

Potential role of proliferation signal inhibitors on atherosclerosis in renal transplant patients

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Abstract

Over the last decade, there has been a decrease in acute graft rejection rates following renal transplantation; however, this has not corresponded with an improvement in long-term outcomes of transplantation. One of the major causes of long-term morbidity and mortality in renal transplant recipients is cardiovascular disease. Immunosuppressive regimens, especially those including steroids and calcineurin inhibitors, have a negative role in the induction of cardiovascular risk factors. The proliferation signal inhibitors (PSIs)/mammalian target of rapamycin (mTOR) inhibitors sirolimus and everolimus have shown considerable promise in reducing acute rejection in renal transplant recipients. Although PSIs are associated with an increase in hyperlipidaemia (hypercholesterolaemia and hypertriglyceridaemia), which is a major risk factor for atherosclerosis and associated cardiovascular disease, recent studies with sirolimus have demonstrated protection from atheroma progression in hyperlipidaemic apolipoprotein E-deficient mice. Here, we summarize the results of pre-clinical and clinical studies with sirolimus and everolimus, with particular emphasis on the beneficial and adverse effects that these drugs exert on the cardiovascular system, and the underlying molecular mechanisms.

Keywords: atherosclerosis; cardioprotective; dyslipidaemia; everolimus; mammalian target of rapamycin inhibitors; proliferation signal inhibitors; renal transplant; sirolimus

Introduction

Cardiovascular disease is currently one of the leading causes of death in renal transplant recipients. Renal transplantation is associated with an increased risk of cardiovascular disease, with patients commonly experiencing cardiovascular complications such as diabetes mellitus, hypertension and hyperlipidaemia. Furthermore, renal insufficiency and dialysis treatment is associated with the progression of advanced atherosclerotic disease [1]. Whereas some immunosuppressive regimens, such as those including steroids or calcineurin inhibitors, may exacerbate the risk of cardiovascular disease, others may have a cardioprotective role.

The proliferation signal inhibitors (PSIs)/mTOR inhibitors everolimus (Certican[®]; Novartis Pharma AG, Basel, Switzerland) and sirolimus (Rapamune[®]; Wyeth Pharmaceuticals, USA) are becoming increasingly used following renal and heart transplantation as a result of their immunosuppressive and anti-proliferative actions. Sirolimus is a macrolide antibiotic produced by *Streptomyces hygroscopicus* [2], which displays potent immunosuppressive, anti-proliferative and anti-migratory properties [3]. Upon binding to the cytosolic immunophilin FK506-binding protein, sirolimus inhibits the FRAP {FKBP-12 rapamycin-associated protein [also known as mTOR]} signalling pathway, thus promoting dephosphorylation and inactivation of p70 ribosomal protein S6 kinase and eukaryotic translation initiation factor 4E-binding protein, accumulation of the growth suppressor p27^{Kip1}, inhibition of cyclin-dependent kinase activity, reduced retinoblastoma protein phosphorylation and inhibition of minichromosome maintenance protein expression [4]. Everolimus also inhibits the phosphatidylinositol-3-kinase signalling pathway, blocking growth factor-mediated interleukin (IL)-2- and IL-15-driven proliferation of T-cells, B-cells and vascular smooth muscle (Figure 1) [5].

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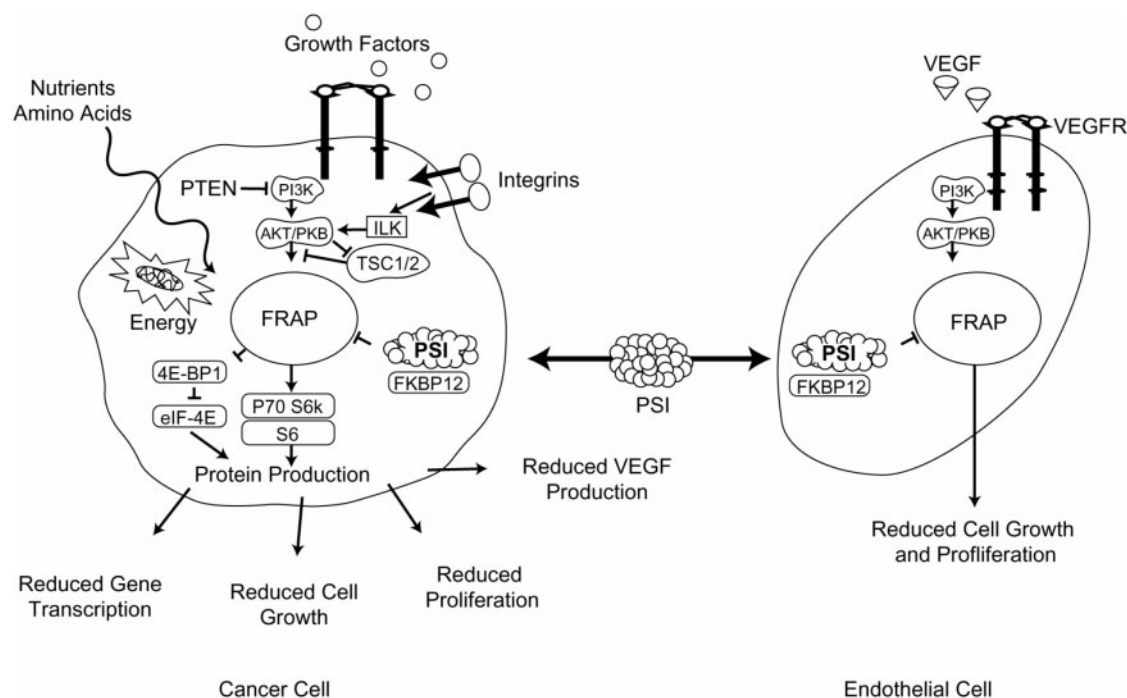


Fig. 1. Mechanism of action of proliferation signal inhibitors. FKBP, FK506-binding protein; FRAP, FKBP-12 rapamycin-associated protein [mTOR]; ILK, integrin-linked kinase; p70 S6k, p70 ribosomal protein S6 kinase; PI3K, phosphatidylinositol 3-phosphate; PKB, protein kinase B; PSI, proliferation signal inhibitor; PTEN, phosphatase and tensin homologue deleted from chromosome 10; TSC, tuberous sclerosis complex; VEGF, vascular endothelial growth factor; VEGFR, vascular endothelial growth factor receptor.

Although sirolimus and everolimus have similar chemical structures, there are key differences in their pharmacokinetic and pharmacodynamic profiles, and in their tolerability. Everolimus has a shorter half-life than sirolimus (28 h vs 62 h, respectively), and the time to reach steady-state is shorter for everolimus (4 days vs 6 days, respectively) [6].

Clinical trials

Clinical trials have demonstrated the efficacy of sirolimus and everolimus therapy in reducing acute graft rejection in kidney [7,8], heart [9,10], liver [11,12] and lung [13,14] transplant recipients. Sirolimus and everolimus also attenuate neointimal thickening and transplant atherosclerosis in animal models of angioplasty, and vessel and cardiac allografts [3,15]. Moreover, clinical trials have demonstrated a significant reduction in binary restenosis, late lumen loss and repeat revascularization rates in selected patient groups receiving sirolimus-eluting stents compared with standard coronary stents [3,16]. Preliminary data from the First Use To Underscore REDuction in restenosis with everolimus I (FUTURE I) trial suggest that these effects are PSI class specific, with lower rates of in-stent late lumen loss in everolimus-eluting stents compared with metallic stents [17]. Up-regulation of p27^{Kip1}, which can reduce both vascular smooth

muscle cell proliferation and migration, might mediate, at least in part, the anti-proliferative and anti-migratory actions of sirolimus, thus contributing to reduced neointimal thickening [3,18].

Adverse events in patients receiving sirolimus-eluting stents include increased risk of late stent thrombosis after cessation of anti-platelet therapy [19,20] and exercise-induced paradoxical coronary vasoconstriction of the adjacent vessel segments (although the vasodilatory response to nitroglycerin is unaffected) [21]. These may be due to drug-induced impaired intimal healing and endothelial dysfunction, respectively. In renal transplant recipients, sirolimus induces dose-dependent hyperlipidaemia [e.g. hypertriglyceridaemia, increased low-density lipoprotein (LDL)-cholesterol and increased apolipoprotein (apo) B-100 and apoC-III circulating levels], which can be reversed after discontinuation of treatment and is well-managed with conventional statin therapy [22–24]. A similar increase in serum cholesterol and triglyceride levels has also been reported in renal transplant recipients receiving everolimus therapy [25]. Suggested mechanisms underlying PSI-induced dyslipidaemia in humans are not clear, and may include increased hepatic secretion of triglycerides in conjunction with cholesterol and apoproteins in very-low-density lipoprotein (VLDL)-cholesterol particles and hypertriglyceridaemia. These may occur as a result of augmented adipose tissue lipase activity and/or

decreased lipoprotein lipase activity in response to altered insulin-mediated signalling [26,27], and it has been proposed that sirolimus indirectly up-regulates expression of the gene *apo CIII*, an important inhibitor of lipoprotein lipase [28]. Furthermore, up-regulation of adipocyte fatty acid-binding protein (aP2) expression in monocytes and macrophages may play a role in the increased accumulation of triglycerides [29].

Recent animal studies

Although the above findings suggest that PSI therapy may promote atherosclerosis, since cholesterol commonly forms part of atherosclerotic plaques in coronary artery disease, recent studies using the well-characterized hyperlipidaemic apoE-null mouse model of atherosclerosis [30] have demonstrated diminished atheroma size following sirolimus administration, either subcutaneously [4,31] or orally [32,33]. Notably, sirolimus did not aggravate hypercholesterolaemia in fat-fed apoE-null mice [4,31–33]. Castro *et al.* found no significant effect of sirolimus (1–4 mg/kg, s.c., q.i.d.) on circulating LDL-cholesterol, high-density lipoprotein (HDL)-cholesterol, VLDL-cholesterol and intermediate-density lipoprotein-cholesterol [4]. By contrast, in another study, Elloso *et al.* q.i.d. reported increased levels of LDL-cholesterol and HDL-cholesterol in the sirolimus-treated group (1–8 mg/kg, s.c., q.i.d.) [31]. Likewise, the effect of sirolimus on circulating triglyceride levels in fat-fed apoE-null mice is controversial, with these levels being unchanged upon oral administration of sirolimus but increased upon intraperitoneal sirolimus administration (3 mg/kg/day) [34,35]. Regardless of these discrepancies, which might be related to differences in high-fat diet composition or doses and route of drug administration, it can be concluded that sirolimus protects from atheroma development in mouse models, even in the presence of severe dyslipidaemia. This may be a class effect of PSIs, and further studies are required to elucidate any beneficial effects of everolimus on the development of atherosclerosis.

Mechanisms of action

Several mechanisms may contribute to sirolimus-dependent atheroprotection regulating both cholesterol homeostasis and inflammatory responses. Firstly, there may be reduced accumulation of cholesterol within the artery wall [36], possibly owing to reduced expression of lipoprotein receptors (LDL receptors, VLDL receptors and CD36) and increased expression of genes involved in cholesterol efflux, including the genes encoding peroxisome proliferator-activated receptor- α , liver X receptor- α and ATP-binding cassette A1 [37]. This results in increased cholesterol efflux from cells, leading to lower intracellular

cholesterol levels. Furthermore, there may also be alterations in endothelial nitric oxide synthase expression, changes in the ratio of matrix metalloproteinases (MMP) to tissue inhibitors of metalloproteinases and MMP-2 and MMP-9 activation [34,35], reduced expression of the positive cell cycle regulator cyclin-dependent kinase-2 and proliferating cell nuclear antigen [4] and pro-inflammatory cytokines [31], and diminished expression of arterial monocyte chemotactic protein-1 [4], which is associated with the attenuation of monocyte chemotaxis and reduced neointimal macrophage counts in sirolimus-fed mice [32]. A study with everolimus has demonstrated that everolimus can cause disruption of cellular lipid homeostasis in mouse peritoneal macrophages at concentrations similar to therapeutic range (0.01 μ M), resulting in increased cholesterol esterification and efflux and reduced triglyceride biosynthesis [38].

Conclusions

Whether the beneficial actions of PSIs/mTORs observed in severely dyslipidaemic mice also operate in renal transplant recipients, and whether they might counteract PSI-induced hyperlipidaemia is, as yet, unknown. Certainly, prospective long-term trials are needed to clarify the impact of sirolimus and everolimus on lipid and lipoprotein patterns and on morbidity and mortality attributable to cardiovascular disease. In short, the intrinsic effect on atherosclerosis prevention and enhanced control of renal function could explain the reduction of cardiovascular morbidity and mortality in PSI-treated patients.

Acknowledgements. Editorial assistance was provided by Ogilvy 4D. Work in the laboratory of V. Andrés is supported in part by grants from the Instituto de Salud Carlos III (Red de Centros RECAVA, C03/01) and from the Spanish Ministry of Education and Science, and the European Regional Development Fund (SAF2004-03057). J.M.C. is partially supported by grants from the Instituto Carlos III (Red de Trasplantes, C05/01), and C.C. is supported by a fellowship from Conicet (Argentina).

Conflict of interest statement. The author received an honorarium from Novartis Pharma AG for speaking at a meeting in Geneva, October 2005, on the role of everolimus in cardiovascular risk management in renal transplantation.

References

1. Parfrey PA, Foley RN. The clinical epidemiology of cardiac disease in chronic renal failure. *J Am Soc Nephrol* 1999; 10: 1606–1615
2. Sehgal SN, Baker H, Vezina C. Rapamycin (AY-22,989), a new antifungal antibiotic. II. Fermentation, isolation and characterization. *J Antibiot* 1975; 28: 727–732
3. Marx SO, Marks AR. Bench to bedside: the development of rapamycin and its application to stent restenosis. *Circulation* 2001; 104: 852–855
4. Castro C, Campistol JM, Sancho D, Sánchez-Madrid F, Casals E, Andrés V. Rapamycin attenuates atherosclerosis induced by dietary cholesterol in apolipoprotein-deficient mice

- through a p27 Kip1-independent pathway. *Atherosclerosis* 2004; 172: 31–38
5. Schuler W, Sedrani R, Cottens S *et al.* SDZ RAD, a new rapamycin derivative: pharmacological properties *in vitro* and *in vivo*. *Transplantation* 1997; 64: 36–42
 6. Neumayer H-H. Introducing everolimus (Certican®) in organ transplantation: an overview of preclinical and early clinical developments. *Transplantation* 2005; 79 [Suppl]: S72–S75
 7. Vitko S, Tedesco H, Eris J *et al.* Everolimus with optimized cyclosporine dosing in renal transplant recipients: 6-month safety and efficacy results of two randomized studies. *Am J Transplant* 2004; 4: 626–635
 8. Kahan BD, Podbielski J, Napoli KL, Katz SM, Meier-Kriesche HU, Van Buren CT. Immunosuppressive effects and safety of a sirolimus/cyclosporine combination regimen for renal transplantation. *Transplantation* 1998; 66: 1040–1046
 9. Eisen HJ, Tuzcu EM, Dorent R *et al.* Everolimus for the prevention of allograft rejection and vasculopathy in cardiac-transplant recipients. *N Engl J Med* 2003; 349: 847–858
 10. Keogh A, Richardson M, Ruygrok P *et al.* Sirolimus in de novo heart transplant recipients reduces acute rejection and prevents coronary artery disease at 2 years – a randomized clinical trial. *Circulation* 2004; 110: 2694–2700
 11. Levy G, Moeller V, Jaffe J *et al.* Safety, tolerability and efficacy of everolimus in de novo liver transplant recipients: 12 and 36 month results. *Liver Transplant* 2006; doi:10.1002/lt.20707
 12. Trotter JF, Wachs M, Bak T *et al.* Liver transplantation using sirolimus and minimal corticosteroids (3-day taper). *Liver Transpl* 2001; 7: 343–351
 13. Snell GI, Valentine VG, Vitulo P *et al.* Everolimus versus azathioprine in maintenance lung transplant recipients: an international, randomized, double-blind clinical trial. *Am J Transplant* 2006; 6: 169–177
 14. Shitrit D, Rahamimov R, Gidon S *et al.* Use of sirolimus and low-dose calcineurin inhibitor in lung transplant recipients with renal impairment: results of a controlled pilot study. *Kidney Int* 2005; 67: 1471–1475
 15. Viklicky O, Zou H, Muller V, Lacha J, Szabo A, Heemann U. SDZ-RAD prevents manifestation of chronic rejection in rat renal allografts. *Transplantation* 2000; 69: 497–502
 16. Morice MC, Serruys PW, Sousa JE *et al.* A randomized comparison of a sirolimus-eluting stent with a standard stent for coronary revascularization. *N Engl J Med* 2002; 346: 1773–1780
 17. Costa RA, Lansky AJ, Mintz GS *et al.* Angiographic results of the first human experience with everolimus-eluting stents for the treatment of coronary lesions (the FUTURE I trial). *Am J Cardiol* 2005; 95: 113–116
 18. Diez-Juan A, Andrés V. Coordinate control of proliferation and migration by the p27Kip1/cyclin-dependent kinase/retinoblastoma pathway in vascular smooth muscle cells and fibroblasts. *Circ Res* 2003; 92: 402–410
 19. Ong AT, McFadden EP, Regar E, de Jaegere PP, van Domburg RT, Serruys PW. Late angiographic stent thrombosis (LAST) events with drug-eluting stents. *J Am Coll Cardiol* 2005; 45: 2088–2092
 20. Nebeker JR, Virmani R, Bennett CL *et al.* Hypersensitivity cases associated with drug-eluting coronary stents: a review of available cases from the Research on Adverse Drug Events and Reports (RADAR) project. *J Am Coll Cardiol* 2006; 47: 175–181
 21. Togni M, Windecker S, Cocchia R *et al.* Sirolimus-eluting stents associated with paradoxical coronary vasoconstriction. *J Am Coll Cardiol* 2005; 46: 231–236
 22. Boots JM, Christiaans MH, van Hooff JP. Effect of immunosuppressive agents on long-term survival of renal transplant recipients: focus on the cardiovascular risk. *Drugs* 2004; 64: 2047–2073
 23. Kahan BD. Efficacy of sirolimus compared with azathioprine for reduction of acute renal allograft rejection: a randomized multicentre study. The Rapamune US Study Group. *Lancet* 2000; 356: 194–202
 24. MacDonald AS. RAPAMUNE Global Study Group. A worldwide, phase III, randomized, controlled, safety and efficacy study of a sirolimus/cyclosporine regimen for prevention of acute rejection in recipients of primary mismatched renal allografts. *Transplantation* 2001; 71: 271–280
 25. Kramer BK, Stahl R *et al.* Graft function, cardiovascular risk factors, and sex hormones in renal transplant recipients on an immunosuppressive regimen of everolimus, reduced dose of cyclosporine, and basiliximab. *Transplant Proc* 2005; 37: 1601–1604
 26. Morrisett JD, Abdel-Fattah G, Hoogeveen R *et al.* Effects of sirolimus on plasma lipids, lipoprotein levels, and fatty acid metabolism in renal transplant patients. *J Lipid Res* 2002; 43: 1170–1180
 27. Hoogeveen RC, Ballantyne CM, Pownall HJ *et al.* Effect of sirolimus on the metabolism of apoB100-containing lipoproteins in renal transplant patients. *Transplantation* 2001; 72: 1244–1250
 28. Tur MD, Garrigue V, Vela C *et al.* Apolipoprotein CIII is upregulated by anticalcineurins and rapamycin: implications in transplantation-induced dyslipidemia. *Transplant Proc* 2000; 32: 2783–2784
 29. Liu Q-Y, Nambi P. Sirolimus upregulates aP2 expression in human monocytes and macrophages. *Transplant Proc* 2004; 36: 3229–3231
 30. Meir KS, Leitersdorf E. Atherosclerosis in the apolipoprotein-E-deficient mouse: a decade of progress. *Arterioscler Thromb Vasc Biol* 2004; 24: 1006–1014
 31. Elloso MM, Azrolan N, Sehgal SN *et al.* Protective effect of the immunosuppressant sirolimus against aortic atherosclerosis in apo E-deficient mice. *Am J Transplant* 2003; 3: 562–569
 32. Pakala R, Stabile E, Jang GJ, Clavijo L, Waksman R. Rapamycin attenuates atherosclerotic plaque progression in apolipoprotein E knockout mice: inhibitory effect on monocyte chemotaxis. *J Cardiovasc Pharmacol* 2005; 46: 481–486
 33. Waksman R, Pakala R, Burnett MS *et al.* Oral rapamycin inhibits growth of atherosclerotic plaque in apoE knock-out mice. *Cardiovasc Radiat Med* 2003; 4: 34–38
 34. Naoum JJ, Woodside KJ, Zhang S, Rychahou PG, Hunter GC. Effects of rapamycin on the arterial inflammatory response in atherosclerotic plaques in Apo-E knockout mice. *Transplant Proc* 2005; 37: 1880–1884
 35. Naoum JJ, Zhang S, Woodside KJ *et al.* Aortic eNOS expression and phosphorylation in Apo-E knockout mice: differing effects of rapamycin and simvastatin. *Surgery* 2004; 136: 323–328
 36. Basso MD, Nambi P, Adelman SJ. Effect of sirolimus on the cholesterol content of aortic arch in ApoE knockout mice. *Transplant Proc* 2003; 35: 3136–3138
 37. Varghese Z, Fernando R, Moorhead JF, Powis SH, Ruan XZ. Effects of sirolimus on mesangial cell cholesterol homeostasis: a novel mechanism for its action against lipid-mediated injury in renal allografts. *Am J Physiol Renal Physiol* 2005; 289: F43–F48
 38. Bellosta S, Arnaboldi L, Canavesi P *et al.* Everolimus affects cholesterol homeostasis in macrophages. 7th Annual Conference on Arteriosclerosis, Thrombosis and Vascular Biology. Denver, Co, USA, 27–29 April 2006.