

Dendritic cells in progressive renal disease: some answers, many questions

A. Richard Kitching^{1,2,3}

¹Department of Medicine, Centre for Inflammatory Diseases, Monash University, Clayton, VIC, Australia, ²Department of Nephrology, Monash Health, Clayton, VIC, Australia and ³Department of Paediatric Nephrology, Monash Health, Clayton, VIC, Australia

Correspondence and offprint requests to: A.R. Kitching; E-mail: Richard.kitching@monash.edu.au

ABSTRACT

Renal disease results from a variety of insults, but whatever its genesis, ongoing inflammation will drive progressive fibrotic disease. Dendritic cells link innate and adaptive immunity by presenting antigens, but they act also in an antigen-independent manner. While systemic dendritic cells (DCs) establish nephritogenic adaptive immunity, DCs are also present in the kidney. The tubulointerstitium is endowed with a network of mononuclear phagocytes, many having dendritic cell characteristics. While the roles of renal DCs are complex, recent evidence demonstrates that in adaptive immune responses affecting the kidney, DCs in the cortical interstitium express the chemokine receptor CX3CR1, are CX3CR1 dependent and are important in ongoing antigen recognition by effector CD4⁺ T cells, leading to progressive disease. Medullary DCs do not share this potent antigen-presenting function and CX3CR1 dependence. Though macrophages have a pathogenic role in antigen-independent renal fibrosis, whether interstitial DCs have any role is not clear. The participation of local and systemic DCs in progressive renal disease varies according to their involvement as antigen-presenting or local innate cells, the nature of the pathogenic process, and the involvement of the glomerulus, the cortical tubulointerstitium and the medulla in disease.

Keywords: chemokine, dendritic cells, glomerulonephritis, macrophage, renal fibrosis

INTRODUCTION

Renal disease is caused by a diverse range of insults, the kidney being affected by multiple immune, inflammatory and metabolic insults, but in progressive disease leading to end-stage kidney disease, interstitial fibrosis and glomerulosclerosis is almost universal. In many cases, fibrotic injury results from ongoing inflammation, with frustrated attempts at healing resulting in dysregulated and persistent matrix deposition. Thus, sclerosis

and atrophy represent failed attempts at resolution, thwarted by the presence of persisting inflammatory and metabolic derangement. The genesis of this inflammation is not hard to conceptualize in antigen-driven forms of renal disease. The kidney can be targeted by virtue of it expressing autoantigens, by antigens being lodged in the kidneys or by antibody-induced disease leading to complement and leukocyte-mediated injury. However, even in diseases that in the past have been viewed as haemodynamic or metabolic in origin, inflammation plays a role in progressive injury. In the kidney, cells central to this process are mononuclear phagocytes, consisting of monocyte/macrophages and dendritic cells (DCs).

DCs are present in most tissues and lymphoid organs in the body, including the kidney. In health, they form a sentinel network to sense and report infectious threats, both to activate adaptive immunity and to allow antigen-specific effectors to localize to sites of inflammation. Immature 'unlicensed' DCs help maintain tolerance by anergizing autoreactive T cells. While DCs have the capacity to secrete pro- and anti-inflammatory cytokines as innate cells, their key specialized function is as professional antigen-presenting cells (APCs). DCs that have been activated by the presence of inflammatory signals induce immunity by migrating to (and by residing in) secondary lymphoid organs. These T helper cells direct adaptive immunity, both humoral and cellular. After the induction of active T cell immunity, local APCs present antigens to specific effector T cells at peripheral sites. In protective immunity, these locally active T cells mediate pathogen control and clearance, but in pathological inflammation and autoimmunity, they contribute to local tissue injury. Similar events occur in recurrent exposure to the antigen, when effector memory T cells recognize an antigen in the periphery presented by APCs to induce delayed-type hypersensitivity-like responses and recruit and activate other inflammatory cells.

There are a number of subtypes of DCs, the details of which have been recently reviewed [1]. DCs reside in peripheral tissues, with the capacity to migrate to secondary lymphoid organs, or they exist within lymph nodes, spleen and other

lymphoid tissue. DCs also exist in the thymus, where they are important in thymic T cell selection, but this aspect of DC function will not be considered further in this review. Although the nomenclature and classification of DCs has been and is complex, the current concepts are that there are two broad types of DCs, plasmacytoid DCs and classical DCs [1]. Plasmacytoid DCs function differently from other DC types, in that they are functionally specialized to sense foreign or altered nucleic acids and produce Type I interferons as well as presenting antigens. Classical DCs reside in secondary lymphoid organs and in peripheral tissues. Different tissues possess DCs with different functions, best suited to their role within that particular organ [1, 2]. During inflammation, blood-borne mononuclear phagocytes are recruited. These cells may arise from monocyte-like precursors and could differentiate into several cell types with a variety of functions [2], including antigen presentation to effector T cells. The cells are known as inflammatory DCs [1, 3]. The murine DC system has been studied more extensively, due to its capacity for genetic alteration, experimental studies and access to tissues. While some markers differ, similar DC types exist between mouse and human. This detailed classification of murine and human DCs has been reviewed recently [1].

This review focuses on the potential role of DCs in progressive renal disease in native kidneys. It will not directly refer to renal transplantation, where although many of the principles may be the same, additional complexities exist, including the presence of donor and recipient DCs, indirect and direct presentation, and altered lymphatic drainage from the graft. Assessing the role of DCs in kidney disease is complicated by the plasticity of DC and monocyte/macrophages and in some instances it is difficult to definitively (or least simply) separate one from another. The overlap between renal macrophages and DCs, collectively termed renal mononuclear phagocytes was reviewed in detail in 2012 [2]. Some of the important points highlighted in this key review [2] included the overlap in cell surface markers, origin and function between the two lineages, and the potential for plasticity. In the normal murine kidney, CD11b+ and F4/80+ cells have often been considered as macrophages, while CD11c+ cells have been classified as DCs. However, most CD11c+ cells in the kidneys are F4/80+ and many are CD11b+. CD11c+ cells derived from the normal mouse kidney collectively exhibit DC-like functions, but in the resting state are not as effective at inducing T cell responses as conventional splenic DCs [4, 5]. One approach to functional classification has been to consider DCs as directors of adaptive immune cells, and macrophages as immune effectors, while recognizing the overlap and plasticity of renal mononuclear phagocytes [6]. Table 1 provides an overview of progressive renal diseases that might be affected by DCs.

RENAL DCs: THEIR ROLE IN STEADY STATE AND IN INFLAMMATION

Within the kidney, DCs reside mainly within the interstitium. Since their first description by Hart and Fabre [7], a number of studies have highlighted their presence and demonstrated their

Table 1. Diseases in which systemic or local DCs could participate in progressive renal disease

1. Induction and perpetuation of nephritogenic immunity
A. Systemically induced immunity to nephritogenic antigens
<i>Relevance to human disease:</i> many diseases, including infection-related nephritis, lupus nephritis and anti-neutrophil cytoplasmic antibody (ANCA)-associated vasculitis.
<i>Modelled by:</i> most models of glomerulonephritis induced by active immunity, including autologous phase accelerated and non-accelerated 'anti-GBM' nephritis, active models of anti-MPO disease, experimental lupus nephritis.
B. Locally induced immunity
<i>Relevance to human disease:</i> possibly some forms of tubulointerstitial nephritis (without glomerular disease), Goodpasture's disease (anti-GBM glomerulonephritis) and primary membranous glomerulonephritis (anti-PLA2R) among others.
<i>Modelled by:</i> transfer of T cells specific for ovalbumin into NOH mice (expressing ovalbumin on podocytes) or RIPmOVA mice (expressing ovalbumin in the proximal tubules), potentially by GBM-specific CD4+ cell transfer (clones/cell lines) into naïve mice.
2. Local recognition of antigen-specific CD4+ cells
A. Glomerular disease
<i>Relevance to human disease:</i> forms of rapidly progressive glomerulonephritis with evidence of effector T-cell participation: anti-GBM disease, Class III/IV lupus nephritis, ANCA-associated vasculitis.
<i>Modelled by:</i> endogenous antigens: transfer of $\alpha 3(IV)NC1$ specific CD4+ cells. Planted antigen models: heterologous globulins (i.e. autologous phase 'anti-GBM' glomerulonephritis), model foreign antigens (e.g. ovalbumin), myeloperoxidase in the cellular component of anti-MPO disease
B. Interstitial disease
<i>Relevance to human disease:</i> tubulointerstitial nephritis (without glomerular disease), potentially the interstitial component of some glomerular diseases.
<i>Modelled by:</i> anti-tubular basement membrane disease, the interstitial component of autologous phase 'anti-GBM' nephritis.
3. T cell-independent effects of DCs
<i>Relevance to human disease:</i> potentially many other causes of progressive tubulointerstitial renal disease, including the tubulointerstitial component of diabetic nephropathy, obstructive nephropathy and hypertensive renal disease.
<i>Modelled by:</i> unilateral ureteric ligation, chronic injury post ischaemia reperfusion, chronic hypertensive models, renal mass reduction models.

function as APCs [4, 5, 8–10]. Two studies, employing multi-photon confocal microscopy in fluorescent reporter mice, one using CX3CR1-GFP mice [8], the other using CD11c-EYFP mice [5], demonstrated the presence of cells with DC-like morphology and probing behaviours within the cortical tubulointerstitium *in vivo*. Further recent studies on the renal mononuclear phagocyte lineages have highlighted the overlap between surface markers, function and origin. Kawakami *et al.* [11] recently described five distinct mononuclear phagocyte populations within the normal mouse kidney based on their relative expression of CD11b and CD11c, with some types being able to stimulate naïve T cells. Fate map tracing using the DC lineage-specific marker Clec9A showed that in the murine kidney some of the cells that were previously considered renal macrophages are derived from DC precursors [12].

DCs are relatively abundant in the tubulointerstitial compartment in health and disease [4, 5, 7, 8, 13–16] and in some diseases, in local germinal centre-like structures [17]. In humans, as in mice, different subsets are present expressing

Table 2. Selected cytokines produced by DCs and their function in renal disease

Cytokine	Effects on T cell polarity	Inflammatory	Direct effects on fibrosis	Role in experimental renal disease
IL-12	Th1	Pro- (indirect)	?	Th1 inducing [36]
IL-1 β	Th17	Pro-	Pro-fibrotic [37]	Several mechanisms [38, 39]
IL-23	Th17	Pro-	?	Th17 maintaining [40, 41]
IL-10	Suppressive	Anti- (usually)	Anti-fibrotic [42]	Protective [43]
Type I interferons		Pro-	? Promotes FSGS [44]	Promoting systemic autoimmunity [45]
TNF		Pro-	Pro-fibrotic [46]	Several mechanisms [47]
TGF- β	Tregs; Th17 (with IL-6 and IL-1 β)	Anti-	Pro-fibrotic [48]	Pro-fibrotic [48] acutely protective [49]
IL-6	Th17	Pro-	? No direct role [50]	Systemic pro-inflammatory [51] Protective acutely [52]

FSGS, focal segmental glomerulosclerosis; Th1, T helper cell 1; Th17, T helper cell 17.

different surface markers. However, the majority of studies show that DCs, though present, are not common in glomeruli [13, 14, 18]. In steady state, DCs are only rarely found in glomeruli [18]. In inflammation, while numbers increased, they remain relatively uncommon [13, 18]. Differences in the presence or number of DCs between glomerular and interstitial compartments complicate our attempts to understand the role of renal DCs in disease. Although we know that effector CD4+ T cells recognize antigens within glomeruli, induce injury and influence innate effector cells within glomeruli [19–21], and that DCs are instrumental in the generation of effector CD4+ cells in lymphoid organs, we are not sure yet whether local DCs are required for the localization and effector function of antigen-specific CD4+ cells within glomeruli. Studies using bone marrow chimeric mice and mice with lineage-specific MHC II deficiency suggest that antigen recognition and T-cell activation leading to severe glomerular injury can occur via MHC II on intrinsic glomerular cells [22, 23], including podocytes [24], but other leukocytes (for example DCs, monocytes, B cells or even neutrophils) within glomeruli could also be important. However, not all studies show a paucity of DCs in glomeruli. Two human studies suggest significant glomerular DC accumulation in disease: one found a significant number of both classical and plasmacytoid DCs in the glomeruli of patients with proliferative lupus nephritis [25], the other, in anti-neutrophil cytoplasmic antibody (ANCA)-associated glomerulonephritis found DCs in glomeruli, but to a lesser extent [16]. Experimentally, one group defined a CD8+ DC that infiltrated glomeruli in autoimmune anti-glomerular basement membrane (GBM) glomerulonephritis in rats to induce effector T cell apoptosis [26]. In addition to the discordance on studies of DC number in glomeruli, *in vivo* studies in mice show that leukocytes, present constitutively in normal glomeruli, migrate a considerable distance bidirectionally within the glomerular capillary loops [21]. These recent findings suggest that even if uncommonly present, DCs (or other leukocytes) within glomeruli might be able to survey large areas of glomerular capillaries.

There is evidence that renal DCs, and DCs in the renal draining lymph node help maintain tolerance in a steady state by capturing low-molecular weight-filtered antigens and peptides [27, 28]. Unlicensed interstitial renal DCs that express low levels of positive costimulatory molecules and do not secrete pro-inflammatory cytokines sample filtered peptides or

peptides derived from small filtered proteins [27]. Constitutive migration of these immature DCs to draining lymph nodes is likely to help maintain tolerance to self-antigens. Further support for ‘resting’ renal DCs being anti-inflammatory comes from observations of the lower capacity of these cells to stimulate naïve T cells [5], that some populations produce IL-10 and can generate foxp3+ inducible regulatory T cells (iTregs) *ex vivo* [11], and that depletion in experimental acute kidney injury [29], or early in the autologous phase of experimental ‘anti-GBM’ glomerulonephritis, worsens tubulointerstitial injury [30], potentially via IL-10.

In the presence of innate inflammation, renal CD11c+ cells become active, express co-stimulatory molecules and migrate from the kidney. They become more effective at presenting soluble and membrane-bound antigens and activating naïve CD4+ and CD8+ cells draining lymph *in vivo* [5, 9, 31, 32]. As well as being more effective at presenting antigen to naïve CD4+ T cells, renal DCs can quickly produce inflammatory cytokines, for example, TNF in ischaemia reperfusion injury [33]. There are however, brakes on T cell activation: PD-1 ligands on DCs limit T-cell activation [9], neutrophil-derived MPO regulates DC function [34] and in humans *ex vivo*, activated primary proximal tubular cells also limit the activation of autologous DCs [35]. Table 2 summarizes some of the cytokines produced by DCs and their possible roles in renal disease.

HOW COULD DCs PROMOTE PROGRESSIVE RENAL DISEASE AND FIBROSIS?

Defining the contribution of DCs to progressive renal disease is complicated by several key issues, discussed above, including the dual roles of DCs in influencing adaptive immunity, as well as acting as innate cytokine-secreting cells. Progression in some diseases is heavily dependent on ongoing adaptive immunity, where the role of DCs is in perpetuating adaptive immunity that directs inflammation. In other conditions, for example, diabetic nephropathy, polycystic kidney disease and obstructive uropathy, adaptive immunity is unlikely to play a role, but innate inflammation is important. Therefore any function for DCs in these conditions is more likely that they act directly as pro- (or even anti-) fibrotic innate immune cells.

Some of the potential roles of DCs in renal disease are summarized in Figure 1, including reference to CX3CR1, discussed below. In influencing renal disease, DCs can act locally or systemically. There is also evidence, detailed below, that within the kidney, DCs may have different primary functions within different compartments of the kidney, specifically within the cortical interstitium and the medulla.

Systemic roles as APCs

DCs in secondary lymphoid organs are important in the induction and maintenance of systemic immunity. An increasing proportion of forms of glomerulonephritis are recognized as resulting from immunity to systemic antigens. Often, as for example, in systemic lupus erythematosus and ANCA-associated vasculitis, the antigens are autoantigens, but in infection-related forms of glomerulonephritis the primary immune responses are systemic and directed against a foreign antigen. Therefore, DCs that drain sites of antigen encounter systemically play a key role in these responses in chronic inflammatory states resulting in progressive renal disease, with the caveat that in established humoral

autoimmunity, antigen-specific B cells may act themselves as potent APCs [53]. Events in the renal draining lymph nodes may be relevant for responses against renal tissue restricted antigens and after immune complex deposition [54–56].

Local roles as APCs

Adaptive immunity initiated by DCs generates antibodies that bind to peripheral targets as effectors, independent of local DCs. However, local DCs do play this APC role in presenting the antigen to CD4+ T cells in the cortical interstitium. Antigen-specific effector CD4+ T cell responses, requiring local MHC II expressing APC are now recognized as relevant to a number of forms of glomerulonephritis. DCs mediate this recognition in the cortical interstitium, though their role in the glomerulus is less clear, as discussed above. Experimentally, we know that CD4+ T cells recognize planted and endogenous antigens in glomeruli. Multiple studies using different antigens, including using transfer of T-cell lines and clones demonstrate that effector T cells mediated glomerular injury in an antigen-specific manner [19, 20, 56–59], but it is not yet clear which

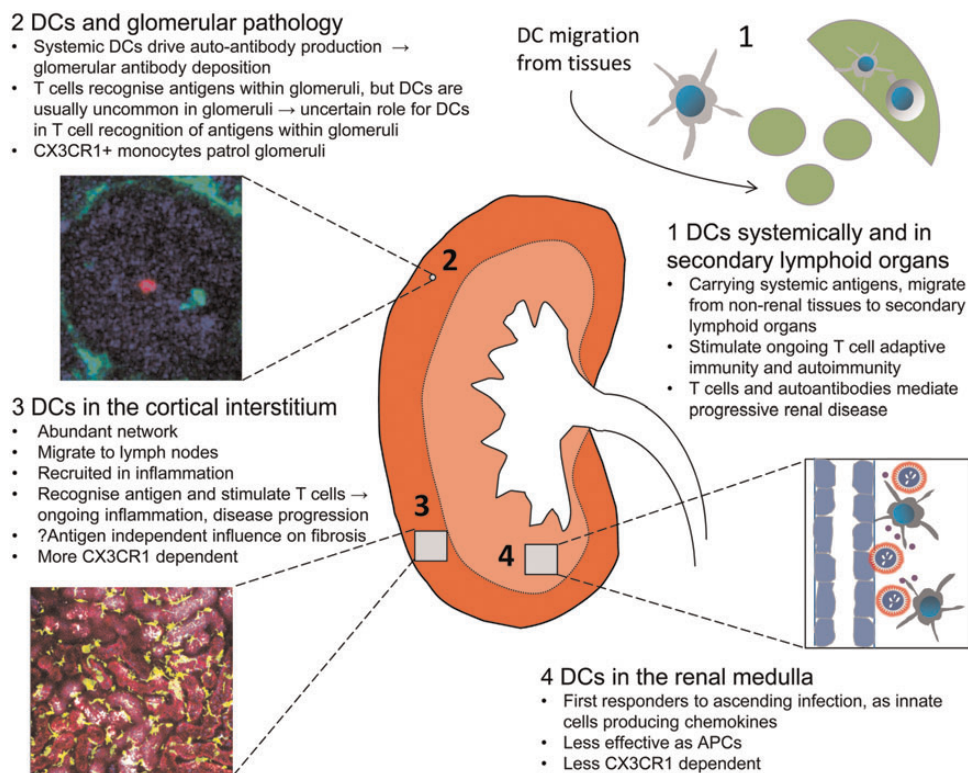


FIGURE 1: The potential roles of dendritic cells (DCs) in progressive renal disease, by location and compartment. (1) Most antigens in renal disease are systemic, so adaptive immune responses are stimulated on an ongoing basis by systemic antigens and DCs from other tissues and within secondary lymphoid organs (spleen and lymph nodes, green cartoon, top right). (2) DCs contribute to ongoing glomerular pathology by directing systemic cellular and humoral immunity, but the role of DCs in the recognition of antigen in glomeruli is unclear. CX3CR1+ monocytes patrol the glomerulus intravascularly. (3) The cortical interstitium is richly endowed with DCs that normally help to maintain tolerance to self-antigens, but in inflammation become potent stimulators of antigen-specific effector CD4+ cells and are dependent on CX3CR1. A direct pro-fibrotic role of DCs has not yet been defined. (4) In the renal medulla, DCs respond innately and rapidly to infection, producing chemokines (purple dots, cartoon, bottom right) that recruit neutrophils (red rimmed cells) in an early response to bacteria within the kidney. Medullary DCs are not CX3CR1 dependent. Illustrative photomicrographs are stills from *in vivo* multiphoton microscopy studies: (2) Glomerulus (Westhorpe C.A., Kitching A.R., Hickey M.J., unpublished image). Glomerular capillaries, blue, antigen-specific CD4+ cell red, CX3CR1+ cells green (within glomerular capillary loop and periglomerular). (3) Normal cortical interstitium (Snelgrove *et al.* [5], unpublished image from work published). Yellow cells are CD11c+ cells with dendritic morphology, epithelial cells are labelled in red via a cell tracker dye.

cell type(s) mediate this antigen-specific recognition within glomeruli.

Local roles as pro-inflammatory and pro-fibrotic innate cells

As well as their specialized antigen-presenting roles, DCs secrete a variety of cytokines that help protect from infectious agents. These cytokines can be pro- or anti-inflammatory, some could have direct pro-fibrotic effects (Table 2). While it is well known that many of these cytokines can contribute to renal disease, or dampen inflammation, it is less clear whether renal DC production of these cytokines makes a significant contribution.

CX3CR1 ON DCs: A MORE SPECIFIC THERAPEUTIC TARGET IN PROGRESSIVE RENAL DISEASE?

CX3CR1, renal cortical DCs and experimental glomerulonephritis

A recent paper from Christian Kurts' group in Bonn, Germany, sheds more light on the role of DCs in the progression of renal inflammation [60]. This group has made important contributions to understanding the biology of renal DCs over the last 10 years [4, 27, 28, 30, 56, 61]. The latest paper suggests that targeting CX3CR1 (the fractalkine receptor), a chemokine receptor expressed by DCs and monocytes, may offer more targeted therapy for progressive renal disease in the future. While chemokines are important in a number of areas of biology, they are perhaps best known for their capacity to recruit cells to tissues, particularly in inflammatory states. Fractalkine (CX3CL1) is somewhat unusual as a chemokine in that it is cell surface anchored [62] and has only one receptor, CX3CR1. CX3CL1 is present in human and experimental glomerular and interstitial renal disease [63–65]. It is expressed on endothelial cells and a variety of other cell types, including tubular epithelial cells and is induced by a variety of inflammatory stimuli [66–68]. Similarly, CX3CR1 has been detected in a variety of human renal diseases, present on CD3+ and CD68+ leukocytes in both the glomerulus and the interstitium [69], increased in fibrotic disease on fibroblasts [70] and in some reports, also DCs [70, 71].

Using a model of glomerulonephritis induced by injecting an anti-basement membrane antibody raised in sheep, which acts as a planted foreign antigen, Hochheiser *et al.* [60] examined the relationship between DC-derived CX3CR1 and disease. CX3CR1-deficient mice developed less severe glomerulonephritis than in other reports that inhibited or deleted CX3CR1 or its ligand CX3CL1 [72–74]. However, unlike some previously published papers, which focussed on impaired monocyte recruitment to glomeruli [72], Hochheiser *et al.* [60] found that interstitial DC influx into the kidney was mediated by CX3CR1, but DCs from other organs did not seem to be particularly CX3CR1 dependent. Recruitment of inflammatory DCs to the kidney was also mediated by CX3CR1. This is likely to be important in the progression of disease mediated by adaptive immunity, as renal DCs migrate

to draining lymph nodes in inflammatory states, to be replaced by inflammatory renal mononuclear phagocytes. As well as potentially acting as macrophages, these recruited cells can differentiate into DCs, ensuring effector CD4+ antigen-specific T cells can recognize antigens locally within the tubulointerstitium.

Medullary DCs behave differently and are less CX3CR1 dependent

Hochheiser *et al.* address, at least to some degree, a key question in the future treatment of immune-mediated disease: how to target injurious inflammatory responses without undue and suppressive effects on host defence. The contention is that, at least in mice, CX3CR1 is an attractive therapeutic target, firstly because it is preferentially expressed in renal DCs (much less so in DCs within other organs) and secondly, because it is required for recruitment of inflammatory DCs. In addition, intriguing data were presented implying that renal host defence might be unimpaired by targeting DCs [60]. CX3CR1+ DCs are enriched in the renal cortical tubulointerstitium and are important in progressive renal disease mediated by adaptive immune responses. Therefore, could inhibiting the function of renal DCs render the host more susceptible to intrarenal infection? Pyelonephritis, the most important and common renal infection, typically results from ascending infection. In this context, DCs in the renal medulla seem to be first responders, with a lesser role for cortical DCs. Hochheiser *et al.* [60] show in experimental murine pyelonephritis that the host defence was unaffected in the absence of CX3CR1. The medullary DCs that are the initial responders to ascending infection are less CX3CR1-dependent and also have less important antigen-presenting functions.

While common forms of renal infection are usually mediated by ascending infection, with initial medullary involvement; some infections do involve the cortex. Another paper published shortly after the Hochheiser paper [75] showed that CX3CR1 in renal cortical DCs is important in host defence in murine systemic candidiasis involving the kidney. Interestingly, clearance from other organs was substantially less affected, supporting a more important role for CX3CR1 in the kidney compared with other organs. Renal cortical candida infection tends to occur more often in people with significant neutropaenia, and therefore may or may not be a significant issue in itself if inhibiting CX3CR1 was to be a strategy employed in humans.

Other CX3CR1-expressing cells: are they relevant to progressive kidney disease and host defence?

Besides DCs, other immune cells also express CX3CR1. There is evidence that these cells, and CX3CR1 expression on these cells, is important in renal disease—in addition to CX3CR1 on DCs [60], though it is uncertain how this might relate to progressive chronic renal disease. CX3CR1 is expressed on circulating and patrolling intravascular monocytes in healthy mice. These monocytes are constitutively present both in the glomerular capillaries [21] and in the interstitial circulation, sense danger and induce acute neutrophil mediated endothelial injury in peritubular capillaries [76]. These

CX3CR1+ monocytes could play roles in infection and inflammation in the kidney, or elsewhere in the body. In human lupus nephritis, CX3CL1 was present in glomeruli, with infiltrating monocytes expressing CX3CR1 [77]. Neutralizing CX3CR1 in non-accelerated ‘anti-GBM’ glomerulonephritis in rats improved glomerular histology, lessened proteinuria and diminished glomerular T cell and macrophage infiltration [73].

Within the interstitium, CX3CR1 mediates monocyte recruitment and promotes injury acutely in experimental ischaemia reperfusion injury [78, 79], which could be relevant chronically. In addition to monocytes, a proportion of human T cells also express CX3CR1 and *in vitro* these T cells can be attracted to proximal tubular cells [80]

These data make the concept targeting CX3CL1–CX3CR1 in progressive renal disease more attractive, as targeting CX3CR1, as well as minimizing T-cell mediated tubulointerstitial disease via DC inhibition, may also have other beneficial effects, including limiting glomerular leukocyte recruitment. However, if patrolling CX3CR1+ monocytes are important in the host defence elsewhere in the body, anti-CX3CR1 therapy might be less specific and have more systemic immunosuppressive effects.

Are DCs critical in all types of progressive renal disease?

The concept that in the future in renal disease, we can limit progressive disease by selectively affecting ‘bad’ or pathogenic renal DCs (those that promote pathological inflammation) while not impairing the function of ‘good’ renal DCs (those that would assist in protecting from ascending infection) is an attractive one. However, renal disease and its progression is complex, as discussed above. We now know that renal DCs are important in antigen-directed T-cell-mediated inflammation [60, 61], but do renal DCs have pro-fibrotic roles in the diverse range of renal diseases non-mediated by adaptive immunity?

DO RENAL DCs HAVE PRO-FIBROTIC ROLES INDEPENDENT OF THEIR ROLES IN ENHANCING ADAPTIVE IMMUNITY?

The active participation of DCs in ongoing T-cell-mediated inflammation means that healing and repair with mature and healthy matrix deposition, and restoration of function, is unlikely to occur. Therefore, if DCs are allowing effector T cells to recognize locally antigens on an ongoing basis (or systemically promoting ongoing adaptive immunity resulting increased antibody deposition or T-cell recruitment), then they are highly likely to be profibrotic. In these settings, the role of DCs in promoting immunity is clearly linked to promoting fibrosis. But do DCs promote fibrosis directly as pro-inflammatory innate cells independent of their roles in adaptive immunity? This is less clear—and finding the answers to this question complicated by DCs’ close relationship and overlap with macrophages, the definition of what is a DC, as well as complexities in experimental depletion strategies throughout the renal mononuclear phagocyte systems [2, 81].

Observations in human renal fibrosis

Many human DC studies focus on inflammation and find that the more inflamed the kidney, the more DCs are present in the interstitium. As chronic inflammation leads to fibrosis, these findings clearly need to be taken into consideration. In studies focussed specifically on human renal fibrosis, renal DC numbers are increased, with increased expression of the co-stimulatory molecule CD86, suggesting increased activation, and increased classical DC (probably inflammatory DCs) production of TGF- β [82]. As might be anticipated, in renal fibrosis, CX3CR1 is upregulated on several cell types, including DCs [70].

Experimental studies in progressive renal injury not driven by innate immunity

Experimentally, most work on a potential role of DCs in ‘innate’ fibrosis uses the murine unilateral ureteric obstruction (UUO) model. There is as yet no absolute clarity in the evidence derived from this model, due at least in part to the complex and overlapping phenotypes of renal mononuclear phagocytes (DCs and monocyte/macrophages), as well as the different techniques used in depletion studies, each with their strengths and weaknesses [81], but there is little hard evidence that DCs themselves contribute innately to fibrosis. It is likely that that a macrophage (or macrophage-like) subset is directly pro-fibrotic and that DCs, although becoming more active, do not play a direct pro-fibrotic role [5, 83, 84]. *In vivo* multiphoton microscopy showed that CD11c+ cells with a dendritic morphology were more active morphologically, tended to cluster around damaged tubules, and showed a higher number of dendrites per cell [5]. The total number of DCs tended not to alter over the first 3 days, potentially due to migration to draining lymph nodes being balanced by an influx of monocyte-like cells then differentiating into CD11c^{hi} inflammatory DCs, though another study found an early increase in DCs with increased early cytokine production [85]. Although the DCs were functionally activated (as they acquired a more potent capacity to stimulate antigen-specific T cells), using the CD11c-DTR system (CD11c+ cells are deleted by the administration of diphtheria toxin), fibrosis at Day 7 was not altered by CD11c+ cell depletion prior or during the process. Depletion at Day 7 did not alter fibrosis at 2 weeks. Therefore, unlike experimental autologous phase ‘anti-GBM’ glomerulonephritis [61], it appears that depleting DCs later in an innate inflammatory response does not diminish fibrotic disease.

CX3CR1’s influence on experimental renal fibrosis has been examined in other settings. CX3CR1-deficient mice were protected in a hypertensive model [86] and in the context of chronic changes after ischaemia reperfusion injury [87]. In this setting the outer medulla was most affected, via diminished macrophage recruitment in the absence of CX3CR1. Whether this is due to CX3CR1’s effect indirectly via DCs or directly via monocytes is not clear.

Clearly the classical antigen-presenting, antigen-specific T-cell-stimulating functions of DCs as APCs are important in the progression of renal disease mediated by adaptive immunity, but it is still unclear whether renal DCs have direct effects

on fibrosis mediated by innate immunity, inflammation and metabolic insults, which maybe more macrophage mediated. Interestingly, in chronic murine adriamycin nephrosis, a model of progressive renal disease, infusions of *ex vivo*-generated anti-inflammatory macrophages, or activated plasmacytoid DCs both reduce chronic inflammation and injury [88, 89].

Can we learn from studies in fibrosis in other organs?

The role of DCs in progressive disease in other tissues has been assessed, but as in the kidney, the situation is unclear. As in human renal disease, DCs accumulate in human fibrotic lung disease, [90]. Experimentally, in toxin-induced liver and lung fibrosis, DCs are more prominent and active [91, 92]. Functionally, in experimental liver disease DCs seem to be responsible for the early increased pro-inflammatory cytokines found [91]. One study found DCs to be pro-fibrogenic [93], but another showed that DCs accelerated regression of fibrosis in a carbon tetrachloride induced model [94]. Most recently a further series of studies in experimental fibrosis (induced by bile duct ligation) suggests that macrophages contribute to fibrosis but DCs do not [95], findings aligned with the emerging view from studies in UUO.

While fibrosis in different organs has a number of common features, we cannot assume that the mediators and mechanisms all are similar. Examples of these organ-specific differences that involve the kidney include the role of the plasminogen, which is protective in experimental pulmonary fibrosis [96], but unexpectedly (yet convincingly) pathogenic in UUO [97, 98], and the pathogenic role of platelet-derived growth factor-C in the kidney not being replicated in the liver [99].

SUMMARY AND FUTURE CHALLENGES

DCs are important cells in kidney health and disease. Understanding their roles in progressive disease is complex, relating to their plasticity and overlap with monocyte/macrophages, the systemic and local roles of DCs, their potentially divergent roles at different stages of disease and the recent recognition of compartment specific functions for DCs. In established inflammation, renal DCs within the cortical interstitium present antigen to activated effector T cells, augmenting and perpetuating inflammation that promotes progressive renal disease. In this setting, local roles for DCs as APCs within glomeruli remain uncertain. Some of the key questions regarding of DCs in progressive renal disease are outlined in Table 3.

In progressive renal disease that features innate inflammation, there is no clear role for DCs, though macrophages (some of which might display some DC-like features) are important. Immunotherapy for renal disease remains in most cases, relatively non-specific, but recent data suggest targeting CX3CL1–CX3CR1 could be a more selective and effective approach, though we still need to understand more about the off-target (non-DC) effects that could potentially be beneficial and/or deleterious. Lastly, studies that attempt to define the role of DCs and identify new therapeutic studies should ideally be performed in well-defined and understood model systems so that their relevance to humans can be clearly assessed.

Table 3. The role of DCs in progressive renal disease: current questions

1. Are renal DCs pro-fibrotic, independent of their effects as APCs. If so, are they best thought of as macrophages or DCs (and to what extent is this distinction important)?
2. Can we dissect out the relative roles of elements of the renal mononuclear phagocyte system (DCs and monocyte/macrophages) in progressive renal disease and target cells with a harmful phenotype?
3. Can we better isolate the systemic and local effects of DCs experimentally to understand their roles?
4. Antigens within glomeruli can be recognized by effector CD4+ cells, but given the paucity of DCs in glomeruli, are they important, and what other cells act as APCs?
5. To what degree are innate pro-fibrotic responses within the kidney 'generic' and how much are their direction and intensity determined by the underlying stimuli?
6. What effect does CX3CR1 inhibition have on cells other than renal cortical DCs and glomerular monocyte/macrophages. Are these effects beneficial, or could there be adverse effects?
7. Can we harness anti-inflammatory and tolerogenic effects of DCs locally and/or systemically to treat renal disease?

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CONFLICT OF INTEREST STATEMENT

None declared.

REFERENCES

1. Merad M, Sathe P, Helft J *et al*. The dendritic cell lineage: ontogeny and function of dendritic cells and their subsets in the steady state and the inflamed setting. *Annu Rev Immunol* 2013; 31: 563–604
2. Nelson PJ, Rees AJ, Griffin MD *et al*. The renal mononuclear phagocytic system. *J Am Soc Nephrol* 2012; 23: 194–203
3. Hespel C, Moser M. Role of inflammatory dendritic cells in innate and adaptive immunity. *Eur J Immunol* 2012; 42: 2535–2543
4. Kruger T, Benke D, Eitner F *et al*. Identification and functional characterization of dendritic cells in the healthy murine kidney and in experimental glomerulonephritis. *J Am Soc Nephrol* 2004; 15: 613–621
5. Snelgrove SL, Kausman JY, Lo C *et al*. Renal dendritic cells adopt a pro-inflammatory phenotype in obstructive uropathy to activate T cells but do not directly contribute to fibrosis. *Am J Pathol* 2012; 180: 91–103
6. Teteris SA, Engel DR, Kurts C. Homeostatic and pathogenic role of renal dendritic cells. *Kidney Int* 2011; 80: 139–145
7. Hart DN, Fabre JW. Major histocompatibility complex antigens in rat kidney, ureter, and bladder. Localization with monoclonal antibodies and demonstration of Ia-positive dendritic cells. *Transplantation* 1981; 31: 318–325
8. Soos TJ, Sims TN, Barisoni L *et al*. CX3CR1+ interstitial dendritic cells form a contiguous network throughout the entire kidney. *Kidney Int* 2006; 70: 591–596
9. Edgton KL, Kausman JY, Li M *et al*. Intrarenal antigens activate CD4+ cells via co-stimulatory signals from dendritic cells. *J Am Soc Nephrol* 2008; 19: 515–526

10. Austyn JM, Hankins DF, Larsen CP *et al.* Isolation and characterization of dendritic cells from mouse heart and kidney. *J Immunol* 1994; 152: 2401–2410
11. Kawakami T, Lichtnekert J, Thompson LJ *et al.* Resident renal mononuclear phagocytes comprise five discrete populations with distinct phenotypes and functions. *J Immunol* 2013; 191: 3358–3372
12. Schraml BU, van Blijswijk J, Zelenay S *et al.* Genetic tracing via DNGR-1 expression history defines dendritic cells as a hematopoietic lineage. *Cell* 2013; 154: 843–858
13. Woltman AM, de Fijter JW, Zuidwijk K *et al.* Quantification of dendritic cell subsets in human renal tissue under normal and pathological conditions. *Kidney Int* 2007; 71: 1001–1008
14. Segerer S, Heller F, Lindenmeyer MT *et al.* Compartment specific expression of dendritic cell markers in human glomerulonephritis. *Kidney Int* 2008; 74: 37–46
15. Fiore N, Castellano G, Blasi A *et al.* Immature myeloid and plasmacytoid dendritic cells infiltrate renal tubulointerstitium in patients with lupus nephritis. *Mol Immunol* 2008; 45: 259–265
16. Wilde B, van Paassen P, Damoiseaux J *et al.* Dendritic cells in renal biopsies of patients with ANCA-associated vasculitis. *Nephrol Dial Transplant* 2009; 24: 2151–2156
17. Chang A, Henderson SG, Brandt D *et al.* In situ B cell-mediated immune responses and tubulointerstitial inflammation in human lupus nephritis. *J Immunol* 2011; 186: 1849–1860
18. Schwarz M, Taubitz A, Eltrich N *et al.* Analysis of TNF-mediated recruitment and activation of glomerular dendritic cells in mouse kidneys by compartment-specific flow cytometry. *Kidney Int* 2013; 84: 116–129
19. Summers SA, Steinmetz OM, Li M *et al.* Th1 and Th17 cells induce proliferative glomerulonephritis. *J Am Soc Nephrol* 2009; 20: 2518–2524
20. Ooi JD, Chang J, Hickey MJ *et al.* The immunodominant myeloperoxidase T-cell epitope induces local cell-mediated injury in antimyeloperoxidase glomerulonephritis. *Proc Natl Acad Sci U S A* 2012; 109: E2615–E2624
21. Devi S, Li A, Westhorpe CL *et al.* Multiphoton imaging reveals a new leukocyte recruitment paradigm in the glomerulus. *Nat Med* 2013; 19: 107–112
22. Li S, Kurts C, Kontgen F *et al.* Major histocompatibility complex class II expression by intrinsic renal cells is required for crescentic glomerulonephritis. *J Exp Med* 1998; 188: 597–602
23. Ruth AJ, Kitching AR, Semple TJ *et al.* Intrinsic renal cell expression of CD40 directs Th1 effectors inducing experimental crescentic glomerulonephritis. *J Am Soc Nephrol* 2003; 14: 2813–2822
24. Goldwisch A, Burkard M, Olke M *et al.* Podocytes are nonhematopoietic professional antigen-presenting cells. *J Am Soc Nephrol* 2013; 24: 906–916
25. Tucci M, Quatraro C, Lombardi L *et al.* Glomerular accumulation of plasmacytoid dendritic cells in active lupus nephritis: role of interleukin-18. *Arthritis Rheum* 2008; 58: 251–262
26. Wu J, Zhou C, Robertson J *et al.* Identification of a bone marrow-derived CD8 α high α dendritic cell-like population in inflamed autoimmune target tissue with capability of inducing T cell apoptosis. *J Leukoc Biol* 2010; 88: 849–861
27. Lukacs-Kornek V, Burgdorf S, Diehl L *et al.* The kidney-renal lymph node-system contributes to cross-tolerance against innocuous circulating antigen. *J Immunol* 2008; 180: 706–715
28. Gottschalk C, Damuzzo V, Gotot J *et al.* Batf3-dependent dendritic cells in the renal lymph node induce tolerance against circulating antigens. *J Am Soc Nephrol* 2013; 24: 543–549
29. Tadagavadi RK, Reeves WB. Endogenous IL-10 attenuates cisplatin nephrotoxicity: role of dendritic cells. *J Immunol* 2010; 185: 4904–4911.
30. Scholz J, Lukacs-Kornek V, Engel DR *et al.* Renal dendritic cells stimulate IL-10 production and attenuate nephrotoxic nephritis. *J Am Soc Nephrol* 2008; 19: 527–537
31. Roake JA, Rao AS, Morris PJ *et al.* Dendritic cell loss from nonlymphoid tissues after systemic administration of lipopolysaccharide, tumor necrosis factor, and interleukin 1. *J Exp Med* 1995; 181: 2237–2247
32. Dong X, Swaminathan S, Bachman LA *et al.* Antigen presentation by dendritic cells in renal lymph nodes is linked to systemic and local injury to the kidney. *Kidney Int* 2005; 68: 1096–1108
33. Dong X, Swaminathan S, Bachman LA *et al.* Resident dendritic cells are the predominant TNF-secreting cell in early renal ischemia-reperfusion injury. *Kidney Int* 2007; 71: 619–628
34. Odobasic D, Kitching AR, Yang Y *et al.* Neutrophil myeloperoxidase regulates T-cell-driven tissue inflammation in mice by inhibiting dendritic cell function. *Blood* 2013; 121: 4195–4204
35. Kassianos AJ, Sampangi S, Wang X *et al.* Human proximal tubule epithelial cells modulate autologous dendritic cell function. *Nephrol Dial Transplant* 2013; 28: 303–312
36. Kitching AR, Tipping PG, Holdsworth SR. IL-12 directs severe renal injury, crescent formation and Th1 responses in murine glomerulonephritis. *Eur J Immunol* 1999; 29: 1–10
37. Jones LK, O'Sullivan KM, Semple T *et al.* IL-1RI deficiency ameliorates early experimental renal interstitial fibrosis. *Nephrol Dial Transplant* 2009; 24: 3024–3032
38. Lan HY, Nikolic-Paterson DJ, Mu W *et al.* Interleukin-1 receptor antagonist halts the progression of established crescentic glomerulonephritis in the rat. *Kidney Int* 1995; 47: 1303–1309
39. Timoshanko JR, Kitching AR, Iwakura Y *et al.* Contributions of IL-1 β and IL-1 α to crescentic glomerulonephritis in mice. *J Am Soc Nephrol* 2004; 15: 910–918
40. Ooi JD, Phoon RK, Holdsworth SR *et al.* IL-23, not IL-12, directs autoimmunity to the Goodpasture antigen. *J Am Soc Nephrol* 2009; 20: 980–989
41. Paust HJ, Turner JE, Steinmetz OM *et al.* The IL-23/Th17 axis contributes to renal injury in experimental glomerulonephritis. *J Am Soc Nephrol* 2009; 20: 969–979
42. Mu W, Ouyang X, Agarwal A *et al.* IL-10 suppresses chemokines, inflammation, and fibrosis in a model of chronic renal disease. *J Am Soc Nephrol* 2005; 16: 3651–3660
43. Kitching AR, Tipping PG, Timoshanko JR *et al.* Endogenous interleukin-10 regulates Th1 responses that induce crescentic glomerulonephritis. *Kidney Int* 2000; 57: 518–525
44. Markowitz GS, Nasr SH, Stokes MB *et al.* Treatment with IFN- α , - β , or - γ is associated with collapsing focal segmental glomerulosclerosis. *Clin J Am Soc Nephrol* 2010; 5: 607–615
45. Braun D, Geraldes P, Demengeot J. Type I interferon controls the onset and severity of autoimmune manifestations in *lpr* mice. *J Autoimmun* 2003; 20: 15–25
46. Guo G, Morrissey J, McCracken R *et al.* Role of TNFR1 and TNFR2 receptors in tubulointerstitial fibrosis of obstructive nephropathy. *Am J Physiol* 1999; 277: F766–F772
47. Karkar AM, Tam FW, Steinkasserer A *et al.* Modulation of antibody-mediated glomerular injury in vivo by IL-1ra, soluble IL-1 receptor, and soluble TNF receptor. *Kidney Int* 1995; 48: 1738–1746
48. Border WA, Okuda S, Languino LR *et al.* Suppression of experimental glomerulonephritis by antiserum against transforming growth factor beta 1. *Nature* 1990; 346: 371–374
49. Wilson HM, Minto AW, Brown PA *et al.* Transforming growth factor-beta isoforms and glomerular injury in nephrotoxic nephritis. *Kidney Int* 2000; 57: 2434–2444
50. Yang J, Chen J, Yan J *et al.* Effect of interleukin 6 deficiency on renal interstitial fibrosis. *PLoS One* 2012; 7: e52415
51. Finck BK, Chan B, Wofsy D. Interleukin 6 promotes murine lupus in NZB/NZW F1 mice. *J Clin Invest* 1994; 94: 585–591
52. Karkar AM, Smith J, Tam FW *et al.* Abrogation of glomerular injury in nephrotoxic nephritis by continuous infusion of interleukin-6. *Kidney Int* 1997; 52: 1313–1320
53. Shlomchik MJ, Craft JE, Mamula MJ. From T to B and back again: positive feedback in systemic autoimmune disease. *Nat Rev Immunol* 2001; 1: 147–153
54. Kurts C, Klebba I, Davey GM *et al.* Kidney protection against autoreactive CD8(+) T cells distinct from immunoprivilege and sequestration. *Kidney Int* 2001; 60: 664–671
55. Bagavant H, Deshmukh US, Wang H *et al.* Role for nephritogenic T cells in lupus glomerulonephritis: progression to renal failure is accompanied by T cell activation and expansion in regional lymph nodes. *J Immunol* 2006; 177: 8258–8265

56. Heymann F, Meyer-Schwesinger C, Hamilton-Williams EE *et al.* Kidney dendritic cell activation is required for progression of renal disease in a mouse model of glomerular injury. *J Clin Invest* 2009; 119: 1286–1297
57. Bolton WK, Chandra M, Tyson TM *et al.* Transfer of experimental glomerulonephritis in chickens by mononuclear cells. *Kidney Int* 1988; 34: 598–610
58. Radeke HH, Tschernig T, Karulin A *et al.* CD4+ T cells recognizing specific antigen deposited in glomeruli cause glomerulonephritis-like kidney injury. *Clin Immunol* 2002; 104: 161–173
59. Ooi JD, Chang J, O'Sullivan KM *et al.* The HLA-DRB1*15:01-restricted Goodpasture's T cell epitope induces GN. *J Am Soc Nephrol* 2013; 24: 419–431
60. Hochheiser K, Heuser C, Krause TA *et al.* Exclusive CX3CR1 dependence of kidney DCs impacts glomerulonephritis progression. *J Clin Invest* 2013; 123: 4242–4254
61. Hochheiser K, Engel DR, Hammerich L *et al.* Kidney dendritic cells become pathogenic during crescentic glomerulonephritis with proteinuria. *J Am Soc Nephrol* 2011; 22: 306–316
62. Bazan JF, Bacon KB, Hardiman G *et al.* A new class of membrane-bound chemokine with a CX3C motif. *Nature* 1997; 385: 640–644
63. Furuichi K, Wada T, Iwata Y *et al.* Upregulation of fractalkine in human crescentic glomerulonephritis. *Nephron* 2001; 87: 314–320
64. Cockwell P, Chakravorty SJ, Girdlestone J *et al.* Fractalkine expression in human renal inflammation. *J Pathol* 2002; 196: 85–90
65. Donadelli R, Zanchi C, Morigi M *et al.* Protein overload induces fractalkine upregulation in proximal tubular cells through nuclear factor kappaB and p38 mitogen-activated protein kinase-dependent pathways. *J Am Soc Nephrol* 2003; 14: 2436–2446
66. Chakravorty SJ, Cockwell P, Girdlestone J *et al.* Fractalkine expression on human renal tubular epithelial cells: potential role in mononuclear cell adhesion. *Clin Exp Immunol* 2002; 129: 150–159
67. Chen YM, Lin SL, Chen CW *et al.* Tumor necrosis factor-alpha stimulates fractalkine production by mesangial cells and regulates monocyte transmigration: down-regulation by cAMP. *Kidney Int* 2003; 63: 474–486
68. Durkan AM, Alexander RT, Liu GY *et al.* Expression and targeting of CX3CL1 (fractalkine) in renal tubular epithelial cells. *J Am Soc Nephrol* 2007; 18: 74–83
69. Segerer S, Hughes E, Hudkins KL *et al.* Expression of the fractalkine receptor (CX3CR1) in human kidney diseases. *Kidney Int* 2002; 62: 488–495
70. Koziolok MJ, Schmid H, Cohen CD *et al.* Potential role of fractalkine receptor expression in human renal fibrogenesis. *Kidney Int* 2007; 72: 599–607
71. Hoffmann U, Bergler T, Segerer S *et al.* Impact of chemokine receptor CX3CR1 in human renal allograft rejection. *Transpl Immunol* 2010; 23: 204–208
72. Chen S, Bacon KB, Li L *et al.* In vivo inhibition of CC and CX3C chemokine-induced leukocyte infiltration and attenuation of glomerulonephritis in Wistar-Kyoto (WKY) rats by vMIP-II. *J Exp Med* 1998; 188: 193–198
73. Feng L, Chen S, Garcia GE *et al.* Prevention of crescentic glomerulonephritis by immunoneutralization of the fractalkine receptor CX3CR1 rapid communication. *Kidney Int* 1999; 56: 612–620
74. Inoue A, Hasegawa H, Kohno M *et al.* Antagonist of fractalkine (CX3CL1) delays the initiation and ameliorates the progression of lupus nephritis in MRL/lpr mice. *Arthritis Rheum* 2005; 52: 1522–1533
75. Lionakis MS, Swamydas M, Fischer BG *et al.* CX3CR1-dependent renal macrophage survival promotes Candida control and host survival. *J Clin Invest* 2013
76. Carlin LM, Stamatides EG, Auffray C *et al.* Nr4a1-dependent Ly6C(low) monocytes monitor endothelial cells and orchestrate their disposal. *Cell* 2013; 153: 362–375
77. Yoshimoto S, Nakatani K, Iwano M *et al.* Elevated levels of fractalkine expression and accumulation of CD16+ monocytes in glomeruli of active lupus nephritis. *Am J Kidney Dis* 2007; 50: 47–58
78. Li L, Huang L, Sung SS *et al.* The chemokine receptors CCR2 and CX3CR1 mediate monocyte/macrophage trafficking in kidney ischemia-reperfusion injury. *Kidney Int* 2008; 74: 1526–1537
79. Oh DJ, Dursun B, He Z *et al.* Fractalkine receptor (CX3CR1) inhibition is protective against ischemic acute renal failure in mice. *Am J Physiol Renal Physiol* 2008; 294: F264–F271
80. Cockwell P, Calderwood JW, Brooks CJ *et al.* Chemoattraction of T cells expressing CCR5, CXCR3 and CX3CR1 by proximal tubular epithelial cell chemokines. *Nephrol Dial Transplant* 2002; 17: 734–744
81. van Blijswijk J, Schraml BU, Reis e Sousa C. Advantages and limitations of mouse models to deplete dendritic cells. *Eur J Immunol* 2013; 43: 22–26
82. Kassianos AJ, Wang X, Sampangi S *et al.* Increased tubulointerstitial recruitment of human CD141hi CLEC9A+ and CD1c+ myeloid dendritic cell subsets in renal fibrosis and chronic kidney disease. *Am J Physiol Renal Physiol* 2013
83. Kitamoto K, Machida Y, Uchida J *et al.* Effects of liposome clodronate on renal leukocyte populations and renal fibrosis in murine obstructive nephropathy. *J Pharmacol Sci* 2009; 111: 285–292
84. Machida Y, Kitamoto K, Izumi Y *et al.* Renal fibrosis in murine obstructive nephropathy is attenuated by depletion of monocyte lineage, not dendritic cells. *J Pharmacol Sci* 2010; 114: 464–473
85. Dong X, Bachman LA, Miller MN *et al.* Dendritic cells facilitate accumulation of IL-17T cells in the kidney following acute renal obstruction. *Kidney Int* 2008; 74: 1294–1309
86. Shimizu K, Furuichi K, Sakai N *et al.* Fractalkine and its receptor, CX3CR1, promote hypertensive interstitial fibrosis in the kidney. *Hypertens Res* 2011; 34: 747–752
87. Furuichi K, Gao JL, Murphy PM. Chemokine receptor CX3CR1 regulates renal interstitial fibrosis after ischemia-reperfusion injury. *Am J Pathol* 2006; 169: 372–387
88. Lu J, Cao Q, Zheng D *et al.* Discrete functions of M2a and M2c macrophage subsets determine their relative efficacy in treating chronic kidney disease. *Kidney Int* 2013; 84: 745–755
89. Zheng D, Cao Q, Lee VW *et al.* Lipopolysaccharide-pretreated plasmacytoid dendritic cells ameliorate experimental chronic kidney disease. *Kidney Int* 2012; 81: 892–902
90. Marchal-Somme J, Uzunhan Y, Marchand-Adam S *et al.* Dendritic cells accumulate in human fibrotic interstitial lung disease. *Am J Respir Crit Care Med* 2007; 176: 1007–1014
91. Connolly MK, Bedrosian AS, Mallen-St Clair J *et al.* In liver fibrosis, dendritic cells govern hepatic inflammation in mice via TNF-alpha. *J Clin Invest* 2009; 119: 3213–3225
92. Bantsimba-Malanda C, Marchal-Somme J, Goven D *et al.* A role for dendritic cells in bleomycin-induced pulmonary fibrosis in mice? *Am J Respir Crit Care Med* 2010; 182: 385–395
93. Blois SM, Piccioni F, Freitag N *et al.* Dendritic cells regulate angiogenesis associated with liver fibrogenesis. *Angiogenesis* 2013
94. Jiao J, Sastre D, Fiel MI *et al.* Dendritic cell regulation of carbon tetrachloride-induced murine liver fibrosis regression. *Hepatology* 2012; 55: 244–255
95. Pradere JP, Kluwe J, De Minicis S *et al.* Hepatic macrophages but not dendritic cells contribute to liver fibrosis by promoting the survival of activated hepatic stellate cells in mice. *Hepatology* 2013; 58: 1461–1473
96. Swaisgood CM, French EL, Noga C *et al.* The development of bleomycin-induced pulmonary fibrosis in mice deficient for components of the fibrinolytic system. *Am J Pathol* 2000; 157: 177–187
97. Edgton KL, Gow RM, Kelly DJ *et al.* Plasmin is not protective in experimental renal interstitial fibrosis. *Kidney Int* 2004; 66: 68–76
98. Zhang G, Kernan KA, Collins SJ *et al.* Plasmin(ogen) promotes renal interstitial fibrosis by promoting epithelial-to-mesenchymal transition: role of plasmin-activated signals. *J Am Soc Nephrol* 2007; 18: 846–859
99. Martin IV, Borkham-Kamphorst E, Zok S *et al.* Platelet-derived growth factor (PDGF)-C neutralization reveals differential roles of PDGF receptors in liver and kidney fibrosis. *Am J Pathol* 2013; 182: 107–117

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