

Review

Germination: an alternative source to promote phytonutrients in edible seeds

Anthony Temitope Idowu,* Oladipupo Odunayo Olatunde,* Ademola Ezekiel Adekoya** and Solomon Idowu***

*Department of Food Technology, Faculty of Agro-Industry, Prince of Songkhla University, Hatyai, Songkhla Province, 90110, Thailand and **Department of Thai Traditional Medicine, Prince of Songkla University, Hat Yai, Songkhla Province, Thailand and ***Department of Community Health and Primary Healthcare, College of Medicine of the University of Lagos, Lagos state, Nigeria

Correspondence to: Anthony Temitope Idowu, Department of Food Technology, Faculty of Agro-Industry, Prince of Songkhla University, Hatyai, Songkhla Province, 90110, Thailand. E-mail: tonitop17@yahoo.com

Received 17 October 2019; Revised 29 November 2019; Editorial decision 9 December 2019.

Abstract

Consumption of less phytonutrient foods has shown to cause different chronic diseases, despite over 50,000 edible plant breed available in various countries around the globe. These edible plants consist of seeds that can be consumed which possessed high health benefits. Moreover, nutritive values such as phytochemicals of edible seeds increased after germination. Therefore, germination has been reported to enhance various bioactive compounds such as anti-diabetic, anti-bacteria, and anti-cancer effects when these seeds are consumed. Consequently, germination can be regarded as a cheap and effective way to enhance the nutritional value of edible seeds.

Key words: edible seeds; water; bioactive compounds; germination; phytonutrients.

Introduction

Germination is a process that involves imbibition of water by a quiescent dry seed which leads to the development and growth of the seed (Nonogaki et al., 2010). In addition, germination is highlighted as a principal bio-processing method in the field of food science and nutrition because it brings about increase in bioactive compounds. According to Bewley et al. (2012), germination incorporates those events which involves the uptake of water by the quiscent dry seeds and terminate with the elongation of the embryonic axis. Watanabe et al. (2004) stated germination as a biological process initiated once dry seeds absorb water which leads to activation of enzymes with a set physical condition desirable for the sprouting of the seeds. Dogra et al. (2013) reported that germination activates seeds from its dormancy which restores the seeds metabolitic activities and leads to biochemical, nutritional, and sensorial changes of the seeds. According to Pimentel et al. (2000), over 50,000 edible plants species in the world have been reported to be available for consumption. It was also reported that edible seeds contain various phytochemicals that provide anti-oxidant, anti-diabetic, and anti-cancer effects.

Many studies have shown that germination can further increase nutritive and bioactive compounds in edible seeds (Saleh et al., 2013; Huang et al., 2014; Fouad and Rehab, 2015). During germination, degradation of macro-nutrients occurs, for example, carbohydrate, protein, and fatty acid are further broken down into glucose, fructose, free amino acid, and organic acids (Shi et al., 2010). Therefore, it is necessary to discuss the activities involved in effecting germination and the resulting changes in bioactive compounds obtained as shown in Figure 1.

Steps to Germination

To facilitate a germination process, there are various steps to carry out as highlighted below:

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (http://creativecommons.org/licenses/ by-nc/4.0/), which permits non-commercial re-use, distribution, and reproduction in any medium, provided the original work is properly cited. For commercial re-use, please contact journals.permissions@oup.com

[©] The Author(s) 2019. Published by Oxford University Press on behalf of Zhejiang University Press.

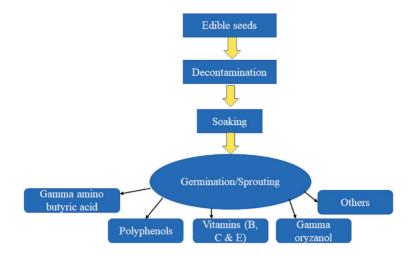


Figure 1. Processing of edible seeds to increase phytonutrient contents.

Decontamination

Although intact seeds are usually germ-free, various steps during harvesting such as hand picking, transportation, and drying make seedcoat to be contaminated with dust and micro-organism. Generally, the seeds are dried to a moisture content of around 8% before storage; thus, the microorganism stained on or even in the seeds could not function. However, when the seeds are soaked in water, the environment becomes easier for the microorganisms to grow and invade the intact seeds leading to spoilage if no decontamination process is applied. Therefore, chemicals including sodium hypochlorite (NaClO) are used to kill the microbes. According to Pajak et al. (2014) and Wu et al. (2012) who reported that NaClO concentration for decontaminating seeds legumes and cereals seeds may vary, but it is usually in small quantity of about 0.5–5% (Selcuk et al., 2008; Bhat et al., 2010). However, excessive decontaminant can be toxic to the seeds and affect the human health when toxic seeds are consumed.

Soaking

The essense of soaking seeds in water is to rehydrate the seeds before germination occur. Parameters such as seed weight/water volume ratio, time, and soaking temperature are very important and should be considered before soaking the seed. In addition, frequent change of water should be implemented, for instance, twice a day, for removal of metabolites in germinated seeds and retard the growth of micro-organisms. Excessive soaking time of rice leads to microbial enrichment and fermentation (Ray et al., 2016). In contrast, insufficient soaking does not support the augmentation of phytochemical content (Chaiyasut et al., 2017).

Sprouting

There are several factors to consider before a seed can sprout. These factors include light, temperature, humidity, water, oxygen, and temperature. Chaiyasut et al. (2017) reported that germination is supported by sufficient oxygen to allow the seeds respire, suitable temperature to allow various metabolic processes during germination. Moongngarm and Saetung (2010) reported that rice seed can germinate anaerobically through a rapid elongation of coleoptile but the radicle could not protude well under this condition. However, once it is switched to aerobic condition, the radicle can continue to elongate, which suggested that oxygen availability is a determinant factor for true germination (Menegus et al., 1991). The soaking time and temperature depend on the seeds and cultivars; however, several

studies have reported that 25–30°C is suitable temperature for germination (Bandara et al., 1991; Capanzana and Buckle, 1997). Germination in the presence of darkness results in stress condition which makes glutamase decarboxylase convert glutamic acid to γ -amino butyric acid (GABA) (Bai et al., 2009). In addition, during sprouting seeds are needed to be sprinkled with water on a daily basis to keep the relative humidity high in order to support their growth.

Influence of Germination on Bioactive Compounds in Edible Seeds

Studies have shown that germination brings about accumulation of different bioactive compounds (Saleh et al., 2013; Gan et al., 2017). These bioactive compounds include GABA, gamma oryzanol, ferulic acid, polyphenols, vitamins, etc.

γ-Amino butyric acid

GABA is regarded as a non-protein amino acid found in plants and animals. GABA content is synthesized through diverse pathways in plant called 'gaba shunt'. It was discovered in plants for a half century ago. Primarily, it is synthesized from l-glutamic acid through glutamate decarboxylase (GAD), a pyridoxal s-phosphate controlled enzyme responsible for the transformation of l-glutamic acid to GABA (Bown and Shelp, 1997). More so, GABA's role is to act as an important depressive neuro-transmitter in the nervous system of mammals and it can also increase insulin secretion from pancreas and regulate blood pressure and heart rate to relieve pain and anxiety, respectively (Adeghate and Ponery, 2002).

Germination has proven to increase GABA content in edible seeds such as adzuki beans, kidney beans, lentil, lupin, sesame seeds, soybeans, pea, brown rice, buckwheat, waxy wheat, and oat (Gani et al., 2012; Gan et al., 2017). GABA content was significantly increased in 18 rice varieties immersed in distilled water at 30°C for 72 h, although an increase of this compound in GABA widely differed among the 18 rice varieties (Roohinejad et al., 2009, 2010). Bown and Shelp (1997) reported that GABA synthesis could be elevated by environmental stress including mechanical and environmental stimulations such as hypoxia, darkness, heat, shock, cold shock, and cystolic acidification. Chung et al. (2009) and Dewar et al. (1997) who reported that hypoxia occurs due to the limited availability of oxygen in water during seed germination result in stress and then GABA content may increase rapidly in the seed in response to hypoxia. In addition, Lin et al. (2015) studied the effect of different germination condition on rice and reported that GABA content increases by 15-fold in a close vessel (limited oxygen) from 0.79 mg/100 g of *Taiwan japonica* 9 brown rice to 12.37 mg/100 g.

Buckwheat sprouts and germinated soybean have been reported to improve GABA from 0.50 to 2.60 and from 0.60 to 37.5 (Martínez-Villaluenga et al., 2006; Lin et al., 2008). In addition, Saikusa et al. (1994) studied the effect of soaking on eight varieties of japonica rice and two hybrid varieties of indica and reported that GABA content was nearly 8 times increased.

Vitamins

Vitamins are organic compounds that are found in both plant and animal sources. They perform vital roles in human health. Conventionally, they are divided into fat-soluble and water-soluble vitamins. The water-soluble vitamins include vitamin B and C, whereas the fat-soluble vitamins include A, D, E, and K. Recent studies have proven that significant increase in the content of some vitamins can be observed as a result of germination.

Vitamin B members comprise of vitamin B_1 (thiamine), vitamin B_2 (riboflavin), vitamin B_3 (niacin), vitamin B_6 (pyridoxine), vitamin B_9 (folate), and vitamin B_{12} (cobalamin). They all have possess vital roles in human health (Pereira and Vicente, 2013). Increase in some vitamins B observed in different edible seeds has been traced to germination.

Shohag et al. (2012) reported significant increase in folate content of soybean and mung bean sprouts when compared with raw seeds by 65%-274% and 78%-326%, respectively, after germination. Also vitamins B₁ and B₆ increased to about 11.8 mg/100 g dry weight in buckwheat sprouts, whereas it was not detected in raw seeds (Kim et al., 2004).

Ascorbic acid (Vitamin C) is derived from fruits and vegetables. Recent studies have proven that germination can increase the content of vitamin C in edible seeds such as buckwheat, lupin, mung bean, soybean, chicken pea, and cowpea when germinated. Previous study reported by Gan et al. (2016) showed that vitamin C content of the green and black mung bean sprouts increased from 13.5 to 24.0 and 10.3 to 21.3 folds when compared with their respective raw seeds after germination for 1-5 days. Accumulation of vitamin C in germinated edible seeds can be newly formed because most seeds have low or non-availability of vitamin C before germination. Furthermore, l-galactono-gamma-lactone dehydrogenase (GLDH) is a major enzyme in ascorbic acid biosynthesis and it helps catalyse the oxidation of l-galactono-1,4-lactone to ascorbic acid. The activity of this enzymes has been observed to increase during soybean germination in parallel with the increase in ascorbic acid content (Wheeler et al., 1998; Xu et al., 2005).

Tocopherols (vitamin E) are fat-soluble vitamins which have four isomers, namely, α -tocopherol, β -tocopherol, γ -tocopherol, and δ -tocopherol. Germination has proven to emphatically alter the content of vitamin E isomers in edible seeds. However, the mechanism is still a mystery.

Gamma-tocopherol, a crucial vitamin E in several edible seeds, has been reported to significantly increase in germinated soybean by 1.55%–164% according to the result of Fernandez-Orozco et al. (2008) when compared with raw seeds, but the vitamin E content in germinated lupin and mung bean reduced (Frias et al., 2005). In general, germination of edible seeds is a useful way to increase vitamins, especially vitamin C which became beneficial in human diets.

Polyphenols

Polyphenols are regarded as a group of small molecules with at least one phenol unit in their structure (Frias et al., 2005). Polyphenols exist in free or bound forms in the plant kingdom. Soluble phenolics are mostly synthesized in the intracellular endoplasmic reticulum of the plants and are stored in the vacuoles while the insoluble phenolics are found in cell wall matrix of plant cells (Agati et al., 2012). The soluble form can be extracted with water, acid, alkali, and enzymatic hydrolysis to quantify their contents in edible plant seeds. Meanwhile, germination has shown through recent studies to alter the level of total phenolics in germinated edible seeds. Gan et al. (2016) reported an increase of 5.0-5.5 folds in phenolic compound of mung bean after 5 days of germination. Increase in soluble phenolics during germination can be attributed to synthesis and conversion of different phenolic compounds (Randhir et al., 2004; Wu et al., 2012; Kim et al., 2013). Glucose is the main pre-cursor for the synthesis of phenolics compounds and several paramount molecular signalling pathways, some of which include glycolysis, propanoid pathway, hydrolysable tannin pathway, oxidative pentose phosphate pathway, and acetate/malonate pathway. They partake in the synthesis and conversion of different phenolic compounds. Bound phenolics have been less explored and reported in germinated seeds. Despite the fact that some germinated edible seeds especially cereal grains contained a lot of bound phenolics (Ti et al., 2014). In addition, some studies reported that bound phenolics first decreased and then increased after germination (Tang et al., 2014). Although in germinated brown rice, it was suggested that bound phenolics content was dependent on conjugation rate and their release.

Germination at an early stage often leads to degradation of carbohydrates and protein followed by an increase in free amino acids and simple sugars and the bound phenolics conjugated with the cell wall components (Wang et al., 2005). Increase in germination time often leads to the proliferation of new plant cells as well as formation of new cell wall. Thereafter, the synthesized soluble phenolics can be secreted to the cell wall which form new bound phenolic. The known phenolic compounds found in germinated edible seeds are phenolic acids such as ferulic acid and courmaric acid.

Generally, phenolic acids are found in fruits and vegetables. Many researchers reported that some germinated edible seeds contained total phenolic content ranged 30–253 mg gallic acid equivalent/100 g fresh weight, whereas common fruits contained about 11.9–386 mg gallic acid equivalent/100 g fresh weight (Lin and Tang, 2007).

Other bioactive compounds

There are other bioactive compounds reported in germinated edible seeds, apart from those listed above. Gamma oryzanol is also a bioactive compound that contains a set of 10 or more compounds with ester bonds between ferulic acid and triterpenes.

Cycloartenyl ferulate, 24- methylene cycloartenyl ferulate, and campestanyl ferulate are the major compounds of gamma oryzanol in a germinated brown rice (Jayadeep and Malleshi, 2011). Gamma oryzanol is associated with decreasing plasma and serum cholesterol (Gerhardt and Gallo, 1998). In addition, oryzanol has also been used to treat hyperlipidaemia, a disorder of menopause in women (Gani et al., 2012).

Numerous phytosterol plants have been found, among which campesterol, beta sitosterol, and stigmasterol are known to be most prevalent (Brufau et al., 2008). Germination of red rice, brown rice, glutinous brown rice, and black rice increased the stigmasterol levels in all the rice types tested, whereas sitosterol and campesterol showed no significant changes (Jung et al., 2013). Phytosterols have been used as nutritional supplements and bio-functional ingredients in foods because it can reduce the cholesterol level, prevent stroke, and display anti-atheromatosis effect (Brufau et al., 2008). Another one is melatonin (N-acetyl-5-methoxytryptamine), which is an indolamine found in plants, animals, bacteria, and fungi. It performs vital physiological functions in various organisms which includes regulation of the circadian rhythm and growth (Gamble et al., 2014). New studies have shown that germination can promote the amount of melatonin in edible seeds, e.g., germinated lentis and kidney beans.

Melatonin reached the highest content of about 2.50 ng/g dry weight and 9.50 ng/g dry weight for lentils and kidney beans, respectively, after germination for 6 days under the dark condition (Aguilera et al., 2015).

D-chiro-inositol (Dcl), a co-enzyme of glycosylphosphatidyl inositol protein, is involved in the insulin signalling pathway and glucose movement; hence, it is regarded as a pivotal insulin mediator with anti-diabetic effect (Adams et al., 2014). During germination, the content of Dcl can be promoted in germinated edible seeds. For instance, Dcl content increased gradually in mung bean by 74% (4.79 mg/g dry weight) when germinated for 80 h and decreased afterwards (Yao et al., 2011).

Conclusion

With the growing phytonutrient deficiencies in human health in many countries across the globe despite the numerous edible seeds available, it is expedient that the process of germination can be employed to improve phytonutrient in these seeds. GABA, vitamins, polyphenols, and other phytochemicals have been reported to increase during germination and has further enhanced the nutritive value of edible seeds around the globe. Thus, germination is undoubtedly an effective way to improve phytonutrients in edible seeds.

Acknowledgements

Acknowledgement goes the Higher Education Research Promotion and the Thailand's Education Hub for Southern Region of Asian Countries Project office of the Higher Education Commission for the scholarship granted to pursue graduate studies.

Author's Contributions

Anthony Temitope Idowu is a research assistant at Prince of Songkla University, Thailand. He conceptualized the research idea and wrote the article. Oladipupo Odunayo Olatunde, Ademola Ezekiel Adekoya, and Solomon Idowu all provided the necessary guide and correction of the article.

Conflict of Interest Statement

None declared.

References

Adams, G. G., et al. (2014). The hypoglycemic effect of pumpkin seeds, Trigonelline (TRG), Nicotinic acid (NA), and D-Chiro-inositol (DCI) in controlling glycemic levels in diabetes mellitus. Critical Reviews in Food Science and Nutrition, 54: 1322–1329.

- Adeghate, E., Ponery, A. S. (2002). GABA in the endocrine pancreas: cellular localization and function in normal and diabetic rats. *Tissue & Cell*, 34: 1–6.
- Agati, G., Azzarello, E., Pollastri, S., Tattini, M. (2012). Flavonoids as antioxidants in plants: location and functional significance. *Plant Science*, 196: 67–76.
- Aguilera, Y., et al. (2015). Estimation of scavenging capacity of melatonin and other antioxidants: contribution and evaluation in germinated seeds. Food Chemistry, 170: 203–211.
- Bai, Q., Chai, M., Gu, Z., Cao, X., Li, Y., Liu, K. (2009). Effects of components in culture medium on glutamate decarboxylase activity and γ-aminobutyric acid accumulation in foxtail millet (*Setaria italica L.*) during germination. *Food Chemistry*, 116: 152–157.
- Bandara, J. M., Vithanege, A. K., Bean, G. A. (1991). Effect of parboiling and bran removal on aflatoxin levels in Sri Lankan rice. *Mycopathologia*, 115: 31–35.
- Bewley, J. D., Bradford, K., Hilhorst, H. (2012). Seeds: Physiology of Development, Germination and Dormancy. Berlin, Germany: Springer Science and Business Media.
- Bhat, R., Sridhar, K., Karim, A. (2010). Microbial quality evaluation and effective decontamination of nutraceutically valued lotus seeds by electron beams and gamma irradiation. *Radiation Physics and Chemistry*, 79: 976–981.
- Bown, A. W., Shelp, B. J. (1997). The metabolism and functions of [gamma]aminobutyric acid. *Plant Physiology*, 115: 1–5.
- Brufau, G., Canela, M. A., Rafecas, M. (2008). Phytosterols: physiologic and metabolic aspects related to cholesterol-lowering properties. *Nutrition Research*, 28: 217–225.
- Capanzana, M., Buckle, K. (1997). Optimisation of germination conditions by response surface methodology of a high amylose rice (Oryza sativa) cultivar. LWT-Food Science and Technology, 30: 155–163.
- Chaiyasut, C., et al. (2017). Optimization of conditions to achieve high content of gamma amino butyric acid in germinated black rice, and changes in bioactivities. Food Science and Technology, 37: 83–93.
- Chung, H.-J., Jang, S.-H., Cho, H. Y., Lim, S.-T. (2009). Effects of steeping and anaerobic treatment on GABA (γ-aminobutyric acid) content in germinated waxy hull-less barley. *LWT-Food Science and Technology*. 42: 1712–1716.
- Dewar, J., Taylor, J., Berjak, P. (1997). Determination of improved steeping conditions for sorghum malting. *Journal of Cereal Science*, 26: 129–136
- Dogra, V., Ahuja, P. S., Sreenivasulu, Y. (2013). Change in protein content during seed germination of a high altitude plant Podophyllum hexandrum Royle. *Journal of Proteomics*, 78: 26–38.
- Fernandez-Orozco, R., Frias, J., Zielinski, H., Piskula, M. K., Kozlowska, H., Vidal-Valverde, C. (2008). Kinetic study of the antioxidant compounds and antioxidant capacity during germination of Vigna radiata cv. emmerald, Glycine max cv. jutro and Glycine max cv. merit. Food Chemistry, 111: 622–630.
- Fouad, A. A., Rehab, F. M. (2015). Effect of germination time on proximate analysis, bioactive compounds and antioxidant activity of lentil (*Lens* culinaris Medik.) sprouts. Acta Scientiarum Polonorum. Technologia Alimentaria, 14: 233–246.
- Frias, J., Miranda, M. L., Doblado, R., Vidal-Valverde, C. (2005). Effect of germination and fermentation on the antioxidant vitamin content and antioxidant capacity of *Lupinus albus L. var. Multolupa. Food Chemistry*, 92: 211–220.
- Gamble, K. L., Berry, R., Frank, S. J., Young, M. E. (2014). Circadian clock control of endocrine factors. *Nature Reviews. Endocrinology*, 10: 466– 475.
- Gan, R.-Y., et al. (2017). Bioactive compounds and bioactivities of germinated edible seeds and sprouts: an updated review. Trends in Food Science and Technology, 59: 1–14.
- Gan, R. Y., Wang, M. F., Lui, W. Y., Wu, K., Corke, H. (2016). Dynamic changes in phytochemical composition and antioxidant capacity in green and black mung bean (*Vigna radiata*) sprouts. *International Journal of Food Science and Technology*, 51: 2090–2098.

- Gani, A., Wani, S., Masoodi, F., Hameed, G. (2012). Whole-grain cereal bioactive compounds and their health benefits: a review. *Journal of Food Pro*cess Technology, 3: 146–156.
- Gerhardt, A. L., Gallo, N. B. (1998). Full-fat rice bran and oat bran similarly reduce hypercholesterolemia in humans. *The Journal of Nutrition*, 128: 865–869.
- Huang, X., Cai, W., Xu, B. (2014). Kinetic changes of nutrients and antioxidant capacities of germinated soybean (*Glycine max L.*) and mung bean (*Vigna radiata L.*) with germination time. Food Chemistry, 143: 268–276.
- Jayadeep, A., Malleshi, N. (2011). Nutrients, composition of tocotrienols, tocopherols, and γ-oryzanol, and antioxidant activity in brown rice before and after biotransformation Nutrientes, composición de tocotrienoles, tocoferoles y γ-oryzanol, y actividad antioxidante del arroz integral antes y después de la biotransformación. CyTA-Journal of Food, 9: 82–87.
- Jung, Y. H., Kim, I. J., Kim, H. K., Kim, K. H. (2013). Dilute acid pretreatment of lignocellulose for whole slurry ethanol fermentation. *Bioresource Tech*nology, 132: 109–114.
- Kim, S.-L., Kim, S.-K., Park, C.-H. (2004). Introduction and nutritional evaluation of buckwheat sprouts as a new vegetable. *Food Research International*, 37: 319–327.
- Kim, Y. B., Thwe, A. A., Kim, Y., Yeo, S. K., Lee, C., Park, S. U. (2013). Characterization of cDNA encoding resveratrol synthase and accumulation of resveratrol in tartary buckwheat. *Natural Product Communications*, 8: 1571–1574.
- Lin, Y. T., Pao, C. C., Wu, S. T., Chang, C. Y. (2015). Effect of different germination conditions on antioxidative properties and bioactive compounds of germinated brown rice. *Biomed Research International*, 2015: 608761.
- Lin, L. Y., Peng, C. C., Yang, Y. L., Peng, R. Y. (2008). Optimization of bioactive compounds in buckwheat sprouts and their effect on blood cholesterol in hamsters. *Journal of Agricultural and Food Chemistry*, 56: 1216–1223.
- Lin, J.-Y., Tang, C.-Y. (2007). Determination of total phenolic and flavonoid contents in selected fruits and vegetables, as well as their stimulatory effects on mouse splenocyte proliferation. *Food Chemistry*, 101: 140–147.
- Martínez-Villaluenga, C., Kuo, Y.-H., Lambein, J., Frías, F., Vidal-Valverde, C. (2006). Kinetics of free protein amino acids, free non-protein amino acids and trigonelline in soybean (*Glycine max L.*) and lupin (*Lupinus angustifolius L.*) sprouts. *European Food Research and Technology*, 224: 177–186.
- Menegus, F., Cattaruzza, L., Mattana, M., Beffagna, N., Ragg, E. (1991). Response to anoxia in rice and wheat seedlings: changes in the pH of intracellular compartments, glucose-6-phosphate level, and metabolic rate. *Plant Physiology*, 95: 760–767.
- Moongngarm, A., Saetung, N. (2010). Comparison of chemical compositions and bioactive compounds of germinated rough rice and brown rice. *Food Chemistry*, 122: 782–788.
- Nonogaki, H., Bassel, G. W., Bewley, J. D. (2010). Germination—still a mystery. *Plant Science*, 179: 574–581.
- Pająk, P., Socha, R., Gałkowska, D., Rożnowski, J., Fortuna, T. (2014). Phenolic profile and antioxidant activity in selected seeds and sprouts. *Food Chemistry*, 143: 300–306.
- Pereira, P. M., Vicente, A. F. (2013). Meat nutritional composition and nutritive role in the human diet. *Meat Science*, 93: 586–592.
- Pimentel, D., Lach, L., Zuniga, R., Morrison, D. (2000). Environmental and economic costs of nonindigenous species in the United States. *BioScience*, 50: 53–66.

- Ray, M., Ghosh, K., Singh, S., Mondal, K. C. (2016). Folk to functional: an explorative overview of rice-based fermented foods and beverages in India. *Journal of Ethnic Foods*, 3: 5–18.
- Roohinejad, S., et al. (2009). Effect of hypocholesterolemic properties of brown rice varieties containing different gamma aminobutyric acid (GABA) levels on Sprague-Dawley male rats. Journal of Food, Agriculture and Environment, 7: 197–203.
- Roohinejad, S., et al. (2010). Effect of pre-germination time of brown rice on serum cholesterol levels of hypercholesterolaemic rats. Journal of the Science of Food and Agriculture, 90: 245–251.
- Saikusa, T., Horino, T., Mori, Y. (1994). Accumulation of γ-aminobutyric acid (GABA) in the rice germ during water soaking. *Bioscience, Biotechnology,* and Biochemistry, 58: 2291–2292.
- Saleh, A. S., Zhang, Q., Chen, J., Shen, Q. (2013). Millet grains: nutritional quality, processing, and potential health benefits. *Comprehensive Reviews* in Food Science and Food Safety, 12: 281–295.
- Selcuk, M., Oksuz, L., Basaran, P. (2008). Decontamination of grains and legumes infected with Aspergillus spp. and Penicillum spp. by cold plasma treatment. Bioresource Technology, 99: 5104–5109.
- Shi, H., Nam, P. K., Ma, Y. (2010). Comprehensive profiling of isoflavones, phytosterols, tocopherols, minerals, crude protein, lipid, and sugar during soybean (*Glycine max*) germination. *Journal of Agricultural and Food Chemistry*, 58: 4970–4976.
- Shohag, M. J., Wei, Y., Yang, X. (2012). Changes of folate and other potential health-promoting phytochemicals in legume seeds as affected by germination. *Journal of Agricultural and Food Chemistry*, 60: 9137–9143.
- Tang, D., Dong, Y., Guo, N., Li, L., Ren, H. (2014). Metabolomic analysis of the polyphenols in germinating mung beans (*Vigna radiata*) seeds and sprouts. *Journal of the Science of Food and Agriculture*, 94: 1639–1647.
- Ti, H., et al. (2014). Dynamic changes in the free and bound phenolic compounds and antioxidant activity of brown rice at different germination stages. Food Chemistry, 161: 337–344.
- Wang, K. H., Lai, Y. H., Chang, J. C., Ko, T. F., Shyu, S. L., Chiou, R. Y. (2005). Germination of peanut kernels to enhance resveratrol biosynthesis and prepare sprouts as a functional vegetable. *Journal of Agricultural and Food Chemistry*, 53: 242–246.
- Watanabe, M., Maeda, T., Tsukahara, K., Kayahara, H., Morita, N. (2004). Application of pregerminated brown rice for breadmaking. *Cereal Chemistry*, 81: 450–455.
- Wheeler, G. L., Jones, M. A., Smirnoff, N. (1998). The biosynthetic pathway of vitamin C in higher plants. *Nature*, 393: 365–369.
- Wu, Z., et al. (2012). Germination dramatically increases isoflavonoid content and diversity in chickpea (*Cicer arietinum L.*) seeds. Journal of Agricultural and Food Chemistry, 60: 8606–8615.
- Xu, M. J., Dong, J. F., Zhu, M. Y. (2005). Effects of germination conditions on ascorbic acid level and yield of soybean sprouts. *Journal of the Science of Food and Agriculture*, 85: 943–947.
- Yao, Y., Cheng, X.-Z., Ren, G.-X. (2011). Contents of D-chiro-Inositol, vitexin, and isovitexin in various varieties of mung bean and its producsssssts. *Agricultural Sciences in China*, 10: 1710–1715.