Endocrine Research

Change in Amniotic Fluid Levels of Multiple Anti-Angiogenic Proteins before Development of Preeclampsia and Intrauterine Growth Restriction

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Context: The cause of preeclampsia remains unknown. Excessive antiangiogenic proteins have been proposed to play a pathogenic role in preeclampsia.

Objective: Our objective was to determine the differences in soluble endoglin (sEndoglin), soluble fms-like tyrosine kinase receptor-1 (sFLT1), leptin, adiponectin, and endothelin 1 concentrations between normal and preeclampsia amniotic fluid (AF). Such results may help us understand the pathophysiology of preeclampsia.

Methods: We performed a nested case-control study. Seventy-one women with preeclampsia were matched to 71 normotensive controls. The preeclamptic women were broken into two subgroups according to the association with fetal intrauterine growth restriction (IUGR). AF concentrations of sEndoglin, sFLT1, leptin, adiponectin, and endothelin 1 were measured by ELISA. Receiver-operating characteristics curve analysis was used to compare the discriminative values of these potential biomarkers. Functional network analysis was performed using MetaCore to reveal the common functions of the interacting proteins.

Results: Increased AF concentrations of sFLT1, sEndoglin, endothelin 1, and leptin were found in women who later developed preeclampsia. sFLT1, sEndoglin, leptin, and adiponectin were significantly higher in the preeclampsia with IUGR than those without IUGR. Leptin has the largest area under the curve (0.753). Network analysis revealed that elevated amniotic proteins are involved in the inflammatory process of the human placenta.

Conclusions: Significant elevation of leptin can be detected in AF 2 months earlier than the appearance of symptoms; thus, it may be used as a predictive marker for preeclampsia. The increase of these antiangiogenic proteins supports the roles of inflammation and oxidative stress in pathogenesis of preeclampsia. (*J Clin Endocrinol Metab* 95: 1431–1441, 2010)

Preeclampsia, which occurs in about 3–5% of all pregnancies, is one of the most common causes for maternal mortality and neonatal death (1, 2). Widespread endothelial dysfunctions triggered by placenta-derived antiangiogenic factors are considered to be major mechanisms of preeclampsia, and symptoms resolve after expulsion of the placenta. Preeclampsia is thought to be caused by the maternal systemic response to poor placen-

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Abbreviations: AF, Amniotic fluid; AUC, area under curve; BMI, body mass index; CV, coefficient of variation; IUGR, intrauterine growth restriction; JNK, c-Jun N-terminal kinase; NF- κ B, nuclear factor- κ B; ROC, receiver-operating characteristic; SEndoglin, soluble endoglin; sFLT1, soluble fms-like tyrosine kinase receptor-1; SMAD, phosphorylated mothers against decapentaplegic; STAT, signal transducer and activator of transcription; VEGFR, vascular endothelial growth factor receptor.

tal formation. Recently some authors proposed that preeclampsia is not only an endothelial disease but a wide systemic inflammatory disease, which results from oxidative stress rather than hypoxia (3, 4). However, the pathophysiology of preeclampsia is not completely clear.

Endoglin, also named CD105, is a coreceptor of TGF- β 1 and -3, which is highly expressed in the cell membranes of vascular endothelium and syncytiotrophoblasts (5, 6). The TGF- β receptors bind TGF- β 1 and -3, and endoglin modulates the action of these ligands, acting as an intracellular signaling mediator and regulating transcriptional responses (7). However, soluble endoglin (sEndoglin) is a placenta-derived soluble form of endoglin. This hypoxiainduced protein is associated with the subsequent development of preeclampsia (8).

Soluble fms-like tyrosine kinase 1 (sFLT1), also known as soluble vascular endothelial growth factor receptor (VEGFR)-1, is a truncated form of the VEGFR-1 that is secreted by the human placenta (9). Circulating sFLT1 binds the proangiogenic proteins, vascular endothelial growth factor and placental growth factor, and prevents their interactions with endothelial cell receptors, thereby inducing endothelial dysfunction (10). Recent studies demonstrated that both circulating and placental sFLT1 expression is essential in the pathogenesis of preeclampsia (11–13). In addition, elevated sFLT1 levels have also been identified in intrauterine growth restriction (IUGR) pregnancies (14).

Endothelin-1, a potent endogenous vasoconstrictor peptide, is involved in the regulation of vascular tone in hypertensive states, and increased blood concentrations of endothelin 1 are associated with preeclampsia (15). Thus, endothelin-1 was proposed to be one of the key links between primary placental disorders and the systemic endothelial dysfunction of preeclampsia (16).

Leptin, normally produced by peripheral adipocytes, is also secreted by human amnion cells into amniotic fluid (AF) (17). The human placenta expresses a high amount of leptin mRNA, and the excessive placental leptin release is thought to be secondary to hypoxia in nonobese preeclamptic women (18). Hypoxia is shown to up-regulate placental leptin gene expression (19), and increased placental leptin synthesis is associated with preeclampsia (20). Furthermore, leptin also plays a key role in T cell activation (21).

Adiponectin is an insulin-sensitizing and antiinflammatory protein released by adipocytes. It suppresses endothelial activation, and positive immunostaining for adiponectin has been observed in endothelial cells of preeclamptic placental tissues (22). The role of adiponectin in preeclampsia remains unclear, and serum levels of adiponectin are found to be increased (18, 23, 24) or decreased (22, 25) in patients with preeclampsia. However, the AF concentrations of adiponectin and preeclampsia have not been determined.

The initiating events in preeclampsia, even before preeclampsia onset, are postulated to be reduced uteroplacental perfusion and poor placentation (4, 26). Thus, we hypothesized that changes in these protein levels in AF might reflect the early pathophysiology of preeclampsia before the appearance of clinical manifestations. In the second trimester of pregnancy, the AF concentrations have been described for sFLT1 (27), endothelin-1 (28), and leptin (29), but those of sEndoglin, and adiponectin are yet to be defined. Our study appears to be the first to simultaneously analyze the concentrations of multiple antiangiogenic proteins in AF in an attempt to understand the pathophysiology of preeclampsia.

Subjects and Methods

Study population

From 2002 to 2008, we prospectively stored AF of 7283 pregnant women who underwent genetic amniocentesis at Chang Gung Memorial Hospital, Lin-Kou Medical Center. The various reasons for amniocentesis included advanced maternal age, high levels of maternal Down syndrome serum markers, fetal ultrasound anomalies, previous history of chromosomal anomalies, a balanced chromosomal translocation carrier in either of the parents, thalassemia carriers, and maternal request.

Preeclampsia was defined by gestational hypertension, proteinuria, and hyperuricemia, with the reversal of hypertension and proteinuria after delivery (30). Gestational hypertension is defined as systolic blood pressure of more than 140 mmHg or diastolic blood pressure of more than 90 mm Hg, beginning after 20 wk of gestation in previously normotensive women. Proteinuria is defined as more than 300 mg protein in a 24-h urine collection or more than 2+ on a voided or more than 1+ on a catheterized random urine sample (31). The time of preeclampsia onset is defined as the time of detection of elevated blood pressure or urinary protein measurement leading to the diagnosis of preeclampsia. Fetal IUGR was defined as women with infant birth weight less than the fifth percentile. Percentiles for growth parameters were derived from a reference Taiwanese population (32).

The flow chart of patient enrollment is summarized in Fig. 1. Patients with obesity, multiple pregnancies, gestational or overt diabetes, chronic hypertension with superimposed preeclampsia, renal disease, or any known metabolic disorders (*e.g.* hyperthyroidism, hyperlipidemia) were excluded to avoid any possible bias such as the association of leptin with diabetes (33) or obesity with preeclampsia (34). The obesity defined as body mass index (BMI) greater than 27 kg/m² during midtrimester amniocentesis. The exact gestational age, fetal growth, and amniotic fluid index of all cases were all ascertained by serial ultrasonography. At amniocentesis, fetuses with growth restriction, oligohydramnios, or polyhydramnios were all excluded. From the same amniotic fluid bank, we used matching criteria of maternal age,

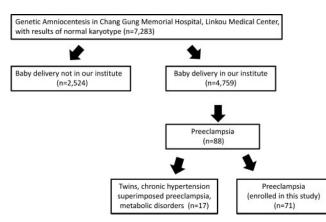


FIG. 1. The flow chart of patient enrollment.

parity, gestational age, and blood pressure during the time of the amniocentesis and BMI to select women who had uncomplicated, normotensive pregnancies free of the aforementioned medical complications and deliveries of healthy babies as the control group. This study was approved by the Ethical Committee of Chang Gung Memorial Hospital (institutional review board no. 93-6366 and 97-2024B).

Procedures of specimen collection

Before amniocentesis was performed, written informed consents were obtained from all participants. In this study, genetic amniocenteses were performed between 16 and 19 gestational weeks at our hospital. Using a transabdominal approach, about 20 ml of AF were aspirated from each patient. After amniocytes were collected via centrifugation for culture and cytogenetic studies, we consecutively stored 4 ml of fresh AF in a -80 C freezer.

Immunoassay procedures

All of the commercially available ELISAs were performed according to the manufacturer's instructions (R&D Systems, Minneapolis, MN) as previously described (35). The AF concentrations of sEndoglin and adiponectin were measured without prior dilution. The minimal detectable limits of the assays for sEndoglin and adiponectin were 0.007 and 0.246 ng/ml, respectively. The intraassay coefficients of variation (CVs) for sEndoglin and adiponectin ELISA were 2.8 and 3.2%, respectively, and the interassay CVs were 6.9 and 7.9%, respectively (n = 20).

Before the AF sFLT1 concentrations were analyzed, AF specimens were diluted to 1:100 in the calibrator diluent buffer that was included in the ELISA kit. The minimal detection limit of the sFLT1 was 3.5 pg/ml, the intraassay CV was 2.6%, and the interassay CVs was 9.8% (n = 20). Before the AF concentrations of leptin were determined, AF samples were diluted to 1:50 with the calibrator diluent buffer. The minimal detection limit of the leptin ELISA was 7.8 pg/ml, the intraassay CVs was 3.3%, and interassay CVs was 5.4% (n = 20).

On the other hand, endothelin-1 in AF needed to be concentrated before the ELISA was performed. Before AF concentration of endothelin-1 were measured, 0.5 ml of each AF was evaporated down in a centrifugal evaporator (minimum drying time 4 h at 37 C) and then reconstituted with 0.125 ml of sample diluent. The minimal detection limit of the endothelin-1 ELISA was 1.0 pg/ml, the intraassay CVs was 4.6%, and the interassay CVs was 6.5% (n = 20).

Statistical analysis

In this study, we used χ^2 tests to compare categorical variables and Pearson's correlation to analyze the relationships of all permutation pairs of antiangiogenic markers. Student *t* tests, one-way ANOVA tests, and *post hoc* Scheffé tests were used to analyze continuous variables (preeclampsia with IUGR, preeclampsia without IUGR, and normal controls). Binary logistic regression with the forward stepwise method was used for multivariate analysis. Receiver-operating characteristic (ROC) curve analysis was used to evaluate the discriminative values of the preeclamptic markers. For all analyses, P < 0.05 was considered statistically significant. All statistics were carried out using SPSS version 12 for Windows (SPSS Inc., Chicago, IL).

Bioinformatics analysis

Procedures of networks analysis to elucidate the biologic processes of differentially expressed proteins in amniotic fluid of the patients who later develop preeclampsia were similar to what we have reported (36). Briefly, we used the analyze networks algorithm in MetaCore (GeneGo, St. Joseph, MI) to build the networks from five input genes: adiponectin, endoglin, endothelin-1, FLT1, and leptin (37, 38). MetaCore is a Web-based computational platform designed for systems biology and drug discovery. It includes a curated database of human protein interactions and metabolism; thus, it is useful for analyzing a cluster of genes in the context of regulatory networks and signaling pathways (39). For the network analysis of a group of genes, MetaCore can be used to calculate the statistical significance (P value) based on the probability of assembly from a random set of nodes (genes) of the same size as the input list (40).

Results

Characteristics of the patients

One hundred forty-two pregnant women who underwent genetic amniocentesis were enrolled in this study. The preeclampsia group consisted of 71 women who subsequently developed preeclampsia, and the control group consisted of 71 women with uncomplicated pregnancies. There was no significant difference in maternal age, BMI, parity, gestational age at amniocentesis, or smoking behavior between groups. The karyotypes of all enrolled cases were normal. As expected with the nature of this disease, women with preeclampsia had higher blood pressures during delivery, lower gestational weeks at delivery, and lower birth weight (all were P < 0.05) than the control women. Likewise, the birth weights of neonates were significant lower in the preeclampsia women with IUGR than the preeclampsia women without IUGR (P < 0.05). The onset of preeclampsia was 26.6 ± 3.1 wk (mean \pm sD), with a range of 21 to 33 wk. Patients' characteristics are summarized in Table 1.

TABLE 1. Characteristics of the entire study population

			Control	
	PE without IUGR (n = 39)	PE with IUGR (n = 32)	Control (n = 71)	P value
Age (yr)	35.4 ± 3.4	34.6 ± 3.9	35.1 ± 1.4	NS
Nulliparity (%) [#]	17 (43.6)	18 (56.3)	37 (52.1)	NS
BMI at AC	21.9 ± 2.2	21.8 ± 2.3	21.2 ± 3.0	NS
BMI at delivery	27.5 ± 3.1	28.3 ± 2.8	27.2 ± 2.8	NS
SBP at AC (mm Hg)	118.2 ± 13.7	116.8 ± 11.6	115.7 ± 12.1	NS
DBP at AC (mm Hg)	73.3 ± 10.2	69.7 ± 8.2	69.6 ± 8.6	NS
SBP at delivery (mm Hg)	171.6 ± 15.8 ^a	168.2 ± 15.3 ^b	125.3 ± 10.0 ^{a,b}	< 0.001*
DBP at delivery (mm Hg)	102.6 ± 8.8 ^c	101.4 ± 10.2 ^d	76.9 ± 8.5 ^{c,d}	< 0.001*
GA at AC (wk)	17.3 ± 0.9	17.5 ± 0.9	17.8 ± 1.0	NS
GA at delivery (wk)	33.6 ± 2.8^{e}	33.3 ± 3.0^{f}	38.6 ± 1.2 ^{e,f}	< 0.001*
Birth weight (g)	2042 ± 620 ^{g, h}	1470 ± 527 ^{g,h}	3109 ± 333 ^{g,h}	< 0.001*
Smoking behavior (%)#	2 (5.1)	1 (3.1)	2 (2.8)	NS

The values shown are mean \pm sD (Student *t* test). The data were analyzed with Scheffé *post hoc* tests and are presented as mean \pm sEM. PE, Preeclampsia; BW, body weight; AC, amniocentesis; SBP, systolic blood pressure; DBP, diastolic blood pressure; GA, gestational age; NS, not significant, defined as $P \ge 0.05$ (*P* values are given only for significant differences).

[#] Data were calculated by χ^2 test; * significant difference by one-way ANOVA test (P < 0.05).

^a P < 0.001; ^b P < 0.001; ^c P < 0.001; ^d P < 0.001; ^e P < 0.001; ^f P < 0.001; ^g P < 0.001; ^h P < 0.001.

Amniotic fluid sEndoglin, sFLT1, endothelin-1, leptin, and adiponectin concentrations between preeclampsia and control groups

The AF concentrations of sEndoglin, sFLT1, endothelin-1, and leptin were significantly higher in the preeclampsia than in the control groups. However, there were no significant differences in adiponectin concentrations between two groups (Table 2). We also analyzed these proteins in only primigravida (labeled as nulliparity in this study) and identified similar results (Supplemental Table 1, published as supplemental data on The Endocrine Society's Journals Online web site at http://jcem.endojournals.org). In women with preeclampsia, there was no significant difference in these AF protein levels between nulliparity and multiparity (Supplemental Table 2).

TABLE 2. AF concentrations of leptin, sEng, sFlt-1, endothelin-1, and adiponectin between the PE and control groups

	PE (n = 71) ^a	Control (n = 71) ^a	P value
Leptin (ng/ml)	7.11 ± 0.46	4.65 ± 0.33	< 0.001
sEndoglin (ng/ml)	0.99 ± 0.82	0.68 ± 0.25	0.001
sFLT1 (pg/ml)	67834 ± 5539	47648 ± 2315	0.001
Endothelin-1 (pg/ml)	57.56 ± 12.55	28 ± 3.43	0.025
Adiponectin (ng/ml)	20.52 ± 2.28	16.09 ± 0.8	NS

PE, Preeclampsia; NS, not significant, defined as $P \ge 0.05$ [*P* values are given only for significant differences (Student t test)].

 $^{\rm a}$ Results are represented as mean \pm sem.

Amniotic fluid antiangiogenic proteins between the preeclampsia with IUGR and the preeclampsia without IUGR groups

One-way ANOVA revealed significant differences (P <0.05) in the AF concentrations of leptin, sEndoglin, sFLT1, and adiponectin among the preeclampsia with and without IUGR and the control groups (Table 3). In post *hoc* Scheffé tests, sEndoglin and leptin were significantly increased in the preeclampsia with IUGR and the preeclampsia without IUGR groups, compared with the control group (all were P < 0.05). There was no significant difference in leptin and sEndoglin between the preeclampsia with IUGR and the preeclampsia without IUGR groups. The AF concentrations of sFLT1 in the preeclampsia with IUGR, but not the preeclampsia without IUGR, were significantly higher than that of the controls (P =0.001). The AF concentrations of adiponectin were significantly higher in the preeclampsia with IUGR group than in either the preeclampsia without IUGR group (P =0.008) or the control group (P = 0.004), despite a lack of a significant difference between the preeclampsia without IUGR and control groups.

Correlations among the amniotic fluid concentrations of sEndoglin, sFLT1, endothelin-1, leptin, and adiponectin

To examine the relationships among the five antiangiogenic proteins in pathogenesis of preeclampsia, we performed Pearson's correlation tests (n = 142) on all of the 10 protein pairs (Fig. 2). There were significantly positive correlations of sEndoglin with sFLT1, leptin, and adiponectin (all were P < 0.001). sFLT1 was positively correlated with leptin, endothelin-1, sEndoglin, and adi-

	PE without $IUGR (n = 39)$	PE with IUGR (n = 32)	Control (n = 71)	P value
Leptin (ng/ml)	6.83 ± 0.68^{a}	7.47 ± 0.68^{b}	4.65 ± 0.33 ^{a,b}	< 0.001
sEndoglin (ng/ml)	$0.99 \pm 0.13^{\circ}$	0.98 ± 0.09^{d}	0.68 ± 0.03 ^{c,d}	0.003
sFLT1 (pg/ml)	59263 ± 7440	78279 ± 8036^{e}	47648 ± 2315 ^e	0.001
Endothelin-1 (pg/ml) Adiponectin (ng/ml)	$\begin{array}{c} 50.01 \pm 17.17 \\ 15.3 \pm 2.25^{f} \end{array}$	66.76 ± 18.53 26.9 ± 4.01 ^{f,g}	28.0 ± 3.43 16.09 ± 0.8^{g}	NS 0.001

TABLE 3. AF concentrations of leptin, sEng. sFlt-1, endothelin-1, and adiponectin among three study groups

The data were analyzed with Scheffé post hoc tests and are presented as mean ± SEM. P values are given only for significant differences (one-way ANOVA). PE, Preeclampsia; NS, not significant, defined as $P \ge 0.0$.

 $^{a}P = 0.002; ^{b}P = 0.01; ^{c}P = 0.02; ^{d}P = 0.02; ^{e}P = 0.001; ^{f}P = 0.008; ^{g}P = 0.004.$

ponectin (all were P < 0.05). Also, there were positive correlations between the leptin vs. adiponectin pair (P <0.001) and the endothelin-1 vs. adiponectin pair (P <0.001). Only two pairs were not significantly correlated: endothelin-1 vs. leptin and endothilin-1 vs. sEndoglin (data not shown).

Multivariate analysis

The association between antiangiogenic proteins and preeclampsia was further studied by a logistic regression model. Among these proteins, leptin (P < 0.001) and sFLT1 (P = 0.008) were factors independently associated with preeclampsia occurrence.

ROC curve analyses

We used the ROC curve analyses to evaluate the usefulness of each of the AF proteins for identifying women with subsequent development of preeclampsia from the control women. When we compared the preeclampsia (n = 71) with the control (n = 71) groups, the areas under curve (AUC) of leptin, sEndoglin, sFLT1, endothelin-1, and adiponectin were 0.753 (P < 0.001), 0.655 (P = $(0.002), 0.655 \ (P = 0.002), 0.614 \ (P = 0.02), and 0.499$ (P = NS), respectively (Fig. 3A). Only AF leptin is considered to have a discriminative value (AUC > 0.7). When the preeclampsia with IUGR (n = 32) and the control (n = 32)71) groups were compared, the AUC of leptin, sEndoglin, sFLT1, endothelin-1, and adiponectin were 0.774 (P <(0.001), 0.676 (P = 0.004), 0.750 (P < 0.001), 0.680 (P =(0.003), and (0.655) (P = (0.012), respectively (Fig. 3B). Both leptin and sFLT1 have potentially discriminative values for the preeclampsia with IUGR cases. However, when the preeclampsia without IUGR (n = 39) and the control (n = 39)71) groups were analyzed, the AUC of leptin, sEndoglin, sFLT1, endothelin-1, and adiponectin were 0.750 (P <(0.001), 0.609 (P = NS), 0.562 (P = NS), 0.611 (P = NS), 0.6and 0.374 (P = NS), respectively (Fig. 3C). Only leptin can discriminate the preeclampsia without IUGR cases from the controls. On the other hand, the combined use of any protein pair did not increase the AUC in ROC analyses (Supplemental Table 3).

Functional network

Using MetaCore algorithm for network analysis, we found that leptin, endoglin, FLT1, endothelin-1, and adiponectin interacted in the network of pathways with

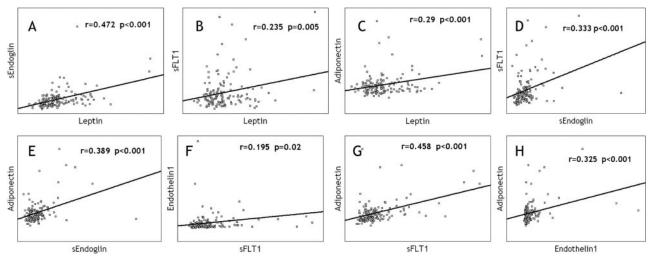


FIG. 2. Significant correlations exist in eight of 10 pairs between amniotic fluid antiangiogenic proteins. Leptin and sEndoglin (A), leptin and sFLT1 (B), leptin and adiponectin (C), sEndoglin and sFLT1 (D), sEndoglin and adiponectin (E), sFLT1 and endothelin 1 (F), sFLT1 and adiponectin (G), and endothelin-1 and adiponectin (H) are shown.

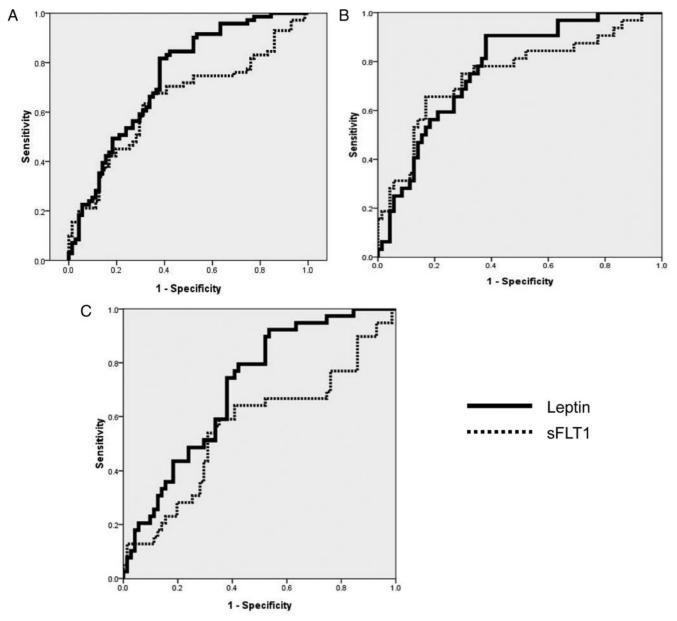


FIG. 3. ROC curve analyses in three comparison groups. ROC curves between the preeclampsia (n = 71) and the control (n = 71) groups (A), ROC curves between the preeclampsia with IUGR (n = 32) and the control (n = 71) groups (B), and ROC curves between the preeclampsia without IUGR (n = 39) and the control (n = 71) groups (C) are shown. AUC and *P* values are described in the text. Only the plots with AUC greater than 0.7 are shown here (leptin and sFLT1).

 $P = 1.14 \times 10^{-12}$ (Fig. 4), indicating that the probability of assembly from random sets of nodes (genes) was very low (40). Through functional analyses using the Disease identification and Tissue localization modules of MetaCore Suites, this network was related to inflammation at $P = 8.26 \times 10^{-10}$ and located to placenta at $P = 3.5 \times 10^{-15}$, respectively. Among the genes of this network, there are several inflammation modulating proteins such as signal transducer and activator of transcription (STAT)-3, SMA- and MAD-related protein 3 (SMAD)-3, nuclear factor- κ B (NF- κ B), and c-Jun N-terminal kinases (JNKs). Results of these anal-

yses further support our hypotheses of these proteins are relevant to pathogenesis of preeclampsia.

Discussion

This study provides the most comprehensive information about multiple AF antiangiogenic protein concentrations in a cohort of 4795 women with complete obstetrical follow-up; among them 88 subsequently developed preeclampsia (Fig. 1). The large amniotic fluid bank, 7283 AF from 2002 to 2008, allowed us to set strict criteria for inclusion of preeclampsia cases, both with IUGR (n = 32)

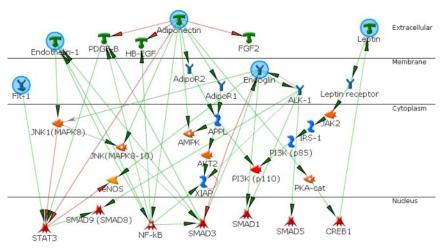


FIG. 4. Functional network analysis of five AF antiangiogenic proteins that are shown as *filled blue circles. Green lines* and *arrowheads* indicate stimulation, whereas *red lines* and *arrowheads* indicate inhibition.

and without IUGR (n = 39), and matched control subjects (n = 71). Unfortunately, we did not have maternal serum collected at the time of amniocentesis; hence, we could not compare protein concentrations between AF and serum. We like to point out that our results were derived from an Asia population. In Asia population, the incidence of pre-eclampsia was around 1.4-2% (41), which is lower than the worldwide incidence of 3-5% (1), and the Taiwanese fetal growth charts (32) are also different from those in Western populations (42).

In our institute, most cases who underwent amniocentesis were because of advanced maternal ages (age > 34 yr). Therefore, in this cohort, about half of the cases who later developed preeclampsia were multipara. Although our institute is a tertiary referral center, most referred cases with severe preeclampsia did not receive amniocentesis in our institute; thus, they were not enrolled in this study. Furthermore, after we excluded the cases who did not deliver in our hospital, our incidence of preeclampsia in the study cohort was 1.9% (88 of 4759), which was within the range of reported incidences in Taiwan (41, 43). So we consider that the excluded cases did not result in bias in the study population.

The main source of antiangiogenic proteins in AF during midtrimester amniocentesis is still unknown. The AF begins to fill in amnion since the 12th day after fertilization. At this stage, the likely mechanism of AF generation is active transport of solute by the amnion into the amniotic space (44). The fetus is not the major producer of AF until midtrimester (45), and fetal urine does not account for the major volume of AF until 20 wk of gestational age (46). Therefore, the AF proteins analyzed in this study are most likely to be derived from the placenta, not the fetus.

Results of this study indicate increased AF levels of leptin, sEndoglin, sFLT1, endothelin-1, and adiponectin were associated with subsequent development of preeclampsia, either with or without IUGR (Tables 2 and 3). Significantly positive correlations were also found among these proteins (Fig. 2), so a network analysis was performed according to the MetaCore database to elucidate common functions of these proteins. Using merely a couple of input genes, Analyze network algorithm of MetaCore was able to reveal additional information that was not present in the original list of root nodes (36, 47). Our network analyses revealed that adiponectin, endothelin-1, FLT1, and endoglin interacted with several inflammation modulating proteins, NF- κ B, SMAD3, JNKs, and STAT3, support-

ing a recent theory that inflammation in the placenta plays a central role in the pathogenesis of preeclampsia (4). In addition to receiving activating signals from adiponectin (38), NF- κ B can be activated by endothelin (48) and vice versa (49). SMAD3 up-regulates endothelin-1 (50) and endoglin-1 (51), but it receives negative feedback from endoglin-1 (52). JNKs may play modulating functions in several pathways: JNKs are activated by adiponectin (53), endoglin-1, and endothelin-1 (54), and JNKs can further activate SMAD3 (55) but inhibit STAT3 (56). STAT3 also moderates the effects from multiple pathways: it can be inhibited by both adiponectin (53) and JNKs (56) but activated by FLT1 (VEGFR-1) (57). Furthermore, this functional network also related endoglin-1 (58), endothelin-1, sFLT1, and adiponectin, all through STAT3 (59) to endothelial nitric oxide synthase, further supporting their roles in regulation of vascular tone and hypertension. Nevertheless, the networks derived from our study remain to be validated by in vitro studies.

Excessive placental leptin secretion (18, 20) and elevated AF concentrations of leptin (60) are associated with preeclampsia development. Leptin is secreted during acute inflammation (61). Inflammatory cytokines, such as TNF- α and IL-6, are significantly correlated with elevated leptin in women with preeclampsia (62). Results of our study further strengthen the importance of leptin in the pathogenesis of preeclampsia, which is also associated with inflammatory processes. Because the elevation of AF leptin was identified 2 months earlier than the appearance of symptoms, our results further suggest that AF leptin may be used as a predictive biomarker for preeclampsia.

Increased plasma sEndoglin concentrations are detected 2 months before the onset of preeclampsia (8). A reduced placental perfusion leading to hypoxia is thought to increase endoglin expression in IUGR pregnancies (63). The low placental oxygenation is believed to be responsible for the growth restriction seen in IUGR pregnancies (64, 65). In our study, the sEndoglin concentrations were significantly elevated in both the preeclampsia with and without IUGR groups, but there was no significant difference between preeclampsia with and without IUGR groups. Although Staff *et al.* (66) reported increased AF concentrations of sEndoglin in term preeclamptic pregnancies, our study is the first report of the association between the elevation of AF sEndoglin in early second trimester and subsequent development of preeclampsia (Tables 2 and 3).

Increased placental production and serum level of sFLT1 are thought to be crucial in the pathogenesis of preeclampsia (11, 67-69). Low oxygen conditions stimulate the secretion of sFLT1, and the increased sFLT1 levels exacerbate the oxidative stress that contributes to vascular dysfunction (70, 71). In addition to preeclampsia, previous studies also found an association between increased sFLT1 and fetal growth restriction (72-74). Levine et al. (8, 12) first demonstrated that maternal plasma concentrations of sEndoglin and sFLT1 increase 5–8 wk before preeclampsia occurrence, although Smith et al. (75) reported that high concentrations of plasma sFLT1 in 10-14 wk of gestation are not associated with the risk of preeclampsia. Park et al. (27) did not detect increased sFLT1 in amniotic fluid in preeclampsia women at the time of the midtrimester amniocentesis. Similarly, we did not detect a significant difference between the preeclampsia without IUGR and the control groups (Table 3). However, there was a significant increase in AF sFLT1 concentrations in women with preeclampsia complicated by IUGR, which, by definition, is severe preeclampsia (Table 3). Placental hypoxia and inflammatory stimulation are thought to be the triggers for sFLT1 release from preeclampsia placentas (4, 76). Results of this study further implicate placenta-derived sFLT1 as an important player in the pathogenesis of preeclampsia.

By analyzing the largest ever reported sample size of AF in this study (n = 142), we found significantly increased AF concentrations of endothelin-1 in the women who subsequently developed preeclampsia (Table 2). These results confirm the findings that increased AF concentrations of endothelin-1 in women who later developed preeclampsia in two previous reports, one with nine cases of preeclampsia (28) and another with 12 cases of IUGR (54). A clear trend is that the AF concentrations of endothelin-1 are highest in severe preeclampsia, followed by the mild preeclampsia, and then the control groups (Table 3). We speculate that the complex assay procedures of endothelin-1, especially the concentrating and reconstituting steps, may add variations to the final endothelin 1 concentrations, resulting in a lack of statistical significance on the difference among endothelin-1 concentrations in Table 3.

This study is the first report of increased AF adiponectin concentrations preceding the appearance of clinical symptoms of preeclampsia, although increased plasma adiponectin concentrations have been reported in preeclamptic women (23). Masuyama *et al.* (77) proposed that the role of adiponectin is to maintain endothelial function, and its deficiency leads to endothelial dysfunction or hypertension. Thus, an elevation of circulating adiponectin concentrations might represent a compensatory response to endothelial dysfunction. Significantly elevated concentrations of adiponectin in the preeclampsia with IUGR than the preeclampsia without IUGR groups (Table 3) further indicate that adiponectin levels may reflect the severity of preeclampsia.

In conclusion, the changes of amniotic antiangiogenic proteins, which occur before the onset of clinical symptoms, may shed new insight to our understanding of the pathophysiology of preeclampsia. Network analyses reveal the role of inflammation in the pathophysiology of preeclampsia, supporting such recent hypotheses that preeclampsia may not only result from a hypoxic placenta but may also be induced from a widespread inflammatory response followed by oxidative stress (3, 4). Among the antiangiogenic proteins, an increase in leptin and sEndoglin may occur in the early pathogenesis process of preeclampsia, but it may not be directly associated with its severity. Both sFLT1 and endothelin-1 may be involved in pathogenesis of preeclampsia and reflect its severity. However, increased levels of adiponectin appear to occur later in the pathophysiology of preeclampsia; thus, it is elevated only in severe preeclampsia.

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References

- 1. Roberts JM, Cooper DW 2001 Pathogenesis and genetics of preeclampsia. Lancet 357:53–56
- 2. Walker JJ 2000 Pre-eclampsia. Lancet 356:1260-1265
- Hung TH, Burton GJ 2006 Hypoxia and reoxygenation: a possible mechanism for placental oxidative stress in preeclampsia. Taiwan J Obstet Gynecol 45:189–200
- 4. Redman CW, Sargent IL 2009 Placental stress and pre-eclampsia: a revised view. Placenta 30(Suppl A):S38–S42
- Cheifetz S, Bellón T, Calés C, Vera S, Bernabeu C, Massagué J, Letarte M 1992 Endoglin is a component of the transforming growth factor-β receptor system in human endothelial cells. J Biol Chem 267:19027–19030
- St-Jacques S, Forte M, Lye SJ, Letarte M 1994 Localization of endoglin, a transforming growth factor-β binding protein, and of CD44 and integrins in placenta during the first trimester of pregnancy. Biol Reprod 51:405–413
- 7. Barbara NP, Wrana JL, Letarte M 1999 Endoglin is an accessory protein that interacts with the signaling receptor complex of multiple members of the transforming growth factor- β superfamily. J Biol Chem 274:584–594
- 8. Levine RJ, Lam C, Qian C, Yu KF, Maynard SE, Sachs BP, Sibai BM, Epstein FH, Romero R, Thadhani R, Karumanchi SA 2006 Soluble endoglin and other circulating antiangiogenic factors in preeclampsia. N Engl J Med 355:992–1005
- 9. Clark DE, Smith SK, Licence D, Evans AL, Charnock-Jones DS 1998 Comparison of expression patterns for placenta growth factor, vascular endothelial growth factor (VEGF), VEGF-B and VEGF-C in the human placenta throughout gestation. J Endocrinol 159:459–467
- Kendall RL, Thomas KA 1993 Inhibition of vascular endothelial cell growth factor activity by an endogenously encoded soluble receptor. Proc Natl Acad Sci USA 90:10705–10709
- Koga K, Osuga Y, Yoshino O, Hirota Y, Ruimeng X, Hirata T, Takeda S, Yano T, Tsutsumi O, Taketani Y 2003 Elevated serum soluble vascular endothelial growth factor receptor 1 (sVEGFR-1) levels in women with preeclampsia. J Clin Endocrinol Metab 88: 2348–2351
- Levine RJ, Maynard SE, Qian C, Lim KH, England LJ, Yu KF, Schisterman EF, Thadhani R, Sachs BP, Epstein FH, Sibai BM, Sukhatme VP, Karumanchi SA 2004 Circulating angiogenic factors and the risk of preeclampsia. N Engl J Med 350:672–683
- Venkatesha S, Toporsian M, Lam C, Hanai J, Mammoto T, Kim YM, Bdolah Y, Lim KH, Yuan HT, Libermann TA, Stillman IE, Roberts D, D'Amore PA, Epstein FH, Sellke FW, Romero R, Sukhatme VP, Letarte M, Karumanchi SA 2006 Soluble endoglin contributes to the pathogenesis of preeclampsia. Nat Med 12: 642–649
- Stepan H, Krämer T, Faber R 2007 Maternal plasma concentrations of soluble endoglin in pregnancies with intrauterine growth restriction. J Clin Endocrinol Metab 92:2831–2834
- 15. Taylor RN, Varma M, Teng NN, Roberts JM 1990 Women with preeclampsia have higher plasma endothelin levels than women with normal pregnancies. J Clin Endocrinol Metab 71:1675–1677
- 16. Fiore G, Florio P, Micheli L, Nencini C, Rossi M, Cerretani D, Ambrosini G, Giorgi G, Petraglia F 2005 Endothelin-1 triggers placental oxidative stress pathways: putative role in preeclampsia. J Clin Endocrinol Metab 90:4205–4210
- 17. Masuzaki H, Ogawa Y, Sagawa N, Hosoda K, Matsumoto T, Mise H, Nishimura H, Yoshimasa Y, Tanaka I, Mori T, Nakao K 1997

Nonadipose tissue production of leptin: leptin as a novel placentaderived hormone in humans. Nat Med 3:1029–1033

- Haugen F, Ranheim T, Harsem NK, Lips E, Staff AC, Drevon CA 2006 Increased plasma levels of adipokines in preeclampsia: relationship to placenta and adipose tissue gene expression. Am J Physiol Endocrinol Metab 290:E326–E333
- Grosfeld A, Andre J, Hauguel-De Mouzon S, Berra E, Pouyssegur J, Guerre-Millo M 2002 Hypoxia-inducible factor 1 transactivates the human leptin gene promoter. J Biol Chem 277:42953–42957
- 20. Mise H, Sagawa N, Matsumoto T, Yura S, Nanno H, Itoh H, Mori T, Masuzaki H, Hosoda K, Ogawa Y, Nakao K 1998 Augmented placental production of leptin in preeclampsia: possible involvement of placental hypoxia. J Clin Endocrinol Metab 83:3225–3229
- Sennello JA, Fayad R, Morris AM, Eckel RH, Asilmaz E, Montez J, Friedman JM, Dinarello CA, Fantuzzi G 2005 Regulation of T cellmediated hepatic inflammation by adiponectin and leptin. Endocrinology 146:2157–2164
- Ichida K, Moriyama T, Morita H, Kondo T, Yoshida S, Ohara N, Maruo T 2007 Plasma adiponectin concentrations and placental adiponectin expression in pre-eclamptic women. Gynecol Endocrinol 23:238–243
- Ramsay JE, Jamieson N, Greer IA, Sattar N 2003 Paradoxical elevation in adiponectin concentrations in women with preeclampsia. Hypertension 42:891–894
- 24. Hendler I, Blackwell SC, Mehta SH, Whitty JE, Russell E, Sorokin Y, Cotton DB 2005 The levels of leptin, adiponectin, and resistin in normal weight, overweight, and obese pregnant women with and without preeclampsia. Am J Obstet Gynecol 193:979–983
- 25. Suwaki N, Masuyama H, Nakatsukasa H, Masumoto A, Sumida Y, Takamoto N, Hiramatrsu Y 2006 Hypoadiponectinemia and circulating angiogenic factors in overweight patients complicated with pre-eclampsia. Am J Obstet Gynecol 195:1687–1692
- 26. Granger JP, LaMarca BB, Cockrell K, Sedeek M, Balzi C, Chandler D, Bennett W 2006 Reduced uterine perfusion pressure (RUPP) model for studying cardiovascular-renal dysfunction in response to placental ischemia. Methods Mol Med 122:383–392
- 27. Park CW, Park JS, Shim SS, Jun JK, Yoon BH, Romero R 2005 An elevated maternal plasma, but not amniotic fluid, soluble fms-like tyrosine kinase-1 (sFlt-1) at the time of mid-trimester genetic amniocentesis is a risk factor for preeclampsia. Am J Obstet Gynecol 193:984–989
- Margarit L, Griffiths A, Tsapanos V, Decavalas G, Gumenos D 2005 Second trimester amniotic fluid endothelin concentration. A possible predictor for pre-eclampsia. J Obstet Gynaecol 25:18–20
- Chan TF, Su JH, Chung YF, Hsu YH, Ych YT, Jong SB, Yuan SS 2003 Amniotic fluid and maternal serum leptin levels in pregnant women who subsequently develop preeclampsia. Eur J Obstet Gynecol Reprod Biol 108:50–53
- Cunningham FG, Grant NF, Leveno KJ, Gilstrap LC, Hauth JC, Wenstrom KD 2001 Hypertensive disorders in pregnancy. In: Williams obstetrics. 21st ed. New York: McGraw-Hill; 567–618
- 31. Sibai B, Dekker G, Kupferminc M 2005 Pre-eclampsia. Lancet 365: 785–799
- Hsieh WS, Wu HC, Jeng SF, Liao HF, Su YN, Lin SJ, Hsieh CJ, Chen PC 2006 Nationwide singleton birth weight percentiles by gestational age in Taiwan, 1998–2002. Acta Paediatr Taiwan 47:25–33
- 33. D'Anna R, Baviera G, Cannata ML, De Vivo A, Di Benedetto A, Corrado F 2007 Midtrimester amniotic fluid leptin and insulin levels and subsequent gestational diabetes. Gynecol Obstet Invest 64:65–68
- 34. Saftlas A, Wang W, Risch H, Woolson R, Hsu C, Bracken M 2000 Prepregnancy body mass index and gestational weight gain as risk factors for preeclampsia and transient hypertension. Ann Epidemiol 10:475
- 35. Wang TH, Chang CL, Wu HM, Chiu YM, Chen CK, Wang HS 2006 Insulin-like growth factor-II (IGF-II), IGF-binding protein-3 (IGFBP-3), and IGFBP-4 in follicular fluid are associated with oocyte maturation and embryo development. Fertil Steril 86:1392–1401

- 36. Wang TH, Chao AS, Chen JK, Chao A, Chang YL, Cheng PJ, Chang SD, Wang HS 2009 Network analyses of differentially expressed proteins in amniotic fluid supernatant associated with abnormal human karyotypes. Fertil Steril 92:96–107
- 37. Chao A, Wang TH, Lee YS, Hong JH, Tsai CN, Chen CK, Tsai CS, Chao AS, Lai CH 2008 Analysis of functional groups of differentially expressed genes in the peripheral blood of patients with cervical cancer undergoing concurrent chemoradiation treatment. Radiat Res 169:76–86
- 38. Tsao TS, Murrey HE, Hug C, Lee DH, Lodish HF 2002 Oligomerization state-dependent activation of NF-κB signaling pathway by adipocyte complement-related protein of 30 kDa (Acrp30). J Biol Chem 277:29359–29362
- 39. Nikolsky Y, Ekins S, Nikolskaya T, Bugrim A 2005 A novel method for generation of signature networks as biomarkers from complex high throughput data. Toxicol Lett 158:20–29
- Mason CW, Swaan PW, Weiner CP 2006 Identification of interactive gene networks: a novel approach in gene array profiling of myometrial events during guinea pig pregnancy. Am J Obstet Gynecol 194:1513–1523
- Lee CJ, Hsich TT, Chiu TH, Chen KC, Lo LM, Hung TH 2000 Risk factors for pre-eclampsia in an Asian population. Int J Gynaecol Obstet 70:327–333
- 42. Alexander GR, Himes JH, Kaufman RB, Mor J, Kogan M 1996 A United States national reference for fetal growth. Obstet Gynecol 87:163–168
- 43. Chen CL, Cheng Y, Wang PH, Juang CM, Chiu LM, Yang MJ, Hung CS, Yang ML 2000 Review of pre-eclampsia in Taiwan: a multiinstitutional study. Zhonghua Yi Xue Za Zhi (Taipei) 63:869–875
- 44. Modena AB, Fieni S 2004 Amniotic fluid dynamics. Acta Biomed 75(Suppl 1):11–13
- 45. Cleveland MG, Bakos MA, Pyron DL, Rajaraman S, Goldblum RM 1991 Characterization of secretory component in amniotic fluid. Identification of new forms of secretory IgA. J Immunol 147:181–188
- 46. Touboul C, Boulvain M, Picone O, Levaillant JM, Frydman R, Senat MV 2008 Normal fetal urine production rate estimated with 3-dimensional ultrasonography using the rotational technique (virtual organ computer-aided analysis). Am J Obstet Gynecol 199:57.e51– 57.e55
- 47. Ekins S, Bugrim A, Brovold L, Kirillov E, Nikolsky Y, Rakhmatulin E, Sorokina S, Ryabov A, Serebryiskaya T, Melnikov A, Metz J, Nikolskaya T 2006 Algorithms for network analysis in systems-ADME/Tox using the MetaCore and MetaDrug platforms. Xenobiotica 36:877–901
- Browatzki M, Schmidt J, Kübler W, Kranzhöfer R 2000 Endothelin-1 induces interleukin-6 release via activation of the transcription factor NF-κB in human vascular smooth muscle cells. Basic Res Cardiol 95:98–105
- 49. Woods M, Wood EG, Bardswell SC, Bishop-Bailey D, Barker S, Wort SJ, Mitchell JA, Warner TD 2003 Role for nuclear factor-κB and signal transducer and activator of transcription 1/interferon regulatory factor-1 in cytokine-induced endothelin-1 release in human vascular smooth muscle cells. Mol Pharmacol 64:923–931
- 50. Rodriguez-Pascual F, Reimunde FM, Redondo-Horcajo M, Lamas S 2004 Transforming growth factor- β induces endothelin-1 expression through activation of the Smad signaling pathway. J Cardiovasc Pharmacol 44(Suppl 1):S39–S42
- 51. Sánchez-Elsner T, Botella LM, Velasco B, Langa C, Bernabéu C 2002 Endoglin expression is regulated by transcriptional cooperation between the hypoxia and transforming growth factor-β pathways. J Biol Chem 277:43799–43808
- Bernabeu C, Conley BA, Vary CP 2007 Novel biochemical pathways of endoglin in vascular cell physiology. J Cell Biochem 102: 1375–1388
- 53. Miyazaki T, Bub JD, Uzuki M, Iwamoto Y 2005 Adiponectin activates c-Jun NH2-terminal kinase and inhibits signal transducer

and activator of transcription 3. Biochem Biophys Res Commun $333{:}79{-}87$

- 54. Guo B, Slevin M, Li C, Parameshwar S, Liu D, Kumar P, Bernabeu C, Kumar S 2004 CD105 inhibits transforming growth factor-β-Smad3 signalling. Anticancer Res 24:1337–1345
- 55. Yamagata H, Matsuzaki K, Mori S, Yoshida K, Tahashi Y, Furukawa F, Sekimoto G, Watanabe T, Uemura Y, Sakaida N, Yoshioka K, Kamiyama Y, Seki T, Okazaki K 2005 Acceleration of Smad2 and Smad3 phosphorylation via c-Jun NH(2)-terminal kinase during human colorectal carcinogenesis. Cancer Res 65: 157–165
- Bogoyevitch MA, Kobe B 2006 Uses for JNK: the many and varied substrates of the c-Jun N-terminal kinases. Microbiol Mol Biol Rev 70:1061–1095
- 57. Lee YK, Shanafelt TD, Bone ND, Strege AK, Jelinek DF, Kay NE 2005 VEGF receptors on chronic lymphocytic leukemia (CLL) B cells interact with STAT 1 and 3: implication for apoptosis resistance. Leukemia 19:513–523
- 58. Toporsian M, Gros R, Kabir MG, Vera S, Govindaraju K, Eidelman DH, Husain M, Letarte M 2005 A role for endoglin in coupling eNOS activity and regulating vascular tone revealed in hereditary hemorrhagic telangiectasia. Circ Res 96:684–692
- 59. Saura M, Zaragoza C, Bao C, Herranz B, Rodriguez-Puyol M, Lowenstein CJ 2006 Stat3 mediates interleukin-6 inhibition of human endothelial nitric-oxide synthase expression. J Biol Chem 281:30057– 30062
- 60. Chan TF, Su JH, Chung YF, Hsu YH, Yeh YT, Yuan SS 2006 Elevated amniotic fluid leptin levels in pregnant women who are destined to develop preeclampsia. Acta Obstet Gynecol Scand 85: 171–174
- 61. Matarese G, Moschos S, Mantzoros CS 2005 Leptin in immunology. J Immunol 174:3137–3142
- 62. Bartha JL, Romero-Carmona R, Escobar-Llompart M, Comino-Delgado R 2001 The relationships between leptin and inflammatory cytokines in women with pre-eclampsia. BJOG 108:1272–1276
- 63. Yinon Y, Nevo O, Xu J, Many A, Rolfo A, Todros T, Post M, Caniggia I 2008 Severe intrauterine growth restriction pregnancies have increased placental endoglin levels: hypoxic regulation via transforming growth factor-β3. Am J Pathol 172:77–85
- 64. McCarthy C, Cotter FE, McElwaine S, Twomey A, Mooney EE, Ryan F, Vaughan J 2007 Altered gene expression patterns in intrauterine growth restriction: potential role of hypoxia. Am J Obstet Gynecol 196:70.e71–70.e76
- 65. Roh CR, Budhraja V, Kim HS, Nelson DM, Sadovsky Y 2005 Microarray-based identification of differentially expressed genes in hypoxic term human trophoblasts and in placental villi of pregnancies with growth restricted fetuses. Placenta 26:319–328
- 66. Staff AC, Braekke K, Johnsen GM, Karumanchi SA, Harsem NK 2007 Circulating concentrations of soluble endoglin (CD105) in fetal and maternal serum and in amniotic fluid in preeclampsia. Am J Obstet Gynecol 197:176.e171–176.e176
- Lam C, Lim KH, Karumanchi SA 2005 Circulating angiogenic factors in the pathogenesis and prediction of preeclampsia. Hypertension 46:1077–1085
- 68. Maynard SE, Min JY, Merchan J, Lim KH, Li J, Mondal S, Libermann TA, Morgan JP, Sellke FW, Stillman IE, Epstein FH, Sukhatme VP, Karumanchi SA 2003 Excess placental soluble fmslike tyrosine kinase 1 (sFlt1) may contribute to endothelial dysfunction, hypertension, and proteinuria in preeclampsia. J Clin Invest 111:649–658
- 69. Shibata E, Rajakumar A, Powers RW, Larkin RW, Gilmour C, Bodnar LM, Crombleholme WR, Ness RB, Roberts JM, Hubel CA 2005 Soluble fms-like tyrosine kinase 1 is increased in preeclampsia but not in normotensive pregnancies with small-for-gestational-age neonates: relationship to circulating placental growth factor. J Clin Endocrinol Metab 90:4895–4903
- Bridges JP, Gilbert JS, Colson D, Gilbert SA, Dukes MP, Ryan MJ, Granger JP 2009 Oxidative stress contributes to soluble fms-like

tyrosine kinase-1 induced vascular dysfunction in pregnant rats. Am J Hypertens 22:564–568

- 71. Gu Y, Lewis DF, Wang Y 2008 Placental productions and expressions of soluble endoglin, soluble fms-like tyrosine kinase receptor-1, and placental growth factor in normal and preeclamptic pregnancies. J Clin Endocrinol Metab 93:260–266
- 72. Stepan H, Geide A, Faber R 2004 Soluble fms-like tyrosine kinase 1. N Engl J Med 351:2241–2242
- 73. Tsatsaris V, Goffin F, Munaut C, Brichant JF, Pignon MR, Noel A, Schaaps JP, Cabrol D, Frankenne F, Foidart JM 2003 Overexpression of the soluble vascular endothelial growth factor receptor in preeclamptic patients: pathophysiological consequences. J Clin Endocrinol Metab 88:5555–5563
- 74. Wallner W, Sengenberger R, Strick R, Strissel PL, Meurer B, Beckmann MW, Schlembach D 2007 Angiogenic growth factors in

maternal and fetal serum in pregnancies complicated by intrauterine growth restriction. Clin Sci (Lond) 112:51–57

- 75. Smith GC, Crossley JA, Aitken DA, Jenkins N, Lyall F, Cameron AD, Connor JM, Dobbie R 2007 Circulating angiogenic factors in early pregnancy and the risk of preeclampsia, intrauterine growth restriction, spontaneous preterm birth, and stillbirth. Obstet Gynecol 109:1316–1324
- 76. Nevo O, Soleymanlou N, Wu Y, Xu J, Kingdom J, Many A, Zamudio S, Caniggia I 2006 Increased expression of sFlt-1 in *in vivo* and *in vitro* models of human placental hypoxia is mediated by HIF-1. Am J Physiol Regul Integr Comp Physiol 291:R1085–R1093
- 77. Masuyama H, Nakatsukasa H, Takamoto N, Hiramatsu Y 2007 Correlation between soluble endoglin, vascular endothelial growth factor receptor-1 and adipocytokines in preeclampsia. J Clin Endocrinol Metab 92:2672–2679