined lognormal priors, and lognormal posterior distributions for Wolfe datasets with pterygote calibration and without Pterygote calibration for the Diptera, Neuroptera, Holometabola and Pterygote nodes. Density is on the *y*-axis and divergence time is on the *x*-axis in all panels.

**Figure S6.** Results of another version of the fossil simulation procedure, for which the initial sampling rate is 50% of the final sampling rate at the Permian/Triassic boundary, and in which occurrences were not clustered into Lagerstätten.

**Figure S7.** Results of another version of the fossil simulation procedure, for which the initial sampling rate is 50% of the final sampling rate at the Permian/Triassic boundary, and in which occurrences were clustered into Lagerstätten.

**Figure S8.** Results of another version of the fossil simulation procedure, for which the initial sampling rate is 10% of the final sampling rate at the Permian/Triassic boundary, and in which occurrences were clustered into Lagerstätten.

**Figure S9.** Results of another version of the fossil simulation procedure, for which the initial sampling rate is 2% of the final sampling rate at the Permian/Triassic boundary, and in which occurrences were not clustered into Lagerstätten.

**Figure S10.** Results of another version of the fossil simulation procedure, for which the initial sampling rate is 2% of the final sampling rate at the Permian/Triassic boundary, and in which occurrences were clustered into Lagerstätten.

**Appendix S1.** The fossils from the Wolfe *et al.* (2016) compendium used in our analyses.