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## MINIREVIEW

# Current progress in the biology of members of the Sporothrix schenckii complex following the genomic era

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## ABSTRACT

Sporotrichosis has been attributed for more than a century to one single etiological agent, *Sporothrix schencki*. Only eight years ago, it was described that, in fact, the disease is caused by several pathogenic cryptic species. The present review will focus on recent advances to understand the biology and virulence of epidemiologically relevant pathogenic species of the *S. schenckii* complex. The main subjects covered are the new clinical and epidemiological aspects including diagnostic and therapeutic challenges, the development of molecular tools, the genome database and the perspectives for study of virulence of emerging *Sporothrix* species.

Keywords: Sporothrix schenckii; Sporothrix brasiliensis; Sporothrix globosa; genome; molecular tools; pathogenesis

## **INTRODUCTION**

Sporotrichosis is an acute or chronic granulomatous subcutaneous mycosis of humans and mammals that has a worldwide distribution and several clinical manifestations (Lopes-Bezerra, Schubach and Costa 2006). Disseminated sporotrichosis is currently associated with a high probability of HIV infection (Lopez-Romero *et al.* 2011). Schenck (1898), at Johns Hopkins Hospital in Baltimore, published the first clinical case of sporotrichosis. Two years later, the etiologic agent was isolated, identified and classified (Hektoen and Perkins 1900), followed by the description of the yeast parasitic phase in 1907 (Lutz and Splendore 1907). For just over a century, this subcutaneous mycosis was assigned to a single etiologic agent, *Sporothrix schenckii*. In 2007, a new benchmark was established in the study of this disease with the description of the so-called S. schenckii complex (Marimon et al. 2007). The authors reported the existence of cryptic pathogenic species within this complex, among which we highlight S. schenckii sensu stricto, S. brasiliensis and S. globosa. According to recent epidemiological data, these are the species of greatest clinical and global epidemiological importance (Chakrabarti et al. 2015).

In Latin America, sporotrichosis is the most prevalent subcutaneous mycosis; however, changes in the epidemiological

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pattern of the disease have been reported. Classically, sporotrichosis is acquired through traumatic inoculation of the fungus into a host's subcutaneous tissue with contaminated plant debris, thorns or soil. This process initiates as an occupational disease that is associated with gardening and rural work. However, the zoonotic transmission by domestic cats (Felix catus) is of clinical and epidemiological importance and is changing the paradigm of this disease. Transmission through contact with sick and apparently healthy cats may derive from bites and scratches, or by contact with nasal secretions (Schubah, Barros and Wanke 2008). Although feline sporotrichosis has been reported in several countries, in Brazil, a major zoonotic outbreak has registered more than 4000 cases of feline sporotrichosis in Rio de Janeiro State (Schubach, Barros and Wanke 2008; Gremiao et al. 2015) and is spreading to other regions (Montenegro et al. 2014). In parallel, the number of human cases is also increasing exponentially and severe clinical manifestations are being reported, also in immunocompetent hosts (Lopes-Bezerra and Nascimento 2012). Therefore, up to now the number of human cases registered in Rio de Janeiro State is far beyond 4000 (personal communication).

Interestingly, feline sporotrichosis was associated with S. brasiliensis (Rodrigues et al. 2013), an emerging species that appears to be geographically restricted to Brazil (Chakrabarti et al. 2015). However, cases of feline sporotrichosis were recently described in Malaysia caused by clade D S. schenckii sensu stricto (Kano et al. 2015). The clinical and epidemiological paradigm is changing because domestic cats play an important role in hosting and spreading species of the S. schenckii complex. Furthermore, S. globosa has been associated with isolated human cases in Europe (de Oliveira et al. 2014) and with endemic areas in Northeast China, where the infection is due to contact with reeds or cornstalks (Yu et al. 2013; Liu, Zhang and Zhou 2014). Therefore, identifying the factors in the adaptation of these fungal pathogens to other environments, thus redefining the route of transmission of sporotrichosis, is of key importance.

The species belonging to the S. schenckii complex are ascomycetous and thermodimorphic fungi with a mycelial saprophytic phase and a yeast parasitic phase (Howard 1961), which can be easily obtained in the laboratory by growing the fungus at 25-28°C and 35-37°C, respectively. In general, the mycelial saprophytic phase is characterized by hyaline, septate and branched hyphae containing thin conidiophores whose apex forms a small vesicle with sympodially arranged conidia (2- $4\,\mu$ m) in a flower-like arrangement. The conidia become detached from the conidiophores, sometimes being arranged side by side in a row bilaterally to the hyphae. The yeast parasitic phase is pleomorphic, with spindle shaped and/or oval cells measuring 2.5–5  $\mu$ m in diameter and resembling a 'cigar' (Travassos and Lloyd 1980). Morphological variations have been reported among cryptic species of the S. schenckii complex (Marimon et al. 2007). Although little is known about the factors that can induce the mycelium to yeast transition, some groups are studying possible regulators of this process in S. schenckii. Experimental evidence points to a calcium/calmodulin kinaseencoding gene in S. schenckii and its possible involvement as an effector of dimorphism in this fungus (Valle-Aviles et al. 2007). Additionally, a homolog of a hybrid histidine kinase, DRK1, known as a global regulator of dimorphism and virulence in dimorphic fungi, was recently reported in S. schenckii (Hou et al. 2013). The SsDrk1 was predicted to be a soluble histidine kinase and to contain three domains: a sensor, a linker and a functional domain. The authors showed by quantitative realtime RT-PCR that SsDRK1 was more highly expressed in the

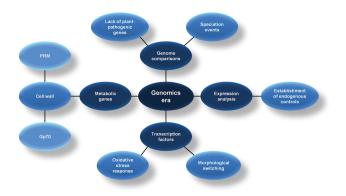


Figure 1. General overview of the main subjects covered by this minireview article about the emerging cryptic species of the S. *schenckii* complex.

yeast phase compared with the mycelial phase, suggesting its involvement in the dimorphic transition. Furthermore, with the genomes of *Sporotrhix* spp. already available (Teixeira *et al.* 2014), other putative regulators of morphogenesis, such as the Efg1 transcription factor, can be further studied within the *S. schenckii* complex.

In the last decade, studies focused on understanding the biology of these fungal pathogens have shed some light on the attributes that allow them to cause sporotrichosis. Furthermore, systematic studies of the clinical and environmental isolates of these fungi have investigated the genetic and phenotypic variability of some pathogenic species, an important topic that will be discussed in this review. The genomic data and the development of genetic tools impact the study of other pathogenic fungi. Therefore, the recent publication of the complete genomes of two pathogenic species of epidemiological relevance, S. schenckii sensu stricto and S. brasiliensis (Teixeira et al. 2014), will leverage other genomic studies and impact basic research into this disease.

In addition, special consideration must be given to the relationship between this pathogen and its human host. In consequence, understanding the biology of the fungus and the factors that contribute to the survival of *Sporothrix* spp. in the diverse environments they can be found, mapping proteins or other factors that are unique to these lower eukaryotes can lead to targets for the development of more selective antifungal drugs. Furthermore, based on recent epidemiological data, molecular methods for clinical diagnosis at the species level are also necessary.

Considering the latest research advances in human and animal sporotrichosis, this minireview will focus on new findings related to genetic variation, genomics and protein expression, as well as the development of genetic tools that will improve our understanding of the biology of these fungal species and contribute to the discovery of molecules related to pathogenesis (Fig. 1).

## THE GENOME IMPACT AND THE DEVELOPMENT OF MOLECULAR TOOLS TO STUDY SPOROTHRIX AND RELATED PATHOGENIC SPECIES

Sporotrichosis is an important mycological disease that is frequently covered in pathophysiology/medical microbiology textbooks for undergraduate healthcare education programs. As mentioned above, S. schenckii, long believed to be the single causative agent of sporotrichosis, is in fact a group of

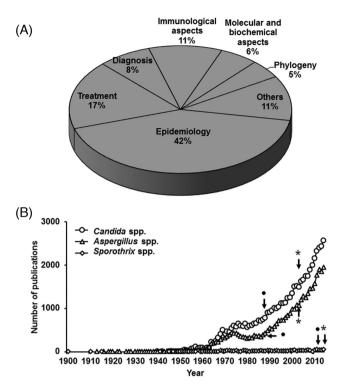


Figure 2. Sporothrix as a publication subject during 1900–2014. An analysis within the PubMed database (http://www.ncbi.nlm.nih.gov/pubmed) searching for the term Sporothrix retrieved 1124 published papers that were classified according the main subject addressed (panel A). In panel B, the number of published papers per year addressing either Sporothrix (open diamonds), Aspergillus (open triangle) or Candida (open circle) is shown. The arrow and closed circle indicate the year where the first report about molecular tools for genetic manipulation was published, while the arrow and asterisk show the year in which the first genome sequence was published.

at least four phylogenetically related species (Marimon et al. 2007). Thus, this infectious disease is now recognized as being caused by several species, some of them present in the same geographical area (Chakrabarti et al. 2015). Despite this advance, we have limited information about the basic biology of these organisms, particularly in fundamental areas such as their ability to survive in the environment without a mammalian host, their virulence factors and the molecular requirements for the immune recognition by the human and feline hosts are poorly understood (Lopez-Romero et al. 2011; Martinez-Alvarez et al. 2014; Tellez et al. 2014). A recent analysis showed that if the keyword 'Sporothrix' is used to search for scientific literature in the PubMed database (http://www.ncbi.nlm.nih.gov/pubmed), 1124 papers are retrieved. This list comprises works published until 2014; of these, 67% covered clinical aspects of these fungi and/or the disease (epidemiology, treatment and diagnosis), 11% of them covered immunological topics, 11% discussed molecular and biochemical aspects, and the remaining 11% studied other subjects related to either Sporothrix or sporotrichosis (Fig. 2A). This number represents the accumulated literature from 1900 to 2014 about this disease and its causative agent, giving an average of only 9.8 publications/year, which is low compared to the amount of literature published within the same period about Candida spp. and Aspergillus spp., the classic models of human fungal pathogens-465.2 publication/year and 340.3 publications/year, respectively (Fig. 2B). Therefore, more efforts are required to understand the clinical and basic biology of members of the S. schenckii complex.

Among the possible explanations for this difference, beside any epidemiological impact, landmark events increased the interest in *Candida* and *Aspergillus* species. The first of them is the development of molecular tools for their genetic manipulation (Durrens *et al.* 1986; Spencer, Spencer and Reynolds 1988), which allowed an exponential publication of papers (Fig. 2B); secondly, the availability of their genomic sequences (Jones *et al.* 2004; Galagan *et al.* 2005) sustained this increased interest (Fig. 2B).

Genomics is the assembly and analysis of the genes and their biological functions encoded within an organism, using bioinformatics, DNA sequencing techniques and recombinant DNA technology (Kotra, Vakulenko and Mobashery 2000). Thus, during the second half of the 1990s, the main aim of some research groups was to sequence the genome of their favorite organism with the aim to reveal fundamental aspects of its evolution, niche adaptation, and in the case of pathogens, the genetic basis of its ability to harm the common host. Although the information we can obtain from a genome sequence is highly valuable and essential for understanding key differences between organisms, such as pathogenic vs. non-pathogenic fungi, this techniques is limited if we compare it with the information generated by other -omics techniques, such as transcriptomics, proteomics and metabolomics (Henry et al. 2014). The combination of all these powerful strategies, based on the information generated by genomics, provides a robust and complete functional analysis of the response of an organism to changes in the surrounding environment, including interactions with members of the same species or the host (Ohashi et al. 2015). Thus, the generation of genomic information is often comparable with the tip of an iceberg: there is a large amount of information about the genome sequence of an organism, but not enough to understand all its biological processes in detail. We recently published the genome sequences of S. schenckii sensu stricto and S. brasiliensis, two causative agents of sporotrichosis in both humans and animals (Teixeira et al. 2014). This has allowed us to understand some basic aspects of their life style and probably boosted the interest of the community and, thus, the number of reports about these pathogens. However, as mentioned, the genome sequences are just the beginning of a thorough study of Sporothrix spp.

The fungal cell wall is often an important topic of study in fungal pathogen research, as the cell wall protects the cell from external insults, displays proteins and molecules associated with host cell damage, and is composed of polysaccharides not synthesized by the human host, allowing the immune system to establish antifungal responses (Netea et al. 2008; Lopes-Bezerra 2011; Diaz-Jimenez et al. 2012; Martinez-Alvarez et al. 2014). Thus far, limited information about the detailed structure, biosynthesis and functional relevance of the cell wall of S. schenckii is available. Early works by Travassos and Lloyd (1980) had identified the main glycoconjugates present in the cell wall of S. schenckii composed by mannose, rhamnose and galactose: a peptidorhamnomanan (PRM), and a peptidorhamnogalactan. The PRM has been of special interest because of its unusual structure and relevance during the immune sensing of S. schenckii cells. The main oligosaccharide found in this glycoconjugate is an O-linked glycan composed of two mannose residues, one glucuronic acid and one or two rhamnose units (Lopes-Alves et al. 1992). The PRM of S. schenckii has the ability to stimulate the production of circulating antibodies in both laboratory animals and infected patients, but immunosuppressive activities have also been related with this molecule (Carlos, Sgarbi and Placeres 1999). The availability of a Sporothrix genome sequence has allowed us to identify candidate genes to

participate not only in the elaboration of PRM but also in the synthesis of different cell wall components (Teixeira et al. 2014), an area that has progressed slowly in the last 10 years. For instance, a significant amount of time and effort have been devoted to isolating a full open reading frame from S. schenckii, using reverse genetics, and this was translated in a handful of studies published between 2001 and 2015 (Kano et al. 2001; Valentin-Berrios et al. 2009; Rodriguez-Caban et al. 2011; Hernandez-Cervantes et al. 2012; Robledo-Ortiz et al. 2012; Lopez-Esparza et al. 2013; Sanchez-Lopez et al. 2015). Now, in the S. schenckii genomics era, this type of project has been simplified to a search in a database, accelerating the generation of information aiming to understand the function of S. schenckii genes (Lopes-Bezerra et al. 2015). Another example of benefits generated by the Sporothrix genome sequence is the study of the antigen Gp70. After PRM, Gp70 is the main cell wall component characterized in S. schenckii (Lopes-Bezerra 2011), and it has received special attention as a potent antigen (Nascimento and Almeida 2005; Ruiz-Baca et al. 2011). Antibodies against Gp70 protect mice from the infection caused by both S. schenckii and S. brasiliensis (Nascimento et al. 2008; de Almeida et al. 2015). Early efforts to identify the polypeptide were unsuccessful (Teixeira et al. 2009), but recent proteomic and genomics approaches identified the gene encoding Gp70 (Castro et al. 2013) and, more importantly, the versatility of this protein in terms of post-translational modifications (Rodrigues et al. 2015).

The Sporothrix genome lacks obvious polysaccharide lyases and polygalacturonase genes, which offers an attractive hypothesis to explain its inability to establish a pathogenic cycle in a plant host (Teixeira *et al.* 2014). However, no biochemical, proteomic or transcriptomic studies have been conducted to discard alternative, non-classical genes that could be activated upon interaction with plant tissues (Couturier *et al.* 2013; Blackman, Cullerne and Hardham 2014).

Traditionally, rRNA has been used as an endogenous control in gene expression analysis techniques like RT-PCR and northern blotting (Rodriguez-Caban *et al.* 2011; Hou *et al.* 2014). However, rRNA is useless when a technique requires mRNA isolation and enrichment, such as microarray analysis or RNA-sequencing techniques, where housekeeping genes with constitutive and constant expression are used as endogenous expression controls to normalize and validate the data (Mantione *et al.* 2014). The Sporothrix genome sequences are allowing us to identify such genes, and we currently have a short list of three candidate genes with constitutive and constant expression that are validated in different growing conditions to use as controls in expression analysis (unpublished results).

The importance of a specific gene in the biology of an organism is usually assessed either by knocking out/down the gene or by its overexpression. Until now, limited attempts of genetic manipulation have been published for *Sporothrix* (Rodriguez-Caban et al. 2011), perhaps in part because few gene sequences were available (limiting the number of possible target genes) but also because a reproducible strategy had not been discovered to transform these organisms (Zhang et al. 2011). We are currently developing stable transformants that allow the expression of foreign genes, gene silencing and targeted gene disruption (unpublished results). As a proof of concept, we have generated a *S. schenc*kii transformant able to express the green fluorescent protein (GPF; Fig. 3). This mutant will allow GFP labeling of specific proteins to localize them within the fungal cell, as in other

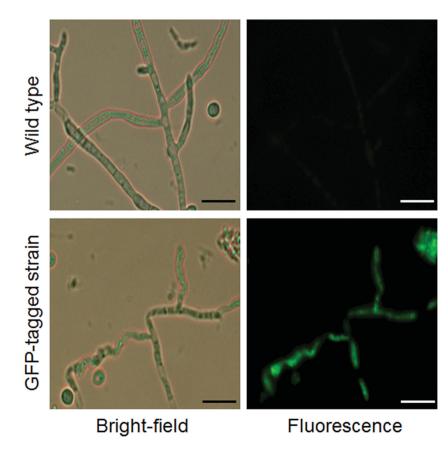


Figure 3. Expression of GFP in S. schenckii. Brightfield and fluorescence microscopy of wild-type and GFP-tagged S. schenckii strains. Scale bar = 10  $\mu$ M.

fungal models (Huh et al. 2003), which is particularly relevant for the large numbers of proteins in the genomes of both S. schenckii and S. brasiliensis with unknown functions (Teixeira et al. 2014). In addition, the expression of GFP will allow us to generate bioluminescent fungal cells and track them within host tissues, as recently reported in *Candida albicans* (Brothers, Newman and Wheeler 2011; Jacobsen et al. 2014), which will provide useful information about the tropisms and dissemination of *Sporothrix* spp.

## DIAGNOSIS, GENETIC VARIABILITY AND DNA-BASED LABORATORY REMARKS

In addition to the classical mycological diagnosis, some authors sought to establish serological methods with different antigens for research purposes and for application in sporotrichosis diagnosis and seroepidemiological surveys, in both humans and cats (Bernardes-Engemann *et al.* 2005; Almeida-Paes *et al.* 2007; Fernandes *et al.* 2011). Serological methods are more accurate for sporotrichosis diagnosis in human clinical specimens (Bernardes-Engemann *et al.* 2005) from special groups of patients, such as pediatric or HIV-positive patients (Bernardes-Engemann *et al.* 2015) or those whose infections display diverse clinical presentations such as arthritis (Orofino-Costa *et al.* 2010) and dacryocystitis (Marques de Macedo *et al.* 2015).

Molecular research is another important tool for the epidemiological characterization of Sporothrix spp. genetic variability and for describing isolates that differ in their interactions with the host. This research has been carried out by studying variations in the Sporothrix mtDNA sequence (Ishizaki et al. 2009), by sequencing highly conserved regions such as the CAL (calmodulin) (Marimon et al. 2007; Oliveira et al. 2011; Rodrigues et al. 2013) and  $\beta$ -tubulin genes (Liu, Zhang and Zhou 2014) and the ITS genomic regions (Zhou et al. 2013; Estrada-Barcenas et al. 2014; Liu, Zhang and Zhou 2014) and by the RAPD technique (Random Amplified Polymorphic DNA; Reis et al. 2009). All of these molecular approaches can contribute to our understanding of the genetic diversity and geographic distribution of the S. schenckii complex species. Another important outcome from genome-based methods is the capacity to compare the molecular profiles of the fungi isolated from environmental niches and healthy (colonized) animals to those from animal and human lesions. These comparisons can trace Sporothrix spp. sources and dissemination routes and the pathways to infection within different hosts.

By using molecular investigative tools, in 2009, Ishizaki et al. proposed the existence of more than one pathogenic species in the S. schenckii complex, based on significant variations detected in mtDNA among isolates from India, Thailand, Brazil, Colombia, Guatemala and Mexico. In a previous study, Marimon et al. (2007), using other molecular markers of genetic variability such as variations in the CAL gene sequence, phylogenetic data and both physiological and morphological information, demonstrated the existence of three new species that had previously identified as S. schenckii. Thus, isolates of the genus were subdivided into five major clades according to their genetic characteristics and associated morpho-physiological data such as differences in conidia, growth rate and assimilation of carbohydrates. The authors further suggested that the observed clades I, III, IV and V do represent different species of this genus. The first three clades were presented as the new species S. brasiliensis, S. globosa and S. mexicana, respectively. Class V was verified to be S. albicans (formerly Sporothrix pallida). The latest understanding of the S. schenckii complex is that it is composed of six distinct species: S. brasiliensis, S. mexicana, S. globosa, S. schenckii sensu stricto, S. luriei and S. albicans. All these species, except S. albicans, have been described in Brazil and the differences in the species' isolation frequency from various environments, including human hosts, and their diverse pathogenicity have been investigated (Rodrigues, de Hoog and de Camargo 2013). Recently, Rodrigues et al. (2013) investigated the CAL and elongation factor 1 alpha-encoding (EF1 $\alpha$ ) genes, and determined that S. brasiliensis is the most prevalent species in the Brazilian epidemics and has at least two different geographical origins. The group was able to reach these conclusions by comparing Sporothrix isolates from five Brazilian states, covering a large area of the South and Southeast Brazilian regions. These authors also verified that S. brasiliensis reveals low genetic diversity compared to S. schenckii sensu stricto. On the other hand, S. globosa and S. mexicana identified as human pathogens in a large collection of Brazilian clinical cases (Rodrigues, de Hoog and de Camargo 2013) were not frequently implicated, appearing in a low number that seems to have remained constant. Another useful method for investigating Sporothrix isolates obtained from different Brazilian areas is the evaluation of polymorphisms at the chromosomal level (Sasaki et al. 2014). This method revealed that closely related species show similar genetic organization through PFGE (pulsed field gel electrophoresis), as previously identified by molecular sequencing. Recently, S. schenckii complex genotyping was greatly simplified by the description of PCR-RFLP (restriction fragment length polymorphism), which allows the quick and relatively low-cost determination of medically significant species (Rodrigues, de Hoog and de Camargo 2014). Finally, a major comparative genome study, including transposable elements, the mitochondrial genome and diverse protein families, was designed to phylogenetically investigate the two major pathogenic species, S. brasiliensis and S. schenckii sensu stricto, leading to the suggestion of a recent event of speciation. The authors concluded that for the Sporothrix lineage to evolve from a plant-associated fungus to a mammalian parasite, a unique ecological shift took place (Teixeira et al. 2014).

Despite these advances, whether the existence of different Sporothrix species or genetically diverse isolates within each species influences the sources of contamination, the dynamics of the pathogens' interactions with the host, the magnitude of the clinical presentations and the host response to treatment are questions yet to be answered. Overall, these studies will have a strong impact on the development of efficient measures of sporotrichosis prevention and control.

## PERSPECTIVES IN THE STUDY OF VIRULENCE AND THE HOST-PATHOGEN INTERPLAY DURING SPOROTRICHOSIS: WHAT HAVE WE LEARNED SO FAR AND WHAT IS TO COME

In the battle between a fungal pathogen and its host, the molecular mechanisms of the host immune response and how this response affects fungal biology and either fungal survival or death are critical areas of interest to researchers. Recent advances in large-scale molecular techniques have demonstrably improved our understanding of the molecular mechanisms employed by both sides during host–pathogen interactions. A better understanding of the mechanisms used by the host to eliminate the pathogen and win this battle against fungi will lead the way to improved diagnosis and treatment of mycosis.

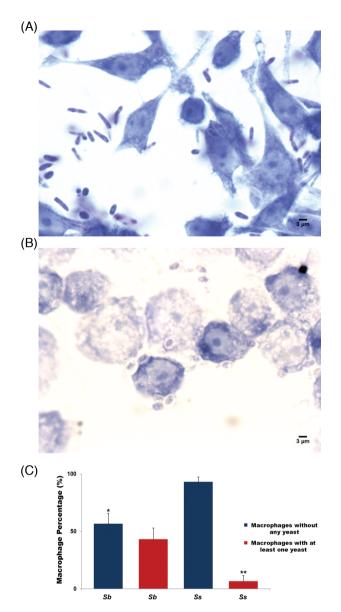
#### The host's perception

When studying an infectious disease such as sporotrichosis, identifying which host factors are required to eliminate the pathogen and prevent the establishment of the disease is vital. Studies that focused on understanding the host's immune response to the cutaneous form of sporotrichosis suggested that cell-mediated immunity to S. *schenckii* is the key to controlling the infection (Carlos *et al.* 1992), with participation of macrophages, dendritic cells and mast cells (Uenotsuchi *et al.* 2006; Romo-Lozano, Hernandez-Hernandez and Salinas 2012; Kusuhara, Qian and Li 2014.)

Although less common, reports show that disseminated sporotrichosis also occurs, especially in alcoholics and immunosuppressed patients, strongly suggesting that defects in the cellmediated immune response make patients more susceptible to the systemic form of the infection (Shiraishi, Nakagaki and Arai 1992; Al-Tawfiq and Wools 1998; López-Romero et al. 2011; Trotter et al. 2014). The same observation seems to be true in mouse models, because immunocompromised animals are more susceptible to S. schenckii than immunocompetent mice (Shiraishi, Nakagaki and Arai 1992). Furthermore, studies using mice with chronic granulomatous disease (CGD), which exhibit defects in the function of NADPH oxidase and the production of superoxide, have shown that CGD mice were unable to eliminate the fungi from the site of inoculation, allowing the fungus to disseminate from the local site of infection to cause fatal systemic mycoses (Kajiwara, Saito and Ohga 2004). These results might also indicate that the production of microbicidal agents, such as reactive oxygen species (ROS) and reactive nitrogen species (RNS), by phagocytic cells plays a key role in eliminating S. schenckii (Fernandes et al. 2000; Kajiwara, Saito and Ohga 2004). Interestingly, this ROS/RNS-mediated killing also seems to be important for the clearance of other fungal pathogens. For example, studies have shown that macrophages, neutrophils and other phagocytic cells can eliminate C. albicans by using oxidative mechanisms, playing a pivotal role in protecting the host against systemic candidiasis (Qian et al. 1994; Jensen, Warner and Balish 1994; Fulurija, Ashman and Papadimitriou 1996; Bogdan, Rollinghoff and Diefenbach 2000; Nathan and Shiloh 2000; Vonk et al. 2006).

Although several studies have focused on the host's immune response to Sporothrix spp. infection, very little is known about the main players and molecular mechanisms involved in innate immune recognition. Host innate immune cells' patternrecognition receptors (PRRs) can recognize fungal organisms mainly through components of the fungal cell wall, also known as pathogen-associated molecular patterns (PAMPs). Once the phagocytic cells recognize the fungal pathogen, these host components of the immune system can activate an effector response, which consists of the production of ROS/RNS and the further induction of adaptive T helper (Th) responses by antigenpresenting cells (Bourgeois and Kuchler 2012; Grimm et al. 2013).

In the case of sporotrichosis, some PRRs have been identified as participants in the recognition/activation process necessary to mount an efficient immune response. Toll-like receptor 2 (TLR2), for example, is a key PRR involved in recognizing S. schenckii, because macrophages from TLR2-deficient mice displayed impaired phagocytosis of S. schenckii cells and lower levels of proinflammatory cytokines (Negrini et al. 2013). Another PRR involved in the recognition/activation process is Toll-like receptor 4 (TLR4), which recognizes components of the S. schenckii yeast phase cell wall, resulting in the production of proinflammatory cytokines and effector molecules, such as nitric oxide (NO) (Sassá et al. 2009). Consistent with this observation,



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Figure 4. Sporothrix brasiliensis is internalized more effectively by J774 macrophages than S. schenckii. Micrograph showing J774 cells cocultured with S. schenckii (A) or S. brasiliensis (B). (C) Graph displaying the percentage of macrophages that have phagocytized S. schenckii (Ss) or S. brasiliensis (Sb). \*P < 0.05 Macrophages without Sb yeasts when compared to macrophages without Ss yeasts. \*\*P < 0.05 Macrophages with Ss yeasts when compared to macrophages without Ss yeasts.

TLR4-deficient mice infected with S. schenckii had difficulty mounting a proinflammatory response (Sassá et al. 2012). In addition to TLRs involved in the recognition of S. schenckii, mannose receptors also seem to play a role in this proinflammatory response (Guzman-Beltran et al. 2012). Interestingly, our recent unpublished results suggest that murine macrophages poorly phagocytize S. schenckii compared to S. brasiliensis (Fig. 4).

#### The pathogen's perception

Many intrinsic characteristics of these fungal pathogens allow them to survive inside the host; among them, we can highlight the ability to adhere to host cells, the secretion of extracellular hydrolytic enzymes, morphogenetic switching and robust fungal stress responses. For instance, fungal pathogens are protected by a highly complex and flexible structure of carbohydrate polymers and proteins (Gow and Hube 2012), which plays a pivotal role in both protecting these pathogens against environmental stresses and mediating the recognition process by immune cells (Levitz 2010). Similar to other pathogenic fungi, *S. schenckii* is mainly immunorecognized by its cell wall. Although the *Sporothrix* spp. cell wall is a rich source of antigens recognized by antibodies raised against this organism, the specific contribution of its components during fungal sensing by immune cells is not known, and PAMPs have not been identified (Carlos, Sgarbi and Placeres 1999; Lima *et al.* 2001, 2004; Carlos *et al.* 2003; Nascimento and Almeida 2005; Lopes-Bezerra 2011; Ruiz-Baca *et al.* 2011).

Among the components of the S. schenckii cell wall that are recognized by the host's immune system, we can highlight the cell wall PRM which is recognized by IgG antibodies present in patient sera (Lloyd and Bitoon 1971; Penha and Lopes-Bezerra 2000) and also by receptors and matrix proteins of host cells (Lima et al. 2001; Figueiredo et al. 2004). Another cell wall component that has been thoroughly studied is the 70 kDa antigen, Gp70 (Lima and Lopes-Bezerra 1997; Nascimento and Almeida 2005; Nascimento et al. 2008), which seems to act also as an adhesin that mediates fungus adhesion to host tissues (Ruiz-Baca et al. 2009; Teixeira et al. 2009). Interestingly, a recent study suggested that the cell surface Gp70 antigen is directly associated with the pathogenicity of Sporothrix spp., because there was a correlation between higher levels of Gp70 expression on the fungal surface and reduced infectivity (Castro et al. 2013). Substantial work has been performed to characterize the Sporothrix spp. cell wall; however, its structure and biochemical composition in the pathogenic species of the S. schenckii complex (Marimon et al. 2007) remain unknown.

As discussed above, cells of the host immune system use ROS as a microbicide to eliminate fungal pathogens (Chauhan and Latge 2006; Brown, Haynes and Quinn 2009); hence, we can hypothesize that the *Sporothrix* species ability to survive host-generated ROS plays an important role in this organism's pathogenicity. Antioxidants such as melanin and catalase contribute to the prevention of cell death following peroxide treatment (Romero-Martinez et al. 2000; Wang et al. 2008), and a recent study has shown that *S. brasiliensis* is more resistant to peroxide stress than *S. schenckii sensu stricto* (Ortega et al. 2015), suggesting that oxidative stress responses allow *Sporothrix* spp. to survive in environments where this fungal pathogen is exposed to ROS, such as inside the phagosome of a phagocytic cell.

The recent sequencing of S. schenckii sensu stricto and S. brasiliensis genomes allows in silico analyses that can identify differences and similarities between proteins identified as virulence determinants in other pathogenic organisms and Sporothrix spp. (Teixeira et al. 2014; Ortega et al. 2015). For example, bioinformatics analyses have generated the hypothesis that the Sporothrix spp. AP1-like transcription factor, which is a key modulator of oxidative stress, is more similar to that of fungal plant pathogens such as Magnaporthe oryzae, Ophiostoma piceae and Grosmannia clavigera than to that of the fungal pathogen C. albicans (Ortega et al. 2015). Once this type of analysis is made, new hypotheses can then be proposed and tested.

## CONCLUSIONS AND PERSPECTIVES ON THE HOST-SPOROTHRIX SPP. STUDIES

Even though significant progress has been made in the study of the immune response during the course of sporotrichosis, much remains to be discovered. Significant insight into the exact nature of the various microenvironments that this fungal pathogen occupies in mammalian hosts can be provided by high-throughput technologies that enable the quantification of fluctuations in the abundance of RNA transcripts (transcriptomic), proteins (proteomic) and other biomolecular components (metabolomic) (Fradin *et al.* 2003; Tierney *et al.* 2012; Muszkieta *et al.* 2013; Reales-Calderon *et al.* 2014). These technologies will aid in painting 'the big picture' of the battle happening between *Sporothrix* spp. and its hosts and in the identification of potential targets that regulate key responses that will give the advantage to either host or pathogen.

In addition, as discussed above, effective molecular tools for the study of *Sporothrix* spp., such as genetic manipulation, are still limited; the development of new or improved techniques will be required in order to study the hypotheses that will arise from high-throughput studies. In this context, the recent sequencing of the *Sporothrix* spp. genome is an important component in developing such tools (Teixeira *et al.* 2014).

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## REFERENCES

- Almeida-Paes R, Pimenta MA, Pizzini CV, et al. Use of mycelialphase Sporothrix schenckii exoantigens in an enzyme-linked immunosorbent assay for diagnosis of sporotrichosis by antibody detection. Clin Vaccine Immunol 2007;14:244–9.
- Al-Tawfiq JA, Wools KK. Disseminated sporotrichosis and Sporothrix schenckii fungemia as the initial presentation of human immunodeficiency virus infection. Clin Infect Dis 1998;26:1403–6.
- Bernardes-Engemann AR, Costa RC, Miguens BR, et al. Development of an enzyme linked immunosorbent assay for the serodiagnosis of several clinical forms of sporotrichosis. *Med Mycol* 2005;**43**:487–93.
- Bernardes-Engemann AR, de Lima Barros M, Zeitune T, et al. Validation of a serodiagnostic test for sporotrichosis: a follow-up study of patients related to the Rio de aneiro zoonotic outbreak. Med Mycol 2015;53:28–33.
- Blackman L, Cullerne D, Hardham A. Bioinformatic characterisation of genes encoding cell wall degrading enzymes in the Phytophthora parasitica genome. BMC genomics 2014;15:785.
- Bogdan C, Rollinghoff M, Diefenbach A. Reactive oxygen and reactive nitrogen intermediates in innate and specific immunity. Curr Opin Immunol 2000;12:64–76.

- Bourgeois C, Kuchler K. Fungal pathogens-a sweet and sour treat for toll-like receptors. Front Cell Infect Microbiol 2012;2:142.
- Brothers KM, Newman ZR, Wheeler RT. Live imaging of disseminated candidiasis in zebrafish reveals role of phagocyte oxidase in limiting filamentous growth. *Eukaryot Cell* 2011;**10**:932–44.
- Brown AJ, Haynes K, Quinn J. Nitrosative and oxidative stress responses in fungal pathogenicity. Curr Opin Microbiol 2009;12:384–91.
- Carlos IZ, Sgarbi DB, Angluster J, et al. Detection of cellular immunity with the soluble antigen of the fungus Sporothrix schenckii in the systemic form of the disease. Mycopathologia 1992;117:139–44.
- Carlos IZ, Sgarbi DB, Placeres MC. Host organism defense by a peptide-polysaccharide extracted from the fungus Sporothrix schenckii. Mycopathologia 1999;144:9–14.
- Carlos IZ, Sgarbi DB, Santos GC, et al. Sporothrix schenckii lipid inhibits macrophage phagocytosis: involvement of nitric oxide and tumour necrosis factor alpha. Scand J Immunol 2003;57:214–20.
- Castro RA, Kubitschek-Barreira PH, Teixeira PA, et al. Differences in cell morphometry, cell wall topography and gp70 expression correlate with the virulence of *Sporothrix brasiliensis* clinical isolates. PLoS One 2013;8:e75656.
- Chakrabarti A, Bonifaz A, Gutierrez-Galhardo MC, et al. Global epidemiology of sporotrichosis. *Med* Mycol 2015;**53**:3–14.
- Chauhan N, Latge JP. Calderone R. Signalling and oxidant adaptation in *Candida albicans* and *Aspergillus fumigatus*. Nat Rev Microbiol 2006;**4**:435–44.
- Couturier M, Roussel A, Rosengren A, et al. Structural and biochemical analyses of glycoside hydrolase families 5 and 26  $\beta$ -(1,4)-mannanases from Podospora anserine reveal differences upon manno-oligosaccharide catalysis. J Biol Chem 2013;**288**:14624–35.
- de Almeida JRF, Kaihami GH, Jannuzzi GP, et al. Therapeutic vaccine using a monoclonal antibody against a 70-kDa glycoprotein in mice infected with highly virulent Sporothrix schenckii and Sporothrix brasiliensis. Med Mycol 2015;53: 42–50.
- de Oliveira MM, Verissimo C, Sabino R, et al. First autochthone case of sporotrichosis by Sporothrix globosa in Portugal. Diagn Micr Infec Dis 2014;**78**:388–90.
- Diaz-Jimenez DF, Perez-Garcia LA, Martinez-Alvarez JA, et al. Role of the fungal cell wall in pathogenesis and antifungal resistance. *Curr Fungal Infect Rep* 2012;**6**:275–82.
- Durrens P, Green P, Arst H, et al. Heterologous insertion of transforming DNA and generation of new deletions associated with transformation in Aspergillus nidulans. Mol Gen Genet 1986;203:544–9.
- Estrada-Barcenas DA, Vite-Garin T, Navarro-Barranco H, et al. Genetic diversity of Histoplasma and Sporothrix complexes based on sequences of their ITS1–5.8S-ITS2 regions from the BOLD System. Rev Iberoam Micol 2014;**31**:90–4.
- Fernandes GF, Lopes-Bezerra LM, Bernardes-Engemann AR, et al. Serodiagnosis of sporotrichosis infection in cats by enzymelinked immunosorbent assay using a specific antigen, SsCBF, and crude exoantigens. Vet Microbiol 2011;147:445–9.
- Fernandes KS, Coelho AL, Lopes Bezerra LM, et al. Virulence of Sporothrix schenckii conidia and yeast cells, and their susceptibility to nitric oxide. Immunology 2000;**101**:563–9.
- Figueiredo CC, Lima OC, Carvalho L, et al. The *in vitro* interaction of *Sporothrix schenckii* with human endothelial cells is modulated by cytokines and involves endothelial surface molecules. *Microb Pathog* 2004;**36**:177–88.

- Fradin C, Kretschmar M, Nichterlein T, et al. Stage-specific gene expression of Candida albicans in human blood. Mol Microbiol 2003;47:1523–43.
- Fulurija A, Ashman RB, Papadimitriou JM. Early inflammatory responses to *Candida albicans* infection in inbred and complement-deficient mice. FEMS Immunol Med Micr 1996;14:83–94.
- Galagan JE, Calvo SE, Cuomo C, et al. Sequencing of Aspergillus nidulans and comparative analysis with A. fumigatus and A. oryzae. Nature 2005;438:1105–15.
- Gow NA, Hube B. Importance of the Candida albicans cell wall during commensalism and infection. *Curr Opin Microbiol* 2012;**15**:406–12.
- Gremiao ID, Menezes RC, Schubach TM, et al. Feline sporotrichosis: epidemiological and clinical aspects. Med Mycol 2015;53:15–21.
- Grimm MJ, Vethanayagam RR, Almyroudis NG, et al. Monocyteand macrophage targeted NADPH oxidase mediates antifungal host defense and regulation of acute inflammation in mice. J Immunol 2013;**190**:4175–84.
- Guzman-Beltran S, Perez-Torres A, Coronel-Cruz C, et al. Phagocytic receptors on macrophages distinguish between different Sporothrix schenckii morphotypes. Microbes Infect 2012;14:1093–101.
- Hektoen L, Perkins CF. Refractory subcutaneous abscesses caused by Sporothrix schenckii. A new pathogenic fungus. J Exp Med 1900;5:77–89.
- Henry VJ, Bandrowski AE, Pepin A-S, et al. OMICtools: an informative directory for multi-omic data analysis. Database 2014, DOI: 10.1093/database/bau069.
- Hernandez-Cervantes A, Mora-Montes HM, Alvarez-Vargas A, et al. Isolation of Sporothrix schenckii MNT1 and the biochemical and functional characterization of the encoded alpha1,2mannosyltransferase activity. Microbiology 2012;158: 2419–27.
- Hou B, Liu X, Zheng F, et al. Molecular cloning, modelling and differential expression of a gene encoding a silent information regulator-like protein from Sporothrix schenckii. Int J Mol Med 2014;33:1415–22.
- Hou B, Zhang Z, Zheng F, et al. Molecular cloning, characterization and differential expression of DRK1 in Sporothrix schenckii. Int J Mol Med 2013;31:99–104.
- Howard DH. Dimorphism of Sporotrichum schenckii. J Bacteriol 1961;81:464–9.
- Huh W-K, Falvo JV, Gerke LC, et al. Global analysis of protein localization in budding yeast. Nature 2003;425: 686–91.
- Ishizaki H, Kawasaki M, Anzawa K, et al. Mitochondrial DNA analysis of Sporothrix schenckii in India, Thailand, Brazil, Colombia, Guatemala and Mexico. Nihon Ishinkin Gakkai Zasshi 2009;**50**:19–26.
- Jacobsen ID, Luttich A, Kurzai O, et al. In vivo imaging of disseminated murine Candida albicans infection reveals unexpected host sites of fungal persistence during antifungal therapy. J Antimicrob Chemoth 2014;69:2785–96.
- Jensen J, Warner T, Balish E. The role of phagocytic cells in resistance to disseminated candidiasis in granulocytopenic mice. J Infect Dis 1994;**170**:900–5.
- Jones T, Federspiel NA, Chibana H, et al. The diploid genome sequence of Candida albicans. P Natl Acad Sci USA 2004;**101**:7329– 34.
- Kajiwara H, Saito M, Ohga S. Impaired host defense against Sporothrix schenckii in mice with chronic granulomatous disease. Infect Immun 2004;72:5073–9.

- Kano R, Nakamura Y, Watanabe S, et al. Identification of *Sporothrix schenckii* based on sequences of the chitin synthase 1 gene. Mycoses 2001;44:261–5.
- Kano R, Okubo M, Siew HH, et al. Molecular typing of Sporothrix schenckii isolates from cats in Malaysia. Mycoses 2015;**58**:220– 4.
- Kotra LP, Vakulenko S, Mobashery S. From genes to sequences to antibiotics: prospects for future developments from microbial genomics. *Microbes Infect* 2000;**2**:651–8.
- Kusuhara M, Qian H, Li X. Mouse bone marrow-derived dendritic cells can phagocytize the *Sporothrix schenc*kii, and mature and activate the immune response by secreting interleukin-12 and presenting antigens to T lymphocytes. *J Dermatol* 2014;**41**:386–92.
- Levitz SM. Innate recognition of fungal cell walls. PLoS Pathog 2010;6:e1000758.
- Lima OC, Bouchara JP, Renier G, et al. Immunofluorescence and flow cytometry analysis of fibronectin and laminin binding to Sporothrix schenckii yeast cells and conidia. Microb Pathog 2004;37:131–40.
- Lima OC, Figueiredo CC, Previato JO, et al. Involvement of fungal cell wall components in adhesion of Sporothrix schenckii to human fibronectin. Infect Immun 2001;**69**:6874–80.
- Lima OC, Lopes-Bezerra LM. Identification of a concanavalin Abinding antigen of the cell surface of Sporothrix schenckii. J Med Vet Mycol 1997;**35**:167–72.
- Liu TT, Zhang K, Zhou X. Molecular identification of Sporothrix clinical isolates in China. J Zhejiang Univ Sci B 2014;15:100–8.
- Lloyd KO, Bitoon MA. Isolation and purification of a peptidorhamnomannan from the yeast form of *Sporothrix schenckii*. Structural and immunochemical studies. *J Immunol* 1971;**107**:663–71.
- Lopes-Alves LM, Mendonca-Previato L, Fournet B, et al. O-Glycosidically linked oligosaccharides from peptidorhamnomannans of Sporothrix schenckii. Glycoconj J 1992;**9**:75–81.
- Lopes-Bezerra LM. Sporothrix schenckii cell wall peptidorhamnomannans. Front Microbiol 2011;2:243.
- Lopes-Bezerra LM, Lozoya-Perez NE, Lopez-Ramirez LA, et al. Functional characterization of Sporothrix schenckii glycosidases involved in the N-linked glycosylation pathway. Med Mycol 2015;**53**:60–8.
- Lopes-Bezerra LM, Nascimento RC. Sporothrix schenckii and general aspects of Sporotrichosis. In: Ruiz-Herrera J (ed.). Dimorphic Fungi: Their Importance as Models for Differentiation and Fungal Pathogenesis. USA: Bentham Science Publishers, 2012, 67– 83.
- Lopes-Bezerra LM, Schubach A, Costa RO. Sporothrix schenckii and sporotrichosis. Rev Bras Cienc 2006;**78**:293–308.
- Lopez-Esparza A, Alvarez-Vargas A, Mora-Montes HM, et al. Isolation of Sporothrix schenckii GDA1 and functional characterization of the encoded guanosine diphosphatase activity. Arch Microbiol 2013;**195**:499–506.
- Lopez-Romero E, Reyes-Montes M del R, Perez-Torres A, et al. Sporothrix schenckii complex and sporotrichosis, an emerging health problem. Future Microbiol 2011;6:85–102.
- Lutz A, Splendore A. A mycosis observed in humans and rats (Original title: 'Sobre uma micose observada em homens e ratos'). *Rev Med São Paulo* 1907;**21**:433–50.
- Mantione KJ, Kream RM, Kuzelova H, et al. Comparing bioinformatic gene expression profiling methods: microarray and RNA-Seq. Med Sci Monit Basic Res 2014;**20**:138–41.
- Marimon R, Cano J, Gene J, et al. Sporothrix brasiliensis, S. globosa, and S. mexicana, three new Sporothrix species of clinical interest. J Clin Microbiol 2007;**45**:3198–206.

- Marques de Macedo P, Sztajnbok DC, Camargo ZP, et al. Dacryocystitis due to Sporothrix brasiliensis: a case report of a successful clinical and serological outcome with low-dose potassium iodide treatment and oculoplastic surgery. Br J Dermatol 2015;**172**:1116–9.
- Martinez-Alvarez JA, Perez-Garcia LA, Flores-Carreon A, et al. The immune response against Candida spp. and Sporothrix schenckii. Rev Iberoam Micol 2014;**31**:62–6.
- Montenegro H, Rodrigues A, Dias M, et al. Feline sporotrichosis due to Sporothrix brasiliensis: an emerging animal infection in Sao Paulo, Brazil. BMC Vet Res 2014;**10**:269.
- Muszkieta L, Beauvais A, Pahtz V, et al. Investigation of Aspergillus 653 fumigatus biofilm formation by various "omics" approaches. Front Microbiol 2013;4:13.
- Nascimento RC, Almeida SR. Humoral immune response against soluble and fractionate antigens in experimental sporotrichosis. FEMS Immunol Med Micr 2005;**43**:241–7.
- Nascimento RC, Espindola NM, Castro RA, et al. Passive immunization with monoclonal antibody against a 70-kDa putative adhesin of Sporothrix schenckii induces protection in murine sporotrichosis. Eur J Immunol 2008;**38**:3080–9.
- Nathan C, Shiloh MU. Reactive oxygen and nitrogen intermediates in the relationship between mammalian hosts and microbial pathogens. P Natl Acad Sci USA 2000;97: 8841–8.
- Negrini Tde C, Ferreira LS, Alegranci P, et al. Role of TLR-2 and fungal surface antigens on innate immune response against Sporothrix schenckii. Immunol Invest 2013;**42**:36–48.
- Netea MG, Brown GD, Kullberg BJ, et al. An integrated model of the recognition of *Candida albicans* by the innate immune system. Nat Rev 2008;**6**:67–78.
- Ohashi H, Hasegawa M, Wakimoto K, et al. Next-generation technologies for multiomics approaches including interactome sequencing. BioMed Res Int 2015;**2015**:104209.
- Oliveira MM, Almeida-Paes R, Muniz MM, et al. Phenotypic and molecular identification of *Sporothrix* isolates from an epidemic area of sporotrichosis in Brazil. Mycopathologia 2011;**172**:257–67.
- Orofino-Costa R, Boia MN, Magalhaes GA, et al. Arthritis as a hypersensitivity reaction in a case of sporotrichosis transmitted by a sick cat: clinical and serological follow up of 13 months. Mycoses 2010;**53**:81–3.
- Ortega I, Soares Felipe MS, Vasconcelos AT, et al. Peroxide sensing and signalling in the Sporothrix schenckii complex: an in silico analysis to uncover putative mechanisms regulating the Hog1 and AP-1 like signalling pathways. *Med Mycol* 2015;**53**:51–9.
- Penha CV, Lopes-Bezerra LM. Concanavalin A-binding cell wall antigens of Sporothrix schenckii: a serological study. Med Mycol 2000;38:1–7.
- Qian Q, Jutila MA, Van Rooijen N, et al. Elimination of mouse splenic macrophages correlates with increased susceptibility to experimental disseminated candidiasis. J Immunol 1994;152:5000–8.
- Reales-Calderon JA, Sylvester M, Strijbis K, et al. Candida albicans induces pro-inflammatory and anti-apoptotic signals in macrophages as revealed by quantitative proteomics and phosphoproteomics. J Proteomics 2013;91:106–35.
- Reis RS, Almeida-Paes R, Muniz M de M, et al. Molecular characterisation 690 of Sporothrix schenckii isolates from humans and cats involved in the sporotrichosis epidemic in Rio de Janeiro, Brazil. Mem I Oswaldo Cruz 2009;**104**:769–74.
- Robledo-Ortiz CI, Flores-Carreon A, Hernandez-Cervantes A, et al. Isolation and functional characterization of Sporothrix

schenckii ROT2, the encoding gene for the endoplasmic reticulum glucosidase II. Fungal Biol 2012;**116**:910–8.

- Rodrigues AM, de Hoog GS, de Camargo ZP. Emergence of pathogenicity in the Sporothrix schenckii complex. Med Mycol 2013;51:405–12.
- Rodrigues AM, de Hoog GS, de Camargo ZP. Genotyping species of the Sporothrix schenckii complex by PCR-RFLP of calmodulin. Diagn Microbiol Infect Dis 2014;**78**:383–7.
- Rodrigues AM, de Melo Teixeira M, de Hoog GS, et al. Phylogenetic analysis reveals a high prevalence of Sporothrix brasiliensis in feline sporotrichosis outbreaks. PLoS Negl Trop Dis 2013;7:e2281.
- Rodrigues AM, Kubitschek-Barreira PH, Fernandes GF, et al. Immunoproteomic analysis reveals a convergent humoral response signature in the Sporothrix schenckii complex. J Proteomics 2015;115:8–22.
- Rodriguez-Caban J, Gonzalez-Velazquez W, Perez-Sanchez L, et al. Calcium/calmodulin kinase1 and its relation to thermotolerance and HSP90 in Sporothrix schenckii: an RNAi and yeast two-hybrid study. BMC Microbiol 2011;**11**:162.
- Romero-Martinez R, Wheeler M, Guerrero-Plata A, et al. Biosynthesis and functions of melanin in Sporothrix schenckii. Infect Immun 2000;**68**:3696–703.
- Romo-Lozano Y, Hernandez-Hernandez F, Salinas E. Mast cell activation by conidia of *Sporothrix schenckii*: role in the severity of infection. *Scand J Immunol* 2012;**76**:11–20.
- Ruiz-Baca E, Mora-Montes HM, Lopez-Romero E, et al. 2Dimmunoblotting analysis of Sporothrix schenckii cell wall. Mem I Oswaldo Cruz 2011;**106**:248–50.
- Ruiz-Baca E, Toriello C, Perez-Torres A, et al. Isolation and some properties of a glycoprotein of 70 kDa (Gp70) from the cell wall of Sporothrix schenckii involved in fungal adherence to dermal extracellular matrix. *Med* Mycol 2009;**47**:185–96.
- Sanchez-Lopez JF, Gonzalez-Ibarra J, Alvarez-Vargas A, et al. Isolation of the GFA1 gene encoding glucosamine-6-phosphate synthase of Sporothrix schenckii and its expression in Saccharomyces cerevisiae. Protein Expres Purif 2015;**110**:57–64.
- Sasaki AA, Fernandes GF, Rodrigues AM, et al. Chromosomal polymorphism in the Sporothrix schenckii complex. PLoS One 2014;9:e86819.
- Sassá MF, Ferreira LS, Ribeiro LC, et al. Immune response against Sporothrix schenckii in TLR-4-deficient mice. Mycopathologia 2012;**174**:21–30.
- Sassá MF, Saturi AE, Souza LF, et al. Response of macrophage Toll-like receptor 4 to a Sporothrix schenckii lipid extract during experimental sporotrichosis. Immunology 2009;**128**:301–9.
- Schenck BR. On refractory subcutaneous abscesses caused by a fungus possibly related to the Sporotricha. Bull Johns Hopkins Hosp 1898;9:286–90.
- Schubach A, Barros MB, Wanke B. Epidemic sporotrichosis. Curr Opin Infect Dis 2008;21:129–33.
- Shiraishi A, Nakagaki K, Arai T. Role of cell-mediated immunity in the resistance to experimental sporotrichosis in mice. Mycopathologia 1992;**120**:15–21.

- Spencer JFT, Spencer DM, Reynolds N. Genetic manipulation of non-conventional yeasts by conventional and non-conventional methods. J Basic Microbiol 1988;28: 321–33.
- Teixeira MM, de Almeida LG, Kubitschek-Barreira P, et al. Comparative genomics of the major fungal agents of human and animal sporotrichosis: Sporothrix schenckii and Sporothrix brasiliensis. BMC Genomics 2014;**15**:943.
- Teixeira PAC, Castro RA, Nascimento RC, et al. Cell surface expression of adhesins for fibronectin correlates with virulence in Sporothrix schenckii. Microbiology 2009;155: 3730–8.
- Tellez MD, Batiste-Duharte A, Portuondo D, et al. Sporothrix schenckii complex biology: environment and fungal pathogenicity. *Microbiology* 2014;**160**:2352–65.
- Tierney L, Linde J, Muller S, et al. An interspecies regulatory network inferred from simultaneous RNA-seq of Candida albicans invading innate immune cells. Front Microbiol 2012;3: 85.
- Travassos LR, Lloyd KO. Sporothrix schenckii and related species of Ceratocystis. Microbiol Rev 1980;44:683–721.
- Trotter JR, Sriaroon P, Berman D, et al. Sporothrix schenckii lymphadentitis in a male with X-linked chronic granulomatous disease. J Clin Immunol 2014;**34**:49–52.
- Uenotsuchi T1, Takeuchi S, Matsuda T, et al. Differential induction of Th1-prone immunity by human dendritic cells activated with Sporothrix schenckii of cutaneous and visceral origins to determine their different virulence. Int Immunol 2006;**18**:1637–46.
- Valentin-Berrios S, Gonzalez-Velazquez W, Perez-Sanchez L, et al. Cytosolic phospholipase A2: a member of the signalling pathway of a new G protein alpha subunit in Sporothrix schenckii. BMC Microbiol 2009;9:100.
- Valle-Aviles L, Valentin-Berrios S, Gonzalez-Mendez RR, et al. Functional, genetic and bioinformatic characterization of a calcium/calmodulin kinase gene in Sporothrix schenckii. BMC Microbiol 2007;7:107.
- Vonk AG, Netea MG, van der Meer JW, et al. Host defence against 765 disseminated Cand/ida albicans infection and implications for antifungal immunotherapy. Expert Opin Biol Th 2006;6:891–903.
- Wang XH, Li RY, Cao CW, et al. Differential mRNA expression of Sporothrix schenckii catalase gene in two growth phases and effect factors. Chin Med J 2008;**121**:2100–2.
- Yu X, Wan Z, Zhang Z, et al. Phenotypic and molecular identification of Sporothrix isolates of clinical origin in Northeast China. Mycopathologia 2013;**176**:67–74.
- Zhang Y, Li G, He D, et al. Efficient insertional mutagenesis system for the dimorphic pathogenic fungus Sporothrix schenckii using Agrobacterium tumefaciens. J Microbiol Meth 2011;84: 418–22.
- Zhou X, Rodrigues AM, Feng P, et al. Global ITS diversity in the Sporothrix schenckii complex. Fungal Divers 2013;66: 153–65.