Forestry 2015; **88**, 521–527, doi:10.1093/forestry/cpv022 Advance Access publication 26 June 2015

Large trees and dense canopies: key factors for maintaining high epiphytic diversity on trunk bases (bryophytes and lichens) in tropical montane forests

Á. Benítez¹, M. Prieto² and G. Aragón^{2*}

¹Sección de Sistemática y Diversidad, Departamento de Ciencias Naturales, Universidad Técnica Particular de Loja, San Cayetano s/n, Loja, Ecuador
²Área de Biodiversidad y Conservación, Departamento de Biología y Geología, Universidad Rey Juan Carlos, Móstoles E-28933, Madrid, España

*Corresponding author. E-mail: gregorio.aragon@urjc.es

Received 15 January 2015

The high richness of epiphytes in moist tropical montane forests is continuously decreasing due to deforestation and habitat loss. Lichens and bryophytes are important components of epiphyte diversity on trunk bases and play an important role in the water balance and nutrient cycling of tropical montane forests. As lichens and bryophytes are very sensitive to microclimatic changes, we hypothesized that their species richness and composition would change with forest alteration. We also expected their response patterns to be different given the capability of lichens to photosynthesize using water vapour. In this study, we assessed the richness and composition of epiphytes (lichens and bryophytes) on the trunk bases of 240 trees in primary and secondary forests of southern Ecuador. We found that diversity was higher in primary forests and lower in monospecific secondary forest stands. Total diversity was negatively affected by habitat loss and by the reduction of canopy cover for bryophytes. Shade epiphytes were replaced by sun epiphytes in open secondary forests. We conclude that lichen and bryophyte diversity of tropical montane forests are negatively affected by the removal of large trees and canopy disruption. The different species compositions of primary and secondary forests and the high number of species exclusive to primary forests indicate that secondary forests are of limited importance in compensating for the loss of non-vascular epiphyte species associated with primary forests.

Introduction

Neotropical montane rain forests are considered 'hot spots' of global biodiversity and are a high conservation priority (Gentry, 1995; Myers et al., 2000; Dirzo and Raven, 2003). Epiphytes constitute an important floristic, structural and functional component in these forests (Barthlott et al., 2001; Gradstein, 2008; Köster et al., 2009); however, this exceptional diversity is threatened by continued deforestation and habitat loss (Churchill et al., 1995; Bruijnzeel and Hamilton, 2000; Gibbs et al., 2010). Forest conversion produces changes that directly influence epiphyte diversity: abiotic conditions are altered, habitat complexity (i.e. tree size, tree species and canopy structure) is reduced, and dispersal is constrained (Werner et al., 2005, 2011; Hietz et al., 2006). In fact, several authors have found a loss of epiphytic diversity (including vascular plants, bryophytes and lichens) in secondary forests and a higher diversity in primary forests (PFs) (Barthlott et al., 2001; Acebey et al., 2003; Krömer and Gradstein 2003; Wolf, 2005; Gradstein 2008; Gradstein and Sporn, 2010).

Non-vascular epiphytes (i.e. bryophytes and lichens) constitute an important fraction of epiphytic organisms in tropical montane forests in terms of diversity, biomass and nutrient cycling (Pócs, 1982; Sipman, 1995; Holz and Gradstein, 2005; Mandl *et al.*, 2010; Gehrig-Downie *et al.*, 2011). Due to their poikilohydric nature, these organisms are tolerant to desiccation (Pardow and Lakatos, 2013), even though their degree of desiccation tolerance varies greatly among species (Proctor *et al.*, 2007; Kranner *et al.*, 2008). In particular, lichens and bryophytes in humid sites in tropical forests, mainly the forest understory and inner parts of the canopy, are highly sensitive to desiccation (Kranner *et al.*, 2008; Pardow and Lakatos, 2013) and may experience photoinhibition when exposed to a small rise in solar radiation (Sillett and Antoine, 2004; Green *et al.*, 2008; Pardow and Lakatos, 2013). As the physiology of these organisms is strongly linked to ambient moisture, solar radiation and temperature (Gignac, 2001; Sillett and Antoine, 2004; Green *et al.*, 2008), forest logging and land use may greatly affect the diversity of non-vascular epiphytic communities.

The canopy disruption caused by forest logging can affect the humidity, temperature and light conditions inside forests, making them unsuitable sites for shade-adapted species (Gradstein, 2008; Gradstein and Sporn, 2010; Normann *et al.*, 2010; Benitez *et al.*, 2012). Open forests are generally drier, warmer and windier compared with closed forests, where moisture content is higher and less variable (Gradstein, 2008). However, these microclimate changes do not necessarily involve a decrease in species

© Institute of Chartered Foresters, 2015. All rights reserved. For Permissions, please e-mail: journals.permissions@oup.com.

Institute of

Chartered Foresters



richness, but rather a replacement in community composition (Holz and Gradstein, 2005; Nöske *et al.*, 2008). The more shade-adapted lichens and bryophytes that are intolerant to desiccation are often replaced by heliophytic species (Ariyanti *et al.*, 2008; Gradstein, 2008; Gradstein and Sporn, 2010; Benítez *et al.*, 2012).

Forest logging may also have immediate negative effects on the persistence of bryophytes and lichens due to the removal of host tree species (Gradstein, 2008). Host tree characteristics, especially tree size, play an important role in lichen and bryophyte colonization (Benítez *et al.*, 2012; Rosabal *et al.*, 2013), probably due to greater bark surface available for colonization on large trees and the creation of additional microhabitats (Fritz *et al.*, 2008; Ranius *et al.*, 2008). Epiphytic diversity may also be influenced by bark roughness, humus and moss cover on the bark surface, stochastic effects of species dispersion, and to lesser extent, bark pH (Sipman and Harris, 1989; Cáceres *et al.*, 2007; Gradstein and Culmsee, 2010; Soto *et al.*, 2012).

As a result of human activities in Ecuador, PFs have often been replaced by secondary vegetation, creating forests with a lessdeveloped canopy structure, smaller trees and less tree diversity. Benítez et al. (2012) found that the diversity of 'shade epiphytes' decreased drastically as a result of such forest disturbance. This could be due to the high percentage of the macrolichen species belonging to the order Peltigerales (\sim 50 per cent), as these species are adapted to within forest conditions, have high water demands and are sensitive to high solar radiation. However, as macrolichens represent less than one-third of all poikilohydric epiphytic species in tropical montane forests, these results should be interpreted with caution when considering epiphytic communities as a whole (bryophytes and lichens). Knowledge of the differences in epiphytic diversity in primary and secondary forests is crucial to evaluate the conservation status of these forests and to design conservation strategies.

The goal of this study was to explore the response of the non-vascular epiphytic community to forest logging in tropical montane rain forests. We hypothesized that differences in species diversity and community composition would be related to differences in forest structure and microclimate caused by the intensity of forest logging. Another objective was to compare the response patterns between bryophytes and lichens, as lichens prefer relatively high light levels (excluding some cyanolichens) (Sillett and Antoine, 2004; Green *et al.*, 2008; Normann *et al.*, 2010) and are generally less negatively affected by drought than bryophytes (Perhans *et al.*, 2009).

Materials and methods

Study area

This study was carried out at two sites in southern Ecuador and included six remnants of tropical montane forests along a disturbance gradient (Table 1). The climate is humid tropical with a mean annual temperature of 20°C, an annual rainfall of ~1900 mm and a relative humidity of ~80 per cent (National Institute of Meteorology and Hydrology, INAMI). The altitude of the studied plots ranged from 2200 to 2800 m a.s.l.

Field work was carried out in three types of forest vegetation varying in age, species composition and tree cover: (1) remnant PF fragments of evergreen tropical montane forests characterized by a dense canopy layer (\sim 75-85 per cent cover) and large trees (35-40 m tall). The main canopy trees were Cinchona macrocalyx Pav. ex DC., Clusia elliptica Kunth, Myrica pubescens Humb. & Bonpl. ex Willd., Podocarpus oleifolius D. Don ex Lamb. and Weinmannia pubescens Kunth. (2) Secondary mixed forest fragments (SF) regrown after selective logging events which took place ca. 40 years earlier (Brown and Lugo, 1990; Holz, 2003). Canopy cover was \sim 60 - 70 per cent, and the main canopy trees were *Melastomataceae* and Lauraceae species (25-30 m tall). (3) Secondary monospecific forests of Alnus acuminata Kunth (MF; 35-40 year old) regrown by natural regeneration after forest clearing (Hofstede and Aquirre, 1999). This tree is a pioneer and native species of the Andes. Monospecific forests are characterized by their uniform structure, absence of understory plants, \sim 50 per cent canopy cover and trees up to 20 m tall. Logging and firewood extraction were the main contemporary human activities in MF, whereas there were no human activities in PF and SF.

Species identification

For species identification, we used >200 taxonomic and floristic papers (e.g. Gradstein *et al.*, 2001; Gradstein and Costa, 2003; Frisch *et al.*, 2006; Cáceres, 2007; Aptroot *et al.*, 2008; Timdal, 2008; Lücking, 2009; Moncada *et al.*, 2013). For species nomenclature, we followed mainly Tropicos.org for bryophytes and MycoBank for lichens.

Experimental design

We sampled two stands of each forest type (PF, SF and MF). We established ten 5×5 m plots in each stand for a total of 60 plots. The distance between the plots in each forest stand was >50 m. In each plot, epiphytic lichens and bryophytes were sampled on the bases of 4 mature trees (total of 240 trees) using 20×30 cm grids. Samples were taken on each tree at three different heights (0–50, 51–150, and 151–200 cm) on the northern and southern exposure for a total of six samples per tree. Species richness was defined as the total number of species found in each plot. For epiphytic composition, we estimated the mean cover of each species (per cent of grid area) per tree

Table 1 Means of the environmental variables in the studied	l primary and secc	ondary montane forests (two st	ands of each forest type) in Ecuador
	i printiary ana sece		

Forest	Location	Canopy cover (%)	Tree diameter (cm)	Elevation (masl)	Slope (°)	Aspect
PF1	4° 33′ 27″ S, 79° 22′ 9″ W	78.0	33.1	2848	26.6	E-SW
PF2	4° 33′ 54″ S, 79° 22′ 13″ W	76.5	34.4	2586	32.8	SW-N
SF1	4° 3′ 9″ S, 79° 9′ 55″ W	67.5	28.2	2688	38.2	NW-NE
SF2	4° 33′ 35″ S, 79° 23′ 21″ W	66.0	26.5	2393	32.6	E-SW
MF1	4° 2′ 36″ S, 79° 10′ 20″ W	51.5	16.2	2377	26.4	E-SW
MF2	3° 59′ 53″ S, 79° 10′ 46″ W	54.0	19.6	2196	18.1	NE-SW

PF, primary forest; SF, mixed secondary forest; MF, monospecific secondary forest of A. acuminata.

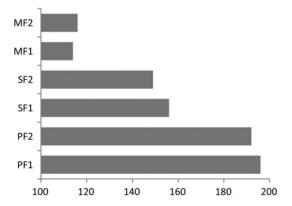
and per plot (as the percentage of four trees). We also measured the following variables at the plot level: canopy cover (per cent), elevation (m a.s.l.), slope (°), aspect (cosine transformed) and mean tree DBH (cm) of the four trees analysed per plot as a proxy for stand structure.

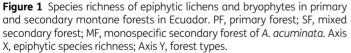
Data analyses

Richness and diversity

We determined the effect of the environmental variables (canopy cover, mean DBH, elevation, aspect and slope) on the following community traits: total species richness, lichen richness, bryophyte richness and species diversity (Simpson inverse and Shannon indices). The Simpson and Shannon indices allow data on species richness and relative abundance to be combined (Gorelick, 2006). The Simpson index was determined by the predominant species, and the Shannon index was based on the assumption that individuals were randomly selected and that all species were represented in the sample (Magurran, 2004). Although host trees have a great influence on epiphyte diversity in temperate regions, the effect of host tree was not explored, as host-specificity does not seem to play an important role in tropical forests with a relatively high diversity of tree species (Sipman and Harris, 1989; Cáceres *et al.*, 2007; Rosabal *et al.*, 2013).

The effects of slope, aspect, elevation, canopy cover and mean tree diameter on species richness, the Shannon index and Simpson inverse index were analysed at the plot level using generalized linear mixed models (GLMMs) (McCullagh and Nelder, 1989; Verbeke and Molenberghs,





1997). Because forest stands were quite far apart (Benitez *et al.*, 2012), stand distance was initially included in the models, but it was later removed as no significant differences were detected. Predictors were included as explanatory variables (fixed factors), and forest and plot were included as random sources of variation. Effects of random factors were tested using the Wald Z-statistic test. We fitted the mixed models using a normal distribution with an 'identity' link function. All GLMM computations were performed using SAS (GLIMMIX version 8 for SAS/STAT).

We measured total species richness, lichen richness and bryophyte richness at the forest level, as the total species identified on 40 trees in each forest. Sampling completeness at the forest level was estimated with *Chao2* species richness estimator, using EstimateS 9.1.0 (Colwell, 2013).

Species composition and community structure

Non-metric multidimensional scaling (NMDS) ordination was performed to detect the main factors influencing epiphytic composition. NMDS analyses were carried out using CRAN software R (R Core Team 2013) with vegan package (Oksanen *et al.*, 2013). For the NMDS analyses, the Bray-Curtis distance was used, as it is one of the most effective measures for community data (McCune *et al.*, 2002). The coefficients of determination (r^2) for the predictor variables were calculated with ordination axes to interpret the relationships between the variables and community composition (1000 permutations).

Bray-Curtis dissimilarity between plots within a forest was calculated as a measure of species replacement. A pairwise PERMANOVA test using Bray-Curtis distance was also performed to assess species similarity among the three types of forest vegetation. Statistical analysis was performed using version 6.1.11 of PRIMER multivariate statistical analysis software (Anderson *et al.*, 2008), allowing 9999 random permutations under the reduced model.

Results

Richness and diversity

A total of 374 epiphytic species (307 lichens and 67 bryophytes) were collected in the 60 plots (Supplementary data) The highest number of species was observed in PFs with 234 species, followed by secondary mixed forests (SF) with 191 species and monospecific secondary forests with 134 species (Figure 1; Table 2). A similar pattern was observed for the richness estimator (*Chao* 2), confirming the occurrence of the highest species richness in PF (Table 2). Fifty-four species were exclusive to PF, exceeding the number of species exclusive to SF (Supplementary data). Species replacement

 Table 2
 Species richness and dissimilarity of bryophytes and lichens at the forest level

	Bryophytes		Lichens	
	Observed species (Chao 2; SE)	Bray-Curtis dissimilarity (%)	Observed species (Chao 2; SE)	Bray-Curtis dissimilarity (%)
PF1	44 (46; 3.42)	70.79	152 (173; 9.25)	76.53
PF2	42 (44; 2.53)	72.51	150 (157; 5.16)	73.26
SF1	35 (36; 1.17)	67.83	121 (128; 5.05)	68.79
SF2	31 (32; 2.13)	69.22	118 (132; 7.71)	67.41
MF1	23 (23: 0.04)	58.04	91 (93: 2.06)	57.85
MF2	26 (27; 2.04)	49.8	86 (92; 3.86)	59.34

Chao 2 estimates of total richness are shown in brackets.

SE, standard error; PF, primary forest; SF, mixed secondary forest; MF, monospecific secondary forest of A. acuminata.

Table 3 Results of the generalized mixed linear models on communitytraits at the plot level including beta coefficients (Coef.) and associatedSEs

Plot level	Coef. (SE)	F-value	P-value
Total richness			
Mean tree diameter	0.009 (0.002)	14.46	0.001
Canopy cover	0.002 (0.002)	1.45	0.235
Elevation	0.032 (0.016)	3.95	0.054
Slope	<-0.001 (0.001)	0.09	0.771
Aspect	<0.001 (0.000)	2.04	0.179
Bryophytes richness			
Mean tree diameter	0.208 (0.070)	8.74	0.005
Canopy cover	0.146 (0.062)	5.62	0.021
Elevation	0.121 (0.501)	0.06	0.811
Slope	0.018 (0.027)	0.42	0.522
Aspect	0.002 (0.004)	0.30	0.591
Lichen richness			
Mean tree diameter	0.430 (0.137)	9.88	0.003
Canopy cover	-0.039 (0.113)	0.12	0.732
Elevation	0.061 (0.053)	3.60	0.052
Slope	-0.019 (0.051)	0.13	0.721
Aspect	0.005 (0.004)	1.63	0.207
Shannon index			
Mean tree diameter	0.017 (0.006)	7.54	0.008
Canopy cover	<-0.001 (0.005)	0.01	0.934
Elevation	0.046 (0.042)	1.17	0.285
Slope	< 0.001 (0.002)	0.02	0.901
Aspect	<-0.001 (0.000)	0.01	0.922
Simpson inverse index			
Mean tree diameter	0.445 (0.173)	6.60	0.013
Canopy cover	0.008 (0.151)	0.01	0.957
Elevation	1.252 (1.230)	1.04	0.313
Slope	0.027 (0.067)	0.16	0.690
Aspect	-0.007 (0.008)	0.73	0.406

Significant values are in bold.

(as a measure of dissimilarity) was also higher in PF for both lichens and bryophytes (Table 2). Analysis of environmental variables showed that tree diameter was the most relevant predictor of species richness at the plot level (Table 3). Canopy cover had a significant effect on bryophyte richness. The random variable forest was non-significant in all cases.

Species composition and community structure

NMDS ordination resulted in a two-dimensional pattern with an average stress of 13.22 and showed a clear separation of the three different forest types. Most of the variability was explained by Axis 1 ($r^2 = 0.69$), followed by Axis 2 ($r^2 = 0.12$, Figure 2). Axis 1 was associated with changes in canopy cover (Axis 1 = -0.926, Axis 2 = +0.378, $r^2 = 0.712$, P = 0.001) and tree diameter (Axis 1 = -0.8333, Axis 3 = +0.553, $r^2 = 0.539$, P = 0.001). The pairwise test revealed significant differences in epiphytic composition between the three types of forest vegetation: PF vs SF (66.40 per cent dissimilarity, P = 0.025), SF vs MF (75.00 per cent dissimilarity, P = 0.015).

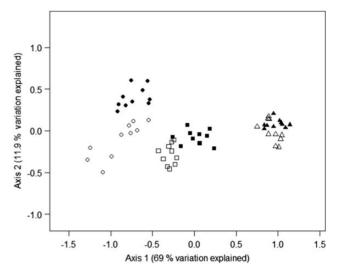


Figure 2 Non-metric multidimensional scaling analysis of species composition for the samples (plots) in the studied primary and secondary montane forests (two stands of each forest type) in Ecuador. PF, primary forest (circle); SF, mixed secondary forest (square); MF, monospecific secondary forest of *Alnus acuminata* (triangle).

Herbertus divergens, Coccocarpia filiformis, C. pellita, Coenogonium eximium and Cryptothecia exilis correlated with a dense canopy and large trees as found in PF, whereas Frullania brasiliensis, F. gibbosa, Metzgeria lechleri, Graphis anfractuosa, G. cinerea, Heterodermia diademata and H. hypochraea correlated with a more open canopy and smaller trees, characteristic of SF and MF (Supplementary data).

Discussion

Our results showed significant changes in non-vascular epiphytic diversity (lichens and bryophytes) related to forest alteration in montane tropical forests. Major shifts in species diversity were caused by changes in canopy cover and tree size. Thus, epiphytic diversity was higher in PF than in the forests with more altered vegetation. In these two forest types, diversity was higher in mixed (SF) than in monospecific (MF) secondary forests. These results are consistent with other studies on epiphyte diversity in tropical montane forests (e.g. Acebey et al., 2003; Wolf, 2005; Werner and Gradstein, 2009) and support the notion that forest alteration leads to species loss in these communities. These data further indicate that species loss is related to the degree of forest alteration (i.e. selective logging, clear-cut, plantation) (Ariyanti et al., 2008; Sporn et al., 2009; Gradstein and Sporn, 2010) and the time since disturbance (Holz and Gradstein, 2005; Gradstein, 2008). As at least one hundred years are needed for the complete recovery of epiphyte diversity in montane forests (Holz and Gradstein, 2005), the maintenance of PFs is crucial in the conservation of tropical rain forest biodiversity (Gibson et al., 2011).

We also found that lichens and bryophytes responded differently to forest disturbance. Species loss in lichens mainly correlated with reduced tree size, whereas species loss in bryophytes was also significantly related to climatic changes (i.e. increase in solar radiation, decrease in air humidity) induced by lower canopy cover in SF and MF. A high, dense canopy promotes optimal climatic

conditions inside forests for the growth of shade epiphytes which have higher water demands and are very sensitive to solar radiation (Sillett and Antoine, 2004; Gradstein, 2008; Benítez et al., 2012; Pardow and Lakatos, 2013). The irradiation in closed forests is converted into heat at the interface of the atmosphere and the canopy, maintaining moist and cool conditions in the forest understory (Hohnwald, 1999, cited in Werner and Gradstein, 2009). Canopy disruption caused by selective logging produces small openings in the canopy (5-10 per cent), which can significantly affect ambient moisture (Zimmerman and Kormos, 2012) and lead to a decrease in the diversity of shade epiphytes, adapted to the moist, shaded interior of the forest (Sipman and Harris, 1989; Acebey et al., 2003; Gradstein, 2008; Gradstein and Sporn, 2010). However, while bryophytes experienced species loss due to high irradiation and evaporation stress in more open habitats (Perhans et al., 2009), total lichen richness was not reduced by these factors. This may be because some of the more shade-adapted species (shade epiphytes) were replaced by lightdemanding species (sun epiphytes) especially in MF where canopy openness was the highest (\sim 50 per cent). Thus, open secondary montane forests can support a high richness of epiphytic lichens, even though there are fewer shade epiphytes (Hietz et al., 2006; Nöske et al., 2008).

Species composition of both bryophytes and lichens was severely altered by the increase in canopy openness, indicating that community composition is a more sensitive indicator of human impact than species richness (Nöske et al., 2008). In general, shade epiphytes are more sensitive to environmental changes, because they are strongly dependent on atmospheric moisture and experience photoinhibition when exposed to greater sunlight than in their normal environment (Gauslaa et al., 2001; Green et al., 2008; Kranner et al., 2008). Ariyanti et al. (2008) found that microclimatic changes related to the loss of shaded cover were responsible for shifts in bryophyte composition. In our study, differences in species composition between the three forest types were particularly noticeable in the higher number of species of the liverwort genus Plagiochila and the lichen genera Coccocarpia, Coenogonium, Herpothallon, Leptogium and Sticta in PFs vs. species of the lichen genera Graphis, Heterodermia or Parmotrema in secondary forests. Biological characteristics of lichens exclusive to PFs are the predominance of the photobiont with a reduction of the mycobiont (Coenogonium) or the presence of cyanobacteria as photobionts, constituting the so-called cyanolichens (Leptogium, Coccocarpia, Sticta) (Green et al., 2008; Benítez et al., 2012). However, some cyanolichen species (e.g. Coccocarpia stellata, Leptogium azureum, L. chloromelum and Sticta weigelii) may also occur in open, relatively dry habitats (Normann et al., 2010; Rosabal et al., 2010). In this sense, and focusing on these cyanolichens, we observed a contrasted vertical and horizontal zonation along the trunks. In the drier and more open sites along our gradient (A. acuminata forests), these species were more common on tree bases (<50 cm) and on northern exposures where light incidence was lower. However, these species in PFs were located at higher elevations (151-200 cm) on both exposures (north and south).

One of the major problems faced by tropical forests is the harvesting of large, long-lived and slow-growing trees (Zimmerman and Kormos, 2012), as they have the greatest bark surface area and the greatest formation of specialized aged bark substrates (e.g. Fritz *et al.*, 2008; Johansson *et al.*, 2009; Király *et al.*, 2013). We suggest that these features, which are absent on younger,

smaller trees, are preferred by epiphyte species, which might explain the high species replacement (measured as dissimilarity) between PF and MF.

Conclusion

Species diversity of non-vascular epiphytes (lichens and bryophytes) growing on the trunk bases of tropical montane forests is negatively affected by forest alteration in two ways: (1) removal of hosts, especially large trees and (2) environmental changes caused by canopy disruption. Opposite to Dent and Wright (2009), who pointed the importance of secondary forests in terms of supporting tropical biodiversity, our analyses showed different species composition of primary and secondary forests and a high number of species found exclusively in PFs, thus suggesting that secondary forests are of limited importance in compensating for the loss of epiphytic species in PFs. Although this study contributes to the knowledge of these organisms and their dynamics in tropical ecosystems, we should consider the constraints related to the number of replicates per forest type. Since the response of lichens and bryophytes to new environmental conditions caused by the increase in canopy openness is related to their morphological and anatomical characteristics (e.g. growth form, thallus thickness, type of photobiont and cortical pigments), more studies on the functional traits of epiphytes are needed to better understand their response to forest disturbance.

Supplementary data

Supplementary data are available at Forestry online.

Acknowledgements

The authors thank X. Yadira González for field assistance, Dr S. Robbert Gradstein, Dr Chris Johnson and anonymous reviewers for their valuable comments, corrections and suggestions, which helped us to improve the manuscript, and Lori De Hond for English revision and comments.

Conflict of interest statement

None declared.

Funding

Financial support for this study was received from the Universidad Técnica Particular de Loja, the Secretaria Nacional de Educación Superior, Ciencia, Tecnología e Innovación of Ecuador and the Ministerio de Ciencia e Innovación of Spain (project EPICON, CGL2010-22049).

References

Acebey, A., Gradstein, S.R. and Krömer, T. 2003 Species richness and habitat diversification of bryophytes in submontane rain forest and fallows of Bolivia. *J. Trop. Ecol.* **19**, 9–18.

Anderson, M.J., Gorley, R.N. and Clarke, K.R. 2008 PERMANOVA+ for PRIMER: Guide to Software and Statistical Methods. PRIMER-E.

Aptroot, A., Lücking, R., Sipman, H.J.M., Umaña, L. and Chaves, J.L. 2008 Pyrenocarpous lichens with bitunicate asci. A first assessment of the lichen biodiversity inventory in Costa Rica. *Bibl. Lichenol.* **97**, 1–162. Ariyanti, N.S., Bos, M.M., Kartawiniata, K., Tjitrosoedirdjo, S.S., Guhadja, E. and Gradstein, S.R. 2008 Bryophytes on tree trunks in natural forests, selectively logged forests and cacao agroforests in Central Sulawesi, Indonesia. *Biol. Conserv.* **141**, 2516–2527.

Barthlott, W., Schmit-Neuerburg, V., Nieder, J. and Engwald, S. 2001 Diversity and abundance of vascular epiphytes: a comparison of secondary vegetation and primary montane rain forest in the Venezuelan Andes. *Pl. Ecol.* **152**, 145–156.

Benitez, A.R., Prieto, M., González, X.Y. and Aragón, G. 2012 Effects of tropical montane forest disturbance on epiphytic macrolichens. *Sci. Tot. Environ.* **441**, 169–175.

Brown, S. and Lugo, A.E. 1990 Tropical secondary forests. *J. Trop. Ecol.* **6**, 1–32.

Bruijnzeel, L.A. and Hamilton, L.S. 2000 Decision time for cloud forests. *IHP Humid Trop. Progr. Ser.* **13**, 1–40.

Cáceres, M.E.S. 2007 Corticolous and microfoliose lichens of northeastern Brazil. *Libri Bot.* **22**, 1–168.

Cáceres, M.E.S., Lücking, R. and Rambold, G. 2007 Phorophyte specificity and environmental parameters versus stochasticity as determinants for species composition of corticolous crustose lichen communities in the Atlantic rain forest of northeastern Brazil. *Mycol Progress* **10**, 190–210.

Churchill, S.P., Balslev, H., Forero, E. and Luteyn, J.L. 1995 *Biodiversity and Conservation of Neotropical Montane Forests*. New York Botanical Garden Bronx.

Colwell, R.K. 2013 Statistical estimation of species richness and share species from samples. http://viceroy.eeb.uconn.edu/estimates/ (accessed on October 2014).

Dent, D.H. and Wright, S.J. 2009 The future of tropical species in secondary forests: a quantitative review. *Biol. Conserv.* **142**, 2833–2843.

Dirzo, R. and Raven, P.H. 2003 Global State of Biodiversity and Loss. *Ann. Rev. Environ. Resour.* **28**, 137–167.

Frisch, A., Kalb, K. and Grube, M. 2006 Contributions towards a new systematics of the lichen family Thelotremataceae. *Bibl. Lichenol.* **92**, 3-556.

Fritz, Ö., Niklasson, M. and Churski, M. 2008 Tree age is a factor for the conservation of epiphytic lichens and bryophytes beech forest. *Appl. Veg.* Sci. **12**, 93–106.

Gauslaa, Y., Ohlson, M., Solhaug, K.A., Bilger, W. and Nybakken, L. 2001 Aspect-dependent high-irradiance damage in two transplanted foliose forest lichens, *Lobaria pulmonaria* and *Parmelia sulcata. Can. J. For. Research* **31**, 1639–1649.

Gehrig-Downie, Ch., Obregón, A., Bendix, J. and Gradstein, S.R. 2011 Epiphyte biomass and canopy microclimate in the tropical lowland cloud forest of French Guiana. *Biotropica* **43**, 591–596.

Gentry, A.H. 1995 Patterns of diversity and floristic composition in neotropical montane forests. In *Biodiversity and Conservation of Neotropical Montane Forests*. Churchill, S.P., Balslev, H., Forero, E. and Luteyn, J.L. (eds). New York Botanical Garden Bronx, pp. 103–126.

Gibbs, H.K., Reusch, A.S., Achard, F., Clayton, M.K., Holmgren, P., Ramankutty, N. *et al.* 2010 Tropical forests were the primary sources of new agricultural lands in the 1980s and 1990s. *Proc. Natl Acad. Sci.* **107**, 16732–16737.

Gibson, L., Lee, T.M., Koh, L.P., Brook, B.W., Gardner, T.A., Barlow, J. *et al.* 2011 Primary forests are irreplaceable for sustaining tropical biodiversity. *Nature* **478**, 378–381.

Gignac, L.D. 2001 Bryophytes as indicators of climate change. *Bryologist* **104**, 410-420.

Gorelick, R. 2006 Combining richness and abundance into a single diversity index using matrix analogues of Shannon's and Simpson's indices. *Ecography* **29**, 525–530.

Gradstein, S.R. 2008 Epiphytes of tropical montane forests – impact of deforestation and climate change. In *The Tropical Mountain Forest. Patterns and Processes in a Biodiversity Hotspot.* Gradstein, S.R., Homeier, J. and Gansert, D. (ed.). University Press, pp. 51–65.

Gradstein, S.R. and Costa, D.P. 2003 The Hepaticae and Anthocerotae of Brazil. *Mem. N. Y. Bot. Gard.* **87**, 1–317.

Gradstein, S.R. and Culmsee, H. 2010 Bryophyte diversity on tree trunks in montane forests of Central Sulawesi. *Trop. Bryol.* **31**, 95–105.

Gradstein, S.R. and Sporn, S.G. 2010 Land-use change and epiphytic bryophyte diversity in the Tropics. *Nova Hedwigia* **138**, 309–321.

Gradstein, S.R., Churchill, S.P. and Salzar, N. 2001 Guide to the bryophytes of tropical America. *Mem. N. Y. Bot. Gard.* **86**, 1–577.

Green, T.G.A., Nash III, T.H. and Lange, O.L. 2008 Physiological ecology of carbon dioxide exchange. In *Lichen Biology*. Nash, T.H. III (ed.). Cambridge University Press, pp. 152–181.

Hietz, P., Buchberger, G. and Winkler, M. 2006 Effect of forest disturbance on abundance and distribution of epiphytic bromeliads and orchids. *Ecotropica* **12**, 103–112.

Hofstede, R. and Aguirre, N. 1999 Biomasa y dinámica del carbono en relación con las actividades forestales en la Sierra del Ecuador. In *El páramo como espacio de mitigación de carbono atmosférico*. Medina, G. and Mena, P. (ed.). Quito, pp. 29–51.

Hohnwald, S. 1999 Beiträge zum Mikroklima interandiner Trockentäler Boliviens. *Geoökodynamik* **20**, 221–229.

Holz, I. 2003 Diversity and Ecology of Bryophytes and Macrolichens in Primary and Secondary Montane Quercus Forests, Cordillera de Talamanca, Costa Rica. PhD Thesis, University of Göttingen.

Holz, I. and Gradstein, R.S. 2005 Cryptogamic epiphytes in primary and recovering upper montane oak forests of Costa Rica – species richness, community composition and ecology. *Pl. Ecol.* **178**, 89–109.

Johansson, V., Bergman, K.O., Lättman, H. and Milberg, P. 2009 Tree and site quality preferences of six epiphytic lichens growing on oaks in Southeastern Sweden. *Ann. Bot. Fenn.* **46**, 496–506.

Király, I., Nascimbene, J., Tinya, F. and Ódor, P. 2013 Factors influencing epiphytic bryophyte and lichen species richness at different spatial scales in managed temperate forests. *Biodiver. Conserv.* **22**, 209–223.

Köster, N., Friedrich, K., Nieder, N. and Barthlott, W. 2009 Conservation of epiphyte diversity in an Andean landscape transformed by human land use. *Conserv. Biol.* **23**, 911–919.

Kranner, I., Beckett, R., Hochman, A. and Nash III, T.H. 2008 Desiccationtolerance in lichens: a review. *Bryologist* **111**, 576–593.

Krömer, T. and Gradstein, S.R. 2003 Species richness of vascular epiphytes in two primary forests and fallows in the Bolivian Andes. *Selbyana* **25**, 190–195.

Lücking, R. 2009 The taxonomy of the genus *Graphis* Staiger (Ascomycota: Ostropales: Graphidaceae). *Lichenologist* **41**, 319–362.

Magurran, A. 2004 Measuring Biological Diversity. Backwell Publishing.

Mandl, N., Lehnert, M., Kessler, M. and Gradstein, S.R. 2010 A comparison of alpha and beta diversity patterns of ferns, bryophytes and macrolichens in tropical montane forests of southern Ecuador. *Biodivers. Conserv.* **19**, 2359–2369.

McCullagh, P. and Nelder, J.A. 1989 *Generalized linear models*. 2nd edn. Monographs on statistics and applied probability 37. Chapman and Hall/ CRC.

McCune, B., Grace, J.B. and Urban, D.L. 2002 Analyses of Ecological Communities. MjM software design Gleneden Beach.

Moncada, B., Coca, F. and Lücking, R. 2013 Neotropical members of *Sticta* (lichenized Ascomycota: Lobariaceae) forming photosymbiodemes, with the description of seven new species. *Bryologist* **116**, 169–200.

Myers, N., Mittermeler, R.A., Mittermeler, C.G., Fonseca, G.A.B. and Kent, J. 2000 Biodiversity hotspots for conservation priorities. *Nature* **403**, 853–858.

Normann, F., Weigelt, P., Gehrig-Downie, C., Gradstein, S.R., Sipman, H.J.M., Obregon, A. *et al.* 2010 Diversity and vertical distribution of epiphytic macrolichens in lowland rain forest and lowland cloud forest of French Guiana. *Ecol. Indic.* **10**, 1111–1118.

Nöske, N.M., Hilt, N., Werner, F.A., Brehm, G., Fiedler, K., Sipman, H.J.M. *et al.* 2008 Disturbance effects on diversity of epiphytes and moths in a montane forest in Ecuador. *Basic Appl. Ecol.* **9**, 4–12.

Oksanen, J., Blanchet, F.G., Kindt, R., Legendre, P., Minchin, P.R., O'Hara, R.B. et al. 2013 Vegan: Community Ecology Package. http://watson.nci.nih.gov/cran_mirror/web/packages/vegan/vegan.pdf (accessed on 19 June 2015).

Pardow, A. and Lakatos, M. 2013 Desiccation tolerance and global change: implications for tropical bryophytes in lowland forests. *Biotropica* **45**, 27–36.

Perhans, K., Appelgren, L., Jonsson, F., Nordin, U., Söderström, B. and Gustafsson, L. 2009 Retention patches as potential refugia for bryophytes and lichens in managed forest landscapes. *Biol. Conserv.* **145**, 1125–1133.

Pócs, T. 1982 Tropical forest bryophytes. In *Bryophyte Ecology*. Smith, A.J.E. (ed.). Champman and Hall, pp. 59–105.

Proctor, M.C.F., Oliver, M.J., Wood, A.J., Alpert, P., Stark, L.R., Cleavitt, N.L. *et al.* 2007 Desiccation-tolerance in bryophytes: a review. *Bryologist* **110**, 595–621.

Ranius, T., Johansson, P., Berg, N. and Niklasson, M. 2008 The influence of tree age and microhabitat quality on the occurrence of crustose lichens associated with old oaks. *J. Veg. Sci.* **19**, 653–662.

Rosabal, D., Burgaz, A.R. and de la Masa, R. 2010 Diversity and distribution of epiphytic macrolichens on tree trunks in two slopes of the montane rainforest of Gran Piedra, Santiago de Cuba. *Bryologist* **113**, 313–321.

Rosabal, D., Burgaz, A.R. and Reyes, O.J. 2013 Substrate preferences and phorophyte specificity of corticolous lichens on five tree species of the montane rainforest of Gran Piedra, Santiago de Cuba. *Bryologist* **116**, 113–121.

Sillett, S.C. and Antoine, M. 2004 Lichens and bryophytes in forest canopies. In *Forest Canopies*. Lowman, M.D. and Rinker, H.B. (ed.). Elsevier Academic Press, pp. 151–174.

Sipman, H.J.M. 1995 Preliminary review of the lichen biodiversity of the Colombian Montane Forests. In *Biodiversity and Conservation on Neotropical Montane Forests*. Churchill, A.P., Balslev, H., Forero, E. and Luteyn, J.L. (ed.). New York Botanical Garden, pp. 313–320.

Sipman, H.J.M. and Harris, R.C. 1989 Lichens. In *Tropical Rain Forest Ecosystems*. Lieth, H. and Werger, M.J.A. (ed.). Elsevier, pp. 303–309.

Soto, E., Lücking, R. and Bolaños, A. 2012 Especificidad de forófito y preferencias microambientales de los líquenes cortícolas en cinco forófitos del bosque premontano de finca Zíngara, Cali, Colombia. *Rev. Bio. Trop.* **60**, 843-856.

Sporn, S.G., Bos, M.M., Hoffstätter-Müncheberg, M., Kessler, M. and Gradstein, S.R. 2009 Microclimate determines community composition but not richness of epiphytic understorey bryophytes of rainforest and cacao agroforests in Indonesia. *Funct. Plant Biol.* **36**, 171–179.

Timdal, E. 2008 Studies on the *Phyllopsora* (Ramalinaceae) in Peru. *Lichenologist* **40**, 337–362.

Verbeke, G. and Molenberghs, G. 1997 *Linear mixed models in practice. A SAS-oriented approach.* Springer.

Werner, F.A. and Gradstein, S.R. 2009 Diversity of dry forest epiphytes along a gradient of human disturbance in the tropical Andes. *J. Veg. Sci.* **20**, 59–68.

Werner, F.A., Homeier, J. and Gradstein, S.R. 2005 Diversity of vascular epiphytes on isolated remnant trees in the montane forest belt of southern Ecuador. *Ecotropica* **11**, 21–40.

Werner, F.A., Homeier, J., Oesker, M. and Boy, J. 2011 Epiphytic biomass of a tropical montane forest varies with topography. *J. Trop. Ecol.* **28**, 23–31.

Wolf, J.H.D. 2005 The response of epiphytes to anthropogenic disturbance of pine-oak forests in the highlands of Chiapas, Mexico. *For. Ecol. Manage.* **212**, 376–393.

Zimmerman, B. and Kormos, C. 2012 Prospects for sustainable logging in tropical forests. *BioScience* **62**, 479–487.