

Children with Classic Congenital Adrenal Hyperplasia Have Elevated Serum Leptin Concentrations and Insulin Resistance: Potential Clinical Implications

EVANGELIA CHARMANDARI, MARTINA WEISE, STEFAN R. BORNSTEIN, GRAEME EISENHOFER, MARGARET F. KEIL, GEORGE P. CHROUSOS, AND DEBORAH P. MERKE

Pediatric and Reproductive Endocrinology Branch (E.C., M.W., M.F.K., G.P.C., D.P.M.), National Institute of Child Health and Human Development, The Warren Grant Magnuson Clinical Center (D.P.M.), and Clinical Neurocardiology Section (G.E.), National Institute of Neurological Disorders and Stroke, National Institutes of Health, Bethesda, Maryland 20892; and Department of Endocrinology (S.R.B.), University of Düsseldorf, 40001 Düsseldorf, Germany

Leptin is secreted by the white adipose tissue and modulates energy homeostasis. Nutritional, neural, neuroendocrine, paracrine, and autocrine factors, including the sympathetic nervous system and the adrenal medulla, have been implicated in the regulation of leptin secretion. Classic congenital adrenal hyperplasia (CAH) is characterized by a defect in cortisol and aldosterone secretion, impaired development and function of the adrenal medulla, and adrenal hyperandrogenism. To examine leptin secretion in patients with classic CAH in relation to their adrenomedullary function and insulin and androgen secretion, we studied 18 children with classic CAH (12 boys and 6 girls; age range 2–12 yr) and 28 normal children (16 boys and 12 girls; age range 5–12 yr) matched for body mass index (BMI). Serum leptin concentrations were significantly higher in patients with CAH than in control subjects (8.1 ± 2.0 vs. 2.5 ± 0.6 ng/ml, $P = 0.01$), and this difference persisted when leptin values were corrected for BMI. When compared with their normal counterparts, children with CAH had significantly lower plasma epinephrine (7.1 ± 1.3 vs. 50.0 ± 4.2 , $P < 0.001$) and free metanephrine concentrations (18.4 ± 2.4 vs. 46.5 ± 4.0 , $P < 0.001$) and higher fasting serum insulin (10.6 ± 1.4 vs. 3.2 ± 0.2 μ U/ml, $P < 0.001$) and testosterone (23.7 ± 5.3 vs. 4.6 ± 0.5 ng/dl, $P = 0.003$) concentrations. Insulin resistance determined by the homeosta-

sis model assessment method was significantly greater in children with classic CAH than in normal children (2.2 ± 0.3 vs. 0.7 ± 0.04 , $P < 0.001$). Leptin concentrations were significantly and negatively correlated with epinephrine ($r = -0.50$, $P = 0.001$) and free metanephrine ($r = -0.48$, $P = 0.002$) concentrations. Stepwise multiple linear regression analysis indicated that serum leptin concentrations were best predicted by BMI in both patients and controls. Gender predicted serum leptin concentrations in controls but not in patients with classic CAH. No association was found between the dose of hydrocortisone and serum leptin ($r = -0.17$, $P = 0.5$) or insulin ($r = 0.24$, $P = 0.3$) concentrations in children with CAH. Our findings indicate that children with classic CAH have elevated fasting serum leptin and insulin concentrations, and insulin resistance. These most likely reflect differences in long-term adrenomedullary hypofunction and glucocorticoid therapy. Elevated leptin and insulin concentrations in patients with CAH may further enhance adrenal and ovarian androgen production, decrease the therapeutic efficacy of glucocorticoids, and contribute to later development of polycystic ovary syndrome and/or the metabolic syndrome and their complications. (*J Clin Endocrinol Metab* 87: 2114–2120, 2002)

CLASSIC CONGENITAL ADRENAL hyperplasia (CAH) due to 21-hydroxylase or 11-hydroxylase deficiency is an autosomal recessive condition in which deletions or mutations of the cytochrome P450 21-hydroxylase or 11-hydroxylase genes result in glucocorticoid and/or mineralocorticoid deficiency or excess, respectively. This leads to increased secretion of ACTH, adrenal hyperplasia, and increased production of androgens and steroid precursors before the enzymatic defect (1, 2). Current treatment is to provide adequate glucocorticoid and, when necessary, mineralocorticoid substitution to prevent adrenal crises and to suppress the abnormal secretion of androgens and steroid precursors from the adrenal cortex. The therapeutic spectrum of glucocorticoids, however, is narrow, and patients are often at risk for developing in tandem iatrogenic Cushing's

syndrome and hyperandrogenism. In addition to impaired adrenocortical function, classic CAH is characterized by compromised adrenomedullary function (3). The latter is owing to developmental defects in the formation of adrenal medulla, leading to a reduction in epinephrine and metanephrine stores and decreased production of catecholamines and their metabolites.

Leptin, the product of the *ob* gene, is a 16-kDa protein secreted by differentiated white adipocytes that signals the size of sc and visceral white adipose tissue depots (4, 5). Leptin acts on the central nervous system to suppress food intake and stimulate energy expenditure by inhibiting appetite and stimulating sympathetic nervous system activity, respectively (5–8). Homozygous defects in the *ob* gene or leptin receptor gene result in obesity, whereas exogenous administration of leptin reduces food intake and body weight (7, 8). Neural, neuroendocrine, paracrine, and autocrine factors have been implicated in the regulation of *ob* gene expression and leptin secretion. Glucocorticoids and insulin increase leptin secretion (9–15), whereas androgens have the

Abbreviations: BMI, Body mass index; CAH, congenital adrenal hyperplasia; CV, coefficient of variation; HOMA, homeostasis model assessment; IR, insulin resistance; PCOS, polycystic ovary syndrome; SW, salt wasting.

opposite effect (16, 17). Catecholamines suppress *ob* gene expression and leptin secretion *in vitro* and *in vivo* via a β -adrenergic receptor mechanism (18–22), while the presence of leptin receptors in the adrenal medulla may suggest a direct interaction between leptin and adrenomedullary function (23). Obesity is associated with high serum concentrations of leptin and impaired β -adrenergic receptor-mediated lipid metabolism and thermogenesis (24, 25), implicating the importance of neuroendocrine modulators on leptin secretion.

The aim of the present study was to examine the effect of chronic adrenomedullary hypofunction, glucocorticoid treatment, and early life hyperandrogenism on leptin secretion. To this end, we determined circulating leptin concentrations in children with classic CAH and healthy children matched for body mass index, and investigated the association between leptin and various hormonal and metabolic parameters in both groups.

Subjects and Methods

Subjects

Eighteen children with classic CAH (12 boys and 6 girls; age range 2–12 yr) and 28 normal children (16 boys and 12 girls; age range 5–12 yr) were studied. Normal children were recruited to participate in the study if they were prepubertal by physical examination, had similar body mass index (BMI) to that of children with CAH, and had no evidence of any associated endocrine or other disorder. No normal subject was receiving medications. All 46 children were prepubertal and their clinical characteristics are summarized in Table 1.

In patients with CAH, clinical and endocrinologic evaluation at presentation revealed that 16 children had classic 21-hydroxylase deficiency [12 salt-wasting (SW), 4 simple virilizing], and two had 11-hydroxylase deficiency. Criteria for classification into the SW phenotype included history of salt-losing crisis with documented hyponatremia, hyperkalemia, and markedly elevated plasma renin activity. The number of adrenal crises requiring hospitalization and the associated biochemical abnormalities were obtained from parental reports and medical records. Patients in whom early virilization was diagnosed at an older age were classified as having the simple virilizing form of the disease. Diagnosis was confirmed by genetic analysis in 89% of patients (14 patients with 21-hydroxylase deficiency and 2 patients with 11-hydroxylase deficiency).

All patients with classic 21-hydroxylase deficiency were on standard doses of hydrocortisone (mean \pm SD, 14.8 \pm 4.2 mg/m² per day) given three times daily and 9 α -fludrocortisone (114 \pm 48 μ g/d) given twice daily (26). The two patients with 11-hydroxylase deficiency were on hydrocortisone only (18.3 and 20.3 mg/m² per day). Six patients with classic 21-hydroxylase deficiency were also receiving GnRH agonists. Suppression of the hypothalamic-pituitary-gonadal axis in these patients was confirmed by a GnRH stimulation test at least 6 months before the study.

The study was approved by the Institutional Review Board at the National Institute of Child Health and Human Development, National

Institutes of Health. Written informed consent was obtained in all cases by a parent, and assent was given by children older than 7 yr.

Methods

All children were seen early in the morning on the day of the study, and standard anthropometric measurements were obtained. Pubertal stage was determined by physical examination by one of two trained observers (D.P.M., M.W.) according to the criteria of Tanner for breast development in females and testicular development in males (27, 28).

An indwelling venous catheter for blood sampling was inserted and all subjects rested in a supine position for a minimum of 30 min before blood samples were collected. All subjects were fasting for at least 9 h before sampling. In patients with CAH, the blood samples were obtained before the administration of the morning medication.

Blood samples for measurement of leptin, insulin, glucose, testosterone, epinephrine, and free metanephrine concentrations were obtained at 0800 h. Samples were centrifuged and separated immediately after collection and were stored at -80 C until assayed.

Assays

Leptin. Serum leptin concentrations were measured using a double-antibody RIA (Esoterix Endocrinology, Calabasas, CA) with a sensitivity of 0.03 ng/ml. The intraassay coefficients of variation (CV) were 9.6% and 6.7% at serum concentrations of 1.5 ng/ml and 4.7 ng/ml, respectively. The interassay CV were 12% and 11% at serum concentrations of 0.77 ng/ml and 6.0 ng/ml, respectively.

Insulin. Insulin was measured using a two-site immunochemiluminometric assay (Esoterix Endocrinology). The sensitivity of the assay was 1.0 μ U/ml. The intraassay CV were 6.2%, 6.6%, 5.2%, and 6.8% at serum concentrations of 3.5, 11, 16, and 35 μ U/ml, respectively. The interassay CV were 9.8%, 7.9%, 9.3%, 7.1%, and 9.0% at serum concentrations of 4.6, 11, 18, 42, and 104 μ U/ml, respectively.

Insulin resistance. Insulin resistance (IR) was estimated using the homeostasis model assessment (HOMA) method as previously described: IR = insulin (μ U/ml) \times glucose (mmol/liter)/22.5 (29).

Epinephrine and free metanephrine. Plasma epinephrine concentrations were quantified by liquid chromatography (30). Plasma free metanephrine concentrations were determined using a different liquid chromatography procedure after extraction onto solid phase ion exchange columns (31). The lowest detection limit for both assays was 1.0 pg/ml. The intraassay CV were 3.0% for epinephrine and 3.3% for free metanephrine. The interassay CV were 9.9% for epinephrine and 5.1% for free metanephrine.

Other measurements. Serum T concentrations were measured by standard RIA (Esoterix Endocrinology). Plasma glucose concentrations were measured by the hexokinase/glucose-6-phosphate dehydrogenase assay (Boehringer, Petersburg, VA).

Statistical analysis

Nonnormally distributed data were logarithmically transformed before statistical analysis. Comparisons between two groups were performed using the two-tailed *t* test. The relation between leptin concentrations and other parameters was investigated by calculation of Pearson's correlation coefficient. Stepwise multiple linear regression analysis was used to investigate independent predictors of serum leptin concentrations in both groups of children. Independent variables tested included body mass index, serum insulin, T concentrations, plasma epinephrine and free metanephrine concentrations, and gender. Gender was included as a dichotomous variable (*M* = 0, *F* = 1). Values are expressed as mean \pm SEM, unless otherwise specified.

Results

There were no significant differences in gender distribution, BMI, height, weight and height-age between children with classic CAH and normal subjects (Table 1). In normal children, there was no significant difference in BMI between

TABLE 1. Clinical characteristics of the 46 children

	CAH	Normal	<i>P</i>
No.	18	28	
% of females	33.3	42.9	NS
Age (yr)	7.2 \pm 0.7	8.9 \pm 0.3	0.03
Bone age (yr)	10.4 \pm 1.0		
BMI (kg/m ²)	18.0 \pm 0.8	16.8 \pm 0.4	NS
Weight (kg)	30.8 \pm 3.6	29.7 \pm 1.4	NS
Height (cm)	125.7 \pm 5.4	131.8 \pm 2.1	NS
Height-age (yr)	8.0 \pm 0.8	8.8 \pm 0.4	NS

Values are mean \pm SEM.

males and females (16.9 ± 0.5 vs. 16.6 ± 0.7). In children with CAH, however, males had significantly higher BMI than females (19.1 ± 1.1 vs. 16.0 ± 0.7 , $P = 0.03$).

Patients with classic CAH, when compared with their normal counterparts, had significantly lower plasma epinephrine (7.1 ± 1.3 vs. 50.0 ± 4.2 pg/ml, $P < 0.001$) and free metanephrine (18.4 ± 2.4 vs. 46.5 ± 4.0 pg/ml, $P < 0.001$) concentrations (Fig. 1, A and B) and significantly higher serum leptin (8.1 ± 2.0 vs. 2.5 ± 0.6 ng/ml, $P = 0.01$) and insulin (10.6 ± 1.4 vs. 3.2 ± 0.2 μ U/ml, $P < 0.001$) concentrations (Fig. 1, C and D). The differences observed in serum leptin and insulin concentrations between the two groups persisted after correction for BMI (Fig. 1, E and F). In addition, children with classic CAH had a significantly higher HOMA index than their normal counterparts (2.2 ± 0.3 vs. 0.7 ± 0.04 , $P < 0.001$) (Fig. 1G). No significant difference in fasting plasma glucose concentrations was observed between groups (81.9 ± 2.5 vs. 85.4 ± 1.3 mg/dl). Leptin concentrations were significantly and negatively correlated with epinephrine and free metanephrine concentrations, both before (epinephrine: $r = -0.50$, $P = 0.001$; free metanephrine: $r = -0.48$, $P = 0.002$) and after (epinephrine: $r = -0.50$, $P = 0.001$; free metanephrine: $r = -0.47$, $P = 0.002$) correction for BMI (Fig. 2).

T concentrations were higher in children with CAH than in normal children (23.7 ± 5.3 vs. 4.6 ± 0.5 ng/dl, $P = 0.003$), regardless of gender. Although prepubertal by physical examination and with T concentrations in the prepubertal range, normal males had significantly lower leptin concentrations than normal females (1.5 ± 0.5 vs. 3.8 ± 1.1 ng/ml, $P = 0.02$). This gender difference in leptin persisted after correction for BMI ($P = 0.008$). In patients with CAH, however, gender differences in leptin concentrations were not observed.

There was no significant correlation between the dose of concurrent hydrocortisone treatment and serum leptin ($r = -0.17$, $P = 0.5$) or insulin ($r = 0.24$, $P = 0.3$) concentrations in patients with classic CAH.

Stepwise multiple linear regression analysis indicated that serum leptin concentrations were best predicted by serum insulin concentrations, BMI, and gender when all patients were considered ($r^2 = 0.73$, $P < 0.001$) (Fig. 3). In normal children, leptin concentrations were independently related to BMI and gender ($r^2 = 0.41$, $P = 0.001$), whereas in patients with CAH, the only independent predictor of leptin concentrations was BMI ($r^2 = 0.64$, $P = 0.001$). In children with CAH, insulin resistance was best predicted by BMI ($r^2 = 0.58$, $P = 0.002$).

Similar results were produced when the patients with 11-hydroxylase deficiency or those receiving GnRH agonist therapy were excluded.

Discussion

Our findings demonstrate clear differences in adrenomedullary function, and leptin and insulin secretion between children with classic CAH and normal subjects matched for BMI. When measured in the morning in a fasting and rested state, children with CAH had significantly higher serum leptin and insulin concentrations, and lower plasma epi-

nephrine and free metanephrine concentrations than their healthy counterparts. These differences are likely to arise as a result of chronic adrenomedullary hypofunction in the former group, which may explain the elevation in both leptin and insulin concentrations, given that the secretion of both hormones is inhibited by catecholamines through β -adrenergic receptors. It is important to consider, however, that although there was no difference in BMI between the two groups, BMI may not be an accurate measure of body fat composition, and differences in body fat distribution might have contributed to the higher leptin and insulin concentrations in patients with CAH (32). It is also unknown whether diurnal variation of leptin occurs in patients with CAH, but a nocturnal rise in leptin is maintained in obesity, hyperinsulinemia (33), and Cushing's syndrome (34).

Impaired adrenomedullary function in patients with CAH was recently described (3). Patients with classic 21-hydroxylase deficiency demonstrate significantly lower (40–80%) plasma epinephrine and metanephrine concentrations as well as urinary epinephrine excretion than normal subjects. This compromise in adrenomedullary function is more pronounced in patients with the most severe, SW form of the disease and is attributed to a combination of decreased intraadrenal cortisol secretion and developmental defects in the formation of adrenal medulla (3).

The sympathetic nervous system and the adrenal medulla play an important role in the *ob* gene expression and leptin secretion. Both short- and long-term stimulation of adipose tissue β -adrenergic receptors inhibits leptin production (18–22, 35–37). In human adipose tissue cultures, incubation with isoproterenol decreases leptin mRNA levels and media concentrations of leptin in a time-dependent fashion, even in the presence of insulin or insulin and dexamethasone (38). Moreover, administration of epinephrine infusion in obese and lean subjects results in a significant reduction in leptin secretion (27%) and mRNA levels (47%), further supporting the concept of a potent inhibitory effect of β -adrenergic receptor stimulation on leptin synthesis and release (20).

Increased leptin secretion in association with long-standing sympathetic nervous system/adrenomedullary hypofunction in humans has been previously reported in Pima Indians, who also had increased incidence of obesity, IR, and diabetes mellitus type II (39, 40). The present study provides additional evidence for the involvement of the adrenal medulla in the regulation of leptin gene expression in humans. Given that leptin regulates negatively its own expression through sympathetic stimulation of β -adrenergic receptors (21, 41–43), patients with long-standing impaired adrenomedullary function may have even higher leptin concentrations than one would expect after an acute and reversible decrease of catecholamine secretion.

The elevated leptin concentrations in children with classic CAH may also be owing to their elevated insulin concentrations, compared with healthy subjects matched for BMI. Both *in vivo* and *in vitro* studies have demonstrated a dose- and time-dependent increase in leptin secretion in response to insulin (10, 14, 15, 44). Leptin, on the other hand, may inhibit insulin secretion via a direct effect on pancreatic β cells (44, 45), indicating that leptin and insulin may function as the afferent limb of a negative feedback loop that provides

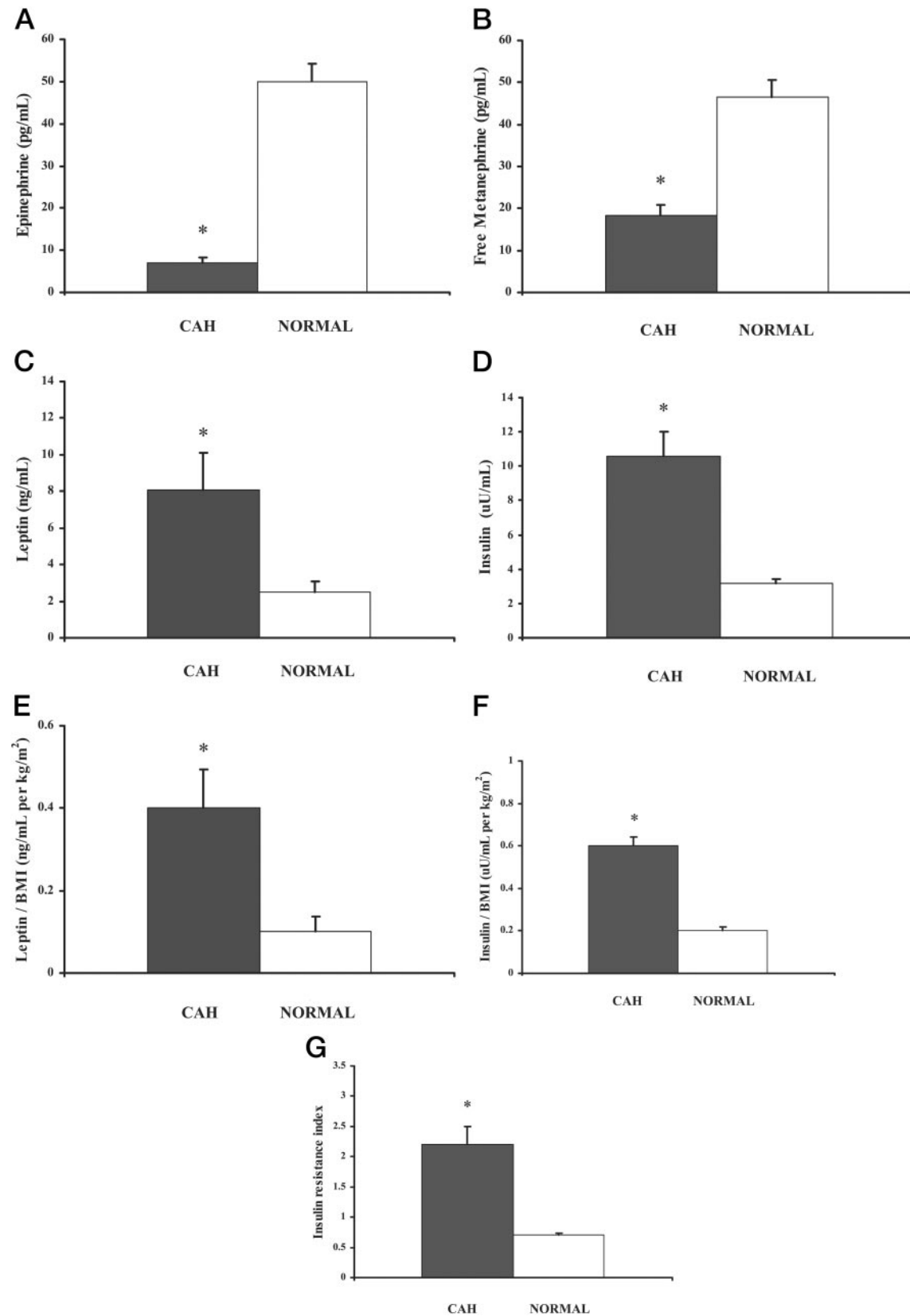


FIG. 1. Children with CAH, compared with normal children matched for BMI, had lower epinephrine (A) and free metanephrine (B) concentrations and higher leptin (C) and insulin (D) concentrations. This difference persisted when leptin and insulin values were corrected for BMI (E and F). IR, estimated using the HOMA, was significantly higher in children with CAH than normal controls (G). Error bars represent SEM. The asterisks indicate significant differences between groups.

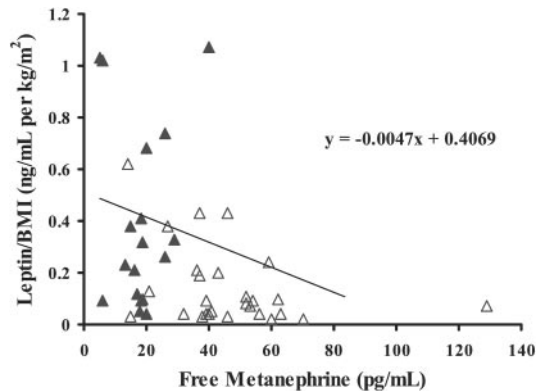


FIG. 2. Serum leptin concentrations corrected for BMI in relation to plasma free metanephrine concentrations in children with CAH (\blacktriangle) and normal controls (\triangle). There is a decline in leptin concentration with increasing catecholamine secretion.

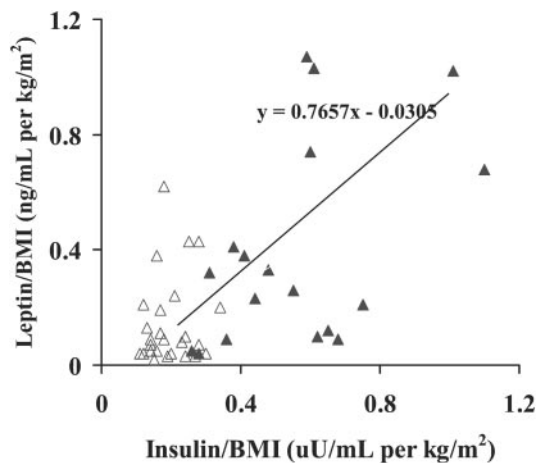


FIG. 3. Serum leptin concentrations corrected for BMI in relation to serum insulin concentrations also adjusted for BMI in children with CAH (\blacktriangle) and normal controls (\triangle). There is an increase in leptin concentration with increasing insulin concentration.

sensory input to the central nervous system about energy balance. Thus, they may interact at several levels to maintain energy homeostasis and body weight.

The significantly higher HOMA index in children with CAH, compared with normal subjects, most likely reflects the long-standing adrenomedullary hypofunction in association with intermittent hypercortisolism and/or adrenal hyperandrogenism. Although a fall in insulin sensitivity has been previously described in adults with nonclassic (late-onset) CAH (46), to our knowledge, there are no reports on alterations in insulin sensitivity/resistance in patients with the classic form of the disease.

Glucocorticoids increase *ob* gene expression and leptin secretion *in vivo*, and this effect appears to be more pronounced in obese than lean subjects (9–12). Although glucocorticoids have direct effects on the regulation of *ob* gene expression, which are independent of insulin concentrations and tissue sensitivity, they also affect leptin concentrations indirectly by inducing short- and long-term (via fat accumulation) IR and hyperinsulinism. The latter has been observed in patients with chronic hypercortisolism due to Cushing's syndrome, in which acute elevation in serum cor-

tisol concentrations did not affect leptin secretion (47), and the observed increase in leptin concentrations was primarily associated with BMI and serum insulin concentrations (48). In our patients with CAH, the concurrent daily dose of hydrocortisone was not correlated significantly with leptin or insulin concentrations. However, this dose hardly reflects the chronic exposure of the patients to glucocorticoids, and possible interpatient differences in hydrocortisone absorption, metabolism, and/or tissue sensitivity to glucocorticoids may have obscured the effect of glucocorticoid treatment on leptin and insulin secretion in these patients.

In addition to the catecholamines and insulin, sex steroids play an important role in the regulation of leptin secretion and, along with gender differences in body composition, contribute significantly to the sexually dimorphic pattern in leptin secretion described in humans (16, 17, 49, 50). Females display a gradual rise in leptin concentrations during puberty in parallel with the rise in estrogen concentrations, whereas males show a pubertal decline in leptin concentrations (51–54). This divergent pattern of leptin secretion in males and females is primarily owing to direct effects of sex steroids on *ob* gene expression and leptin secretion (16, 17, 49) but also to an indirect effect of the BMI by shifting the relative contribution to the increase of BMI toward muscle mass in the male and fat mass in the female (52). *In vitro* studies of cultured human adipocytes revealed a suppressive effect of T and its biologically active metabolite dihydrotestosterone on leptin production, suggesting a direct effect of T at the fat cell level before its aromatization to E2 (49). In the present study, the sexual dimorphism in serum leptin concentrations was observed in normal children but not in children with classic CAH, and this may be because of the significant differences in androgen concentrations between males and females in the former but not the latter group. The fact that leptin concentrations were higher in children with CAH than in normal subjects despite their higher T concentrations indicates that insulin secretion and/or adrenomedullary function may play a more potent role than androgens in the regulation of *ob* gene expression and leptin production.

The elevated leptin and insulin concentrations in children with classic CAH may bear a number of adverse effects on the course of the disease and therefore have important implications for the management of these patients: Hyperleptinemia decreases cortisol production by decreasing the expression of steroidogenic acute regulatory protein and may predispose patients with CAH to more frequent adrenal crises (55, 56). Both hyperleptinemia and hyperinsulinism may alter the activity of enzymes participating in adrenal steroidogenesis and may result in further increases in androgen production (23, 57, 58), imposing greater difficulty on their management. Furthermore, hyperinsulinism and insulin resistance play an important role in the development of polycystic ovary syndrome (PCOS) (59), metabolic syndrome-related atherosclerotic cardiovascular disease in adult life (59, 60), and adrenal incidentalomas (61–63). Although reports on cardiovascular morbidity and mortality in patients with classic CAH are lacking, there is considerable evidence to suggest increased incidence of PCOS in females with CAH (64, 65).

We conclude that children with classic CAH have signif-

icantly higher serum leptin and insulin concentrations and increased IR index, compared with their healthy counterparts. These differences most likely reflect long-term differences in adrenomedullary function, androgen concentrations, and exposure to glucocorticoids. Both elevated leptin and insulin concentrations play a role in enhancing adrenal androgen production and may expose these patients to an increased risk for developing PCOS and the cardiovascular complications of IR. Further studies are required to determine the regulation of insulin and glucose in patients with CAH, who may benefit from prevention and treatment of their IR.

Acknowledgments

We are indebted to the patients and their families for participating in this study; to Donna Peterson for assistance in data management; and to the members of the 9 West nursing staff at the Warren Grant Magnuson Clinical Center who assisted in carrying out these studies.

Received October 2, 2001. Accepted January 18, 2002.

Address all correspondence and requests for reprints to: Evangelia Charmandari, M.D., Pediatric and Reproductive Endocrinology Branch, National Institute of Child Health and Human Development, National Institutes of Health, 10 Center Drive, Building 10, Suite 9D42, Bethesda, Maryland 20892-1583. E-mail: charmane@mail.nih.gov.

D.P.M. is a Commissioned Officer in the United States Public Health Service.

References

- White PC, Speiser PW 2000 Congenital adrenal hyperplasia due to 21-hydroxylase deficiency. *Endocr Rev* 21:245–291
- New MI, Wilson RC 1999 Steroid disorders in children: congenital adrenal hyperplasia and apparent mineralocorticoid excess. *Proc Natl Acad Sci USA* 96:12790–12797
- Merke DP, Chrousos GP, Eisenhofer G, Weise M, Keil MF, Rogol AD, Van Wyk JJ, Bornstein SR 2000 Adrenomedullary dysplasia and hypofunction in patients with classic 21-hydroxylase deficiency. *N Engl J Med* 343:1362–1368
- Zhang Y, Proenca R, Maffei M, Barone M, Leopold L, Friedman JM 1994 Positional cloning of the mouse obese gene and its human homologue. *Nature* 372:425–432
- Halaas JL, Gajiwala KS, Maffei M, Cohen SL, Chait BT, Rabinowitz D, Lallone RL, Burley SK, Friedman JM 1995 Weight-reducing effects of the plasma protein encoded by the obese gene. *Science* 269:543–546
- Stephens TW, Basinski M, Bristow PK, Bue-Valleskey JM, Burgett SG, Craft L, Hale J, Hoffmann J, Hsiung HM, Kriauciunas A, et al. 1995 The role of neuropeptide Y in the antiobesity action of the obese gene product. *Nature* 377:530–532
- Campfield LA, Smith FJ, Guisez Y, Devos R, Burn P 1995 Recombinant mouse OB protein: evidence for a peripheral signal linking adiposity and central neural networks. *Science* 269:546–549
- Pelleymounter MA, Cullen MJ, Baker MB, Hecht R, Winters D, Boone T, Collins F 1995 Effects of the obese gene product on body weight regulation in ob/ob mice. *Science* 269:540–543
- De Vos P, Saladin R, Auwerx J, Staels B 1995 Induction of ob gene expression by corticosteroids is accompanied by body weight loss and reduced food intake. *J Biol Chem* 270:15958–15961
- Wabitsch M, Jensen PB, Blum WF, Christoffersen CT, Englaro P, Heinze E, Rascher W, Teller W, Tornqvist H, Hauner H 1996 Insulin and cortisol promote leptin production in cultured human fat cells. *Diabetes* 45:1435–1438
- Sliker LJ, Sloop KW, Surface PL, Kriauciunas A, LaQuier F, Manetta J, Bue-Valleskey J, Stephens TW 1996 Regulation of expression of ob mRNA and protein by glucocorticoids and cAMP. *J Biol Chem* 271:5301–5304
- Miell JP, Englaro P, Blum WF 1996 Dexamethasone induces an acute and sustained rise in circulating leptin levels in normal human subjects. *Horm Metab Res* 28:704–707
- Cusin I, Sainsbury A, Doyle P, Rohner-Jeanrenaud F, Jeanrenaud B 1995 The ob gene and insulin. A relationship leading to clues to the understanding of obesity. *Diabetes* 44:1467–1470
- Malmstrom R, Taskinen MR, Karonen SL, Yki-Jarvinen H 1996 Insulin increases plasma leptin concentrations in normal subjects and patients with NIDDM. *Diabetologia* 39:993–996
- Saad MF, Khan A, Sharma A, Michael R, Riad-Gabriel MG, Boyadjian R, Jinagouda SD, Steil GM, Kamdar V 1997 Physiological insulinemia acutely modulates plasma leptin. *Diabetes* 47:544–549
- Luukkkaa V, Pesonen U, Huhtaniemi I, Lehtonen A, Tilvis R, Tuomilehto J, Koulu M, Huupponen R 1998 Inverse correlation between serum testosterone and leptin in men. *J Clin Endocrinol Metab* 83:3243–3246
- Watanobe H, Suda T 1999 A detailed study on the role of sex steroid milieu in determining plasma leptin concentrations in adult male and female rats. *Biochem Biophys Res Commun* 259:56–59
- Deng C, Moinat M, Curtis L, Nadakal A, Preitner F, Boss O, Assimacopoulos-Jeannet F, Seydoux J, Giacchino JP 1997 Effects of beta-adrenoreceptor subtype stimulation on obese gene messenger ribonucleic acid and on leptin secretion in mouse brown adipocytes differentiated in culture. *Endocrinology* 138:548–552
- Sivitz WI, Fink BD, Morgan DA, Fox JM, Donohue PA, Haynes WG 1999 Sympathetic inhibition, leptin, and uncoupling protein subtype expression in normal fasting rats. *Am J Physiol* 277:E668–E677
- Carulli L, Ferrari S, Bertolini M, Tagliafico E, Del Rio G 1999 Regulation of ob gene expression: evidence for epinephrine-induced suppression in human obesity. *J Clin Endocrinol Metab* 84:3309–3312
- Commins SP, Marsh DJ, Thomas SA, Watson PM, Padgett MA, Palmiter R, Gettys TW 1999 Norepinephrine is required for leptin effects on gene expression in brown and white adipose tissue. *Endocrinology* 140:4772–4778
- Bottner A, Haidan A, Eisenhofer G, Kristensen K, Castle AL, Scherbaum WA, Schneider H, Chrousos GP, Bornstein SR 2000 Increased body fat mass and suppression of circulating leptin levels in response to hypersecretion of epinephrine in phenylethanolamine-N-methyltransferase (PNMT)-overexpressing mice. *Endocrinology* 141:4239–4246
- Glasow A, Haidan A, Hilbers U, Breidert M, Gillespie J, Scherbaum WA, Chrousos GP, Bornstein SR 1998 Expression of Ob receptor in normal human adrenals: differential regulation of adrenocortical and adrenomedullary function by leptin. *J Clin Endocrinol Metab* 83:4459–4466
- Webber J, Taylor J, Greathead H, Dawson J, Buttery PJ, Macdonald IA 1994 A comparison of the thermogenic, metabolic and haemodynamic responses to infused adrenaline in lean and obese subjects. *Int J Obes Relat Metab Disord* 18:717–724
- Schiffelers SL, Saris WH, Boomsma F, van Baak MA 2001 Beta (1)- and beta (2)-adrenoceptor-mediated thermogenesis and lipid utilization in obese and lean men. *J Clin Endocrinol Metab* 86:2191–2199
- Merke DP, Keil MF, Jones JV, Fields J, Hill S, Cutler GB Jr 2000 Flutamide, testosterone, and reduced hydrocortisone dose maintain normal growth velocity and bone maturation despite elevated androgen levels in children with congenital adrenal hyperplasia. *J Clin Endocrinol Metab* 85:1114–1120
- Marshall W, Tanner J 1969 Variation in pattern of pubertal changes in girls. *Arch Dis Child* 44:291–303
- Marshall W, Tanner J 1970 Variation in pattern of pubertal changes in boys. *Arch Dis Child* 45:13–23
- Matthews DR, Hosker JP, Rudenski AS, Naylor BA, Treacher DF, Turner RC 1985 Homeostasis model assessment: insulin resistance and beta-cell function from fasting plasma glucose and insulin concentrations in man. *Diabetologia* 28:412–419
- Eisenhofer G, Goldstein DS, Stull R, Keiser HR, Sunderland T, Murphy DL, Kopin IJ 1986 Simultaneous liquid-chromatographic determination of 3,4-dihydroxyphenylglycol, catecholamines, and 3,4-dihydroxyphenylalanine in plasma, and their responses to inhibition of monoamine oxidase. *Clin Chem* 32:2030–2033
- Lenders JW, Eisenhofer G, Armando I, Keiser HR, Goldstein DS, Kopin IJ 1993 Determination of metanephrines in plasma by liquid chromatography with electrochemical detection. *Clin Chem* 39:97–103
- Vgontzas AN, Papanicolaou DA, Bixler EO, Hopper K, Lotsikas A, Lin HM, Kales A, Chrousos GP 2000 Sleep apnea and daytime sleepiness and fatigue: relation to visceral obesity, insulin resistance, and hypercytokinemia. *J Clin Endocrinol Metab* 85:1151–1158
- Saad MF, Riad-Gabriel MG, Khan A, Sharma A, Michael R, Jinagouda SD, Boyadjian R, Steil GM 1998 Diurnal and ultradian rhythmicity of plasma leptin: effects of gender and adiposity. *J Clin Endocrinol Metab* 83:453–459
- Weise M, Abad V, Considine RV, Nieman L, Rother KI 1999 Leptin secretion in Cushing's syndrome: preservation of diurnal rhythm and absent response to corticotropin-releasing hormone. *J Clin Endocrinol Metab* 84:2075–2079
- Nonogaki K 2000 New insights into sympathetic regulation of glucose and fat metabolism. *Diabetologia* 43:533–549
- Mantzoros CS, Qu D, Frederich RC, Susulic VS, Lowell BB, Maratos-Flier E, Flier JS 1996 Activation of beta (3) adrenergic receptors suppresses leptin expression and mediates a leptin-independent inhibition of food intake in mice. *Diabetes* 45:909–914
- Kosaki A, Yamada K, Kuzuya H 1996 Reduced expression of the leptin gene (ob) by catecholamine through a G (S) protein-coupled pathway in 3T3-L1 adipocytes. *Diabetes* 45:1744–1749
- Halleux CM, Servais I, Reul BA, Detry R, Brichard SM 1998 Multihormonal control of ob gene expression and leptin secretion from cultured human visceral adipose tissue: increased responsiveness to glucocorticoids in obesity. *J Clin Endocrinol Metab* 83:902–910
- Fox C, Esparza J, Nicolson M, Bennett PH, Schulz LO, Valencia ME, Ra-

- vussin E 1999 Plasma leptin concentrations in Pima Indians living in drastically different environments. *Diabetes Care* 22:413–417
40. Tataranni PA, Cizza G, Snitker S, Gucciardo F, Lotsikas A, Chrousos GP, Ravussin E 1999 Hypothalamic-pituitary-adrenal axis and sympathetic nervous system activities in Pima Indians and Caucasians. *Metabolism* 48:395–399
 41. Trayhurn P, Duncan JS, Rayner DV, Hardie LJ 1996 Rapid inhibition of ob gene expression and circulating leptin levels in lean mice by the beta 3-adrenoceptor agonists BRL 35135A and ZD2079. *Biochem Biophys Res Commun* 228:605–610
 42. Gettys TW, Harkness PJ, Watson PM 1996 The beta 3-adrenergic receptor inhibits insulin-stimulated leptin secretion from isolated rat adipocytes. *Endocrinology* 137:4054–4057
 43. Rayner DV, Simon E, Duncan JS, Trayhurn P 1998 Hyperleptinaemia in mice induced by administration of the tyrosine hydroxylase inhibitor alpha-methyl-p-tyrosine. *FEBS Lett* 429:395–398
 44. Fried SK, Ricci MR, Russell CD, Laferrere B 2000 Regulation of leptin production in humans. *J Nutr* 130:3127S–3131S
 45. Emilsson V, Liu YL, Cawthorne MA, Morton NM, Davenport M 1997 Expression of the functional leptin receptor mRNA in pancreatic islets and direct inhibitory action of leptin on insulin secretion. *Diabetes* 46:313–316
 46. Speiser PW, Serrat J, New MI, Gertner JM 1992 Insulin insensitivity in adrenal hyperplasia due to nonclassical steroid 21-hydroxylase deficiency. *J Clin Endocrinol Metab* 75:1421–1424
 47. Cizza G, Lotsikas AJ, Licinio J, Gold PW, Chrousos GP 1997 Plasma leptin levels do not change in patients with Cushing's disease shortly after correction of hypercortisolism. *J Clin Endocrinol Metab* 82:2747–2750
 48. Widjaja A, Schurmeyer TH, Von zur Muhlen A, Brabant G 1998 Determinants of serum leptin levels in Cushing's syndrome. *J Clin Endocrinol Metab* 83:600–603
 49. Wabitsch M, Blum WF, Muehe R, Braun M, Hube F, Rascher W, Heinze E, Teller W, Hauner H 1997 Contribution of androgens to the gender difference in leptin production in obese children and adolescents. *J Clin Invest* 100:808–813
 50. Palmert MR, Radovick S, Boepple PA 1998 The impact of reversible gonadal sex steroid suppression on serum leptin concentrations in children with central precocious puberty. *J Clin Endocrinol Metab* 83:1091–1096
 51. Garcia-Mayor RV, Andrade MA, Rios M, Lage M, Dieguez C, Casanueva FF 1997 Serum leptin levels in normal children: relationship to age, gender, body mass index, pituitary-gonadal hormones, and pubertal stage. *J Clin Endocrinol Metab* 82:2849–2855
 52. Blum WF, Englaro P, Hanitsch S, Juul A, Hertel NT, Muller J, Skakkebaek NE, Heimann ML, Birkett M, Attanasio AM, Kiess W, Rascher W 1997 Plasma leptin levels in healthy children and adolescents: dependence on body mass index, body fat mass, gender, pubertal stage, and testosterone. *J Clin Endocrinol Metab* 82:2904–2910
 53. Roemmich JN, Clark PA, Berr SS, Mai V, Mantzoros CS, Flier JS, Weltman A, Rogol AD 1998 Gender differences in leptin levels during puberty are related to the subcutaneous fat depot and sex steroids. *Am J Physiol* 275:E543–E551
 54. Horlick MB, Rosenbaum M, Nicolson M, Levine LS, Fedun B, Wang J, Pierson Jr RN, Leibel RL 2000 Effect of puberty on the relationship between circulating leptin and body composition. *J Clin Endocrinol Metab* 85:2509–2518
 55. Pralong FP, Roduit R, Waeber G, Castillo E, Mosimann F, Thorens B, Gaillard RC 1998 Leptin inhibits directly glucocorticoid secretion by normal human and rat adrenal gland. *Endocrinology* 139:4264–4268
 56. Cherradi N, Capponi AM, Gaillard RC, Pralong FP 2001 Decreased expression of steroidogenic acute regulatory protein: a novel mechanism participating in the leptin-induced inhibition of glucocorticoid biosynthesis. *Endocrinology* 142:3302–3308
 57. L'Allemand D, Penhoat A, Lebrethon MC, Ardevol R, Baehr V, Oelkers W, Saez JM 1996 Insulin-like growth factors enhance steroidogenic enzyme and corticotropin receptor messenger ribonucleic acid levels and corticotropin steroidogenic responsiveness in cultured human adrenocortical cells. *J Clin Endocrinol Metab* 81:3892–3897
 58. Biason-Lauber A, Zachmann M, Schoenle EJ 2000 Effect of leptin on CYP17 enzymatic activities in human adrenal cells: new insight in the onset of adrenarche. *Endocrinology* 141:1446–1454
 59. Dunaif A 1997 Insulin resistance and the polycystic ovary syndrome: mechanism and implications for pathogenesis. *Endocr Rev* 18:774–800
 60. Arslanian SA, Lewy VD, Danadian K 2001 Glucose intolerance in obese adolescents with polycystic ovary syndrome: roles of insulin resistance and beta-cell dysfunction and risk of cardiovascular disease. *J Clin Endocrinol Metab* 86:66–71
 61. Seppel T, Schlaghecke R 1994 Augmented 17 alpha-hydroxyprogesterone response to ACTH stimulation as evidence of decreased 21-hydroxylase activity in patients with incidentally discovered adrenal tumours ('incidentaltomas'). *Clin Endocrinol (Oxf)* 41:445–451
 62. Reincke M, Fassnacht M, Vath S, Mora P, Allolio B 1996 Adrenal incidentaltomas: a manifestation of the metabolic syndrome? *Endocr Res* 22:757–761
 63. Barzon L, Scaroni C, Sonino N, Fallo F, Gregianin M, Macri C, Boscaro M 1998 Incidentally discovered adrenal tumors: endocrine and scintigraphic correlates. *J Clin Endocrinol Metab* 83:55–62
 64. Hague WM, Adams J, Rodda C, Brook CG, de Bruyn R, Grant DB, Jacobs HS 1990 The prevalence of polycystic ovaries in patients with congenital adrenal hyperplasia and their close relatives. *Clin Endocrinol (Oxf)* 33:501–510
 65. Ghizzoni L, Viridis R, Vottero A, Cappa M, Street ME, Zampolli M, Ibanez L, Bernasconi S 1996 Pituitary-ovarian responses to leuprolide acetate testing in patients with congenital adrenal hyperplasia due to 21-hydroxylase deficiency. *J Clin Endocrinol Metab* 81:601–606